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## Cellular and molecular mechanisms of hepatic fibrogenesis

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#### INTRODUCTION

Progressive accumulation of fibrillar extracellular matrix (ECM) in the liver is the consequence of reiterated liver tissue damage due to infective (mostly hepatitis B and C viruses), toxic/drug-induced, metabolic and autoimmune causes and the relative chronic activation of the wound healing reaction. The process may result in clinically evident liver cirrhosis and hepatic failure. Cirrhosis is defined as an advanced stage of fibrosis, characterized by the formation of regenerative nodules of liver parenchyma that are separated by and encapsulated in fibrotic septa and associated with major angio-architectural changes<sup>1</sup>. While millions of patients worldwide are affected by chronic liver diseases potentially leading to cirrhosis, only a minority (~25-30%) are likely to develop significant fibrosis and cirrhosis. This is particularly true for chronic hepatitis due to HCV infection, whose prevalence is predicted to peak between the years 2010 to 2015. In recent years, attention has been focused on «non alcoholic fatty liver disease» (NAFLD) and in particular on «non-alcoholic steatohepatitis» (NASH). The occurrence of NASH is associated with progressive fibrosis and cirrhosis in a high percentage of patients (up to 50%). Considering that, in industrialized countries, NASH affects 3% of the general population and 20-45% of obese patients, it becomes clear that this clinical entity represents a major health problem.

Independently of the etiology, liver cirrhosis is the most common non-neoplastic cause of death among hepatobiliary and digestive diseases, both in the United States of America and Europe. In addition, this condition is largely associated with primary liver cancer, with a further increase in the relative mortality rate<sup>2,3</sup>. In general terms, the following clinical features have been shown to be predictors of the development of significant fibrosis, or at

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least, of a faster progression to cirrhosis: *a*) male gender (for groups of age < 50 years), *b*) age at infection (hepatitis virus, particularly HCV), *c*) obesity, diabetes mellitus and, more in general, the clinical features of the so-called «metabolic syndrome», *d*) daily alcohol intake, independently of the major cause of hepatocellular damage, and *e*) hepatic iron content. In addition, individual factors are likely to affect several aspects of the fi-brogenic process (i.e: differences in handling a metabolic/toxic load, in the immune system reactions towards infectious agents and autoantigens, and in the evolution of the chronic wound healing reaction). In addition, there are markedly different rates of fibrosis progression for apparently similar clinical features<sup>4,5</sup>.

In general, in those chronic liver diseases (CLD) evolving towards cirrhosis, a significant accumulation of fibrillar ECM is observed only after a clinical course lasting several years and even decades.

#### DEFINITION OF DIFFERENT TYPES OF FIBROGENESIS

Although cirrhosis is the common result of progressive fibrogenesis, there are distinct patterns of fibrotic development, related to the underlying disorders causing the fibrosis<sup>6</sup>. Biliary fibrosis, due to the co-proliferation of reactive bile ductules and periductular (myo)fibroblastlike cells at the portal-parenchymal interface, tends to follow a portal to portal direction. This leads to the formation of portal-portal septa surrounding liver nodules, where the central vein and its connections with the portal tract are preserved until late stages. In contrast, the chronic viral hepatitis pattern of fibrosis is considered the results of portal-central (vein) bridging necrosis, thus originating portal-central septa. In addition, this form of fibrogenic evolution is characterized by the presence of «interface» hepatitis and development of portal to portal septa and septa ending blind in the parenchyma, and by rapid derangement of the vascular connections with the portal system (early portal hypertension). The so-called central to central (vein) form of fibrogenic evolution is in general secondary to venous outflow problems (e.g.

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chronic heart failure) and is characterized by the development of central to central septa and «reversed lobulation». Finally, a peculiar type of fibrosis development (pericellular/sinusoidal) is observed in alcoholic and metabolic liver diseases (e.g. NASH), in which the deposition of fibrillar matrix is concentrated around the sinusoids (capillarization) and around groups of hepatocytes (chickenwire pattern). These different patterns of fibrogenic evolution are related to different factors and particularly: *a*) the topographic localization of tissue damage, and *b*) the relative concentration of pro-fibrogenic factors, and 3. the prevalent pro-fibrogenic mechanism(s). In addition, these different patterns imply the participation of different cellular effectors of the fibrogenic process.

#### CELLULAR EFFECTORS OF HEPATIC FIBROGENESIS

As the liver becomes fibrotic, there are both quantitative and qualitative changes in the composition of the hepatic ECM. The total content of collagenous and noncollagenous components increases 3-5-fold, accompanied by the shift in the type of ECM in the subendothelial space from the normal low-density basement membrane-like matrix to interstitial type matrix containing fibril-forming collagens. These quantitative and qualitative changes in the composition of ECM, in addition to their mechanical and physical implications, contribute to the formation of a new biochemical environment. Indeed, each ECM component has the ability to modulate cell growth, migration, gene expression, and other important cellular functions directly by interaction with cell adhesion molecules and, indirectly, by functioning as a biologic reservoir for proinflammatory and pro-fibrogenic mediators in their active or inactive forms7.

