



REVISIÓN

Role of altered immune cells in liver diseases: a review



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Abstract Immune cells play an important role in controlling liver tumorigenesis, viral hepatitis, liver fibrosis and contribute to pathogenesis of liver inflammation and injury. Accumulating evidence suggests the effectiveness of natural killer (NK) cells and Kupffer cells (KCs) against viral hepatitis, hepatocellular damage, liver fibrosis, and carcinogenesis. Activation of natural killer cells provides a novel therapeutic strategy to cure liver related diseases. This review discusses the emerging roles of immune cells in liver disorders and it will provide baseline data to scientists to design better therapies for treatment.

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PALABRAS CLAVE

Hígado;
Cirrosis;
Células NK;
Células dendríticas;
Células de Kupffer

Revisión sobre el papel de los inmunocitos alterados en las enfermedades hepáticas

Resumen Los inmunocitos o células inmunitarias desempeñan un papel importante en el control de la carcinogénesis hepática, la hepatitis vírica, la fibrosis hepática y contribuyen a la patogénesis de la inflamación y la lesión hepáticas. La creciente evidencia sugiere la efectividad de los linfocitos citolíticos naturales (NK, *natural killer*) y las células de Kupffer (KC, Kupffer cells) frente a la hepatitis vírica, la lesión hepatocelular, la fibrosis hepática y la carcinogénesis.

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La activación de linfocitos citolíticos naturales ofrece una nueva estrategia terapéutica para curar enfermedades relacionadas con el hígado. Esta revisión trata de las nuevas funciones de los inmunocitos en los trastornos hepáticos y ofrecerá datos básicos a los científicos para diseñar mejores terapias para el tratamiento.

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Introduction

Liver is the metabolic centre of organism controlled by central nervous system. Its anatomic localization and specific tissue structure indicates its defensive role in organism. Approximately 80% of liver cells are hepatocytes. Non-hepatocytes include 40% endothelial cells 20% Kupffer cells, 20% lymphocytes, 20% stellate cells, and biliary cells. Natural killer cells make up 50% of liver population that reside in liver sinusoids. The multitude of cells makes this organ play active role in peripheral immune tolerance of the organism using transforming growth factor- β and haemopoietic cells. The fifth most common cancer in the world is liver cancer, 90% of which is hepatocellular carcinoma (HCC)¹ and the better understanding of liver's immunological processes will provide insight into the role of immune tolerance mechanisms and its contribution in the development of autoimmune diseases and chronic viral infections of liver.

Natural killer cells inhibit liver fibrosis, viral infection and tumor cells growth. The liver immune system is properly equipped with liver immune cells that achieve the critical task of protection against metastatic cells, pathogens, and foreign antigens by coordinating with anti-microbial components (inflammatory cytokines, chemokines, acute phase proteins, complement). Liver plays its role as a buffer between systematic circulation and the contents of gut and about 80% of blood is supplied from gut into liver through portal vein. This blood is rich in harmless environmental antigens, microflora of gut, and dietary elements. Liver must endure immunogenic load by providing immunosurveillance for malignant cells and pathogenic infections.^{2,3} Innate immune cells of liver such as KCs, monocytes, dendritic cells (DCs), NK cells, natural killer T cells (NKT) cells, and neutrophils produce cytokine and initiate inflammation.⁴ We will briefly discuss the potential roles of immune cells in the pathogenesis of liver related disorders.

Natural killer (NK) and natural killer T (NKT) cells

The lymphocytes present in liver are enriched in NK and NKT cells that are key regulators of antitumor defenses, antiviral defenses, and pathogenesis of chronic liver disease. These cells account for 25-40% of total intrahepatic lymphocytes. NK cells have some peculiar functional and phenotypic properties such as specific cytokine profiles, and TRAIL-dependent cytotoxicity.

Decline in NK/NKT cells greatly increases tumor metastasis of liver and enhancement of NK/NKT cells weakens it,^{5,6} both type of cells produce significant quantity of

cytokines that stimulate adaptive immune response and help in removal of food antigens, toxins, and pathogens. The role of NKT cells play dominant role as anti-tumor agent by inhibiting liver fibrosis through suppression of hepatic stellate cells activation.^{7,8} NK and NKT cells play an important role in pathogenesis of liver inflammation and injury, liver fibrosis and tumorigenesis. Several studies demonstrate that NK and NKT cells control viral hepatitis.⁹⁻¹² The mechanisms behind this enrichment and peculiar characteristics of liver NK cells are still not completely elucidated however, effects may be attributed to both cross talk between NK cells and other liver cell types as well as high hepatic expression levels of diverse NK cell-recruiting chemokines.^{13,14} NK cells produce diverse cytokines e.g, interferon-gamma (IFN- γ) and kill target cells (Fig. 1).

The opposing signals of stimulatory and inhibitory receptors on NK cells and their association with analogous ligands bound to target cells determines the potential of NK cells to kill target cells.¹⁵ Alterations that occur as a result of signal expression of receptors of NK cells and their interaction with ligands present on liver cells contribute to pathogenesis of liver diseases.¹⁶ NK cells are activated by cytokines e.g, IFN and interleukins (ILs), IFN- α , IFN- β , IL-12, IL-15, IL-18 and several other related cytokines during acute HCV infection.¹⁷ These activated NK cells act as key contributors in prevention of hepatitis C virus (HCV) either by stimulating adaptive immunity or by killing HCV infected liver cells.^{18,19} IFN- α acts as powerful NK cell activator and cures viral hepatitis by suppressing tumor formation and liver fibrosis. The anti-tumor, anti-fibrotic, and anti-viral effects of IFN- α therapy are stimulated by activation of NK cells. In addition to IFN- α , there are other NK cell activators like IL-12 and IL-18 that have been proved effective against liver carcinogenesis in animal models.^{20,21}

NK cells provide protection against HCV infection because lot of evidences have suggested that selective impairment of NK cell is cause of chronic HCV infection.²² NK cells also increase NK cell ADCC (antibody-dependent cell-mediated cytotoxicity) and target cancers and tumor cells.²³ Another study demonstrated that NK cell cytotoxicity against tumor cells is increased by blockade of NK cell inhibitory receptors and monoclonal antibodies such as rituximab, IPH-2101 that block killer-cell immunoglobulin-like receptor to treat hematological cancers, this strategy is currently in phase II clinical trials.^{24,25} Therefore, blockage of NK cell inhibitory receptors, targeting of NK cells to increase ADCC, and activation of NK cells by cytokines all form the basis of therapeutic strategies against hepatocellular carcinoma (HCC). Adoptive transfer of activated NK cells isolated from donor

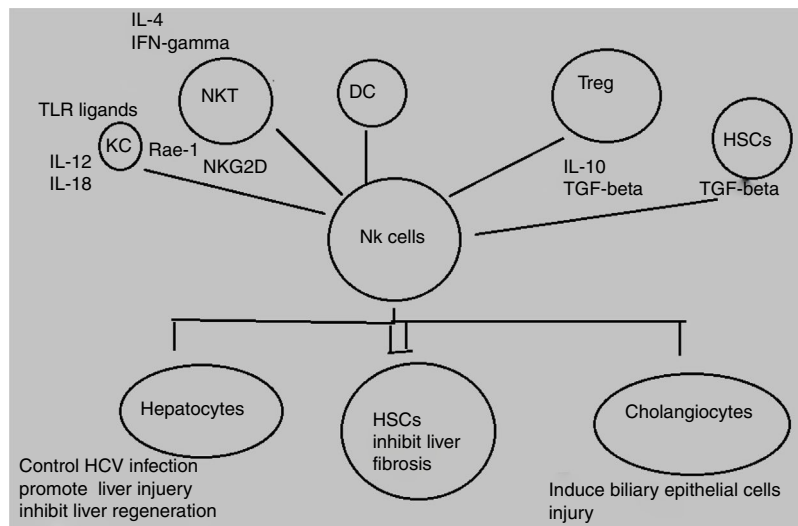


Figure 1 Animal model study has proved that immunopathology and immunodefense of liver cells is greatly influenced by NK cells which impede tumor cell growth, liver fibrosis, and viral infection and also inhibit liver regeneration but increase hepatocellular damage. Activating NK cells can kill HCV-infected hepatocytes leading to control of HCV infection.¹⁴

