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Clinical and microbiological profile of infectious keratitis in an area of Madrid, Spain



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ABSTRACT

Introduction: To study antibiotic susceptibility in bacterial keratitis (BK), its profile over 10 years and its influence on ophthalmological practice.

Methods: Retrospective review of BK with positive corneal scraping over a 10-year period. Risk factors for keratitis, visual acuity (VA), empirical topical treatment, corneal infection characteristics and outcomes were analyzed for BK due to *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Streptococcus pneumoniae*, *Pseudomonas aeruginosa* and *Propionibacterium acnes*.

Results: 389 positive corneal scrapings were collected. All Gram-positive bacteria were susceptible to vancomycin. *P. aeruginosa* demonstrated >90% sensitivity to the most-commonly-used topical antibiotics. Susceptibility to methicillin was 90.2% for *S. aureus* and 66.3% for *S. epidermidis*. The results of 215 patients were available. 1.9% required enucleation and 2.8% required surgical treatments. Final VA improved after treatment in keratitis due to *S. aureus* ($p = 0.026$) and *S. epidermidis* ($p = 0.005$). There was a correlation between *S. aureus* resistance to methicillin ($p = 0.002$) and levofloxacin ($p = 0.043$) and enucleation (20% and 10%, respectively) compared with a 0% rate of enucleation in *S. aureus*-susceptible keratitis.

Conclusions: BK due to *S. pneumoniae* is very aggressive irrespective of antibiotic sensitivity. *S. aureus* was frequently isolated in patients with systemic diseases. It causes severe keratitis and remains moderately resistant to methicillin and levofloxacin. For this reason, keeping vancomycin in empirical regimens is believed to be necessary.

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Perfil clínico y microbiológico de la queratitis infecciosa en un área de Madrid, España

RESUMEN

Palabras clave:

Susceptibilidad a antibióticos

Queratitis bacteriana

Infecciones corneales

Queratitis

Enucleación

Introducción: Estudiar la susceptibilidad antibiótica en queratitis bacteriana (QB), el perfil temporal a lo largo de 10 años y su influencia en la clínica ocular.

Métodos: Revisión retrospectiva durante un periodo de 10 años de QB con raspado corneal positivo. Se analizaron los factores de riesgo de queratitis, la agudeza visual (AV), el tratamiento empírico tópico, las características de la infección corneal y el resultado clínico para QB por *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Streptococcus pneumoniae*, *Pseudomonas aeruginosa* y *Propionibacterium acnes*.

Resultados: Se recogieron 389 raspados corneales positivos. Todas las bacterias grampositivas fueron susceptibles a la vancomicina. *P. aeruginosa* presentó sensibilidad mayor del 90% a los antibióticos tópicos más comúnmente utilizados. La susceptibilidad a la meticilina fue del 90,2% para *S. aureus* y del 66,3% para *S. epidermidis*. Los resultados clínicos estaban disponibles para 215 pacientes. El 1,9% requirieron

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enucleación y el 2,8% tratamientos quirúrgicos. La AV final mejoró después del tratamiento en queratitis por *S. aureus* ($p=0,026$) y por *S. epidermidis* ($p=0,005$). Hubo correlación entre la resistencia de *S. aureus* a la meticilina ($p=0,002$) y levofloxacino ($p=0,043$) y enucleación (20 y 10%, respectivamente) en comparación con una tasa de enucleación del 0% en *S. aureus* susceptible.

Conclusiones: Las QB por *S. pneumoniae* son muy agresivas independientemente de la sensibilidad antibiótica. *S. aureus* se aisló con frecuencia en pacientes con enfermedades sistémicas, causa queratitis severa y permanece moderadamente resistente a la meticilina y a levofloxacino; debido a esto, consideramos necesario mantener la vancomicina en la pauta empírica.

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Introduction

Bacterial infectious keratitis (BK) is a common reason of consulting in ophthalmology and is associated with high morbidity. All cases of moderate to severe keratitis require a detailed laboratory work-up, which ensures that if there is partial or no response to initial therapy, the antimicrobial treatment can be modified based on the results of culture and susceptibility tests. Generally, broad-spectrum antibiotics are used as empiric first-line treatment for presumed BK after obtaining appropriate corneal scrapes. The drugs chosen as initial therapy are either commercially available quinolone or a combination of fortified antibiotics, topical solutions prepared from parenteral antibiotics by reconstituting them with sterile injection water¹ or Balanced Salt Solution (BSS)² with one agent largely directed against Gram-positive and the other against Gram-negative organisms.

The maintenance of the effectiveness of the empiric therapy requires a low resistance rate of the bacteria that can cause keratitis. Longitudinal epidemiologic studies provide clinicians with vital information on the changing microbiological pattern of keratitis in their specific area concerning causative organisms and their antibiotic sensitivities,^{3–5} which is critical for choosing the most suitable empirical regimen. These descriptive studies are gaining importance because monotherapy quinolone is currently being proposed as the main empirical option for bacterial keratitis although caution seems advisable when using monotherapy for any serious bacterial corneal ulcer.⁶

The main purpose of this study is to know the causative bacteria of corneal infection in our area, and their susceptibility profile. Secondary purpose are to analyze the temporal profile of the antibiotic susceptibility to the most relevant topical antibacterial agents, to describe the clinical presentation of the most frequent bacterial keratitis groups and to study if antibiotic resistance has any association with the clinical outcome. Significant trends in these factors over the 10 years of the study were sought to help to select the most convenient empiric initial regimen for BK in our area.

Methods

A retrospective audit was performed of the isolate records of different episodes of patients with symptoms and biomicroscopic signs of BK who had a positive corneal scrape from January 2006 to December 2015 at a tertiary Hospital in Madrid, Spain. All the corneal smears and cultures were typically indicated in our hospital in cases of corneal infiltrates with at least one of the following criteria: dense infiltrate, epithelial ulcer of central location, association with anterior chamber cells 1+ (10 cells or greater in a 1-mm beam), absence or partial response to broad spectrum antibiotic therapy and/or any infiltrate with clinical features suggestive of fungal, amoebic or mycobacterial keratitis. As well, samples were obtained usually in case of infiltrates or epithelial ulcers in relation with contact lens (CL) users.