The cellular source of connective tissue components in fibrotic liver has been controversial for many years, with some elements of controversy still remaining. Among other cell types potentially involved in the abnormal progressive deposition of fibrillar ECM, HSC have received much attention, perhaps due to the possibility to isolate these cells from liver tissues with a relatively high purity. Consequently, most of the present knowledge on the cell and molecular biology of hepatic fibrosis derives from in vitro studies employing culture activated HSC isolated from rat, mouse or human liver. Regardless, it is now evident that distinct ECM-producing cells, each with a distinct localization and a characteristic immunohistochemical and/or electron microscopic phenotype, are likely to contribute to liver fibrosis6. These include: fibroblasts and myofibroblasts of the portal tract, smooth muscle cells localized in vessel walls, and myofibroblasts localized around the centrolobular vein. It is now evident that the relative participation of these different cell types is dependent on the development of distinct patterns of fibrosis. Major efforts are currently made in order to identify and characterize the origin of the different cell types responsible of the fibrogenic process. By employing cell identification markers<sup>6</sup>, three populations of ECM producing cells can be identified: a) septal myofibroblasts, present in the inner part of fibrous septa, expressing a panel of cell markers identical to portal myofibroblasts; b) activated HSC, located in capillarized sinusoids adjacent to expanded portal tracts, and c) interface myofibroblasts, located at the edge of fibrous septa. This last population presents with an expression profile intermediate between portal myofibroblasts and activated HSC. It is conceivable that interface myofibroblasts derive from activated HSC recruited at the site of active fibrogenesis, where there is high level of cell damage, extracellular matrix degradation by gelatinases and inflammatory infiltration. In addition, preliminary reports suggest that bone marrow-derived stromal cells recruited at the site of liver injury could contribute to this population of fibrogenic cells8.

#### **PRO-FIBROGENIC MECHANISMS**

Some of the emerging profibrogenic mechanisms for the fibrogenic evolution of CLDs will be summarized below. In general, fibrosis should be regarded to as an alteration of the process of chronic wound healing. This process, which is highly efficient in the presence of a single acute tissue insult, leads to progressive scarring when tissue damage is chronic. In other terms, deposition of fibrillar matrix rather then organized tissue regeneration becomes the best option in order to maintain tissue continuity. Hepatic fibrogenesis is characterized by the following key features: a. the persistence of hepatocellular/cholangiocellular damage with variable degree of necrosis and apoptosis; b. a complex inflammatory infiltrate including mononuclear cells and immunocompetent cells; c. the activation of different types of ECM-producing cells (HSC, portal myofibroblasts, etc.) with marked proliferative, synthetic and contractile features; d. marked changes in the quality and quantity of the hepatic ECM associated with very limited or absent possibilities of remodeling and regeneration<sup>9</sup>.

#### Inflammation

One of the first steps in tissue repair is the recruitment of inflammatory cells in order to neutralize possible infectious agents and to remove the necrotic tissue. In this phase of the process, HSC are recruited at the site of injury in order to synthesize and secrete ECM components. Recruitment and proliferation of HSC is under the control of soluble factors secreted by the cells of the inflammatory infiltrate. However, in the presence of reiterated tissue damage, HSC secrete several chemokines thus becoming a site of amplification and chronic organization of the inflammatory infiltrate<sup>10</sup>. Monocyte chemotactic protein-1 (MCP-1) is the most prominent chemotactic factor secreted by chronically activated HSC and it Proinflammatory cytokines such as interleukin-1, tumor necrosis factor-

alpha and interferon- $\gamma$  have been shown to be strong stimulators of gene and protein expression of MCP-1 in HSC<sup>11,12</sup>. The exposure of HSC to soluble mediators that may potentially affect their pro-fibrogenic role has represented a key area of investigation in the last decade. It is important to stress that exposure to these mediators, generically defined as «inflammatory», may be time-limited or chronically present according to the nature, extent and reiteration of parenchymal damage.

A pivotal profibrogenic mechanism operated by infiltrating cells, such as inflammatory cells, is the synthesis and release of soluble factors playing a biological role on HSC. Consolidated experimental evidence suggests that two polypeptide growth factors, namely platelet-derived growth factor (PDGF) and transforming growth factor-B1 (TGF $\beta$ 1), greatly contribute to the profibrogenic role of HSC<sup>13</sup>. TGF $\beta_1$  belongs to the superfamily of receptors comprising a large number of structurally related polypeptide growth factors, each capable of regulating different arrays of cellular processes including cell proliferation, lineage determination, differentiation, motility, adhesion and death, thereby playing a prominent role in the development, homeostasis and repair of all tissues in organisms<sup>14</sup>. Members of the TGFB cytokine family initiate signalling through their interaction with heteromeric type I/II TGFB receptors, which propagate signals downstream through phosphorylation of cytoplasmic mediators of the receptor-regulated Smad family<sup>15,16</sup>. Upon activation, a Smad2/3-Smad4 complex will translocate to the nucleus where they are involved in the regulation of transcription factors of TGF $\beta_1$  target genes such as collagen type I<sup>17,18</sup>. In many fibrogenic diseases abnormal accumulation of ECM proteins is associated with increased expression of the TGFB and TGFB receptors. Established evidence indicates that this growth factor play a multiple role in hepatic fibrogenesis. In activated HSC, TGFB induces a strong and consistent up-regulation of the genes encoding for fibrillar collagens (particularly collagen type I and collagen type III) and other ECM components. In addition, TGF $\beta$  induces a down-regulation of the gene encoding for matrix metalloproteinase (MMP)-1 (collagenase) associated with an up-regulation of the gene for tissue inhibitor of metalloproteinase (TIMP)-1. In the general economy of the normal wound-healing process, the newly deposited fibrillar ECM is continuously subjected to degradation and remodelling towards the normal liver ECM. Along these lines, one of the key roles of TGF $\beta$  in the chronically activated wound healing process is the inhibition of fibrillar ECM degradation and, in other terms, the promotion of progressive ECM accumulation and scarring<sup>19-23</sup>.