(cadaveric liver) into recipient (HCC infected individual) is under phase I clinical trial.²⁶

NK cells control liver fibrosis in humans because of cytotoxic nature of NK cells that kill hepatic stellate cells (HSCs) based on FasL, TRAIL, NKG2A, NKG2D, and Nkp46.^{27–29} In case of HCV infection, several studies have shown that the activation of NK cells controls HCV because of reportedly good treatment response associated with higher NK cell cytotoxicity and expression of activating receptors in contrast to, higher expression of inhibitory receptors associated with poor treatment outcome.^{30,31} The potential of NK cells in the control of hepatitis A virus (HAV) infection has been indicated in one study based on HAV-infected fibroblasts.³² Likewise, another study reported the pro-inflammatory cytokine production by NK cells in hepatitis E virus (HEV) infected patients as compared to those with resolved HEV infection.³³ In HCC patients, circulating and intrahepatic NK cells are functionally impaired and tumor-infiltrating NK cells mediated reduced expression of perforin and granzymes as well as reduced cytotoxic potential was observed in healthy controls as compared to HCC patients.³⁴ Similarly, frequency of NK cells and cytotoxic function is reduced in human peripheral blood of alcoholic liver disease (ALD) patients as compared to healthy controls.³⁵ An increased number of NK cells was observed in the livers of nonalcoholic steatohepatitis (NASH) patients as compared to healthy controls and simple steatosis. The study further elucidated an increase in NKG2D dependent activation of NK cells and increase in expression of NKG2D ligands and apoptotic hepatocytes in NASH patients.³⁶ An autoimmune disorder, primary sclerosing cholangitis (PSC) is also associated with tumor necrosis factor- α (TNF- α)-mediated-increased-functionally-impaired NK cell numbers in peripheral blood of patients. Another study revealed two SNPs in NKG2D gene responsible for the development of cholangiocarcinoma.^{37,38} The number of NK cells, perforin expression, and cytotoxicity are increased in the blood and liver of PBC patients.³⁹

Innate like T cells or NKT cells play an important immunoregulatory role in cancer, infectious diseases, and autoimmune diseases of liver.^{40,41} Several studies have shown the migration of NKT cells to liver sinusoids in knock-in *Cxcr6^{gfp/+}* mice administered with galactosylceramide (α GalCer) injection.⁴² The activation and enrichment of NKT cells in liver sinusoids reflects the active participation of these cells in the mechanism of controlling the prevention or induction of inflammation in the liver in various immunological responses.^{43,44} Type 1 NKT cells have been shown to play a pathogenic role in variety of liver disorders such as PBC, con A-induced hepatitis, NAFLD, and *ischemia-reperfusion injury* (IRI), whereas, type 1 NKT cells play protective role in acute liver injury. Type I NKT cell-dependent inhibition of macrophage inflammatory protein-2, KC and TNF- α production inhibited liver injury as well as neutrophil infiltration in model of acute CCl_4 -induced fibrosis and mouse model of cholestasis and biliary obstruction.⁴⁵ From accumulating evidence, it can be inferred that in case of acute liver injury the activation of type 1 NKT cells may be protective whereas, the same cells promote liver injury in chronic conditions.⁴⁶ Likewise, in another study α GalCer-mediated activation of type1 NKT cells resulted in prevention of liver injury by induction of neutrophil apoptosis through STAT-1-dependent mechanism whereas, this activation can also promote neutrophil infiltration and hepatitis in a STAT-6 dependent manner.⁴⁷ The role of NKT cells in humans has not been studied properly. However, an increase in the quantity of pro-inflammatory cytokines especially IL-1, IL-6, IL-8, osteopontin (OPN), and TNF α has been observed in the sera and liver biopsies of human liver with alcoholic hepatitis.⁴³ Likewise, another study revealed that patients with alcoholic hepatitis had higher frequencies of IL-22-producing cells and increased IL-17 plasma levels.^{48,49}

In NAFLD, Tim-3/Gal-9 signaling pathway or KCs activates and results in apoptosis of type 1 NKT cells in liver that cause insulin resistance and steatosis.⁵⁰ Similarly, an increase in number of $\text{CD3}^+\text{CD56}^+$ cells and the expression

Table 1 Function of NK and NKT cells in liver diseases.⁵⁸

Peripheral cells	Liver cells	Cytokines	Functions
NK cells	Tumor, Kupffer cells, HSCs, hepatocytes IFN-alpha, IFN-gamma, IL-12, IL-18, IL-2, IL-15	IFN-gamma	Liver injury ↑
		TNA-alpha	Liver fibrosis ↓
		IL-4, IL-5, IL10, IL-13, IL-22	Liver regeneration ↓ Liver tumor ↓ Hepatitis viral infection ↓
NKT cells	DC, HSCs, IL-12, IL18	IFN-gamma, IL-10, IL-13, IL-22, IL-17, IL-2, TGF-beta, TNF-alpha, GM-CSF	Liver injury ↑ Liver fibrosis ↑↓ Liver regeneration ↓↑ Liver tumor ↑↓ Hepatitis viral infection ↓ Autoimmune disease ↑ Microbial pathogens ↑↓

of CD1d in NASH patients indicates the prominent role of NKT cells in NAFLD.^{51,52} In the same way, autoimmune hepatitis (AIH), is also characterized by secretion of IL-17 from type I NKT cells. Some recent studies have reflected the association of liver disorders' severity with high levels of IL-17 in the portal areas of liver biopsies and serum of the patients infected with AIH, NASH, and primary biliary cirrhosis (PBC) as compared to control subjects.⁵³⁻⁵⁵ In HBV and HCV infection, NKT cells are reported to control the replication of both viruses during early stages of infection whereas, the cells may also cause liver injury through different mechanisms such as the induction of hepatocyte apoptosis, lysis of hepatocytes, inhibition of hepatocyte proliferation, and production of pro-inflammatory cytokines.⁵⁶⁻⁵⁸ Table 1 summarizes the role of NKT cells and NK cells in different liver diseases.

Interaction of NK cells with the activated hepatic stellate cells (HSCs)

HSCs play an important role in liver fibrosis.⁵⁹ The critical step in fibrosis involves two events i.e, activation of HSCs and transdifferentiation into major extracellular matrix-producing cell in fibrotic liver known as myofibroblasts. LPS (lipopolysaccharide) like aberrant stimuli induces the production of chemokines and cytokines from HSCs in both rodents and humans that leads to regulation of hepatic inflammatory and immune responses via their own gene expression.^{60,61} Key factors involved in HSC activation belong to platelet-derived growth factor (PDGF) and transforming growth factor beta1 (TGF-beta1) family.^{62,63} Animal model based study revealed the effective role NK cells in inhibition of liver fibrosis by generating anti-fibrotic cytokine IFN- γ and killing hepatic stellate cells.¹⁶

Hepatocyte damage activates HSCs that cause decrease in NK cell inhibition and increase in NK cell stimulation. Increased amount of retinoic acid produced by early activated HSCs increase the expression of RAE-1 that acts as

ligand for activation of NK cell receptor NKG2D.⁶⁴ RAE-1 combines with MICA and stimulate killing of activated HSCs by NK cells.⁶⁵ This mechanism increases liver fibrosis regression.²⁷ The role of another activating receptor NKp30 has been reported in recent studies.⁶⁶ Activation of HSCs downregulates MHC-1 resulting in increased killing of NK cells and decreased engagement of inhibitory NK cell receptors.²⁸ Another study based on mice model demonstrated that decreasing the expression of inhibitory Ly49 receptor by siRNA mediated silencing increases HSC killing by NK cells and improves liver fibrosis. Inflammatory cytokines especially IFN- α and IFN- γ can further influence this process.⁶⁷ IFN- α increases NK-cell mediated HSC killing by increasing the expression of TRAIL receptor on surface of HSCs. NK cell-derived IFN- γ induce HSC apoptosis and cell cycle arrest and produce antifibrotic effects.^{68,69} Concentration of central regulator in chronic liver disease, transforming growth factor-beta (TGF- β), is increased during chronic liver injury. TGF- β signalling acts as an active participant from early progression of disease to cirrhosis and cancer because of its apoptotic and cytostatic effects in hepatocytes.⁷⁰ Downregulation of NKG2D and 2B4 surface expression lead to suppression of antifibrotic function of NK cells.⁷¹ A recent study has elucidated that activating inhibitory killer immunoglobulin-related receptors iKIR knockdown stimulates NK cells and promotes their antifibrogenic activity in human co-cultures and mice.²⁸ Previously, HSCs were discussed in context of liver fibrosis only whereas, recent studies elucidates its role in liver inflammation via navigation of T lymphocytes into parenchyma, production of inflammatory cytokines, and response towards external signals. Further studies are still required to explore their role in development of hepatitis as well as novel therapeutic target.^{72,73}