Specimens were collected in the cornea unit, emergency room or surgery room by corneal scraping procedure and direct inoculation onto the appropriate culture media by the 24 h on call microbiologist at any day or night time. Gram stains were immediately performed on smears, whereas blood, chocolate blood agar plate with hemin and vitamin K1 and Sabouraud agar plates were inoculated. Incubation time for plates varied between 7 and 10 days from blood and chocolate agar plates, and up to 4 weeks for Sabouraud agar plates. Blood and chocolate agar plates were incubated in carbon dioxide environments at 35 °C, whereas two Sabouraud agar plates were incubated in oxygen environments at 25 °C and 35 °C. A second blood agar plate with hemin and vitamin K1 was incubated in an anaerobic environment at 35 °C for 14 days.

To exclude accidental contaminants, the minimum criterion for a positive culture was the growth of at least 3 colonies on one solid medium with similar morphology to the Gram stain, if this was positive. Cultures that isolated multiple organisms were analyzed separately. Antibiotic resistance was determined by broth microdilution to determinate the Minimum Inhibitory Concentrations. The antibiotics tested for the isolated microorganisms are described in Table 1. The interpretations for sensitive, intermediate and resistant were in accordance with EUCAST 4.0/2014 standards.⁷

Risk factors for keratitis, initial and final visual acuity (VA) (final VA was the VA on the last visit before medical discharge for the episode), topical empirical treatment, characteristics of the corneal infection and clinical outcome were analyzed from patients with a positive corneal scrape for *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Streptococcus pneumoniae*, *Pseudomonas aeruginosa* and *Propionibacterium acnes* keratitis. Influence of antibiotic susceptibility to clinical outcome (enucleation, corneal surgery and number of days of antibiotic treatment) was analyzed.

The statistical analysis was performed using SPSS 11.5.⁸ Causative bacteria of keratitis and their susceptibility profile were described with their relative frequency and counts. Lineal by lineal association test was used to analyze the temporal profile of the antibiotic susceptibility to the most relevant topical antibacterial agents between 3 periods, over the 10 years of the study. For the description of the keratitis episodes clinical data, quantitative variables were described with their mean and the standard deviation (SD) and the days of treatment variable with median and interquartile range. The qualitative variables were described with their relative frequency and counts. Finally, to study if antibiotic resistance has any effect in the clinical outcome, a comparative statistical analysis has been done: the Fisher Test was used in 2 × 2 cross tables, in any other case Chi-square was considered. For quantitative variable (days of treatment) Mann-Whitney test was used. To compare the two related samples of visual acuity (VA), the Wilcoxon Test was used.

Table 1
Antibiotics tested for isolated microorganisms.

Isolated bacteria	Isolated bacteria
<i>Abiotrophia defectiva</i>	P CTX MER VAN ERI CLI LEV RI
<i>Acinetobacter baumannii</i>	CF CTX IMI MER DOR COL G TO AK TIG CIP LEV T/S NIT
<i>Alcaligenes faecalis</i>	TIC A/S P/T CF CTZ CFP C/C AZT IMI MER COL G TO AK NET MIN CIP LEV TS FOS
<i>Arthrobacter aurescens</i>	OX A/C CX VAN G TO AK ERI CLI TE CIP LEV T/S FOS
<i>Corynebacterium macginleyi</i>	P AM CFX CTX CFP MER VAN TEI ERI CLI TE CIP T/S RI LEV
<i>Corynebacterium propinquum</i>	P CTX CFP S VAN ERI CLI TE CIP T/S RI
<i>Corynebacterium pseudodiphthericum</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Corynebacterium spp.</i>	P CXT CFP MER VAN ERI CLI TE CIP T/S RI
<i>Corynebacterium tuberculosis</i>	P MER VAN ERI CLI CIP T/S RI
<i>Eikenella corrodens</i>	BLA AMIX
<i>Enterobacter cloacae</i>	TIG AM A/C P/T CF CFZ CFX CX CTX CTZ CFP C/C C/C AZT ETP IMI COL G TO AK AL CIP T/S NIT
<i>Enterococcus faecalis</i>	P AM OX CX A/C VAN TEI DAP G TO AK 100 G5 ERI CLI Q/D LNZ MIN TE CIP LEV T/S FOS NIT RI
<i>Escherichia coli</i>	AMX A/C P/T CF CFX CX CTX CTZ CFP C/C C/C AZT ETP IMI MER COL G TO AK MIN NAL CIP T/S FOS NIT
<i>Granulicatella adiacens</i>	P CTX MER VAN ERI CLI LEV RI
<i>Haemophilus influenzae</i>	BLA P AM AMX A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Lactobacillus gasseri</i>	TIC A/S P/T CF CTZ CFP AZT IMI MER COL G TO AK NET MIN NAL CIP LEV T/S FOS NIT
<i>Moraxella lacunata</i>	BLA P AM AMX A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI LEV TE CIP T/S C RI
<i>Paenibacillus spp.</i>	P AM OX A/C VAN G 100 G5 ERI CLI LEV TS FOS
<i>Propionibacterium acnes</i>	P AMX PIP A/C P/T CX IMI CLI C MTR
<i>Proteus mirabilis</i>	TIG AM A/C P/T CF CFZ CFX CX CTX CTZ CFP C/C C/C AZT ETP IMI COL G TO AK NAL CIP T/S NIT
<i>Pseudomonas aeruginosa</i>	TIC A/S P/T CF CTZ CFP AZT IMI MER COL G TO AK NET MIN NAL CIP LEV T/S FOS NIT
<i>Serratia marcescens</i>	TIG AM A/C P/T CF CFZ CFX CX CTX CTZ CFP AZT ETP IMI COL G TO AK NAL CIP T/S NIT
<i>Staphilococcus hominis</i>	P AM OX A/C VAN TEI DAP G TO AK 100 G5 ERI CLI Q/D LNZ AFU MUP MIN LEV T/S FOS NIT RI
<i>Staphylococcus aureus</i>	P AM OX A/C CX VAN TEI DAP G TO AK 100 G5 ERI CLI Q/D LNZ AFU MUP MIN TE CIP LEV T/S FOS NIT RI
<i>Staphylococcus capitis</i>	P AM OX A/C CX VAN TEI DAP G TO AK ERI CLI LNZ AFU MUP TE CIP LEV T/S FOS NIT RI
<i>Staphylococcus cohnii</i>	P OX A/C CX VAN TEI DAP G TO AK ERI CLI LNZ AFU MUP TE CIP LEV T/S FOS NIT RI
<i>Staphylococcus epidermidis</i>	P AM OX A/C VAN TEI DAP G TO AK 100 G5 ERI CLI Q/D LNZ AFU MUP MIN LEV T/S FOS NIT RI
<i>Staphylococcus haemolyticus</i>	P AM OX A/C VAN TEI DAP G TO AK 100 G5 ERI CLI Q/D LNZ AFU MUP TE CIP MIN LEV T/S FOS NIT RI
<i>Staphylococcus intermedius</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Staphylococcus warneri</i>	P AM OX A/C VAN TEI DAP G TO AK 100 G5 ERI CLI Q/D LNZ AFU MUP MIN LEV T/S FOS NIT RI
<i>Stenotrophomonas maltophilia</i>	AMX A/C P/T CF CFX CX CTX CTZ CFP C/C C/C AZT ETP IMI MER COL G TO AK MIN CIP T/S FOS
<i>Streptococco gordonii</i>	P AM AC CFX CTX CFR CFP MER VAN ERI CL CLI T/S
<i>Streptococcus agalactiae</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE LEV T/S C RI
<i>Streptococcus anginosus</i>	TIC A/S P/T CF CTZ CFP AZT IMI MER COL G TO AK NET MIN NAL CIP LEV T/S FOS NIT
<i>Streptococcus constellatus</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Streptococcus mitis</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Streptococcus oralis</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Streptococcus pneumoniae</i>	P AM A/C CFX CTX CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Streptococcus pyogenes</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Streptococcus salivarius</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Streptococcus sanguinis</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI
<i>Streptococcus viridans</i>	P AM A/C CFX CTX CFI CFP MER VAN TEI ERI CL CLI TE CIP LEV T/S C RI