Among other polypeptide growth factors, platelet derived growth factor (PDGF) is the most potent mitogen and chemoattractant for HSC. More specifically, the PDGF-BB dimeric isoform has shown to be the most potent factor in stimulating growth and this is associated with the predominant expression of PDGF $\beta$  receptor<sup>24-26</sup>. These receptors, which have intrinsic tyrosine kinase activity, dimerize, become autophosphorylated upon binding to

their ligand. In this conformation the receptor becomes highly operative as a docking site for many intracellular signalling molecules such as Grb2 which recruits mSos, followed by Ras activation and Erk translocation to the nucleus where it will increase the expression of *c-fos*, a transcription factor, necessary for PDGF-induced mitogenesis, chemotaxis and cell survival<sup>27,28</sup>.

#### **Oxidative stress**

When chronic liver injury is not clearly associated with an abundant inflammatory infiltrate, other soluble agents may sustain the activation of HSC through pathways that are specific for a particular type of damage. Evidence for oxidative stress has been detected in almost all the clinical and experimental conditions of CLDs with different etiology and fibrosis progression rate, often in association with decreased antioxidant defenses. As already proposed for atherosclerosis and chronic degenerative diseases of CNS, oxidative stress-related molecules such as reactive oxygen intermediates (ROI) and reactive aldehydes, may act as mediators able to modulate tissue and cellular events responsible for the progression of liver fibrosis<sup>29</sup>. In alcoholic liver injury, for example, acetaldehyde, the main metabolite of ethanol, is able to increase gene transcription and synthesis of different ECM components in activated HSC<sup>30,31</sup>. In addition to acetaldehyde, products of lipid peroxidation generated by exposure to ethanol or the production of iron overload may also perpetuate HSC activation<sup>32</sup>. Along these lines, stimulation of lipid peroxidation or exposure to 4-hydroxynonenal (4-HNE), a highly reactive aldehydic end-product of lipid peroxidation, increases procollagen I gene expression in activated human HSC<sup>32,33</sup>.

Inflammation and oxidative stress are tightly linked together. In a recent study, we demonstrated that interfering with the mechanisms of inflammatory cell recruitment as observed in MCP-1 knock-out mice, limits the generation of intrahepatic oxidative stress<sup>34</sup>. On the other hand, some types of leukocytes have been shown to counter-regulate the development of fibrosis. Depletion of NK cells worsens matrix accumulation, due to the fact that this cell type induces programmed cell death of activated HSC<sup>35</sup>.

# Derangement of the epithelial-mesenchymal interaction

Cholangiopathies are progressive liver disorders representing a major cause of chronic cholestasis both in adults and children. Both bile duct proliferation and ductular metaplasia are associated with profound changes in the surrounding mesenchymal cells and extracellular matrix. It is likely that at least in the early phases, ECM-producing cells other than HSC are primarily involved, whereas HSC become subsequently involved when proliferating bile ducts tend to invade lobular areas. It is still unclear whether the changing epithelial phenotype directly in-

duces an alteration in portal mesenchymal cells and ECM or whether the epithelial cell changes are induced by modifications in ECM. Cytokines and proinflammatory mediators, released in the portal spaces, likely contribute to these processes by activating fibrogenesis, stimulating apoptotic and proliferative responses, damaging the peribiliary circulation, increasing the expression of histocompatibility antigens in cholangiocytes and by altering the transport functions of the epithelium. An emerging concept is that bile duct epithelial cells are active participants in inflammatory diseases and, in pathologic conditions, secrete proinflammatory and chemotactic cytokines, such as IL-6, TNF $\alpha$ , IL-8, and MCP-1, together with growth factors able to activate mesenchymal cells and matrix production (ET-1, PDGF-BB, TGF $\beta$ 2, CTGF)<sup>36-40</sup>. These mediators, either released in the portal spaces by immune cells, macrophages and mesenchymal cells or produced by the epithelium itself, may have profound effects on epithelial cell function. Accordingly, several lines of evidence suggest that «activated» cholangiocytes play an active role in stimulating the fibrogenic response, through an extensive cross-talk with portal fibroblasts/myofibroblasts and HSC. It is very relevant that this close association between bile duct proliferation and mesenchymal activation is present also in cholangiocarcinomas, a group of neoplasms frequently characterized by a strong desmoplastic reaction.