The progression as well as onset of hepatocellular carcinoma has been studied in genetically engineered mouse model that showed persistent deregulation of numerous NK cell-related genes in the early stages of the disease. This study suggested the association of early onset of

hepatocarcinogenesis with disruption of NK cell-mediated immune surveillance.⁷⁴ It has been observed that the number of intrahepatic NK cells decreases or impairs in HCC patients specifically during post-surgical recurrence.³⁴ Another study demonstrated that NK cells from peripheral blood mononuclear cells (PBMCs) and tumor infiltrating lymphocytes (TILs) in HCC patients was associated with defective cytokine secretion and cytotoxicity compared to healthy donors whereas, diminished activity of NK cells was observed during the development and invasion of HCC.^{75,76} A change in distribution of NK sub-populations, reduction in CD56-dim NK subset has also been observed in HCC patients.^{77,78} Higher number of NK cells associated with high levels of activating and reduced levels of inhibitory NK receptors play an important role in HCC control. Unlike this, some researchers reported that NK cells efficiently killed different cell lines and eliminated metastases and small HCC lesions in vivo.⁷⁹

Dendritic Cells (DCs)

DCs are rare, bone marrow-derived antigen-presenting cells that play a dominant role in the regulation and induction of immune reactivity. Hepatic dendritic cells (HDCs) are localized in portal areas and modulates hepatic immune responses by presenting antigens to lymphocytes. HDCs are classified into myeloid or classical (mHDCs) and plasmacytoid (PDCA-1+; pHDCs) moreover, mHDCs are further classified into DC103+ /CD11b- type 1 (mHDC1) and DC103- /CD11b+ type 2 (mHDC2) cells.⁸⁰ In healthy livers, HDCs are characterized by high production of kinurenin, IL-10, IL-27, and low capacity to stimulate T-lymphocytes and endocytose antigens leading to a tolerogenic environment in healthy livers.⁸¹ According to recent study, murine CD103+ DCs have been reported to provide protection against steatosis progression towards steatohepatitis.⁸² Altered dendritic cell function leads to immunological changes in hepatic fibrosis. DCs also influence pathogenesis of liver fibrosis. Study based on mouse model demonstrated the role of dendritic cells in altered hepatic immunity during fibrosis and their contribution in regulation of inflammatory milieu within the fibrotic liver. Raised level of inflammatory mediators produced in the fibrotic liver is overturned by decline in dendritic cells. DCs induce T cells, NK cells, and HSCs to trigger proliferation, inflammation and immune responses after liver injury. The immunogenic and proinflammatory impacts of fibrotic dendritic cells were contingent on their secretion of tumor necrosis factor- α . Thus, regulation of DCs may be an effective therapeutic strategy against fibro-inflammatory liver disease.⁸³ DCs play an important role in development of primary biliary cirrhosis⁸⁴ and nonalcoholic fatty liver disease because it has been demonstrated that decrease in DCs decrease the severity of NAFDL.⁸⁵ Likewise, another study suggested indirect involvement of dendritic cells triggering antitumor effects.⁸⁶ DCs have been proved effective therapeutic agents against cancer.⁸⁷

Ninomaya *et al.* demonstrated that immature function and phenotype was exhibited by peripheral blood-derived dendritic cells propagated by hepatocellular carcinoma infected patients compared to normal controls.⁸⁷ Another study has reported the identification of follicular dendritic

cell neoplasm in the liver.⁸⁸ Several other studies suggest the participation of dendritic cell in hepatic granuloma formation and granulomatous liver disease.⁸⁹

In patients suffering from HCC, DCs stimulated allo-genic T cells at lower pace in allogenic mixed leukocytes reaction as compared to DCs from normal healthy controls and liver cirrhosis. Moreover, decreased amounts of IL-12 and lower levels of HLA DR was expressed by DCs of HCC patients compared to normal controls ($p < 0.05$). The prevalence of immature DCs because of high levels of inflammatory cytokines show defective DCs maturation during hepatocarcinogenesis. These findings suggest that the maturation of DCs can act as effective DC-based immune therapy.⁸⁷ Another study have suggested the administration of DCs into cancer nodules to control HCC.⁹⁰

Neutrophils

Neutrophils are most abundant type of white blood cells in human body that have the potential to regulate immune response. Neutrophil recruitment to the liver is mediated by different adhesion molecules during sepsis/endotoxemia and sterile inflammation. Neutrophils migrate to the site of inflammation and mediate hepatocyte injury by producing reactive oxygen species, pro-inflammatory mediators, elastase etc. and after the clearance of inflammation, apoptosis kill neutrophils leading to stimulation of an active program that resolve inflammation.⁹¹ Sustained virological response in association with reduced neutrophil level has been observed in interferon treated HCV patients, likewise, in HBV infected transgenic mice, the inhibition of neutrophil elastase led to improvement in liver injury.⁹² In addition to this, distinct neutrophil subsets have been reported in the peripheral blood of HCC patients. Accumulating evidence suggest that neutrophil dysfunction leads to poor liver cirrhosis outcomes.⁹²⁻⁹⁴ Liu ZX *et al.*, reported that neutrophils kill hepatocytes after expressing Fas ligand via an apoptosis-induced mechanism.⁹⁵ hepatic E-selectin induced infiltration of neutrophils into the liver and its role in pathogenesis of human alcoholic liver disease have been elucidated in a recent study.⁹⁶ Although, the role of immune cells hasn't been studied but it is a known fact that leukocytes present in HCC microenvironment regulates tumor growth.⁹⁷ Another study reported that intratumoral neutrophil-to-CD8⁺ T cell ratio act as better predictor of outcome.^{98,99}

Macrophages

Many experimental models of liver fibrosis demonstrate the role of liver macrophages or kupffer cells in modulating inflammatory response leading to activation of hepatic stellate cells and release of proinflammatory chemokines and cytokines. Macrophages contribute to both fibrosis regression and progression therefore, interactions and differentiation of macrophages with other hepatic cell types in injured liver acts as potential novel target for future therapeutics to combat liver fibrosis (Fig. 2).

Kupffer cells are macrophages that reside in the liver. These cells are activated by CD14/TLR4 receptor complex because of elevated intestinal translocation of

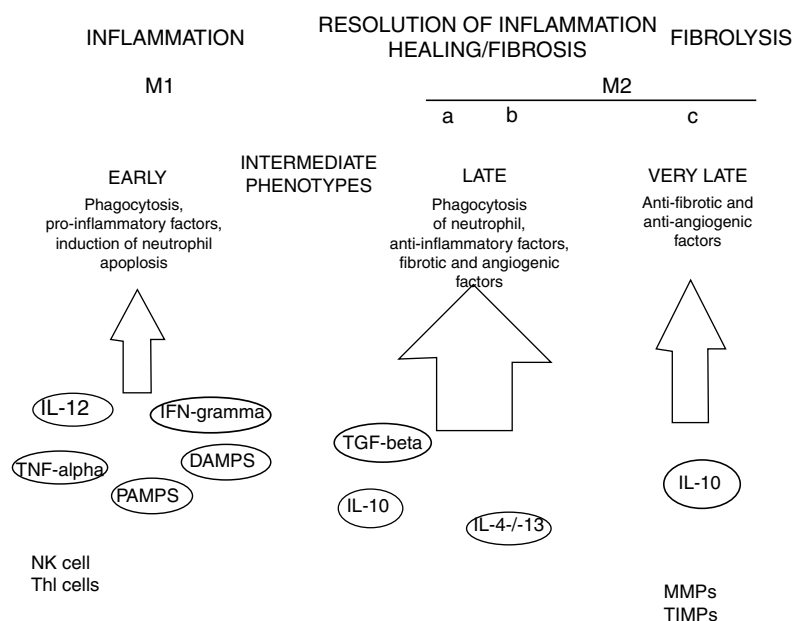


Figure 2 Role of macrophages in renal fibrosis: Activation of parenchymal cells that occurs as a result of tissue damage leads to activation of innate immunity. This involves monocytes recruitment and their differentiation into diverse macrophage phenotypes that rely on local tissue environment. Necrotic cells and pathogens secrete factors that trigger activation of immune receptors such as toll-like receptors as a result of which macrophages are polarized towards 'M1' proinflammatory macrophage. In contrast to this, phagocytic uptake of inflammatory signals especially apoptotic cells supports polarization of macrophages towards anti-inflammatory or profibrotic 'M2' phenotypes. Secretion of proteases by fibrolytic macrophages digests extracellular matrix proteins.⁹⁹

