



ORIGINAL ARTICLE

Losartan prevents mesenteric vascular bed alterations in high-fat diet fed rats



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Abstract Dysfunction of perivascular adipose tissue of mesenteric bed participates in the pathophysiology of high blood pressure linked to metabolic syndrome. Thus, it might consider a new therapeutic objective to take account in cardiovascular and metabolic diseases. Besides its antihypertensive effect, there is a growing interest on the pleiotropic actions of losartan, an angiotensin II type 1 (AT₁) receptor antagonist. The aim of the study was to analyze the actions of losartan treatment on adiposity index and prostanoids release from mesenteric vascular bed and its relationship with blood pressure as well as homeostasis model of assessment of insulin resistance (HOMA-IR) in Sprague-Dawley rats under a high-fat (HF) diet for 8 weeks. Four groups were used: control (C), HF diet (HF, 50%, w/w bovine fat), losartan-treated (CL8, 30 mg/kg/body weight/day in the drinking water) and losartan-treated HF diet (HFL, both treatments). A high-fat diet incremented systolic blood pressure, HOMA-IR, adiposity of mesenteric vascular bed and the release of vasoconstrictor prostanoids such as thromboxane (TX) B₂ and prostaglandin (PG) F_{2α} as well as PGE₂, an inflammatory prostanoid in a context of insulin resistance and hypertension. We found a positive correlation between adiposity index and systolic blood pressure. Also, both parameters are positive correlated with the HOMA IR index. Moreover, we also found that these prostanoids release correlate with systolic blood pressure as well as with mesenteric vascular bed adiposity index. Losartan treatment prevented all these alterations and normalized the PGI₂/TXA₂ ratio in high-fat fed rats. We conclude that losartan may play beneficial actions on perivascular adipose tissue alterations and endothelial dysfunction through restoration of normal balance of vasoactive substances in this model.

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PALABRAS CLAVE

Nutrición;
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 Obesidad;
 Hipertensión;
 Losartán;
 Prostanoides

Losartán previene las alteraciones del lecho vascular mesentérico en ratas alimentadas con dieta alta en grasas

Resumen La disfunción del tejido adiposo perivascular del lecho mesentérico posee una participación en la fisiopatología de la hipertensión arterial relacionada con el síndrome metabólico. Por lo tanto, podría considerarse como un nuevo blanco terapéutico en las enfermedades cardiovasculares y metabólicas. Además de su efecto antihipertensivo, existe un interés creciente en las acciones pleiotrópicas de losartán, antagonista del receptor de angiotensina II. El objetivo del estudio fue analizar las acciones de losartán sobre el índice de adiposidad y la liberación de prostanoides del lecho vascular mesentérico y su relación con la presión arterial, así como en el índice HOMA-IR (modelo de evaluación homeostático de la resistencia a la insulina) en ratas con dieta alta en grasas. Observamos que la dieta alta en grasas incrementó la adiposidad del lecho vascular mesentérico y la liberación de prostanoides vasoconstrictores como tromboxano (TX) B₂ y prostaglandina (PG) F₂ α , así como la PGE₂, un prostanoides inflamatorio en el contexto de resistencia a la insulina e hipertensión. También encontramos una correlación positiva entre el índice de adiposidad y la presión arterial sistólica y ambos parámetros se correlacionan positivamente con el índice HOMA IR. Adicionalmente observamos que la liberación de estos prostanoides se correlaciona con la presión arterial sistólica, así como con el índice de adiposidad del lecho vascular mesentérico. El tratamiento con losartán previno todas estas alteraciones y normalizó la relación PGI₂/TXA₂ en ratas alimentadas con una dieta alta en grasa. Concluimos entonces que losartán puede ejercer acciones beneficiosas sobre las alteraciones del tejido adiposo perivascular y la disfunción endotelial a través de la restauración del equilibrio normal de sustancias vasoactivas en este modelo experimental.

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Introduction

Dysfunction of perivascular adipose tissue (PVAT) plays a role in the pathogenesis of hypertension associated to metabolic syndrome and cardiovascular diseases.¹⁻⁴ In this way, ectopic fat deposition in PVAT could be relevant to trigger insulin resistance. Also, a local renin-angiotensin-aldosterone system (RAAS) in adipose tissue is involved in metabolic syndrome development.⁵⁻⁷ In this context, pleiotropic effects of drugs used for the treatment of hypertension associated to metabolic diseases, such as losartan, an angiotensin II type 1 (AT₁) receptor blocker, still generate great interest.⁸

PVAT has an active contribution in the regulation of vascular function. The physiological anti-contractile and anti-inflammatory effects of PVAT are diminished in hypertension.^{9,10} From its first mention by Leonardo da Vinci, mesenteric vascular bed remained almost without clinical relevance.¹¹ It is formed by resistance arteries surrounded by PVAT, mainly of white visceral adipose tissue.^{12,13} On the other hand, PVAT of the mesenteric arteries has a local RAS with high density of AT₁ receptors.^{14,15}

In addition, mesenteric vascular bed is a source of prostanoids derived from the action of cyclooxygenases that includes prostaglandins (PGs) and thromboxanes (TXs) which participate in vascular tone regulation. These agents are synthesized and released from endothelial and smooth muscle cells as well as in adipose tissue of mesenteric vascular bed.¹⁶ Therefore dysfunctional PVAT on those vessels as a result of a high-fat diet could be a possible

link between metabolic diseases, high blood pressure and vascular complications. Concomitantly, we have previously demonstrated alterations in prostanoids release in experimental models of metabolic syndrome.^{17,18} However, little is known about the role of RAS in the development of PVAT on mesenteric vascular bed. Thus, the aim of this study was to analyze the effects of losartan on the adiposity and prostanoids release from mesenteric vascular bed and its relationship with blood pressure and insulin resistance in rats under a high-fat diet.