Results

Isolates from patients with bacterial keratitis

Of a total of 389 positive bacterial corneal scrapings, Gram-positive bacteria ($n=305$, 78.4%) were the most common group of organisms; predominantly Gram-positive cocci of the genera *Staphylococcus* and *Streptococcus*. Gram-negative bacteria grew in 85 cultures (21.9%), being *Pseudomonas aeruginosa* ($n=38$; 9.7%) the most common isolate. The Gram-positive bacilli were the second group in frequency, being *P. acnes* ($n=32$, 8.2%) and *Corynebacterium macginleyi* ($n=27$, 6.9%) the most common isolates (Table 2).

Susceptibility profile of the most common bacterial pathogens from ocular infections against relevant antibacterial agents

The most frequently isolated bacteria were selected for the analysis of the results, only studied in *S. aureus*, *S. epidermidis*, *S. pneumoniae*, *P. aeruginosa* and *P. acnes*.

In vitro susceptibility of five of the most common bacterial pathogens against relevant antibacterial agents is described in Table 3. All Gram-positive cocci bacteria were susceptible to vancomycin. 51 isolates were *S. aureus* and 104 were *S. epidermidis*: 9.8% of the *S. aureus* were methicillin-resistant (MRSA) and 33.7% were methicillin-resistant *S. epidermidis* (MRSE). Their resistance rate was close to 20% for levofloxacin. All 22 isolates

of *S. pneumoniae* were sensitive to vancomycin, penicillin, levofloxacin, and amoxicillin. Nearly of the entirety of *P. aeruginosa* colonies ($n=38$) were sensitive to the most common antipseudomonal agents. All *P. acnes* analyzed ($n=10$) were sensitive to penicillin, amoxicillin-clavulanic acid, piperacillin/tazobactam, imipenem, and clindamycin.

The temporal profile of resistances

Time tracking was divided into 3 periods: 2005 to 2009 (58 isolations); 2010 to 2012 (95 isolations) and from 2013 to 2015 (234 isolations) to study the antibiotic susceptibility over time of the bacteria with some resistance. The rate of antibiotic susceptibility *P. aeruginosa* remains stable through the period of the study. It has not been possible to demonstrate differences because the sample size is small when making etiological groups (Tables 4–5).

Clinical data

Clinical outcome data were available for 215 events of *S. aureus*, *S. epidermidis*, *S. pneumoniae*, *P. aeruginosa* and *P. acnes* keratitis. The rate of coinfection (isolation of more than one microorganism in the same corneal scrap) was 5.3%, being more frequent in cases of *S. epidermidis* (22%) and *P. acnes* (30.2%) keratitis.

Table 2

Isolated bacteria in corneal scrapps in patients with bacterial keratitis from January 2006 to December 2015. Each isolated bacterium is counted separately and the total number of isolates is indicated. The associations are described at the end of the table.

Isolated bacteria	Nº (%)
Gram-positive cocci	225
<i>Staphylococci</i> (173)	
<i>Staphylococcus epidermidis</i>	104 (27.01)
<i>Staphylococcus aureus</i>	51 (13.24)
<i>Staphylococcus capitis</i>	6 (1.55)
<i>Staphylococcus warneri</i>	4 (1.03)
<i>Staphylococcus haemolyticus</i>	4 (1.03)
<i>Staphylococcus intermedius</i>	2 (0.51)
<i>Staphylococcus cohnii</i>	1 (0.25)
<i>Staphylococcus hominis</i>	1 (0.25)
<i>Streptococci</i> (45)	
<i>Streptococcus pneumoniae</i>	22 (5.71)
<i>Streptococcus sanguinis</i>	6 (1.55)
<i>Enterococcus faecalis</i>	6 (1.55)
<i>Streptococcus oralis</i>	6 (1.55)
<i>Streptococcus mitis</i>	2 (0.51)
<i>Streptococcus salivarius</i>	1 (0.25)
<i>Streptococcus constellatus</i>	1 (0.25)
<i>Streptococcus anginosus</i>	1 (0.25)
<i>Streptococcus agalactiae</i>	1 (0.25)
<i>Streptococcus viridans</i>	1 (0.25)
<i>Streptococcus pyogenes</i>	1 (0.25)
<i>Streptococcus gordoni</i>	1 (0.25)
<i>Abiotrophia defectiva</i>	2 (0.51)
<i>Granulicatella adiacens</i>	1 (0.25)
<i>Gram-negative cocci</i>	2
<i>Moraxella catarrhalis</i>	2 (0.51)
Gram-negative cocobacilli	19
<i>Moraxella lacunata</i>	9 (2.33)
<i>Moraxella osloensis</i>	2 (0.51)
<i>Moraxella nonliquefaciens</i>	1 (0.25)
<i>Haemophilus influenzae</i>	7 (1.81)
Gram-positive bacilli	78
<i>Propionibacterium acnes</i>	32 (8.31)
<i>Corynebacterium</i> (43)	
<i>Corynebacterium macginleyi</i>	27 (7.01)
<i>Corynebacterium spp.</i>	5 (1.29)
<i>Corynebacterium pseudodiphthericum</i>	4 (1.03)
<i>Corynebacterium tuberculostearicum</i>	3 (0.77)
<i>Corynebacterium propinquum</i>	2 (0.51)
<i>Corynebacterium kroppenstedtii</i>	1 (0.25)
<i>Corynebacterium accolens</i>	1 (0.25)
<i>Paenibacillus spp.</i>	2 (0.51)
<i>Lactobacillus gasseri</i>	1 (0.25)
<i>Gram-negative bacilli</i>	64
<i>Pseudomonas aeruginosa</i>	38 (9.7)
<i>Serratia marcescens</i>	9 (2.33)
<i>Escherichia coli</i>	4 (1.03)
<i>Serratia liquefaciens</i>	2 (0.51)
<i>Proteus mirabilis</i>	2 (0.51)
<i>Enterobacter cloacae</i>	2 (0.51)
<i>Alcaligenes faecalis</i>	2 (0.51)
<i>Moxarella catarrhalis</i>	1 (0.25)
<i>Stenotrophomonas maltophilia</i>	1 (0.25)
<i>Eikenella corrodens</i>	1 (0.25)
<i>Citrobacter koseri</i>	1 (0.25)
<i>Acinetobacter baumannii</i>	1 (0.25)
Gram-positive cocobacilli	1
<i>Arthrobacter aurescens</i>	1 (0.25)
Total isolated bacteria	389 (100%)
Polybacterial infections	
<i>Staphylococcus epidermidis + Streptococcus sanguinis</i>	3
<i>Staphylococcus epidermidis + Streptococcus salivarius</i>	1