lipopolysaccharides that may contribute to alcohol-induced liver injury.¹⁰⁰ Kupffer cells are necessary for initiating inflammatory responses and sensing tissue injury while infiltrating Ly-6C⁺ monocyte-derived macrophages that are associated with fibrosis and chronic inflammation. Additionally, proliferation of recruited or local macrophages may lead to their accumulation in injured liver. A recent study has reported the association of M2 macrophages with liver inflammation because of proinflammatory cytokines secreted by M2 macrophages in the liver of HCV infected mice.¹⁰¹

A study published in Nature demonstrates that initial response to liver injury is provided by Kupffer cells which produce cytokines and chemokines such as tumor necrosis factor (TNF) α , CCL2, CCL5, IL-1 β leading to recruitment of other immune cells like monocytes. Dramatic expansion of hepatic macrophages owing to massive influx of monocytes into the liver is observed in both chronic and acute liver injuries.^{102,103}

Several other evidences suggest the involvement of macrophages in the induction and resolution of fibrosis. Slight alterations in the pattern of MMP (matrix metalloproteinase) can significantly influence outcomes with MMP1 and MMP13 exhibiting potent antifibrotic activity and macrophage-derived MMP12 increasing fibrosis regression.¹⁰⁴ MMPs promote degradation of extracellular matrix leading to fibrosis regression. MMP9 degrade basement membrane and permits recruited fibroblasts and inflammatory cells to enter injury sites. Secretions of several other factors promote myofibroblast apoptosis and remove cellular debris thus leading to negative regulation of

fibrosis. Macrophage-mediated modifications in the extracellular matrix protein can also influence myofibroblasts' survival and terminate progressive fibrosis.¹⁰⁵ Due to expression of Arg-1, macrophages deplete an essential amino acid that is required for proliferation of myofibroblasts and CD4⁺ T cells thereby helps in down-regulation of profibrotic immune responses.¹⁰⁶ All such findings are concomitant with recent evidences that have proved macrophages important component of resolution of fibrosis.

Tumor associated macrophages (TAMs) promote HCC invasion, metastasis, growth, and angiogenesis. Furthermore, TAMs interact with cancer and stromal cells within tumor microenvironment to suppress antitumor immune response. In HCC, CCL2, M-CSF, TGF β , and VEGF are recruited by TAMs and release number of chemokines, cytokines, and growth factors. In particular, OPN, TNF α , IL-6, and MMPs play an eminent role in metastasis and invasion TGF β and IL-6 favor tumor growth, whereas, the suppression of antitumor response is promoted by TGF β and IL-10.¹⁰⁷

Liver sinusoidal endothelial cells (LSECs)

Liver sinusoidal endothelial cells are highly specialized endothelial cells that are involved in transport lipoproteins, lipids, and nutrients. These cells represent the interface between blood cells and hepatic stellate cell or hepatocytes. LSECs act as permeable barrier and possess highest endocytosis capacity of human cells. LSECs maintain low portal pressure by regulating hepatic vascular tone. LSECs inhibit fibrosis development, intrahepatic vasoconstriction

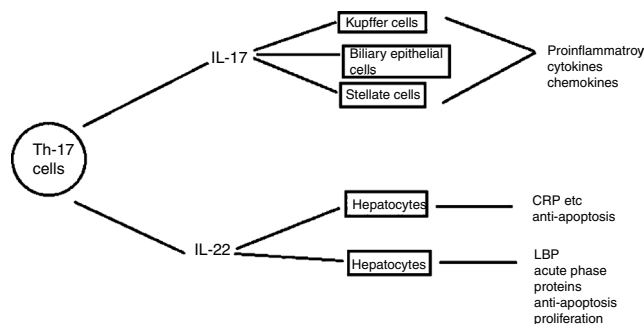


Figure 3 Role of IL-17 in the pathogenesis of liver diseases: IL-17 cells produced by Th17 stimulates different types of liver nonparenchymal cells such as Kupffer cells, monocytes/mDC, biliary epithelial cells, stellate cells to produce chemokines and cytokines that ultimately induce liver inflammation. IL-17 activates hepatic stellate cells thereby, promoting liver fibrogenesis. IL-17 can promote hepatocyte survival by producing acute phase proteins such as C-reactive proteins, LPS-binding protein and myeloid dendritic cells also.¹¹⁰

and maintain hepatic stellate cell quiescence. LSECs are the key contributors of progression and initiation of chronic liver diseases such as hepatocellular carcinoma, and liver lesions associated with infection and inflammation. LSECs have been found to promote vasoconstriction and angiogenesis because they lose their protective properties after getting capillarized. LSEC injury has been reported in diverse liver diseases specifically non-alcoholic fatty liver disease (NAFLD). LSEC progenitors and/or LSECs detect alteration in shear stress occurring as a result of surgery, and they interact with inflammatory cells and platelets.^{108,109}

Th17 cells

Subtype of CD4⁺ T-helper cells known as Th17, which produce IL-17 and IL-22 that triggers host defense against autoimmunity and infections, has recently been discovered. Th17 cells differentiation in response to two cytokines IL-6 and TGF- β that are present in injured liver contributes to hepatic inflammation (Fig. 6).

Fig. 3

Several other studies have demonstrated the activated Th17 cells and Th17-related cytokines in different liver diseases. However, recruitment of Th17 cells specifically CCR4 or CCR6 promoted by chemokine receptors and chemokines might present novel therapeutic targets mediating with migration or differentiation of TH17 in liver disease.¹¹¹ Several other evidences have elucidated the involvement of Th17 cells in variety of human liver diseases such as viral hepatitis,¹¹² primary biliary cirrhosis (PBC),¹¹³ autoimmune hepatitis,¹¹⁴ non-alcoholic steatohepatitis (NASH), hepatocellular carcinoma (HCC), and induced liver injury.¹¹¹

The number of Th17 cells has been found higher in HCC tissues than in non-tumor tissues.¹¹⁵ Several other studies suggested that proinflammatory cytokines produced by HCC tumor-activated monocytes stimulate the proliferation of functional Th17 cells within the tumor tissues.¹¹⁶ In short, the role of Th17 cells in HCC is still to be explored because it has been shown to either inhibit tumor growth by stimulation of cytotoxic T-cell response or promote tumor growth by stimulating angiogenesis or inhibit tumor growth. TH17 cells can also promote the growth of HCC by producing IL-22 that is involved on liver tumor cell proliferation.¹¹⁰

Th22 cells

Host immunity against pathogenic invasion is modulated by Th22 cells. A study demonstrated the role of Th22 cells as dominant inducers of tissue inflammation. Blood, fresh HCC (human hepatocellular carcinoma) tissues and adjacent HCC tissues were collected from HCC infected individuals and healthy individuals. Flow cytometry analysis exhibited elevated levels of serum IL-22, Th22 cells, and Th17 cells in HCC infected patients.¹¹⁷

Th22 cells protect host against chronic hepatitis B.¹¹⁸ A study showed an increase in population of Th22/Th17 cells and related cytokines in drug induced liver injury with hepatocellular injury type.¹¹⁹