Material and methods

Ethical approval

The experimental protocol was previously approved by the local Comité Institucional para el Cuidado y Uso de Animales de Laboratorio (CICUAL; Facultad de Farmacia y Bioquímica; Universidad de Buenos Aires; Resolution N° 1881-1999) according to the International Principles for Research on Animals. All the animals were housed with a 12 h light/dark cycle, controlled temperature (22 ± 2 °C) and adequate humidity.

Animal's protocol and diet

Twenty-four male Sprague-Dawley rats (weighing 180–210 g at the beginning) were studied for 9 weeks. They were

randomly divided into four groups ($n=6$ each group): control group (C) were fed standard rodent diet (SD, 3.3 kcal/g; with 2% fiber, 3% fat, 6% minerals, 20% proteins and 69% starch and vitamins supplements; Commercial Rodents Purina Chow, Asociación Cooperativas Argentinas SRL, Buenos Aires, Argentina) and water to drink; high-fat diet group (HF), which received 50% (w/w) bovine fat (BF, 9 kcal/g; 99% total fat: 77% saturated fat and 19% trans-fat, and necessary amounts of carbohydrates, protein, fiber, sodium, vitamins and minerals supplements; Quick-food S.A. Provincia de Santa Fe, Argentina) added to 50% (w/w) SD and water to drink; losartan-control group (CL), which received losartan (30 mg/kg/bodyweight/day, highest available commercial grade was purchased from Droguería Saporiti S.A.C.I.F.I.A, Buenos Aires, Argentina) dissolved in the drinking water and fed SD; and losartan-high-fat diet group (HFL), which received losartan (same dose) in the water and 50% (w/w) BF added to 50% (w/w) SD. All animals were given free access to water and food, ad libitum. Body weight, food and water intake were monitored throughout the entire period in all experimental groups. Dietary and pharmacological treatments began at the same time. The dose of losartan was chosen according to previous studies.^{19–21}

Blood analysis

At the end of the study period, rats were fasted for 5 h, weighed and blood samples were collected from the retro-ocular sinus under light anesthesia.^{22,23} Plasma glucose levels were measured by glucose meter (Roche Accu-Chek®, Germany); plasma triglyceride levels were evaluated using commercial kits (enzymatic methods, TG Color Wiener Laboratories, S.A.I.C, Rosario, Argentina) and insulin levels by rat/mouse insulin ELISA kit (Merck Millipore, USA). The following equation was used to determine the homeostasis model of assessment of insulin resistance (HOMA-IR) = glucose (mM) \times insulin (mIU/L)/22.5.²⁴

Adiposity index and blood pressure

Rats were weighed prior to dietary and pharmacological manipulation and at the end of the study. The entire mesenteric vascular bed that includes mesenteric blood vessels with PVAT was dissected and weighed from each animal. We calculated the mesenteric vascular bed adiposity index as its weight/body weight \times 100. For two weeks prior to the end of the experimental period, all animals were trained to the procedure of systolic blood pressure (SBP) measurement, which was performed by indirect method of tail cuff plethysmography (Tektronix Inc., Portland, OR, USA).

Prostanoid release measurement

The mesenteric vascular bed dissected tissue was embedded with Krebs solution (mM; KCl 4.7, NaCl 118.0, NaH₂PO₄ 1.0, MgSO₄ 1.2, CaCl₂ 2.6, NaHCO₃ 25.0, glucose 11.1), and incubated during 60 min at 37 °C. Then, the media was acidified (pH 3.5) with 1 M formic acid and extracted with chloroform to measure prostanoid release. Dried extracted

chloroform samples were suspended in the mobile phase and injected into the Reversed-phase HPLC system (BBS Hypersil C18, Thermo Electron Co., Bellefonte, PA, USA). The following prostanoids standards were run together with the samples: 6-keto PGF₁ α (stable metabolite of PGI₂ or prostacyclin), PGE₂, PGF₂ α and TXB₂ (stable metabolite of TXA₂) (Sigma Chemical Co., Saint Louis, MO, USA). The results were expressed as nanograms of prostanoid per milligram of wet tissue weight.

Statistics

Statistical analysis was performed using InfoStat software program, version 2018 (Córdoba, Argentina), by means of two-way ANOVA and Tukey's post hoc test. For correlation analysis, Pearson's correlation coefficients (r) of the data points from experimental rats were calculated by linear regression. A $P < 0.05$ was considered statistically significant. All results are expressed as the mean \pm SEM.

Results

Caloric intake, body weight and metabolic parameters

As shown in Table 1, caloric intake was significantly higher in HF compared to C rats ($P < 0.01$). Losartan did not alter total calories intake in CL and HFL groups compared to C and HF, respectively. At the end of period study, high-fat diet produced an increase in body weight in HF compared to C rats ($P < 0.05$). HF fed rat exhibited increased triglyceridaemia, glycaemia and insulinaemia as well as the HOMA-IR with respect to C group ($P < 0.01$; Table 1). These results indicate that insulin resistance model in metabolic syndrome was effectively induce by high-fat diet. Losartan treatment ameliorated all these alterations in HFL compared to HF ($P < 0.01$; Table 1).

Effects of losartan on mesenteric vascular bed adiposity and its relationship with systolic blood pressure

High-fat diet produced increments on mesenteric vascular bed adiposity index and SBP in HF rats compared to C ($P < 0.01$; Table 2). Losartan treatment not only prevented SBP rise as expected ($P < 0.01$; Table 2), but also prevented the increase of mesenteric vascular bed adiposity index in HFL compared to HF ($P < 0.01$; Table 2). A positive correlation was found between adiposity index of mesenteric vascular bed and SBP ($r = 0.87$, $R^2 = 0.80$, $P < 0.01$; Fig. 1a). Also, significant correlations were found between SBP and HOMA-IR ($r = 0.90$, $R^2 = 0.80$, $P < 0.01$; Fig. 1b) as well as between mesenteric vascular bed adiposity index and HOMA-IR ($r = 0.93$, $R^2 = 0.86$, $P < 0.01$; Fig. 1c).

Effects of losartan on prostanoids release from mesenteric vascular bed

High-fat diet significantly increased vasoconstrictor prostanoids in HF fed rats compared to C group: TXB₂

Table 1 Caloric intake, body weight and metabolic parameters.