#### Adipokines

This group of cytokines produced by adipose tissue is believed to play a role in nonalcoholic steatohepatitis (NASH), that develops in the presence of an excess of fat. Leptin is a hormone produced by adipocytes, that regulates food intake via actions on the hypothalamus. Different groups have provided compelling in vivo evidence for the pro-fibrogenic action of leptin in rodents<sup>41-</sup> <sup>45</sup>. Injection of leptin during acute and chronic intoxication results in a marked upregulation of the expression of type I procollagen and transforming growth factor- $\beta$ 1, a key pro-fibrogenic cytokine. This finding was followed by the demonstration that scarring is reduced in fa/fa rats or ob/ob mice chronically exposed to thioacetamide. These studies also confirmed that the expression of pivotal profibrogenic mediators is limited in the absence of leptin signaling.

The pro-fibrogenic action of leptin depends, at least in part, on a direct effect on HSC, which express functional leptin receptors and are directly responsive to leptin. Incubation of HSC with recombinant leptin stimulates mRNA and protein expression of type I procollagen, potentiates the effects of TGF $\beta$ , and up-regulates expression of the tissue inhibitor of metalloproteinase (TIMP)-1, thus blocking collagen degradation<sup>44,46-47</sup>. Moreover, leptin is a mitogen and a survival factor for activated HSC, and limits their apoptosis, resulting in an increase in the number of fibrogenic cells, and induces secretion of pro-inflammatory chemokines such as MCP-1, via activation

of nuclear factor- $\kappa$ B<sup>48</sup>. Finally, a recently-identified activity of leptin on fibrogenic cells is the induction of vascular endothelial growth factor, one of the most potent inducers of neovessel formation, via oxygen-independent activation of hypoxia-inducible factor 1 $\alpha$ , a master switch of the angiogenic response<sup>48</sup>. These data suggest that leptin may be an effector of the increased fibrogenesis observed in obese patients<sup>49</sup>.

Adiponectin is a recently identified protein that is predominantly, but not exclusively, by the adipose tissue. It circulates at high levels in the bloodstream, representing one of the main plasma proteins. Adiponectin is considered a major determinant of the sensitivity to insulin, acting at different sites of glucose metabolism, including liver, muscle, and fat itself. Experimental data also show that administration of recombinant adiponectin ameliorates metabolic derangements and liver damage in mouse models of alcoholic and nonalcoholic hepatitis<sup>50</sup>. Thus, adiponectin may block the development of fibrosis limiting hepatic damage. More recently, a direct antifibrogenic action of adiponectin has been demonstrated in animals undergoing toxic liver damage<sup>51</sup>, a condition independent of deranged metabolism. In addition, a balance between the biology of leptin and that of adiponectin seems to take place in stellate cells<sup>52</sup>. Adiponectin's effects are mediated by two receptors, known as AdipoR1 and AdipoR253, and at least some of the metabolic effect of adiponectin are dependent on receptor-mediated activation of AMPdependent protein kinase. However, the contribution of the different receptor isoforms and/or AMP-dependent kinase to the antifibrogenic effects of adiponectin has not yet been elucidated. The emerging biology of adiponectin make this molecule a very appealing target for future studies in NASH and other liver diseases.

Other adipokines are possibly implicated in the fibrogenic process. Resistin contributes to insulin resistance in rodents, but its metabolic effects in humans are still uncertain<sup>54</sup>. Recent evidence obtained in our laboratory indicates that resistin modulates the biology of human HSC inducing a pro-inflammatory phenotype. In addition, like reported for other «adipokines» the expression of resistin is detectable in liver tissue, especially in conditions of fibrosis<sup>55</sup>.

#### Renin-angiotensin-aldosterone system

The renin-angiotensin system is another pivotal player in the pathogenesis of liver fibrosis. Exposure of fibrogenic cells to angiotensin II mediates key biological actions involved in hepatic tissue repair, including proliferation, infiltration of inflammatory cells, and collagen synthesis<sup>56</sup>. Activated HSCs express all components of the renin-angiotensin system, and the autocrine effects of Ang II are mediated by activation of NADPH oxidase<sup>57,58</sup>. Blockade of the renin-angiotensin system attenuates fibrosis development in different experimental models of chronic liver injury<sup>59</sup>. In addition, aldosterone antagonists also have a direct antifibrogenic action<sup>60</sup>. Interfering with the reninangiotensin system is therefore a very promising strategy to prevent fibrosis progression in chronic liver diseases, and controlled clinical trials are under way.