Other T cells

Many pathological conditions of liver are driven by regulatory T cells (Tregs) and other T cell such as Th1 and Th2. Proliferation of HSCs is increased by Th1 cells via IFN- γ /STAT1 pathway and is attenuated by Tregs. Wen J *et al.*, found decrease in the frequency of Tregs and increased frequency of Th1, Th2 and Th17 in the peripheral blood of biliary atresia patients.¹²⁰

Cytokines (such as IL-6, IL-22, IL-33, TGF- β , and TNF- α)

All nucleated cell types in our body e.g, monocytes/macrophages, fibroblasts, epithelial cells, lymphocytes etc. produce regulatory peptides known as cytokines. Recently published data have demonstrated a very curious relationship between liver repair and injury.¹²¹ Some cytokines mediate the liver tissue regeneration after injury whereas, the same mediators also mediate apoptosis and necrosis of liver cells, fibrosis, cholestasis, and hepatic inflammation. An example of this phenomenon is ischemia-preconditioning induced protection followed by ischemia-induced injury observed in liver tissues whereas, apoptosis driven ischemia-reperfusion-based liver injury has also been reported.¹²²

A study revealed the correlation of interferon gamma (IFN-gamma), tumor necrosis factor alpha (TNF-alpha), C-reactive protein (CRP), interleukin-1 (IL-1 beta), and interleukin-6 (IL-6) with different hepatic diseases. When serum level of these mediators was investigated in 264 patients infected with chronic liver diseases and 128 individuals in non-cirrhotic stage of chronic hepatic diseases (CHD), a significant increase in serum level of tumor necrosis factor alpha (TNF-alpha), interleukin-1 (IL-1 beta), and interleukin-6 (IL-6) was observed in cirrhotic group of CHD patients. Thus, increased concentration of cytokines in serum is an indicator of cirrhotic liver disease. Therefore, the study suggests that cytokine level increases as a result of liver dysfunction.^{123,124} Cytokines specifically IL-6 are key factors of liver regeneration.¹²²

According to another study inflammatory chemokines or cytokines are key players of alcoholic liver disease. Kupffer cells lead to generation of TNF- α through TLR4 (Toll like receptor-4) thus, playing a significant role during early stage of alcoholic liver disease.

Immunotherapeutic strategies especially cytokine therapy may serve as a substitute to liver transplantation¹²⁵ by restoring normal functioning of liver via regeneration of healthy tissue remnants. TNF- α ¹²⁶ and IL-22¹²⁷ may also form the basis of several novel approaches of treating alcoholic liver disease.

Growth factors (VEGF, PDGF, FGF, and HGF)

Vascular endothelial growth factor (VEGF) and platelet derived growth factor (PDGF) trigger hepatic fibrosis and inflammation.¹²⁸ Abralde JG *et al.*, indicated a significant increase of VEGF concentration in cirrhotic rats.¹²⁹ Likewise, another study suggested that VEGF level was elevated in HCC patients.¹³⁰

Platelet derived growth factor (PDGF) family consists of four members PDGF-A, B, C, and D. PDGF is a potent mitogen to hepatic stellate cells among all polypeptide growth factors.¹³¹ Overexpression of PDGF and its receptors has been observed in liver cirrhosis and activity of PDGF was shown to be increased with degree of liver fibrosis.¹³² Several factors like chemicals, viruses, or mechanical damage to liver cells can induce Kupffer cells to secrete PDGF.¹³³ In response to binding of PDGF with its specific receptors present on membranes of hepatic stellate cells, PDGF activates transcription factors and corresponding signal molecules leading to activation of hepatic stellate cells and downstream target genes.¹³⁴ PDGF has been found to decrease ECM degradation by inhibiting the activity of collagenase and increase the expression expression of TIMP-1, MMP-2, and MMP-9.¹³⁵ Isoform PDGF-D is potent enough to activate hepatic stellate cells and induce fibrogenic and mitogenic effects, therefore, plays dominant role in liver fibrosis.¹³⁶

There are many evidences which highlight the function of fibroblast growth factor in progression of hepatocellular carcinoma (HCC) and metastasis. The expression of FGF2 has been detected in the liver tissues of HCC infected individuals. Likewise, the serum concentration of FGF2 was considerably higher in patients with liver cirrhosis, chronic hepatitis, and HCC compared with healthy individuals.^{137,138}

Hepatocyte growth factor is well known for its morphogenic, motogenic, tumor suppressor, and mitogenic activities.^{139,140} Endogenous HGF repair injured lungs, liver and kidneys etc. It also exerts protective effects on organs through anti-inflammatory and anti-apoptotic signals. Significant increase in concentration of HGF has been observed during organ diseases and infusion of anti-HGF antibody has been shown to accelerate tissue destruction in mice model. Therefore, endogenous HGF is necessary for disease control while inadequate secretion of HGF cause organ failure.¹⁴¹

Collagen-producing cells

Deposition of collagen causes the disruption of liver and lead to cirrhosis.¹⁴² The major collagen producing-cells found in injured liver are portal fibroblasts, myofibroblasts, and activated hepatic cells that are activated by fibrogenic cytokines specifically leptin, angiotensin II, and TGF- β 1.¹⁴³ Hepatic stellate cells (HSCs) are the main matrix-producing cells which play an important role in liver fibrosis. Liver injury activates HSCs which differentiates to fibrogenic myofibroblast-like cells.¹⁴⁴ Activated fibroblasts and other cell types of fibroblast lineage like vascular myofibroblasts or portal fibroblasts have been proved as dominant mediators of liver fibrosis.¹⁴⁵ Collagen 1 has been reported to promote HCC cell proliferation by regulating integrin β 1/FAK signaling.¹⁴⁶

Conclusion

Hepatic immune cells specifically natural killer cells perform many roles after activation, like secretion of cytokines and killing tumor cells and viral-infected cells, and also play important roles in regeneration, liver injury, and fibrosis. The characterization of intrahepatic immune cell functions is offering new opportunities and new therapeutic targets for curbing liver disorders. Although findings of many researchers are promising but further studies are still needed to translate these findings into clinical practice for therapeutics.

Conflict of interest

Authors declare no conflict of interest.

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References

1. Hudspeth K, et al. The role of natural killer cells in autoimmune liver disease: a comprehensive review. *Journal of autoimmunity*. 2013;46:55–65.
2. Janeway CA. The immune system evolved to discriminate infectious nonself from noninfectious self. *Immunology today*. 1992;13:11–6.