Table 2 (Continued)

Isolated bacteria	Nº (%)
<i>Staphylococcus epidermidis + Stenotrophomonas maltophilia</i>	1
<i>Staphylococcus epidermidis + Serratia marcescens</i>	1
<i>Staphylococcus epidermidis + Staphylococcus aureus</i>	1
<i>Staphylococcus epidermidis + Serratia liquefaciens</i>	1
<i>Staphylococcus epidermidis + Staphylococcus warneri</i>	1
<i>Staphylococcus epidermidis + Pseudomonas aeruginosa</i>	2
<i>Staphylococcus epidermidis + Moraxella lacunata + Staphylococcus aureus</i>	1
<i>Staphylococcus epidermidis + Corynebacterium spp.</i>	1
<i>Staphylococcus epidermidis + Corynebacterium pseudodiphthericum + Corynebacterium macginleyi</i>	1
<i>Staphylococcus aureus + Staphylococcus capitis</i>	1
<i>Staphylococcus aureus + Proteus mirabilis</i>	1
<i>Staphylococcus aureus + Corynebacterium macginleyi</i>	1
<i>Pseudomonas aeruginosa + Escherichia coli</i>	1
<i>Pseudomonas aeruginosa + Staphylococcus warneri</i>	1
<i>Moxarella catarrhalis + Stenotrophomonas maltophilia</i>	1

Risk factors for keratitis

Corneal traumatism or a corneal foreign body were rare factors in our area (2% and 1% respectively). *P. aeruginosa* was the most prevalent bacteria in contact lens wearers (44.7% in 38 cases of *P. aeruginosa* keratitis analyzed), however the most isolated bacteria in patients with a therapeutic contact lens were *S. pneumoniae* (9.1% of 22 cases analyzed) and *S. aureus* (4% of 50 cases analyzed). *S. pneumoniae* was the most frequently associated with corneal surgery, mainly in penetrating keratoplasty (3 of 22 cases) and radial keratotomy (2 of 22 cases). The presence of an ocular surface disease (Stevens Johnson syndrome, ocular pemphigoid, Sjögren syndrome, bullous keratopathy) was recorded in 22.5% of the Gram-positive cocci but only in 7.9% of the *P. aeruginosa*. The presence of meibomitis presented in 35% of 20 patients with *S. aureus* keratitis, in 28.6% of 21 patients with *S. epidermidis*, and in 15.4% of 26 patients with *P. aeruginosa*. Palpebral malposition was associated mainly with *S. epidermidis* (17.4%). The main risk factors in patients with anaerobic BK (*P. acnes*) in 18 analyzed cases were contact lenses (41%), previous ocular surgery (16%) and herpetic keratitis (14%). Systemic diseases were associated mainly with *S. aureus* keratitis: 25% diabetes mellitus, 20% rheumatoid arthritis, 25% immunodepression; 15% dialysis and 15% cognitive deficiency in 20 cases analyzed.

Characteristic of the corneal infection

Keratitis caused by coinfection with other microorganism as *Herpesvirus*, *Acanthamoeba* spp., mycobacteria or fungus, and *S. epidermidis* keratitis with the simultaneously isolation of other bacteria were exclude for this analysis.

The presence of more than one corneal infiltrate (multiple infiltrates) was present in 4 (22.2%) of *P. acnes*, 10% of *P. aeruginosa* and

Table 3

In vitro susceptibility of the four most common bacterial pathogens isolated in corneal scrapps against relevant antibacterial agents from the total of analyzed cases [penicillin (P), oxacillin (OX), ceftazidime (CTZ), cefepime (CFP), imipenem (IMI), meropenem (MER), aztreonam (AZT), colistin (COL), levofloxacin (LEV), ciprofloxacin (CIP), gentamicin (GEN), tobramycin (TO), amikacin (AK), vancomycin (VAN) and rifampycin (RIF)]. Period of study 2005–2015.

Antibiotics	<i>S. aureus</i> (n = 51)	<i>S. epidermidis</i> (n = 104)	<i>S. pneumoniae</i> (n = 22)	<i>P. aeruginosa</i> (n = 38)	<i>P. acnes</i> (n = 32)
P	21.6% (n = 51)	15.8% (n = 101)	100% (n = 22)		100% (n = 26)
OX	90.2% (n = 51)	66.3% (n = 101)			
CTZ				94.7% (n = 38)	
CFP				94.7% (n = 38)	
IMI				94.7% (n = 38)	100% (n = 26)
MER				100% (n = 38)	
AZT				97.1% (n = 34)	
COL				100% (n = 35)	
LEV	80.4% (n = 51)	84.2% (n = 101)	100% (n = 22)	94.1% (n = 34)	
CIP				94.7% (n = 38)	
AK				100% (n = 38)	
G		79.0% (n = 100)		92.1% (n = 38)	
TO		67.4% (n = 92)			
LNZ	98.0% (n = 50)				
RIF	95.9% (n = 49)	97.8% (n = 92)			
VAN	100% (n = 51)	100% (n = 101)	100% (n = 22)		

Table 4

Temporal profile of antibiotic susceptibility in *S. aureus* and *S. epidermidis* isolated in corneal scrapps through 3 periods: 2005 to 2009; 2010 to 2012 and from 2013 to 2015.