#### Nuclear hormone receptors

A number of studies has recently suggested that antidiabetic thiazolidinediones (TZD), or «glitazones» may represent a possible novel pharmacological treatment for liver fibrosis. TZDs are employed for the treatment of insulin resistance in patients with type 2 diabetes and are selective ligands for the nuclear transcription factor peroxisome proliferator-activated receptor  $(PPAR)\gamma^{61}$ . PPARy is expressed in quiescent HSC and its abundance and/or transcriptional activity decreases along the activation process that accompanies the acquisition of fibrogenic properties<sup>62-64</sup>. More important, exposure of HSC to PPARy ligands, including different glitazones, reverts most features of the activated phenotype of HSC. In these cells, PPAR $\gamma$  activation reduces the expression of interstitial collagens and other matrix proteins, downregulates the ability to proliferate and migrate in response to PDGF, blocks the secretion of pro-inflammatory chemokines such as monocyte chemoattractant protein-1, and induces apoptosis<sup>62-66</sup>. In addition, daily intragastric administration of rosiglitazone or pioglitazone, started at the same time as injury, leads to a marked reduction of fibrotic tissue accumulation and fibrogenic cell proliferation<sup>66</sup>. These data provided compelling in vivo evidence supporting an antifibrogenic role of thiazolidinediones, although recent data suggest that administration of these drugs may be less effective if started after the onset of injury<sup>67</sup>.

#### Other mechanisms

Recently, different groups have reported the biological effects of proteins of the hepatitis C virus (HCV) on cultured HSC. A number of different approaches have been used, including incubation with recombinant proteins of the envelope, adenoviral-mediated overexpression of HCV proteins or exposure to conditioned media of hepatocyte-like cells expressing the HCV replicon<sup>68-70</sup>. Exposure to HCV proteins induces increased expression of extracellular matrix and ECM-regulating cytokines (collagens, TGFB, CTGF) and up-regulation of MMP-2, an index of HSC activation. In addition, down-regulation of fibrolytic matrix-metalloproteinases (e.g. MMP-1), induction of ROS generation, and up-regulation of pro-inflammatory cytokines have been described. This represents a novel area of investigation linking fibrogenesis and viral hepatitis.

The cannabinoid system comprises an additional group of mediators that have been recently implicated in the regulation of fibrogenesis. Anandamide, a lipid mediator, efficiently induces necrosis in activated HSCs, but not hepatocytes<sup>71,72</sup>. In addition, interference with cannabinoid

receptors leads to differential results. Knock-out of CB2 is associated with increased fibrosis, demonstrating that CB2 activation triggers antifibrotic pathways<sup>73</sup>. In contrast, animals deficient for CB1 have lower accumulation of scar tissue, indicating a pro-fibrogenic action for this receptor<sup>74</sup>. These results may have a relevance ion consideration of the availability of rimonabant, a selective CB1 blocker, that has been proposed as a therapy for the metabolic syndrome.

#### Future directions

The increasing knowledge on the pathogenesis of hepatic fibrosis has led to important changes in the clinical interpretation of this phenomenon. Firstly, the need of an accurate and effective monitoring of the fibrotic progression of chronic liver diseases and of the effectiveness of the currently proposed treatments has become an impellent need. Moreover, the identification of the genes involved in the progression of liver fibrosis would hopefully lead to the establishment of prognostic markers indicating a faster progression of fibrogenic chronic liver diseases. Along these lines, several ongoing studies are addressing the relevance of gene expression and/or gene polymorphisms in defined subset of patients. Finally, novel profibrogenic pathways are being elucidated, leading to new approaches for antifibrotic treatments.

#### REFERENCES

- 1. Friedman SL. Liver fibrosis from bench to bedside. J Hepatol. 2003;38:38S-53S.
- 2. El-Serag HB. Hepatocellular carcinoma and hepatitis C in the United States. Hepatology. 2002; 36 Suppl 1:74-83.
- Befeler AS, Di Bisceglie AM. Hepatocellular carcinoma: diagnosis and treatment. Gastroenterology. 2002;122:1609-19.
- Poynard T, Bedossa P, Opolon P. Natural history of liver fibrosis progression in patients with chronic hepatitis C. The OB-SVIRC, METAVIR, CLINIVIR, and DOSVIRC groups. Lancet. 1997;349:825-32.
- Pinzani M, Romanelli RG, Magli S. Progression of fibrosis in chronic liver diseases: time to tally the score. J Hepatol. 2001;34:764-7.
- Cassiman D, Roskams T. Beauty is in the eye of the beholder: emerging concepts and pitfalls in hepatic stellate cell research. J Hepatol. 2002;37:527-35.
- 7. Schuppan D, Ruehl M, Somasundaram R, Hahn EG. Matrix as a modulator of hepatic fibrogenesis. Semin Liver Dis. 2001;21:351-72.
- Forbes SJ, Russo FP, Rev V, Burra P, Rugge M, Wright NN, et al. A significant proportion of myofibroblasts are of bone marrow origin in human liver cirrhosis. Gastroenterology. 2004;1 26:955-63.
- 9. Pinzani M. Liver fibrosis. Springer Semin Immunopathol. 2000; 21:475-90.
- 10. Marra F. Hepatic stellate cells and the regulation of liver inflammation. J Hepatol. 1999;31:1120-30.
- Marra F, Valente AJ, Pinzani M, Abboud HE. Cultured human liver fat-storing cells produce monocyte chemotactic protein 1. Regulation by proinflammatory cytokines. J Clin Invest. 1993;92:1674-80.
- Marra F, DeFranco R, Grappone C, Milani S, Pastacaldi S, Pinzani M, et al. Increased expression of monocyte chemotactic protein-1 during active hepatic fibrogenesis: correlation with monocyte infiltration. Am J Pathol. 1998;152:423-30.