3. Takeuchi O, Akira S. Pattern recognition receptors and inflammation. *Cell*. 2010;140:805–20.
4. Bieghs V, Trautwein C. The innate immune response during liver inflammation and metabolic disease. *Trends in immunology*. 2013;34:446–52.
5. Wiltrout RH, et al. Role of organ-associated NK cells in decreased formation of experimental metastases in lung and liver. *The Journal of Immunology*. 1985;134:4267–75.
6. Vermijlen D, et al. Hepatic natural killer cells exclusively kill splenic/blood natural killer-resistant tumor cells by the perforin/granzyme pathway. *Journal of leukocyte biology*. 2002;72:668–76.
7. Wang H, Yin S. Natural killer T cells in liver injury, inflammation and cancer. *Expert review of gastroenterology & hepatology*. 2015;9:1077–85.
8. Radaeva S, et al. Natural killer cells ameliorate liver fibrosis by killing activated stellate cells in nkg2d-dependent and tumor necrosis factor-related apoptosis-inducing ligand-dependent manners. *Gastroenterology*. 2006;130:435–52.
9. O'Leary JG, et al. T cell- and B cell-independent adaptive immunity mediated by natural killer cells. *Nature immunology*. 2006;7:507–16.
10. Syn WK, et al. Accumulation of natural killer T cells in progressive nonalcoholic fatty liver disease. *Hepatology*. 2010;51:1998–2007.
11. Kakimi K, et al. Natural killer T cell activation inhibits hepatitis B virus replication in vivo. *The Journal of experimental medicine*. 2000;192:921–30.
12. Ghosh S, et al. Natural Killer cells contribute to hepatic injury and help in viral persistence during progression of HBeAg-negative chronic HBV infection. *Clinical Microbiology and Infection*. 2016.
13. Maghazachi AA. Role of chemokines in the biology of natural killer cells, in *The Chemokine System in Experimental and Clinical Hematology*. Springer; 2010. p. 37–58.
14. Tian Z, Chen Y, Gao B. Natural killer cells in liver disease. *Hepatology*. 2013;57:1654–62.
15. Raulet DH. Roles of the NKG2D immunoreceptor and its ligands. *Nature Reviews Immunology*. 2003;3:781–90.
16. Gao B. Natural killer group 2 member D, its ligands, and liver disease: good or bad? *Hepatology*. 2010;51:8–11.
17. Amadei B, et al. Activation of natural killer cells during acute infection with hepatitis C virus. *Gastroenterology*. 2010;138:1536–45.
18. Pelletier S, et al. Increased degranulation of natural killer cells during acute HCV correlates with the magnitude of virus-specific T cell responses. *Journal of hepatology*. 2010;53:805–16.
19. Fehniger TA, et al. CD56bright natural killer cells are present in human lymph nodes and are activated by T cell-derived IL-2: a potential new link between adaptive and innate immunity. *Blood*. 2003;101:3052–7.
20. Subleski JJ, et al. Enhanced antitumor response by divergent modulation of natural killer and natural killer T cells in the liver. *Cancer research*. 2006;66:11005–12.
21. Chang CJ, et al. Combined GM-CSF and IL-12 gene therapy synergistically suppresses the growth of orthotopic liver tumors. *Hepatology*. 2007;45:746–54.
22. Holder KA, Russell RS, Grant MD. Natural killer cell function and dysfunction in hepatitis C virus infection. *BioMed research international*. 2014;2014.
23. Houot R, et al. Targeting immune effector cells to promote antibody-induced cytotoxicity in cancer immunotherapy. *Trends in immunology*. 2011;32:510–6.
24. Kohrt HE, et al. Anti-KIR antibody enhancement of anti-lymphoma activity of natural killer cells as monotherapy and in combination with anti-CD20 antibodies. *Blood*. 2014;123:678–86.
25. Alici E. IPH-2101, a fully human anti-NK-cell inhibitory receptor mAb for the potential treatment of hematological cancers. *Current opinion in molecular therapeutics*. 2010;12:724–33.
26. Safety Study of Liver Natural Killer Cell Therapy for Hepatoma Liver Transplantation (MIAMINK), U.S. National Institutes of Health ClinicalTrials.gov Identifier: NCT01147380.
27. Gur C, et al. NKp46-mediated killing of human and mouse hepatic stellate cells attenuates liver fibrosis. *Gut*. 2011. p. gutjnl-2011-301400.
28. Muhanna N, et al. Amelioration of hepatic fibrosis by NK cell activation. *Gut*. 2011;60:90–8.
29. Morishima C, et al. Decreased NK cell frequency in chronic hepatitis C does not affect ex vivo cytolytic killing. *Hepatology*. 2006;43:573–80.
30. Golden-Mason L, et al. Increased natural killer cell cytotoxicity and NKp30 expression protects against hepatitis C virus infection in high-risk individuals and inhibits replication in vitro. *Hepatology*. 2010;52:1581–9.
31. Werner JM, et al. Innate immune responses in hepatitis C virus-exposed healthcare workers who do not develop acute infection. *Hepatology*. 2013;58:1621–31.
32. Baba M, et al. Cytolytic activity of natural killer cells and lymphokine activated killer cells against hepatitis A virus infected fibroblasts. *Journal of clinical & laboratory immunology*. 1993;40:47–60.
33. Das R, Tripathy A. Increased expressions of NKp44, NKp46 on NK/NKT-like cells are associated with impaired cytolytic function in self-limiting hepatitis E infection. *Medical microbiology and immunology*. 2014;203:303–14.
34. Cai L, et al. Functional impairment in circulating and intra-hepatic NK cells and relative mechanism in hepatocellular carcinoma patients. *Clinical Immunology*. 2008;129:428–37.
35. Laso FJ, et al. Chronic alcohol consumption is associated with an increased cytotoxic profile of circulating lymphocytes that may be related with the development of liver injury. *Alcoholism: Clinical and Experimental Research*. 2010;34:876–85.
36. Kahraman A, et al. Major histocompatibility complex class I-related chains A and B (MIC A/B): A novel role in nonalcoholic steatohepatitis. *Hepatology*. 2010;51:92–102.
37. Bo X, et al. Tumour necrosis factor α impairs function of liver derived T lymphocytes and natural killer cells in patients with primary sclerosing cholangitis. *Gut*. 2001;49:131–41.
38. Melum E, et al. Cholangiocarcinoma in primary sclerosing cholangitis is associated with NKG2D polymorphisms. *Hepatology*. 2008;47:90–6.
39. Chuang Y-H, et al. Increased killing activity and decreased cytokine production in NK cells in patients with primary biliary cirrhosis. *Journal of autoimmunity*. 2006;26:232–40.
40. Bendelac A, Savage PB, Teyton L. The biology of NKT cells. *Annu. Rev. Immunol.* 2007;25:297–336.
41. Godfrey DI, Stankovic S, Baxter AG. Raising the NKT cell family. *Nature immunology*. 2010;11:197–206.
42. Geissmann F, et al. Intravascular immune surveillance by CXCR6+ NKT cells patrolling liver sinusoids. *PLoS biology*. 2005;3:e113.
43. Gao B, Bataller R. Alcoholic liver disease: pathogenesis and new therapeutic targets. *Gastroenterology*. 2011;141:1572–85.
44. Kumar V. NKT-cell subsets: promoters and protectors in inflammatory liver disease. *Journal of Hepatology*. 2013;59:618–20.
45. Wintermeyer P, et al. Invariant natural killer T cells suppress the neutrophil inflammatory response in a mouse model of cholestatic liver damage. *Gastroenterology*. 2009;136:1048–59, e2.