Antibiotics	2005–2009		2010–2012		2013–2015		p	
	R	S	R	S	R	S		
LEV	<i>S. epidermidis</i>	3 (30%)	7 (70%)	6 (20.7%)	23 (79.3%)	7 (12.3%)	50 (87.7%)	0.3
	<i>S. aureus</i>	3 (30%)	7 (70%)	1 (8.3%)	11 (91.7%)	6 (20.7%)	23 (79.3%)	0.433
OX	<i>S. epidermidis</i>	6 (40%)	9 (60%)	12 (41.4%)	17 (58.6%)	16 (28.1%)	41 (71.9%)	0.398
	<i>S. aureus</i>	2 (20%)	8 (80%)	1 (8.3%)	11 (91.7%)	2 (6.9%)	27 (93.1%)	0.477
ERI	<i>S. epidermidis</i>	8 (53.3%)	7 (46.7%)	14 (48.3%)	15 (52.7%)	32 (56.1%)	25 (43.9%)	0.787
	<i>S. aureus</i>	1 (10%)	9 (90%)	4 (33.3%)	8 (66.7%)	8 (28.6%)	20 (71.4%)	0.414
LNZ	<i>S. epidermidis</i>	0	15 (100%)	0	29 (100%)	1 (1.8%)	56 (98.2%)	0.677
	<i>S. aureus</i>	0	10 (100%)	1 (8.3%)	11 (91.7%)	0	28 (100%)	0.199
RIF	<i>S. epidermidis</i>	0	15 (100%)	2 (6.9%)	27 (93.1%)	0	48 (100%)	0.109
	<i>S. aureus</i>	1 (10%)	9 (90%)	1 (9.1%)	10 (90.9%)	0	28 (100%)	0.248
MIN	<i>S. epidermidis</i>	0	11 (100%)	0	26 (100%)	0	9 (100%)	
	<i>S. aureus</i>	0	2 (100%)	0	9 (100%)	0	9 (100%)	

Table 5

Temporal profile of antibiotic susceptibility in *P. aeruginosa* through 3 periods: 2005 to 2009; 2010 to 2012 and from 2013 to 2015.

Antibiotics	2005–2009		2010–2012		2013–2015		p
	R	S	R	S	R	S	
LEV	0	2 (100%)	0	12 (100%)	2 (10%)	18 (90%)	0.475
CTZ	0	6 (100%)	1 (8.3%)	11 (91.7%)	1 (5%)	19 (95%)	0.755
CFP	0	6 (100%)	1 (8.3%)	11 (91.7%)	0	20 (100%)	0.329
IMI	0	6 (100%)	0	12 (100%)	1 (5%)	19 (95%)	0.63
MER	0	6 (100%)	0	12 (100%)	0	20 (100%)	
AZT	0	2 (100%)	1 (8.3%)	11 (91.7%)	0	20 (100%)	0.389
CIP	0	6 (100%)	0	12 (100%)	2 (10%)	18 (90%)	0.387
AK	0	6 (100%)	0	12 (100%)	0	20 (100%)	
G			1 (8.3%)	11 (91.7%)	2 (10%)	18 (90%)	0.876
COL	0	3 (100%)	0	12 (100%)	0	20 (100%)	

S. epidermidis, and 6% of *S. aureus* infections. Four (22.2%) *P. acnes*, two (4%) *S. aureus* and two (2.3%) *S. epidermidis* keratitis showed epithelial defect but not stromal infiltrate as initial presentation. The infiltrate position in *P. aeruginosa* was central or paracentral in 80% of patients, while in *Staphylococcus* spp. and *P. acnes* infections; the position most commonly found was peripheral (57 and 56% of cases respectively). *S. pneumoniae* locates in the center equally as peripheral cornea. The bacteria associated with larger corneal infiltrate (analyzed in 100 patients) were *S. pneumoniae* and the bacteria associated with smaller infiltrates were *S. epidermidis* (Fig. 1). *S. pneumoniae* was also the bacteria most associated with hypopyon (37.5%), followed by *P. aeruginosa* (23.5%), *S. aureus* (16.7%), *P. acnes* (11.1%) and *S. epidermidis* (2.3%).

At the time of diagnosis, 25% of *S. pneumoniae* and 5% of *S. aureus* keratitis presented as corneal perforation. Stromal thinning was reported in 66.7%, 33.3%, 33%, 19.5% and 13.3% of *S. pneumoniae*, *S. aureus*, *P. aeruginosa*, *S. epidermidis*, and *P. acnes* keratitis respectively. *S. epidermidis* was the only tested bacteria that did not present leucoma in all patients (only 75%).

Antimicrobial regimen

Most frequently antibiotics used were the combination of vancomycin and ceftazidime topical eye drops in 54 (25.1%) of cases analyzed. The second more used antimicrobial regimen were quinolone topical eye drops monotherapy in 26 cases (12.1%) (LEV,

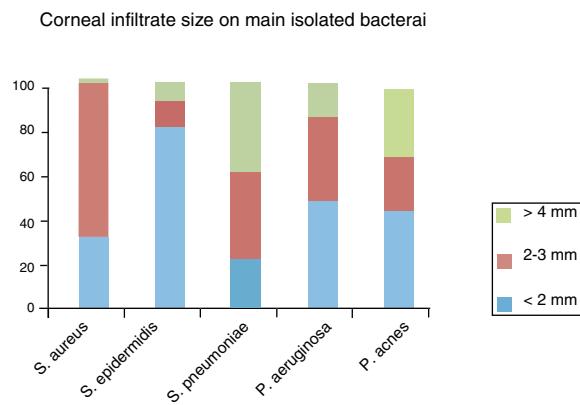


Fig. 1. Corneal infiltrate size on main isolated bacteria: from small infiltrates (<2 mm) 24 were caused by *S. epidermidis*, 9 polymicrobial infection, 6 *Pseudomonas*, 5 *P. acnes*, 3 *S. aureus* and 2 *S. pneumoniae*. From medium size infiltrates (>2 and <4 mm) 6 were caused by *S. aureus*, 6 polymicrobial infection, 4 *S. pneumoniae*, 3 *Pseudomonas*, 3 *S. epidermidis* and 3 *P. acnes*. From infiltrates >4 mm, 4 were caused by *S. pneumoniae*, 4 by *P. acnes*, 4 Polymicrobial infections, 2 *Pseudomonas* and 2 *S. epidermidis*.