- Pinzani M, Marra F. Cytokine receptors and signaling in hepatic stellate cells. Semin Liver Dis. 2001;21:397-416.
- Branton MH, and Kopp JB. TGFβ and fibrosis. Microbes Infect. 1999;1:1349-65.
- 15. Piek E, Heldin CH, Ten Dijke P.Specificity, diversity, and regulation in TGF-beta superfamily signaling. FASEB J. 1999;13:2105-24.
- Massague J. How cells read TGF-beta signals. Nat Rev Mol Cell Biol. 2000;1:169-78.
- Inagaki Y, Truter S, Ramírez F. Transforming growth factorbeta stimulates alpha 2(I) collagen gene expression through a cis-acting element that contains an Sp1-binding site. J Biol Chem. 1994;269:14828-34.
- Inagaki Y, Nemoto T, Nakao A, Ten Dijke P, Kobayashi K, Takehara K, et al. Interaction between GC box binding factors and Smad proteins modulates cell lineage-specific alpha 2(I) collagen gene transcription. J Biol Chem. 2001;276:16573-9.
- Nakatsukasa H, Nagy P, Evarts RP, Hsia CC, Marsden E, Thorgeirsson SS. Cellular distribution of transforming growth factor-beta 1 and procollagen types I, III, and IV transcripts in carbon tetrachloride-induced rat liver fibrosis. J Clin Invest. 1990;85:1833-43.
- Castilla A, Prieto J, Fausto N. Transforming growth factors beta 1 and alpha in chronic liver disease. Effects of interferon alfa therapy. N Engl J Med. 1991;324:933-40.
- Bachem MG, Meyer D, Melchior R, Sell KM, Gressner AM. Activation of rat liver perisinusoidal lipocytes by transforming growth factors derived from myofibroblast-like cells. A potential mechanism of self perpetuation in liver fibrogenesis. J Clin Invest. 1992;89:19-27.
- 22. Sanderson N, Factor V, Nagy P, Kopp J, Kondaiah P, Wakefield L, et al. Hepatic expression of mature transforming growth factor beta 1 in transgenic mice results in multiple tissue lesions. Proc Natl Acad Sci USA. 1995;92:2572-6.
- 23. George J, Roulot D, Koteliansky VE, Bissell DM. In vivo inhibition of rat stellate cell activation by soluble transforming growth factor beta type II receptor: a potential new therapy for hepatic fibrosis. Proc Natl Acad Sci USA. 1999;96:12719-24.
- Pinzani M, Gesualdo L, Sabbah GM, Abboud HE. Effects of platelet-derived growth factor and other polypeptide mitogens on DNA synthesis and growth of cultured rat liver fat-storing cells. J Clin Invest. 1989;84:1786-93.
- Pinzani M, Milani S, Grappone C, Weber FL Jr, Gentilini P, Abboud HE. Expression of platelet-derived growth factor in a model of acute liver injury. Hepatology. 1994;19:701-7.
- Pinzani M, Milani S, Herbst H, DeFranco R, Grappone C, Gentilini A, et al. Expression of platelet-derived growth factor and its receptors in normal human liver and during active hepatic fibrogenesis. Am J Pathol. 1996;148:785-800.
- 27. Marra F, Arrighi MC, Fazi M, Caligiuri A, Pinzani M, Romanelli RG, et al. ERK activation differentially regulates PDGF's actions in hepatic stellate cells, and is induced by in vivo liver injury. Hepatology. 1999;30:951-8.
- Marra F, Pinzani M, DeFranco R, Laffi G, Gentilini P. Involvement of phoshatidylinositol 3-kinase in the activation of extracellular signal-regulated kinase by PDGF in hepatic stellate cells. FEBS Lett. 1995;376:141-5.
- 29. Parola M, Robino G. Oxidative stress-related molecules and liver fibrosis J Hepatol. 2001;35:297-306.
- Casini A, Cunningham M, Rojkind M, Lieber CS. Acetaldehyde increases procollagen type I and fibronectin gene transcription in cultured rat fat-storing cells through a protein synthesis-dependent mechanism. Hepatology. 1991;13:758-65.
- Casini A, Ceni E, Salzano R, Biondi P, Parola M, Galli A, et al. Neuthrophil-derived superoxide anion induces lipid peroxidation and stimulate collagen synthesis in human hepatic stellate cells. Role of nitric oxide. Hepatology. 1997;25:361-7.
- 32. Pietrangelo A. Metals, oxidative stress, and hepatic fibrogenesis. Semin Liver Dis. 1996;16:13-30.
- Parola M, Robino G, Marra F, Pinzani M, Bellomo G, Leonarduzzi G, et al. HNE interacts directly with JNK isoforms in human hepatic stellate cells. J Clin Invest. 1998;102:1942-50.
- Zamara E, Galastri S, Aleffi S, Petrai I, Aragno M, Mastrocola R, et al. Prevention of severe toxic liver injury and oxidative stress in MCP-1-deficient mice. J Hepatol. 2007;46:230-8.
- 35. Radaeva S, Sun R, Jaruga B, Nguyen VT, Tian Z, Gao B. Natural killer cells ameliorate liver fibrosis by killing activated

stellate cells in NKG2D-dependent and tumor necrosis factorrelated apoptosis-inducing ligand-dependent manners. Gastroenterology. 2006;130:435-52.