46. Duwaerts CC, et al. Cross-activating invariant NKT cells and kupffer cells suppress cholestatic liver injury in a mouse model of biliary obstruction. *PLoS One*. 2013;8:e79702.
47. Wang H, et al. Invariant NKT cell activation induces neutrophil accumulation and hepatitis: Opposite regulation by IL-4 and IFN- γ . *Hepatology*. 2013;58:1474–85.
48. Støy S, et al. Cytotoxic T lymphocytes and natural killer cells display impaired cytotoxic functions and reduced activation in patients with alcoholic hepatitis. *American Journal of Physiology-Gastrointestinal and Liver Physiology*. 2015;308:G269–76.
49. Bandyopadhyay K, Marrero I, Kumar V. NKT cell subsets as key participants in liver physiology and pathology. *Cellular and Molecular Immunology*. 2016;13:337.
50. Kremer M, et al. Kupffer cell and interleukin-12-dependent loss of natural killer T cells in hepatosteatosis. *Hepatology*. 2010;51:130–41.
51. Syn W-K, et al. NKT-associated hedgehog and osteopontin drive fibrogenesis in non-alcoholic fatty liver disease. *Gut*. 2012. p. gntjnl-2011-301857.
52. Wolf MJ, et al. Metabolic activation of intrahepatic CD8+ T cells and NKT cells causes nonalcoholic steatohepatitis and liver cancer via cross-talk with hepatocytes. *Cancer cell*. 2014;26:549–64.
53. Halder RC, et al. Type II NKT cell-mediated anergy induction in type I NKT cells prevents inflammatory liver disease. *Journal of Clinical Investigation*. 2007;117:2302.
54. Corrigan M, et al. Autoimmune hepatitis: an approach to disease understanding and management. *British medical bulletin*. 2015;114.
55. Lafdil F, et al. Th17 cells and their associated cytokines in liver diseases. *Cellular and Molecular Immunology*. 2010;7:250.
56. Malhi H, Gores GJ. Cellular and molecular mechanisms of liver injury. *Gastroenterology*. 2008;134:1641–54.
57. Vilarinho S, et al. Blockade of NKG2D on NKT cells prevents hepatitis and the acute immune response to hepatitis B virus. *Proceedings of the National Academy of Sciences*. 2007;104:18187–92.
58. Gao B, Radaeva S, Park O. Liver natural killer and natural killer T cells: immunobiology and emerging roles in liver diseases. *Journal of leukocyte biology*. 2009;86:513–28.
59. Baeck C, Tacke F. Balance of inflammatory pathways and interplay of immune cells in the liver during homeostasis and injury. *EXCLI journal*. 2014;13:67.
60. Fujita T, et al. Hepatic stellate cells relay inflammation signaling from sinusoids to parenchyma in mouse models of immune-mediated hepatitis. *Hepatology*. 2015.
61. Harvey SA, et al. The transcriptomic response of rat hepatic stellate cells to endotoxin: implications for hepatic inflammation and immune regulation. *PLoS One*. 2013;8:e82159.
62. Gressner A, Weiskirchen R. Modern pathogenetic concepts of liver fibrosis suggest stellate cells and TGF- β as major players and therapeutic targets. *Journal of cellular and molecular medicine*. 2006;10:76–99.
63. Liedtke C, et al. Experimental liver fibrosis research: update on animal models, legal issues and translational aspects. *Fibrogenesis & tissue repair*. 2013;6:1.
64. Radaeva S, et al. Retinoic acid signaling sensitizes hepatic stellate cells to NK cell killing via upregulation of NK cell activating ligand RAE1. *American Journal of Physiology-Gastrointestinal and Liver Physiology*. 2007;293:G809–16.
65. Krizhanovsky V, et al. Senescence of activated stellate cells limits liver fibrosis. *Cell*. 2008;134:657–67.
66. Mantovani S, et al. NKp30 isoforms in patients with chronic hepatitis C virus infection. *Immunology*. 2015;146:234–42.
67. Jeong WI, et al. STAT1 inhibits liver fibrosis in mice by inhibiting stellate cell proliferation and stimulating NK cell cytotoxicity. *Hepatology*. 2006;44:1441–51.
68. Ahlenstiel G, et al. Early changes in natural killer cell function indicate virologic response to interferon therapy for hepatitis C. *Gastroenterology*. 2011;141:1231–9, e2.
69. Stegmann KA, et al. Interferon- α -induced TRAIL on natural killer cells is associated with control of hepatitis C virus infection. *Gastroenterology*. 2010;138:1885–97, e10.
70. Fabregat I, et al. TGF- β signalling and liver disease. *The FEBS journal*. 2016;283:2219–32.
71. Sun C, et al. TGF- β 1 down-regulation of NKG2D/DAP10 and 2B4/SAP expression on human NK cells contributes to HBV persistence. *PLoS Pathog*. 2012;8:e1002594.
72. Mizuhara H, et al. T cell activation-associated hepatic injury: mediation by tumor necrosis factors and protection by interleukin 6. *The Journal of experimental medicine*. 1994;179:1529–37.
73. Reifart J, et al. Modulating CD4+ T cell migration in the postischemic liver: hepatic stellate cells as new therapeutic target? *Transplantation*. 2015;99:41–7.
74. Coulouarn C, et al. Genomic modeling of tumor onset and progression in a mouse model of aggressive human liver cancer. *Carcinogenesis*. 2011;32:1434–40.
75. Jinushi M, et al. Impairment of natural killer cell and dendritic cell functions by the soluble form of MHC class I-related chain A in advanced human hepatocellular carcinomas. *Journal of Hepatology*. 2005;43:1013–20.
76. Wu Y, et al. Monocyte/macrophage-elicited natural killer cell dysfunction in hepatocellular carcinoma is mediated by CD48/2B4 interactions. *Hepatology*. 2013;57:1107–16.
77. He L, Wang X, Montell DJ. Shining light on Drosophila oogenesis: live imaging of egg development. *Current opinion in genetics & development*. 2011;21:612–9.
78. Une Y, Kawata A, Uchino J. Adopted immunochemotherapy using IL-2 and spleen LAK cell-randomized study. *Nihon Geka Gakkai Zasshi*. 1991;92:1330–3.
79. Lygidakis N, et al. Hepatocellular carcinoma: surgical resection versus surgical resection combined with pre-and post-operative locoregional immunotherapy-chemotherapy. A prospective randomized study. *Anticancer research*. 1995;15:543–50.
80. Bernsmeier C, Albano E. Liver dendritic cells and NAFLD evolution: A remaining open issue. *Elsevier*; 2017.
81. Thomson AW, Knolle PA. Antigen-presenting cell function in the tolerogenic liver environment. *Nature reviews. Immunology*. 2010;10:753.
82. Heier E-C, et al. Murine CD103+ dendritic cells protect against steatosis progression towards steatohepatitis. *Journal of Hepatology*. 2017;66:1241–50.
83. Connolly MK, et al. In liver fibrosis, dendritic cells govern hepatic inflammation in mice via TNF- α . *The Journal of clinical investigation*. 2009;119:3213–25.
84. Kaji K, et al. B7-2 positive cells around interlobular bile ducts in primary biliary cirrhosis and chronic hepatitis C. *Journal of gastroenterology and hepatology*. 1997;12:507–12.
85. Almeda-Valdes P, et al. The Role of Dendritic Cells in Fibrosis Progression in Nonalcoholic Fatty Liver Disease. *BioMed research international*. 2015;2015.
86. Péron J-M, et al. FLT3-ligand administration inhibits liver metastases: role of NK cells. *The Journal of Immunology*. 1998;161:6164–70.
87. Ninomiya T, et al. Dendritic cells with immature phenotype and defective function in the peripheral blood from patients with hepatocellular carcinoma. *Journal of Hepatology*. 1999;31:323–31.
88. Shek T, et al. Intra-abdominal follicular dendritic cell tumour: a rare tumour in need of recognition. *Histopathology*. 1998;33:465–70.
89. Yoneyama H, et al. Regulation by chemokines of circulating dendritic cell precursors, and the formation of portal