CIP or MOX) and the combination of tobramycin with a quinolone topical eye drops (LEV, CIP or MOX) in 24 patients (11.2% of cases). A triple empiric therapy (vancomycin plus ceftazidime plus quinolone topical eye drops) was used in 5.3% of the cases only for moderate to big infiltrates. We did not find evidence of an association between the size of the corneal infiltrate and the empiric antibiotic regimen used ($\chi^2 = 5.49$, $p = 0.24$).

Visual acuity

The worst visual acuity at diagnosis was in patients with *S. pneumoniae* keratitis. Final visual acuity was significantly higher than visual acuity at diagnosis in patients with *S. aureus* keratitis ($p = 0.026$) and in patients with *S. epidermidis* ($p = 0.005$) ("Wilcoxon

test"). Changes between initial and final visual acuity were no significant in patients with *P. aeruginosa*, *S. pneumoniae* or *P. acnes* (Fig. 2a-e).

Correlation analysis

Clinical response variables were correlated (enucleation, corneal surgery and number of days of antibiotic treatment) with susceptibility to empirical antibiotic used between bacterias with antibiotic resistance; *S. aureus* and *S. epidermidis*. This correlation was not analyzed in cases of *P. aeruginosa*, *P. acnes* or *S. pneumoniae* keratitis because they are usually sensitive to commonly antibiotics used.

The antibiotics P, AMP, OXA and LEV were analyzed in 50 cases of *S. aureus* keratitis and in 37 cases of *S. epidermidis* keratitis. Oxacillin resistance *S. aureus* (methicillin-resistant *S. aureus*, MRSA) was significantly associated with worse prognosis; one (20%) of the 5 MRSA cases suffered enucleation and no case of the 45 MSSA cases analyzed suffered enucleation ($p = 0.002$), and frequency of corneal surgery was also higher in patients with MRSA (one of five patients, 20%) compared to methicillin-sensitive *S. aureus* (MSSA) (one of 44 patients, 2.2%) ($p = 0.054$). Patients with *S. aureus* keratitis and resistance to LEV treatment were also associated with greater risk of enucleation: 1 of 10 patients (10%) in MRSA cases and no patient of 40 MSSA analyzed cases ($p = 0.043$).

Corneal surgery frequency was higher in *S. epidermidis* resistant to rifampicin with 1 of 2 cases (50%) in resistant bacteria compared with 3 of 77 cases (3.9%) in susceptible bacteria ($p = 0.003$). Surgery was more frequent in *S. epidermidis* resistant to levofloxacin: 2 of 14 (14.3%) in MRSE cases needed corneal surgery compared with 2 of 72 cases (2.8%) in MSSE cases, but without statistic significance ($p = 0.061$). Patients with this bacteria and LEV resistance were also associated with increased treatment time (50.57 days) compared to 26.43 days of *S. epidermidis* LEV sensitive treatment (Mann-Whitney U test, Z: -2.56, $p = 0.009$).

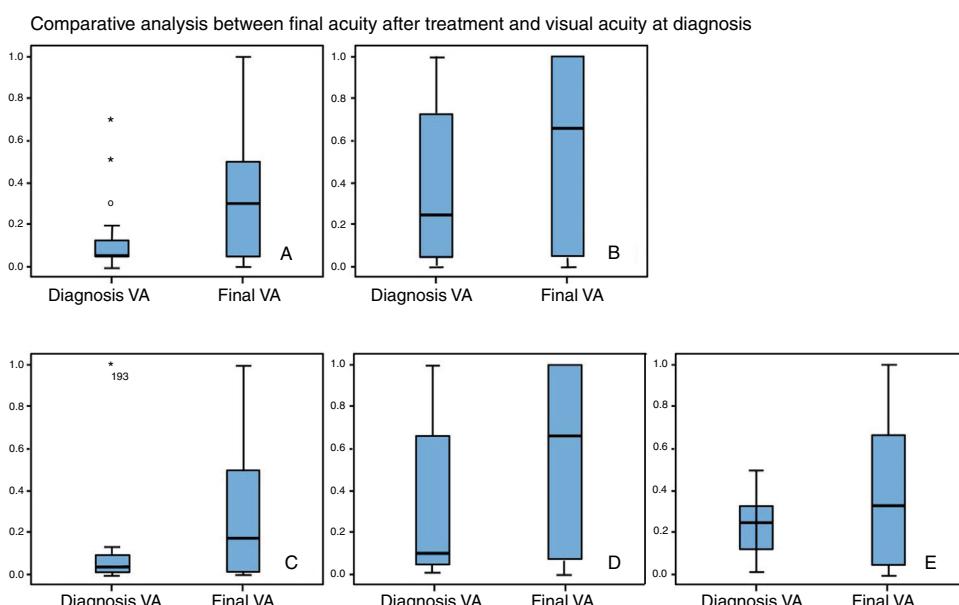


Fig. 2. A: *S. aureus* ($n = 17$ cases): The average visual acuity (VA) at diagnosis was 0.16 (DE = 0.22) and final VA was 0.36 (DE = 0.34). The final VA was significantly higher than VA at diagnosis (Z Wilcoxon -2.826, $p = 0.005$). B: *S. epidermidis* ($n = 31$ cases): The average VA at diagnosis was 0.42 (DE = 0.37) and final VA was 0.59 (DE = 0.44). The final VA was significantly higher than VA at diagnosis (Z Wilcoxon -2.826, $p = 0.005$). C: *S. pneumoniae* ($n = 8$ cases): The average VA at diagnosis was 0.14 (DE = 0.32) and final VA was 0.33 (DE = 0.38). The improvement of AV was 1.84 lines (Z Wilcoxon -1.841, $p = 0.066$). D: *P. aeruginosa* ($n = 12$ cases): The average VA at diagnosis was 0.33 (DE = 0.35) and final VA was 0.50 (DE = 0.41), without significant improvement (1.53 lines) (Z Wilcoxon -1.53; $p = 0.12$). E: *P. acnes* ($n = 13$ cases): The average VA at diagnosis was 0.27 (DE = 0.22) and final VA was 0.43 (DE = 0.39), without significant improvement (1.53 lines) (Z Wilcoxon -1.58; $p = 0.113$).