	C	CL	HF	HFL
Caloric intake (Kcal/day)	73.7 ± 1.2	74.9 ± 1.2	103.8 ± 2.5*	102.7 ± 1.6*
Body weight (g)	424.4 ± 19.2	421.0 ± 22.5	477.1 ± 8.5#	441.5 ± 16.1
Triglycerides (mg/dl)	71 ± 3	73 ± 6	166 ± 9*	88 ± 4***
Glycemia (mg/dl)	108 ± 4	114 ± 6	148 ± 4*	129 ± 3***
Insulinemia (ng/ml)	1.2 ± 0.1	1.0 ± 0.1	4.9 ± 0.5*	2.3 ± 0.2****
HOMA-IR	0.11 ± 0.01	0.09 ± 0.01	0.57 ± 0.06*	0.23 ± 0.05**,#

Results are expressed as mean ± SEM. Control (C); high-fat diet (HF); losartan-control (CL); losartan-high-fat diet (HFL).

* $P < 0.01$ vs. C, CL.

$P < 0.05$ vs. C, CL.

** $P < 0.01$ vs. HF.

*** $P < 0.01$ vs. HF, C.

**** $P < 0.01$ vs. HF, C, CL.

Table 2 Mesenteric vascular bed adiposity and systolic blood pressure.

	Mesenteric vascular bed adiposity index (%)	Systolic blood pressure (mm Hg)
C	0.7 ± 0.1	118 ± 2
CL	0.7 ± 0.1	111 ± 2##
HF	2.0 ± 0.1*	150 ± 2*
HFL	1.2 ± 0.1****	115 ± 2**

Results are expressed as mean ± SEM. Control (C); high-fat diet (HF); losartan-control (CL); losartan-high-fat diet (HFL).

* $P < 0.01$ vs. C, CL.

$P < 0.05$ vs. C.

**** $P < 0.01$ vs. HF, C, CL.

** $P < 0.01$ vs. HF, C.

($P < 0.01$; Fig. 2a) and $\text{PGF}_{2\alpha}$ ($P < 0.01$; Fig. 2b). Losartan prevented these increases in HFL compared to HF rats: TXB_2 ($P < 0.01$; Fig. 2a) and $\text{PGF}_{2\alpha}$ ($P < 0.01$; Fig. 2b). In addition, positive correlations were found between release of both prostanoids and SBP (TXB_2 ; $r = 0.93$, $R^2 = 0.87$, $P < 0.01$; Fig. 3a and $\text{PGF}_{2\alpha}$; $r = 0.95$, $R^2 = 0.89$, $P < 0.01$; Fig. 3b) as well as with mesenteric vascular bed adiposity index (TXB_2 : $r = 0.89$, $R^2 = 0.80$, $P < 0.01$; Fig. 4a and $\text{PGF}_{2\alpha}$: $r = 0.90$, $R^2 = 0.82$, $P < 0.01$; Fig. 4b).

As it is shown in Fig. 5a, PGE_2 release increased in HF group compared to C ($P < 0.01$) and losartan prevented this effect ($P < 0.01$, Fig. 5a). A positive correlation was found between PGE_2 and SBP ($r = 0.90$, $R^2 = 0.81$, $P < 0.01$; Fig. 5b) and also between PGE_2 and adiposity index of mesenteric vascular bed ($r = 0.88$, $R^2 = 0.80$, $P < 0.01$; Fig. 5c).

On the other hand, the prostacyclin (PGI_2)/thromboxane (TXA_2) release ratio (measured as their stable metabolites) was significantly reduced in HF fed rats compared to C group ($P < 0.01$; Fig. 6a). Losartan treatment was able to prevent this reduction in HFL compared to HF ($P < 0.01$; Fig. 6a). Moreover, negative correlations were found between $\text{PGI}_2/\text{TXA}_2$ release ratio and SBP ($r = -0.91$, $R^2 = 0.82$, $P < 0.01$; Fig. 6b) as well as with mesenteric vascular bed adiposity index ($r = -0.84$, $R^2 = 0.72$, $P < 0.01$; Fig. 6c).

Discussion

Our results demonstrate the preventive effect of losartan treatment on the mesenteric vascular bed adiposity deposition as well as on vasoconstrictor (TXB_2 and $\text{PGF}_{2\alpha}$) and inflammatory (PGE_2) prostanoids release, in a context of hypertension and insulin resistance produced by a high-fat diet. Furthermore, we showed that losartan improved the $\text{PGI}_2/\text{TXA}_2$ ratio, a marker of endothelial dysfunction.

High-fat diet fed rats is a suitable animal model that resembles pathophysiological features of human metabolic syndrome.²⁵ In agreement with previous reports,^{17,18} we found higher triglyceridaemia, glycaemia, insulinaemia and insulin resistance; increased visceral adiposity and elevated blood pressure. Losartan treatment attenuated such characteristics in HF rats. Previously, Mourant et al.²⁶ reported losartan effects on higher levels of plasma glucose, triglycerides and insulin; except for the fact that losartan treatment was started after 12 weeks of treatment with high-fat diet. In our protocols, dietary and losartan were co-administered from the beginning of the experiments to establish their prevention by pharmacological inhibition of RAS with losartan. On the other hand, they did not measure blood pressure in their study.

Regarding the results on adiposity index, we have found different selection criteria from different authors in defining the fat areas. Some studies reported the sum of epididymal (male rat model), retroperitoneal, perirenal or/and mesenteric fat pads.²⁷⁻²⁹ Mourant et al.²⁶ had reported losartan effect on the ratio of visceral fat/gastrocnemius muscle considered as an index of body composition but they did not specify where they took visceral fat pads. Wang et al.³⁰ found that losartan treatment exhibited a significantly decreased in epididymal, retroperitoneal and mesenteric fat pad weights in SHR rat model. Our report shows the preventive effect of losartan on the mesenteric vascular bed adiposity increase.