- Morland CM, Fear J, McNab G, Joplin R, Adams DH. Promotion of leukocyte transendothelial cell migration by chemokines derived from human biliary epithelial cells in vitro. Proc Assoc Am Physicians. 1997;109:372-82.
- 37. Marra F, DeFranco R, Grappone C, Milani S, Pastacaldi S, Pinzani M, et al. Increased expression of monocyte chemotactic protein-1 during active hepatic fibrogenesis: correlation with monocyte infiltration. Am J Pathol. 1998;152:423-30.
- Caligiuri A, Glaser S, Rodgers RE, Phinizy JL, Robertson W, Papa E, et al. Endothelin-1 inhibits secretin-stimulated ductal secretion by interacting with ETA receptors on large cholangiocytes. Am J Physiol. 1998;275:835G-46G.
- Grappone C, Pinzani M, Parola M, Pellegrini G, Caligiuri A, DeFranco R, et al. Cholangiocytes contribute to cholestatic fibrosis in the rat through platelet-derived growth factor. J Hepatol. 1999;31:100-9.
- Kinnman N, Goria O, Wendum D, Gendron MC, Rey C, Poupon R, et al. Hepatic stellate cell proliferation is an early platelet-derived growth factor-mediated cellular event in rat cholestatic liver injury. Lab Invest. 2001;81:1709-16.
   Ikejima K, Honda H, Yoshikawa M, Hirose M, Kitamura T,
- Ikejima K, Honda H, Yoshikawa M, Hirose M, Kitamura T, Takei Y, Sato N. Leptin augments inflammatory and profibrogenic responses in the murine liver induced by hepatotoxic chemicals. Hepatology. 2001;34:288-97.
   Honda H, Ikejima K, Hirose M, Yoshikawa M, Lang T,
- 42. Honda H, Ikejima K, Hirose M, Yoshikawa M, Lang T, Enomoto N, et al. Leptin is required for fibrogenic responses induced by thioacetamide in the murine liver. Hepatology. 2002;36:12-21.
- 43. Ikejima K, Takei Y, Honda H, Hirose M, Yoshikawa M, Zhang YJ, et al. Leptin receptor-mediated signaling regulates hepatic fibrogenesis and remodeling of extracellular matrix in the rat. Gastroenterology. 2002;122:1399-410.
- 44. Saxena NK, Ikeda K, Rockey DC, Friedman SL, Anania FA. Leptin in hepatic fibrosis: evidence for increased collagen production in stellate cells and lean littermates of ob/ob mice. Hepatology. 2002;35:762-71.
- Leclercq IA, Farrell GC, Schriemer R, Robertson GR. Leptin is essential for the hepatic fibrogenic response to chronic liver injury. J Hepatol. 2002;37:206-13.
- 46. Cao Q, Mak KM, Ren C, Lieber CS. Leptin stimulates tissue inhibitor of metalloproteinase-1 in human hepatic stellate cells: respective roles of the JAK/STAT and JAK-mediated H2O2dependant MAPK pathways. J Biol Chem. 2003;279:4292-304.
- 47. Saxena NK, Titus MA, Ding X, Floyd J, Srinivasan S, Sitaraman SV, Anania FA. Leptin as a novel profibrogenic cytokine in hepatic stellate cells: mitogenesis and inhibition of apoptosis mediated by extracellular regulated kinase (Erk) and Akt phosphorylation. FASEB J. 2004;18:1612-4.
- Aleffi S, Petrai I, Bertolani C, Parola M, Colombatto S, Novo E, et al. Up-regulation of pro-inflammatory and pro-angiogenic cytokines by leptin in human hepatic stellate cells. Hepatology. 2005;42:1339-48.
- 49. Marra F. Leptin and liver tissue repair: Do rodent models provide the answers? J Hepatol. 2007;46:12-8.
- Xu A, Wang Y, Keshaw H, et al. The fat-derived hormone adiponectin alleviates alcoholic and nonalcoholic fatty liver diseases in mice. J Clin Invest. 2003;112:91-100.
- Kamada Y, Tamura S, Kiso S, et al. Enhanced carbon tetrachloride-induced liver fibrosis in mice lacking adiponectin. Gastroenterology. 2003;125:1796-807.
- 52. Ding X, Saxena NK, Lin S, Xu A, Srinivasan S, Anania FA. The roles of leptin and adiponectin: a novel paradigm in adipocytokine regulation of liver fibrosis and stellate cell biology. Am J Pathol. 2005;166:1655-69.
- Yamauchi T, Kamon J, Ito Y, et al. Cloning of adiponectin receptors that mediate antidiabetic metabolic effects. Nature. 2003;423:762-9.
- Steppan CM, Lazar MA. The current biology of resistin. J Intern Med. 2004;255:439-47.
- 55. Bertolani C, Sancho-Bru P, Failli P, Bataller R, Aleffi S, De-Franco R, et al. Resistin as an intrahepatic cytokine: overexpression during chronic injury and induction of proinflammatory actions in hepatic stellate cells. Am J Pathol. 2006;169: 2042-53.