Discussion

We analyzed the BK treated in the cornea unit of a major public hospital in an urban area of Madrid. Because our study has been done retrospectively we cannot affirm that we performed a corneal scrape of all the suspected infectious keratitis and this could have some influence on our results. Another study limitation could be the impossibility of ensuring a correct therapeutic compliance in all patients. We found a low antibiotic resistance index except in the case of *Staphylococcus* spp. where we observed the influence of resistance on the clinical prognosis (index of enucleation).

S. epidermidis was the bacteria most commonly isolated in corneal smears, agreeing with the reports from other series.^{9,10} Although this bacteria has been considered a commensal bacteria, we saw some severe and disabling keratitis; moreover, 33.7% were MRSE and levofloxacin resistant *S. epidermidis* and those were related to a prolonged antibiotic treatment.

Infectious keratitis caused by MRSA is an increasing problem around the world^{11,12} and is of great concern because it is related to fluoroquinolones resistance¹³ and responds poorly to conventional antibiotic treatment.¹⁴ Our rate of MRSA is 9.8% and was significantly associated with worse clinical prognosis. The susceptibility rate of MRSA to vancomycin is reportedly still 100%^{15–19} and thus vancomycin is highly valued for the treatment of MRSA infections. Vancomycin (2 µg/mL) had the lowest MIC90 values (µg/mL) for ocular *S. aureus* and coagulase-negative *Staphylococcus* isolates recovered from the eye.²⁰ To treat ocular infections, a topical application of vancomycin solution, or as a 1% or 5% ophthalmic ointment, has proven to be useful for the treatment of external ocular MRSA or MRSE infections.^{21–24} The solution reached high corneal tissue concentrations that significantly exceeded the MIC90 (2–10 mg/ml) for most key Gram-positive corneal pathogens.²⁴ We consider topical vancomycin an irreplaceable antibiotic as initial empirical combined therapy of severe BK in patients predisposed to MRS keratitis (previous surgery, previous methicillin-resistant staphylococcus (MRS) ocular infection, immunosuppression or dialysis-patients). *Staphylococcus* spp. kept a low sensitivity rate to beta-lactams or levofloxacin and we found significant association between the MIC of levofloxacin and methicillin and the prognosis of *Staphylococcus* spp. BK. Moreover the patients affected by *S. aureus* keratitis suffered from systemic debilitating conditions, which could compromise the defense host mechanism against infections.

Ulcers from *S. pneumoniae* occurred in patients wearing therapeutic contact lens and showed the largest abscesses and a high rate of perforation as initial presentation. All the *S. pneumoniae* isolated showed susceptibility to penicillin, levofloxacin or vancomycin. Regarding fluorquinolones, moxifloxacin²⁵ appears to be more effective against *S. pneumoniae*. On the other hand, enhanced penetration of levofloxacin into the ocular tissue in addition to the higher concentration in the tear film combined with its lower MICs against streptococci, may allow for higher MIC ratios and a more immediate cure.²⁶ An experimental study evaluating the chemotherapeutic efficacy of topical antibiotic *in vivo* showed that the combination of gentamicin and vancomycin was most effective against penicillin-resistant pneumococci.²⁷ Due to the lack of penicillin resistance in our hospital we could consider a combination of penicillin and levofloxacin as an appropriate regimen for this aggressive bacteria.

P. aeruginosa was the most frequently isolated Gram-negative bacillus (9.7%). This report shows low levels of resistance of *P. aeruginosa* over the period 2005 to 2015. In Europe, there may be a rate of resistance of *P. aeruginosa* to aminoglycosides and ciprofloxacin, which is currently reported at 11%.²⁵ *P. aeruginosa* are often associated with the largest ulcers²⁸ and significantly poorer visual acuity than patients with other bacterial ulcers.²⁹ In our study final visual

acuity was 0.5 for *P. aeruginosa* keratitis, but was lower for *S. pneumoniae*, and *S. aureus*. Gram-positive cocci in our series affected patients with other ocular conditions, which could explain this difference. While many options are available for susceptible *P. aeruginosa*, colistin³⁰ or the synergistic activity between a combination of meropenem/ciprofloxacin are possible treatments for the more resistant strains.³¹ Our prefer topical final regimen for this aggressive keratitis is the combination of ceftazidime plus ciprofloxacin plus tobramycin or amikacine.

P. acnes, isolated in 8.2%, affected patients with previous surgery, herpetic keratitis or contact lens wearers. This commensal bacteria showed susceptibility to the tested antibiotics and caused peripheral and small infiltrates more frequently, but is also capable to produce largest infiltrates with hypopyon and stromal thinning.

Microbial keratitis requires prompt and appropriate management to ensure the best visual outcome for the patient. Culturing allows sensitivity testing to a range of agents so that treatment modifications can be made in an informed manner if the clinical response to initial treatment is inadequate. In our urban community the most common bacteria causing moderate and severe keratitis were *Staphylococcus*, *Streptococcus*, and *Pseudomonas* species. Because of the level of methicillin and levofloxacin resistance found in our study for *Staphylococcus* spp., we propose to continue the empirical treatment used in our hospital with vancomycin, combined with ceftazidime or fluorquinolones, as an effective regimen for severe central bacterial keratitis when we suspect a MRS. With our standard practice the rate of enucleation or need of surgery remain below 5% in bacterial keratitis.

Conflict of interest

The authors declare that they have no conflict of interest.