PVAT has been proposed to impact microvascular function and it could be involved in the pathogenesis and progression of insulin resistance and hypertension.^{31,32} Accordingly, we have found significant correlations between adiposity index of mesenteric vascular bed with insulin resistance and SBP as well. The development of high blood pressure associated with metabolic diseases is multifactorial. One of the

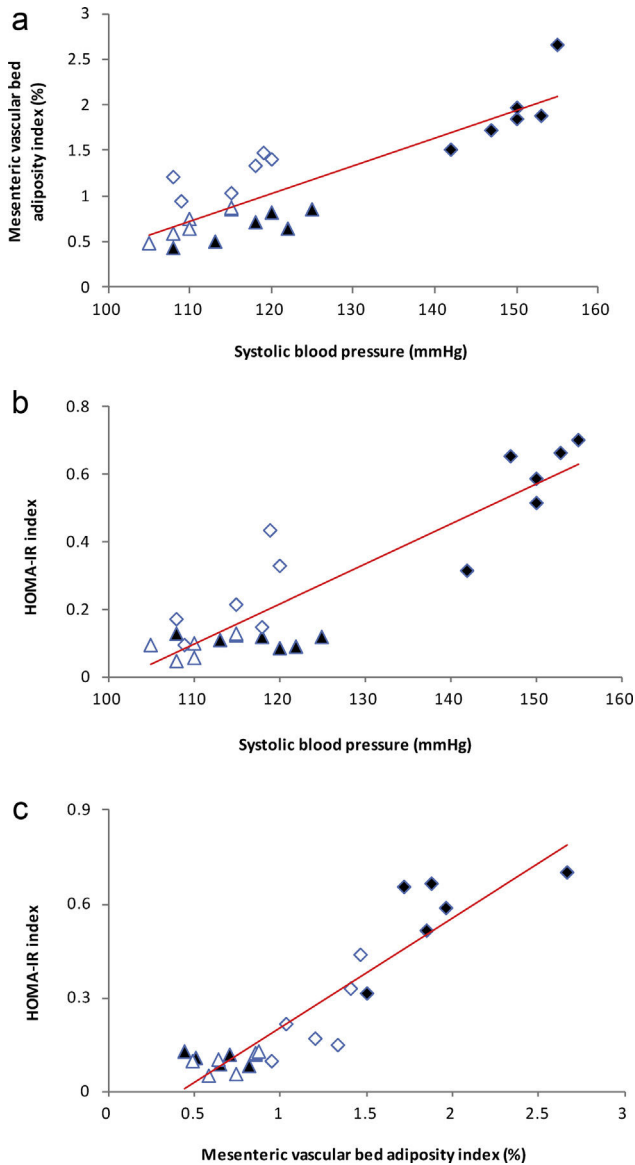


Figure 1 (a) Linear regression of systolic blood pressure against mesenteric vascular bed adiposity index: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r=0.87$, $R^2=0.80$, $P<0.01$. (b) Linear regression of systolic blood pressure against HOMA-IR index: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r=0.90$, $R^2=0.80$, $P<0.01$. (c) Linear regression of mesenteric vascular bed adiposity index against HOMA-IR index: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r=0.93$, $R^2=0.86$, $P<0.01$.

possible mechanisms involved could be due to PVAT dysfunction caused by a high-fat diet which produces modifications in both the amount and expression of vasoactive substances,³³ thus contributing to susceptibility of the vessels to develop vascular diseases. Within vasoactive substances (vasodilator and vasoconstrictor) implicated in the regulation of vascular tone and resistance can be included, among others, nitric oxide (NO),³⁴ angiotensin II,¹⁴ PGs and TXs.^{35–37}

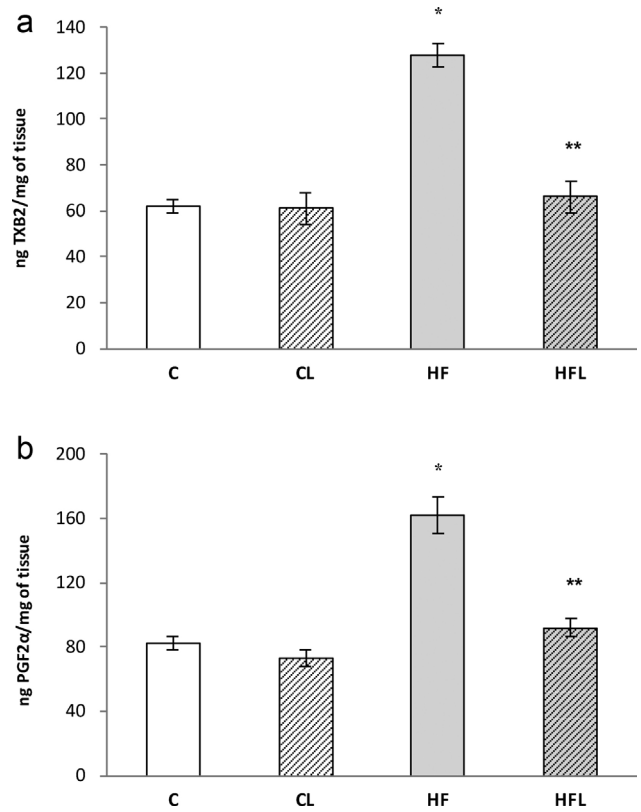


Figure 2 (a) Release of TXB₂ in control (C), high-fat diet (HF), losartan-control (CL), losartan- high-fat diet (HFL). * $P<0.01$ vs. C, CL; ** $P<0.01$ vs. HF. (b) Release of PGF_{2α} in control (C), high-fat diet (HF), losartan-control (CL), losartan- high-fat diet (HFL). * $P<0.01$ vs. C, CL; ** $P<0.01$ vs. HF.