- Bataller R, Gines P, Nicolas JM, Gorbig MN, García-Ramallo E, Gasull X, et al. Angiotensin II induces contraction and proliferation of human hepatic stellate cells. Gastroenterology. 2000;118:1149-56.
- 57. Bataller R, Sancho-Bru P, Gines P, Lora JM, Al-Garawi A, Sole M, et al. Activated human hepatic stellate cells express the renin-angiotensin system and synthesize angiotensin II. Gastroenterology. 2003;125:117-25.
- Bataller R, Schwabe RF, Choi YH, Yang L, Paik YH, Lindquist J, et al. NADPH oxidase signal transduces angiotensin II in hepatic stellate cells and is critical in hepatic fibrosis. J Clin Invest. 2003;112:1383-94.
- Yoshiji H, Kuriyama S, Yoshii J, Ikenaka Y, Noguchi R, Nakatani T, et al. Angiotensin-II type 1 receptor interaction is a major regulator for liver fibrosis development in rats. Hepatology. 2001;34:745-50.
- Caligiuri A, De Franco RM, Romanelli RG, Gentilini A, Meucci M, Failli P, et al. Antifibrogenic effects of canrenone, an antialdosteronic drug, on human hepatic stellate cells. Gastroenterology. 2003;124:504-20.
- 61. Yki-Jarvinen H. Thiazolidinediones. N Engl J Med. 2004;351: 1106-18.
- Galli A, Crabb D, Price D, Ceni E, Salzano R, Surrenti C, Casini A. Peroxisome proliferator-activated receptor gamma transcriptional regulation is involved in platelet-derived growth factor-induced proliferation of human hepatic stellate cells. Hepatology. 2000;31:101-8.
  Marra F, Efsen E, Romanelli RG, Caligiuri A, Pastacaldi S,
- 63. Marra F, Efsen E, Romanelli RG, Caligiuri A, Pastacaldi S, Batignani G, et al. Ligands of peroxisome proliferator-activated receptor gamma modulate profibrogenic and proinflammatory actions in hepatic stellate cells. Gastroenterology. 2000;119:466-78.
- Miyahara T, Schrum L, Rippe R, Xiong S, Yee HF Jr, Motomura K, et al. Peroxisome proliferator-activated receptors and hepatic stellate cell activation. J Biol Chem. 2000;275:35715-22.
- Planaguma A, Claria J, Miquel R, López-Parra M, Titos E, Masferrer JL, et al. The selective cyclooxygenase-2 inhibitor

SC-236 reduces liver fibrosis by mechanisms involving nonparenchymal cell apoptosis and PPAR-gamma activation. FASEB J. 2005;19:1120-2.

- 66. Galli A, Crabb DW, Ceni E, Salzano R, Mello T, Svegliati-Baroni G, et al. Antidiabetic thiazolidinediones inhibit collagen synthesis and hepatic stellate cell activation in vivo and in vitro. Gastroenterology. 2002;122:1924-40.
- Leclercq IA, Sempoux C, Starkel P, Horsmans Y. Limited therapeutic efficacy of pioglitazone on progression of hepatic fibrosis in rats. Gut. 2006;55:1020-9.
- Bataller R, Paik YH, Lindquist JN, Lemasters JJ, Brenner DA. Hepatitis C virus core and nonstructural proteins induce fibrogenic effects in hepatic stellate cells. Gastroenterology. 2004;126:529-40.
- Mazzocca A, Sciammetta SC, Carloni V, Cosmi L, Annunziato F, Harada T, et al. Binding of hepatitis C virus envelope protein E2 to CD81 up-regulates matrix metalloproteinase-2 in human hepatic stellate cells. J Biol Chem. 2005;280:11329-39.
- Schulze-Krebs A, Preimel D, Popov Y, Bartenschlager R, Lohmann V, Pinzani M, et al. Hepatitis C virus-replicating hepatocytes induce fibrogenic activation of hepatic stellate cells. Gastroenterology. 2005;129:246-58.
- Siegmund SV, Seki E, Osawa Y, Uchinami H, Cravatt BF, Schwabe RF. Fatty acid amide hydrolase determines anandamide-induced cell death in the liver. J Biol Chem. 2006;281: 10431-8.
- Siegmund SV, Uchinami H, Osawa Y, Brenner DA, Schwabe RF. Anandamide induces necrosis in primary hepatic stellate cells. Hepatology. 2005;41:1085-95.
- Julien B, Grenard P, Teixeira-Clerc F, Van Nhieu JT, Li L, Karsak M, et al. Antifibrogenic role of the cannabinoid receptor CB2 in the liver. Gastroenterology. 2005;128:742-55.
- 74. Teixeira-Clerc F, Julien B, Grenard P, Tran Van Nhieu J, Deveaux V, Li L, et al. CB1 cannabinoid receptor antagonism: a new strategy for the treatment of liver fibrosis. Nat Med. 2006; 12:671-6.