References

- Lin J, Tsai Y, Fu Y. The fixed combination of fortified vancomycin and amikacin ophthalmic solution—VA solution in vitro study of the potency and stability. Cornea. 2005;24:717–21.
- Karmpataki V, Papanikolaou T, Giannousis M, Goulas A, Mandraveli K, Kilmpasani M, et al. Stability and antibacterial potency of ceftazidime and vancomycin eyedrops reconstituted in BSS against *Pseudomonas aeruginosa* and *Staphylococcus aureus*. Acta Ophthalmol. 2009;87:555–8.
- Chang VS, Dhaliwal DK, Raju L, Kowalski RP. Antibiotic resistance in the treatment of *Staphylococcus aureus* keratitis: a 20-year review. Cornea. 2015;34:698–703.
- Hsiao CH, Sun CC, Yeh LK, Ma DH, Chen PY, Lin HC, et al. Shifting trends in bacterial keratitis in Taiwan: a 10-year review in a Tertiary-Care Hospital. Cornea. 2016;35:7–313.
- Ruiz Caro JM, Cabrejas L, de Hoz MR, Mingo D, Duran SP. Clinical features and microbiological in bacterial keratitis in a tertiary referral hospital. Arch Soc Esp Oftalmol. 2017.
- Sharma N, Arora T, Jain V, Agarwal T, Jain R, Jain V, et al. Gatifloxacin 0.3% versus fortified tobramycin-cefazolin in treating nonperforated bacterial corneal ulcers: randomized, controlled trial. Cornea. 2016;35:56–61.
- The European Committee on Antimicrobial Susceptibility Testing (EUCAST) Available from: <http://www.eucast.org>.
- <https://www.ibm.com/analytics/us/en/technology/spss>.
- Hernandez-Camarena JC, Graue-hernandez EO, Ortiz-Casas M, Ramirez-Miranda A, Navas A, Pedro-Aguilar L, et al. Trends in microbiological and antibiotic sensitivity patterns in infectious keratitis: 10-year experience in Mexico City. Cornea. 2015;34:778–85.
- Chirinos-Saldana P, Bautista de Lucio VM, Hernandez-Camarena JC, Navas A, Ramirez-Miranda A, Vizuet-Garcia L, et al. Clinical and microbiological profile of infectious keratitis in children. BMC Ophthalmol. 2013;13:54.
- Wroblewski KJ, Pasternak JF, Bower KS, Schallhorn SC, Hubickey WJ, Harrison CE, et al. Infectious keratitis after photorefractive keratectomy in the United States army and navy. Ophthalmology. 2006;113:5–520.
- Solomon R, Donnenfeld ED, Perry HD, Rubinfeld RS, Ehrenhaus M, Wittpenn JR Jr, et al. Methicillin-resistant *Staphylococcus aureus* infectious keratitis following refractive surgery. Am J Ophthalmol. 2007;143:629–34.
- Willcox MD. Review of resistance of ocular isolates of *Pseudomonas aeruginosa* and *staphylococci* from keratitis to ciprofloxacin, gentamicin and cephalosporins. Clin Exp Optom. 2011;94:8–161.

14. Sotozono C, Inagaki K, Fujita A, Koizumi N, Sano Y, Inatomi T, et al. Methicillin-resistant *Staphylococcus aureus* and methicillin-resistant *Staphylococcus epidermidis* infections in the cornea. Cornea. 2002;21 Suppl.:S94–101.
15. Maple PA, Hamilton-Miller JM, Brumfitt W. World-wide antibiotic resistance in methicillin-resistant *Staphylococcus aureus*. Lancet. 1989;1:537–40.
16. Cimolai N. Ciprofloxacin and multi-resistant staphylococci. Lancet. 1997;349:1030.
17. Maffett M, O'Day DM. Ciprofloxacin-resistant bacterial keratitis. Am J Ophthalmol. 1993;115:6–545.
18. Marangon FB, Miller D, Muallem MS, Romano AC, Alfonso EC. Ciprofloxacin and levofloxacin resistance among methicillin-sensitive *Staphylococcus aureus* isolates from keratitis and conjunctivitis. Am J Ophthalmol. 2004;137:8–453.
19. Elsahn AF, Yildiz EH, Jungkind DL, Abdalla YF, Erdurmus M, Cremona FA, et al. In vitro susceptibility patterns of methicillin-resistant *Staphylococcus aureus* and coagulase-negative staphylococcus corneal isolates to antibiotics. Cornea. 2010;29:5–1131.
20. Miller D, Chang JS, Flynn HW, Alfonso EC. Comparative in vitro susceptibility of besifloxacin and seven comparators against ciprofloxacin- and methicillin-susceptible/nonsusceptible staphylococci. J Ocul Pharmacol Ther. 2013;29:339–44.
21. Goodman DF, Gottsch JD. Methicillin-resistant *Staphylococcus epidermidis* keratitis treated with vancomycin. Arch Ophthalmol. 1988;106:1–1570.
22. Fleischer AB, Hoover DL, Khan JA, Parisi JT, Burns RP. Topical vancomycin formulation for methicillin-resistant *Staphylococcus epidermidis* blepharoconjunctivitis. Am J Ophthalmol. 1986;101:7–283.
23. Sotozono C, Fukuda M, Ohishi M, Yano K, Origasa H, Saiki Y, et al. Vancomycin ophthalmic ointment 1% for methicillin-resistant *Staphylococcus aureus* or methicillin resistant *Staphylococcus epidermidis* infections: a case series. BMJ. 2013;3.
24. Cahane M, Ben Simon GJ, Barequet IS, Grinbaum A, Diamantstein-Weiss L, Goller O, et al. Human corneal stromal tissue concentration after consecutive doses of topically applied 3.3% vancomycin. Br J Ophthalmol. 2004;88:22–4.
25. Ramakrishnan R, Ramesh S, Bharathi MJ, Amuthan M, Viswanathan S. Comparative in-vitro efficacy of fluoroquinolones against *Streptococcus pneumoniae* recovered from bacterial keratitis as determined by E-test. Indian J Pathol Microbiol. 2010;53:276–80.
26. Morrissey I, Burnett R, Viljoen L, Robbins M. Surveillance of the susceptibility of ocular bacterial pathogens to the fluoroquinolone, gatifloxacin and other antimicrobials in Europe during 2001/2002. J Infect. 2004;49:109–14.
27. Guzek JP, Cline DJ, Row PK, Wessels IF, Beeve S, Ispirescu S, et al. Rabbit *Streptococcus pneumoniae* keratitis model and topical therapy. Invest Ophthalmol Vis Sci. 1998;39:7–2012.
28. Kaye S, Tuft S, Neal T, Tole D, Leeming J, Figueiredo F, et al. Bacterial susceptibility to topical antimicrobials and clinical outcome in bacterial keratitis. Invest Ophthalmol Vis Sci. 2010;51:8–362.
29. Sy A, Srinivasan M, Mascarenhas J, Lalitha P, Rajaraman R, Ravindran M, et al. *Pseudomonas aeruginosa* keratitis: outcomes and response to corticosteroid treatment. Invest Ophthalmol Vis Sci. 2012;53:267–72.
30. Jain R, Murthy SI, Motukupally SR, Jain M. Use of topical colistin in multiple drug-resistant *Pseudomonas aeruginosa* bacterial keratitis. Cornea. 2014;33:7–923.
31. Sueke H, Kaye SB, Neal T, Hall A, Tuft S, Parry CM. An in vitro investigation of synergy or antagonism between antimicrobial combinations against isolates from bacterial keratitis. Invest Ophthalmol Vis Sci. 2010;51:5–4151.