An ectopic fat deposition by high-fat diet may contribute to development of hypertension through increased activity of local RAAS in visceral adipose tissue linked to insulin resistance.³⁸ RAAS activation in PVAT could promote increased angiotensin II, exacerbating insulin resistance.³⁹ Microvascular insulin resistance may be a shared pathophysiological mechanism between hemodynamic and metabolic consequences of visceral adiposity dysfunction.⁴⁰

There is evidence that supports the role of insulin resistance in the endothelial dysfunction. The local vascular effect of insulin beyond systemic effects, is modulated by production of the vasodilator NO via activation of insulin receptor substrate-1 (IRS-1)/phosphatidylinositol 3-kinase (PI3 kinase) AKT/endothelial NO synthase (eNOS) pathway. Also, increased angiotensin II levels due to PVAT dysfunction can favor microvascular vasoconstriction through AT₁ receptor by stimulating the secretion of vasoconstrictor prostanoids.⁴¹ The activation of local RAAS in PVAT produce alterations in this signaling resulting in decreased beneficial vascular effects of metabolic insulin.^{42–44}

An upregulated expression of cyclooxygenase (COX) in visceral fat that drive production of vasoconstrictor prostanoids and reactive oxygen species is an important hallmark in the pathogenesis of endothelial dysfunction observed in obesity-related conditions such as metabolic syndrome, hypertension and atherosclerosis.⁴⁵ Angiotensin II regulates COX-2 expression and prostanoid release via AT₁

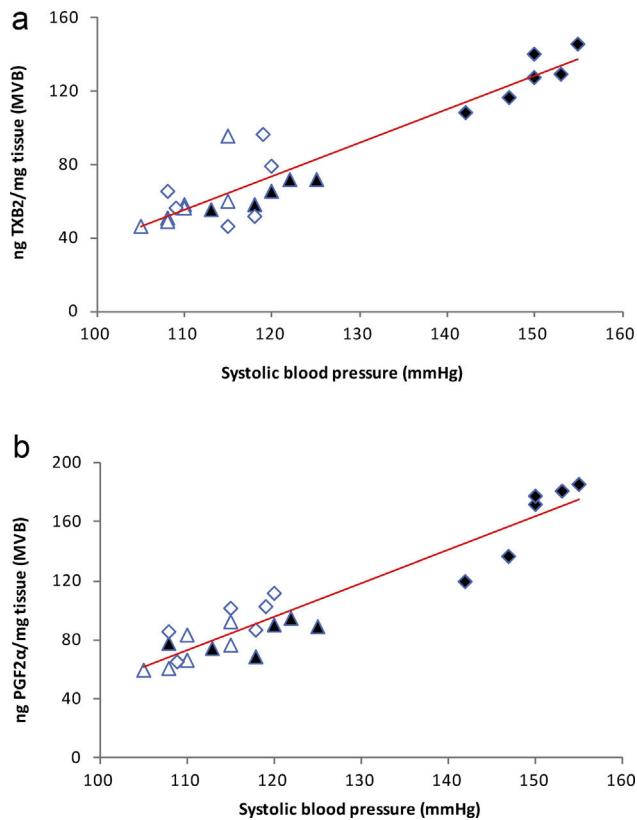


Figure 3 (a) Linear regression of systolic blood pressure against TXB₂ release: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r=0.93$, $R^2=0.87$, $P<0.01$. (b) Linear regression of systolic blood pressure against PGF_{2 α} release: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r=0.95$, $R^2=0.89$, $P<0.01$.

receptors activation.^{46,47} It has been reported that losartan reduces COX-2 mRNA up regulation and also acts as a competitive antagonist of the thromboxane A₂ receptor.⁴⁸

In accordance with previous studies,^{18,19} we found increases of PGE₂, PGF_{2 α} and TXB₂ induced by a high-fat diet at 8 weeks. An increase of pro-inflammatory and contractile substances in PVAT could be associated with the development of insulin resistance, as well as endothelial dysfunction. We have found that adiposity index correlates positively not only with the release of PGE₂, but also with vasoconstrictors prostanoids release (PGF_{2 α} , TXB₂), suggesting a pathophysiological connection. Present results demonstrated a preventive action of losartan on the release of those prostanoids induced by high-fat diet.

It has been reported that losartan reduced TXA₂, PGE₂ and PGF_{2 α} release from second, third and fourth branches of mesenteric artery cleaned of fat in streptozotocin rat model.⁴⁸ Ishida et al.⁴⁹ found losartan treatment reduced abnormal release of PGE₂ and PGF_{2 α} in stimulated rings of the superior mesenteric artery without PVAT. Moreover, Matsumoto et al.⁵⁰ described that losartan suppressed endothelium stimulated release of prostanoids in mesenteric arteries rings. Wanderer et al.⁵¹ found that losartan reduces a PGF₂-induced vasoconstriction in basilar artery ring segments after subarachnoid hemorrhage. As far as we know, our results provide new data regarding the preventive effect

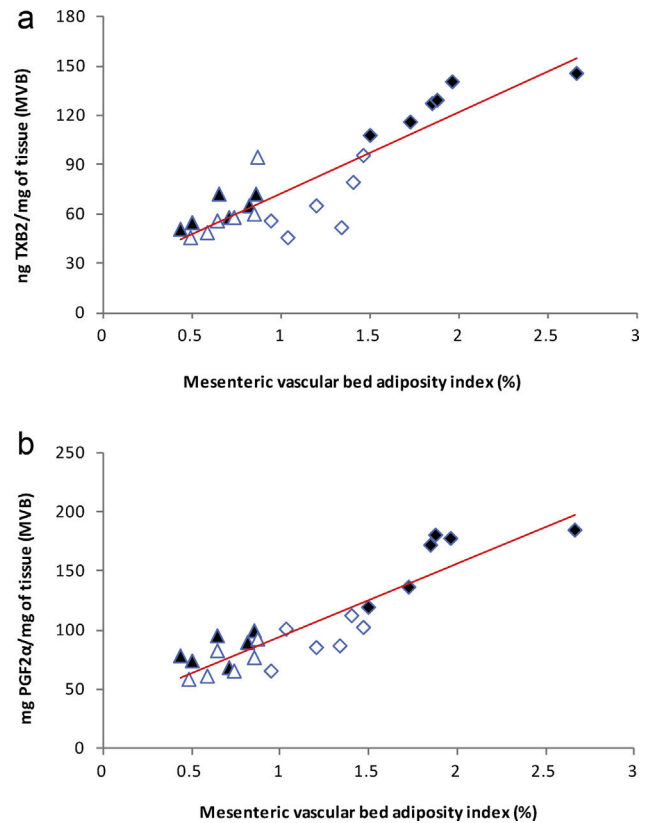


Figure 4 (a) Linear regression of mesenteric vascular bed adiposity index against TXB₂ release: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r=0.89$, $R^2=0.80$, $P<0.01$. (b) Linear regression of mesenteric vascular bed adiposity index against PGF_{2 α} release: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r=0.90$, $R^2=0.82$, $P<0.01$.

of losartan on prostanoids release from the entire mesenteric vascular bed in a metabolic syndrome model induced by high-fat diet.