- tract-associated lymphoid tissue, in a granulomatous liver disease. *The Journal of experimental medicine*. 2001;193:35–50.
90. Kumagi T, et al. Administration of dendritic cells in cancer nodules in hepatocellular carcinoma. *Oncology reports*. 2005;14:969–73.
 91. Xu R, et al. The role of neutrophils in the development of liver diseases. *Cellular and Molecular Immunology*. 2014;11:224.
 92. Mookerjee RP, et al. Neutrophil dysfunction in alcoholic hepatitis superimposed on cirrhosis is reversible and predicts the outcome. *Hepatology*. 2007;46:831–40.
 93. Gungor G, et al. Neutrophil gelatinase-associated lipocalin in prediction of mortality in patients with hepatorenal syndrome: a prospective observational study. *Liver International*. 2014;34:49–57.
 94. Tritto G, et al. Evidence of neutrophil functional defect despite inflammation in stable cirrhosis. *Journal of Hepatology*. 2011;55:574–81.
 95. Liu ZX, et al. Neutrophil depletion protects against murine acetaminophen hepatotoxicity. *Hepatology*. 2006;43:1220–30.
 96. Bertola A, Park O, Gao B. Chronic plus binge ethanol feeding synergistically induces neutrophil infiltration and liver injury in mice: a critical role for E-selectin. *Hepatology*. 2013;58:1814–23.
 97. Wilson C, et al. NFκB1 is a suppressor of neutrophil-driven hepatocellular carcinoma. *Nature Communications*. 2015;6.
 98. Li Y-W, et al. Intratumoral neutrophils: a poor prognostic factor for hepatocellular carcinoma following resection. *Journal of Hepatology*. 2011;54:497–505.
 99. Lech M, Anders H-J. Macrophages and fibrosis: How resident and infiltrating mononuclear phagocytes orchestrate all phases of tissue injury and repair. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*. 2013;1832:989–97.
 100. Enomoto N, et al. Kupffer Cell Sensitization by Alcohol Involves Increased Permeability to Gut-Derived Endotoxin. *Alcoholism: Clinical and Experimental Research*. 2001;25(s2):515–45.
 101. Ohtsuki T, et al. M2 macrophages play critical roles in progression of inflammatory liver disease in hepatitis C virus transgenic mice. *Journal of virology*. 2016;90:300–7.
 102. Ju C, Tacke F. Hepatic macrophages in homeostasis and liver diseases: from pathogenesis to novel therapeutic strategies. *Cellular & molecular immunology*. 2016;13:316–27.
 103. Krenkel O, Tacke F. Liver macrophages in tissue homeostasis and disease. *Nature Reviews Immunology*. 2017;17:306–21.
 104. Fallowfield JA, et al. Scar-associated macrophages are a major source of hepatic matrix metalloproteinase-13 and facilitate the resolution of murine hepatic fibrosis. *The Journal of Immunology*. 2007;178:5288–95.
 105. Issa R, et al. Mutation in collagen-1 that confers resistance to the action of collagenase results in failure of recovery from CCl4-induced liver fibrosis, persistence of activated hepatic stellate cells, and diminished hepatocyte regeneration. *The FASEB Journal*. 2003;17:47–9.
 106. Pesce JT, et al. Arginase-1-expressing macrophages suppress Th2 cytokine-driven inflammation and fibrosis. *PLoS Pathog*. 2009;5:e1000371.
 107. Capece D, et al. The inflammatory microenvironment in hepatocellular carcinoma: a pivotal role for tumor-associated macrophages. *BioMed Research International*. 2013;2013.
 108. Poisson J, et al. Liver sinusoidal endothelial cells: physiology and role in liver diseases. *Journal of Hepatology*. 2016.
 109. Miyao M, et al. Pivotal role of liver sinusoidal endothelial cells in NAFLD/NASH progression. *Laboratory Investigation*. 2015;95:1130–44.
 110. Lafdil F, et al. Th17 cells and their associated cytokines in liver diseases. *Cellular & molecular immunology*. 2010;7:250–4.
 111. Hammerich L, Heymann F, Tacke F. Role of IL-17 and Th17 cells in liver diseases. *Clinical and Developmental Immunology*. 2010;2011.
 112. Sun H, et al. Increased Th17 cells contribute to disease progression in patients with HBV-associated liver cirrhosis. *Journal of viral hepatitis*. 2012;19:396–403.
 113. Rong G, et al. Imbalance between T helper type 17 and T regulatory cells in patients with primary biliary cirrhosis: the serum cytokine profile and peripheral cell population. *Clinical & Experimental Immunology*. 2009;156:217–25.
 114. Longhi MS, et al. Inhibition of interleukin-17 promotes differentiation of CD25+ cells into stable T regulatory cells in patients with autoimmune hepatitis. *Gastroenterology*. 2012;142:1526–35, e6.
 115. Zhang J-P, et al. Increased intratumoral IL-17-producing cells correlate with poor survival in hepatocellular carcinoma patients. *Journal of Hepatology*. 2009;50:980–9.
 116. Kuang D-M, et al. Activated monocytes in peritumoral stroma of hepatocellular carcinoma foster immune privilege and disease progression through PD-L1. *Journal of Experimental Medicine*. 2009;206:1327–37.
 117. Qin S, et al. Th22 cells are associated with hepatocellular carcinoma development and progression. *Chinese Journal of Cancer Research*. 2014;26:135–41.
 118. Cobleigh MA, Robek MD. Protective and pathological properties of IL-22 in liver disease: implications for viral hepatitis. *The American journal of pathology*. 2013;182:21–8.
 119. Lai R, et al. Protective effect of Th22 cells and intrahepatic IL-22 in drug induced hepatocellular injury. *Journal of Hepatology*. 2015;63:148–55.
 120. Wen J, et al. Interactions between Th1 cells and Tregs affect regulation of hepatic fibrosis in biliary atresia through the IFN-γ/STAT1 pathway. *Cell death and differentiation*. 2017;24:997.
 121. Tilg H. Cytokines and liver diseases. *Canadian journal of gastroenterology= Journal canadien de gastroenterologie*. 2001;15:661–8.
 122. Galun E, Axelrod JH. The role of cytokines in liver failure and regeneration: potential new molecular therapies. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*. 2002;1592:345–58.
 123. Tilg H, et al. Serum levels of cytokines in chronic liver diseases. *Gastroenterology*. 1992;103:264–74.
 124. Goral V, Atayan Y, Kaplan A. Relation between pathogenesis of liver cirrhosis, hepatic encephalopathy and serum cytokine levels 嗶 what is the role of tumor necrosis factor 嗶 alpha? *Journal of Chinese clinical medicine*. 2010;5.
 125. Chen X, et al. Cytokine and human leukocyte antigen (HLA) profile for graft-versus-host disease (GVHD) after organ transplantation. *European journal of medical research*. 2016;21:38.
 126. Kawaratani H, et al. The effect of inflammatory cytokines in alcoholic liver disease. *Mediators of inflammation*. 2013;2013.
 127. Gao B. Hepatoprotective and anti-inflammatory cytokines in alcoholic liver disease. *Journal of gastroenterology and hepatology*. 2012;27(s2):89–93.
 128. Desouky M. VEGF and PDGF in liver cirrhosis and their relation to echocardiographic parameters and Carotid Intima-Media Thickness. *Life Science Journal*. 2013;10.
 129. Abraldes JG, et al. Mild increases in portal pressure upregulate vascular endothelial growth factor and endothelial nitric oxide synthase in the intestinal microcirculatory bed, leading to a hyperdynamic state. *American Journal of Physiology-Gastrointestinal and Liver Physiology*. 2006;290:G980–7.
 130. Ghonaim MM, El-Edel RH. Significance of Serum Vascular Endothelial Growth Factor in Chronic Liver Disease and Hepatocellular Carcinoma: An Exploratory Study. *Ibnosina Journal of Medicine and Biomedical Sciences*. 2013;5:288–95.

131. Martin IV, et al. Platelet-derived growth factor (PDGF)-C neutralization reveals differential roles of PDGF receptors in liver and kidney fibrosis. *The American journal of pathology*. 2013;182:107–17.
132. Cao S, et al. Neuropilin-1 promotes cirrhosis of the rodent and human liver by enhancing PDGF/TGF- β signaling in hepatic stellate cells. *The Journal of clinical investigation*. 2010;120:2379–94.
133. Borkham-Kamphorst E, et al. Dominant-negative soluble PDGF- β receptor inhibits hepatic stellate cell activation and attenuates liver fibrosis. *Laboratory investigation*. 2004;84:766–77.
134. Pinzani M. PDGF and signal transduction in hepatic stellate cells. *Frontiers in bioscience: a journal and virtual library*. 2002;7:d1720–6.
135. Czochra P, et al. Liver fibrosis induced by hepatic overexpression of PDGF-B in transgenic mice. *Journal of hepatology*. 2006;45:419–28.
136. Borkham-Kamphorst E, et al. Pro-fibrogenic potential of PDGF-D in liver fibrosis. *Journal of hepatology*. 2007;46:1064–74.
137. Tsunematsu H, et al. Fibroblast growth factor-2 enhances NK sensitivity of hepatocellular carcinoma cells. *International journal of cancer*. 2012;130:356–64.
138. Gauglhofer C, et al. Up-regulation of the fibroblast growth factor 8 subfamily in human hepatocellular carcinoma for cell survival and neoangiogenesis. *Hepatology*. 2011;53:854–64.
139. Ishiki Y, et al. Direct evidence that hepatocyte growth factor is a hepatotrophic factor for liver regeneration and has a potent antihepatitis effect in vivo. *Hepatology*. 1992;16:1227–35.
140. Fjuiwara K, et al. Stimulation of liver growth by exogenous human hepatocyte growth factor in normal and partially hepatectomized rats. *Hepatology*. 1993;18:1443–9.
141. Nakamura T, Mizuno S. The discovery of hepatocyte growth factor (HGF) and its significance for cell biology, life sciences and clinical medicine. *Proceedings of the Japan Academy, Series B*. 2010;86:588–610.
142. McGee JD. Collagen deposition in liver disease. *Annals of the rheumatic diseases*. 1977;36 Suppl 2:29.
143. Bataller R, Brenner DA. Liver fibrosis. *The Journal of clinical investigation*. 2005;115:209–18.
144. Moreira RK. Hepatic stellate cells and liver fibrosis. *Archives of pathology & laboratory medicine*. 2007;131:1728–34.
145. Knittel T, et al. Rat liver myofibroblasts and hepatic stellate cells: different cell populations of the fibroblast lineage with fibrogenic potential. *Gastroenterology*. 1999;117:1205–21.
146. Zheng X, et al. Collagen I promotes hepatocellular carcinoma cell proliferation by regulating integrin β 1/FAK signaling pathway in nonalcoholic fatty liver. *Oncotarget*. 2017;8:95586.