A role for vasoconstrictor prostanoids in hypertension associated metabolic syndrome may be related to increased production of oxygen-derived free radicals determining abnormalities of vascular function that affects expression of eNOS and bioavailability of NO. Moreover, oxidative stress linked to the activation of protein kinase C and the NADPH oxidase regulates prostanoids activity in endothelial dysfunction.⁵²⁻⁵⁴ Bayorh et al.⁵⁵ reported losartan attenuation of oxidative stress induced by glutathione depletion in Sprague-Dawley rats.

Endothelial dysfunction is one of the trigger factors in the pathogenesis of hypertension associated to metabolic syndrome and cardiovascular diseases. We have also demonstrated a decrease in the PGI₂/TXA₂ ratio, suggesting an endothelial dysfunction in mesenteric vascular bed due to a pro-inflammatory and contractile state of the PVAT exposed to a high-fat diet. Losartan treatment prevented the PGI₂/TXA₂ ratio alteration induced by high-fat diet.

Finally, our findings attribute, at least in part, endothelial dysfunction in mesenteric vascular bed to alterations on prostanoids release in the context of the triad: adiposity, HOMA IR and hypertension.

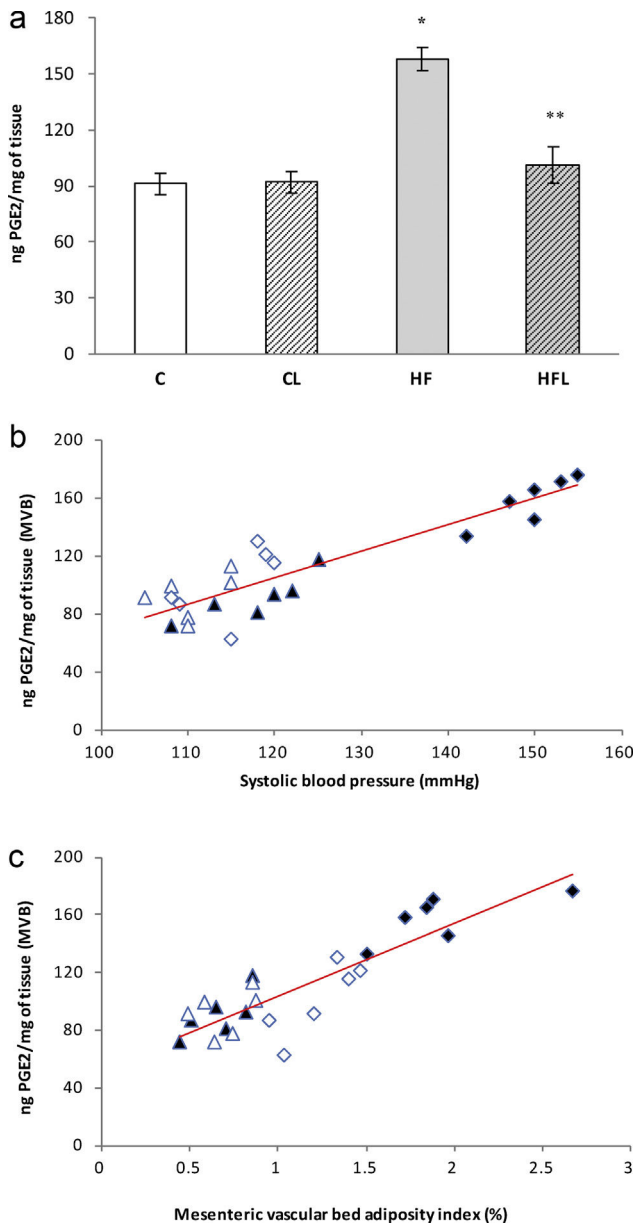


Figure 5 (a) Release of PGE₂ in control (C), high-fat diet (HF), losartan-control (CL), losartan-high-fat diet (HFL). * $P < 0.01$ vs. C, CL; ** $P < 0.01$ vs. HF. (b) Linear regression of systolic blood pressure against PGE₂ release: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r = 0.90$, $R^2 = 0.81$, $P < 0.01$. (c) Linear regression of mesenteric vascular bed adiposity index against PGE₂ release: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r = 0.88$, $R^2 = 0.80$, $P < 0.01$.

Conclusion

PVAT dysfunction produce alterations in the release of both vasoconstrictor and vasodilator factors that play a fundamental role in the endothelial dysfunction in hypertension associated to metabolic syndrome. This work has focused mainly on the prostanoids of the mesenteric vascular bed to point out their relevance in vascular function. One of the new potential therapeutic targets to be considered is PVAT.

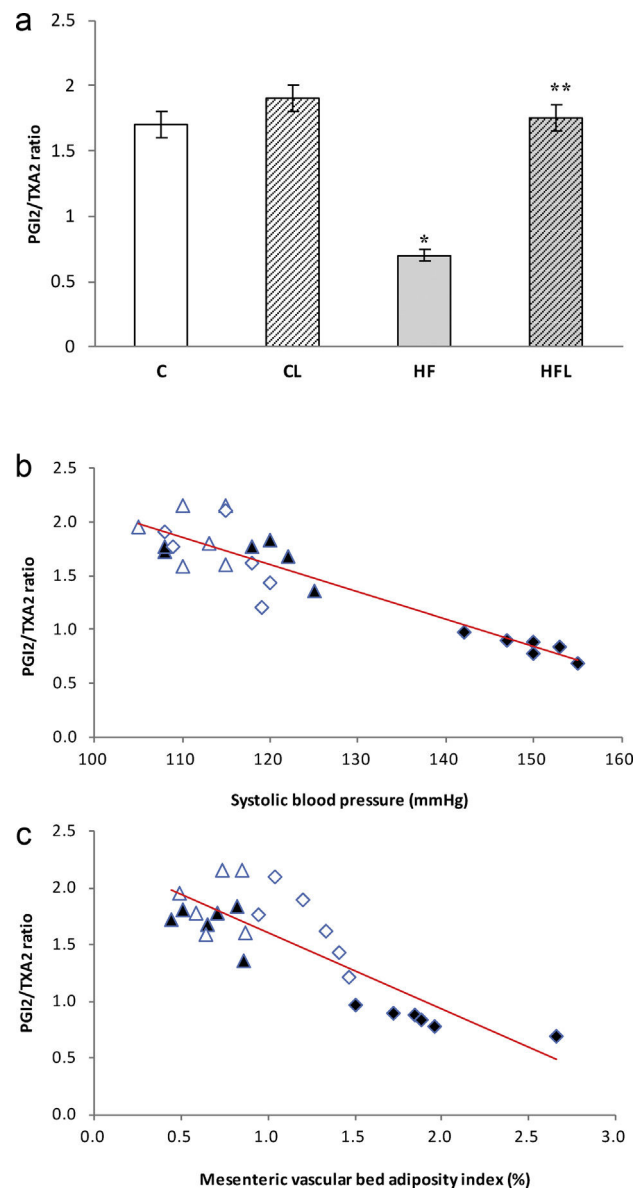


Figure 6 (a) PGI₂/TXA₂ release ratio in control (C), high-fat diet (HF), losartan-control (CL), losartan-high-fat diet (HFL). * $P < 0.01$ vs. C, CL; ** $P < 0.01$ vs. HF. (b) Linear regression of systolic blood pressure against PGI₂/TXA₂ release ratio: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r = 0.91$, $R^2 = 0.82$, $P < 0.01$. (c) Linear regression of mesenteric vascular bed adiposity index against PGI₂/TXA₂ release ratio: Control (▲), high-fat diet (◆), losartan-control (△), losartan-high-fat diet (◇). $r = 0.84$, $R^2 = 0.72$, $P < 0.01$.

Our experimental findings provide more evidence to choose losartan in the treatment of hypertension in patients considered to be at high risk for developing metabolic diseases. We demonstrated that losartan treatment prevents vasoconstrictor and inflammatory prostanoids release as well as mesenteric vascular bed adiposity increase induced by a high-fat diet. We have shown that losartan plays multiple positive actions/effects beyond its antihypertensive effect.

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Conflict of interest

All authors have declared no conflict of interest.

References

- Meijer R, Serne EH, Smulders YM, van Hinsbergh VW, Yudkin JS, Eringa EC. Perivascular adipose tissue and its role in type 2 diabetes and cardiovascular disease. *Curr Diab Rep.* 2011;11:211–7.
- Van de Voorde J, Boydens C, Pauwels B, Decaluwé K. Perivascular adipose tissue, inflammation and vascular dysfunction in obesity. *Curr Vasc Pharmacol.* 2014;12:403–11.
- Lian X, Gollasch M. A clinical perspective: contribution of dysfunctional perivascular adipose tissue (PVAT) to cardiovascular risk. *Curr Hypertens Rep.* 2016;18:82.
- Agabiti-Rosei C, Paini A, De Ciuceis C, Withers S, Greenstein A, Heagerty AM, et al. Modulation of vascular reactivity by perivascular adipose tissue (PVAT). *Curr Hypertens Rep.* 2018;20:44.
- Yudkin JS, Eringa E, Stehouwer CD. "Vasocrine" signaling from perivascular fat: a mechanism linking insulin resistance to vascular disease. *Lancet.* 2005;365:1817–20.
- Despres JP, Lemieux I. Abdominal obesity and metabolic syndrome. *Nature.* 2006;444:881–7.
- Putnam K, Shoemaker R, Yiannikouris F, Cassis LA. The renin-angiotensin system: a target of and contributor to dyslipidemias, altered glucose homeostasis, and hypertension of the metabolic syndrome. *Am J Physiol Heart Circ Physiol.* 2012;302:H1219–30.
- Sivasubramanian S, Kumarasamy B. Pleiotropic effects of losartan in hypertensive patients with dyslipidemia. *J Clin Diagn Res.* 2017;11:FC05–8.
- Szasz T, Bomfim GF, Webb RC. The influence of perivascular adipose tissue on vascular homeostasis. *Vasc Health Risk Manag.* 2013;9:05–16.
- van Dam AD, Boon MR, Berbée JFP, Rensen PCN, van Harmelen V. Targeting white, brown and perivascular adipose tissue in atherosclerosis development. *Eur J Pharmacol.* 2017;816:82–92.
- Byrnes KG, Walsh D, Lewton-Brain P, McDermott K, Coffey JC. Anatomy of the mesentery: historical development and recent advances. *Semin Cell Dev Biol.* 2019;92:4–11.
- Hildebrand S, Stümer J, Pfeifer A. PVAT and its relation to brown, beige, and white adipose tissue in development and function. *Front Physiol.* 2018;9:70.
- Frontini A, Cinti S. Distribution and development of brown adipocytes in the murine and human adipose organ. *Cell Metab.* 2010;11:253–6.
- Gálvez-Prieto B, Bolbrinker J, Stucchi P, de Las Heras AI, Merino B, Arribas S, et al. Comparative expression analysis of the renin-angiotensin system components between white and brown perivascular adipose tissue. *J Endocrinol.* 2008;197:55–64.
- Cassis LA, Fettinger MJ, Roe AL, Shenoy UR, Howard G. Characterization and regulation of angiotensin II receptors in rat adipose tissue. *Adv Exp Med Biol.* 1996;396:39–47.
- Mendizábal Y, Llorens S, Nava E. Vasoactive effects of prostaglandins from the perivascular fat of mesenteric resistance arteries in WKY and SHROB rats. *Life Sci.* 2013;93:1023–32.
- Peredo HA, Lee HJ, Donoso AS, Andrade V, Sánchez Eluchans N, Puyó AM. A high fat plus fructose diet produces a vascular prostanoid alterations in the rat. *Auton Autocoid Pharmacol.* 2015;34:35–40.
- Lee HJ, Cantú SM, Donoso AS, Choi MR, Peredo HA, Puyó AM. Metformin prevents vascular prostanoid release alterations induced by a high-fat diet in rats. *Auton Autocoid Pharmacol.* 2017;37:37–43.
- Peredo HA, Mayer MA, Carranza A, Puyó AM. Pioglitazone and losartan modify hemodynamic and metabolic parameters and vascular prostanoids in fructose-overloaded rats. *Clin Exp Hypertens.* 2008;30:159–69.
- Boshra V, El Wakeel G, Nader M. Effect of celecoxib on the anti-hypertensive effect of losartan in a rat model of renovascular hypertension. *Can J Physiol Pharmacol.* 2011;89:103–7.
- Smith PM, Hindmarch CC, Murphy D, Ferguson AV. AT1 receptor blockade alters nutritional and biometric development in obesity-resistant and obesity-prone rats submitted to a high fat diet. *Front Psychol.* 2014;5:832.
- Institute for Laboratory Animal Research. Guide for the care and use of laboratory animals. Washington (DC): National Academies Press.
- Nowland MH, Hugunin KMS, Rogers KL. Effects of short-term fasting in male Sprague-Dawley rats. *Compar Med.* 2011;61:138–47.
- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentration in man. *Diabetologia.* 1985;28:412–9.
- Wang H, Wang Q, Liang C, Su M, Wang X, Li H, et al. Acupuncture regulating gut microbiota in abdominal obese rats induced by high-fat diet. *Evid Based Complement Alternat Med.* 2019;2019:4958294.
- Mourad AA, Heeba GH, Taye A, El-Moselhy MA. Comparative study between atorvastatin and losartan on high fat diet-induced type 2 diabetes mellitus in rats. *Fundam Clin Pharmacol.* 2013;27:489–97.
- Rosselli MS, Burgueño AL, Carabelli J, Schuman M, Pirola CJ, Sookoian S. Losartan reduces liver expression of plasminogen activator inhibitor-1 (PAI-1) in a high fat-induced rat nonalcoholic fatty liver disease model. *Atherosclerosis.* 2009;206:119–26.
- Oliveira-Junior SA, Dal Pai M, Guizoni DM, Torres BP, Martinez PF, Campos DHS, et al. Effects of AT1 receptor antagonism on interstitial and ultrastructural remodeling of heart in response to a hypercaloric diet. *Physiol Rep.* 2019;7:e13964.
- Sabry MM, Mahmoud MM, Shoukry HS, Rashed L, Kamar SS, Ahmed MM. Interactive effects of apelin, renin-angiotensin system and nitric oxide in treatment of obesity-induced type 2 diabetes mellitus in male albino rats. *Arch Physiol Biochem.* 2019;125:244–54.
- Wang T, Lian G, Cai X, Lin Z, Xie L. Effect of prehypertensive losartan therapy on AT1R and ATRAP methylation of adipose tissue in the later life of high-fat-fed spontaneously hypertensive rats. *Mol Med Rep.* 2018;17:1753–61.
- Houben AJ, Eringa EC, Jonk AM, Serne EH, Smulders YM, Stehouwer CD. Perivascular fat and the microcirculation: relevance to insulin resistance diabetes, and cardiovascular disease. *Curr Cardiovasc Risk Rep.* 2012;6:80–90.
- Levy BI, Schiffrin EL, Mourad JJ, Agostini D, Vicaut E, Safar ME, et al. Impaired tissue perfusion: a pathology common to hypertension, obesity, and diabetes mellitus. *Circulation.* 2008;118:968–76.
- Guzik TJ, Marvar PJ, Czesnikiewicz-Guzik M, Korb R. Perivascular adipose tissue as a messenger of the brain-vessel axis: role

- in vascular inflammation and dysfunction. *J Physiol Pharmacol*. 2007;58:591–610.
34. Gil-Ortega M, Stucchi P, Guzman-Ruiz R, Cano V, Arribas S, Gonzalez MC, et al. Adaptive nitric oxide overproduction in perivascular adipose tissue during early diet-induced obesity. *Endocrinology*. 2010;151:3299–306.
 35. Chang L, Villacorta L, Li R, Hamblin M, Xu W, Dou C, et al. Loss of perivascular adipose tissue on peroxisome proliferator-activated receptor gamma deletion in smooth muscle cells impairs intravascular thermoregulation and enhances atherosclerosis. *Circulation*. 2012;126:1067–78.
 36. Ozen G, Topal G, Gomez I, Ghorreshi A, Boukais K, Benyahia C, et al. Control of human vascular tone by prostanoids derived from perivascular adipose tissue. *Prostaglandins Other Lipid Mediat*. 2013;107:13–7.
 37. Meyer MR, Fredette NC, Barton M, Prossnitz ER. Regulation of vascular smooth muscle tone by adipose-derived contracting factor. *PLoS One*. 2013;8:e79245.
 38. Dorresteyn JA, Visseren FL, Spiering W. Mechanisms linking obesity to hypertension. *Obes Rev*. 2012;13:17–26.
 39. Engeli S, Negrel R, Sharma AM. Physiology and pathophysiology of the adipose tissue renin–angiotensin system. *Hypertension*. 2000;35:1270–7.
 40. Karaca Ü, Schram MT, Houben AJ, Muris DM, Stehouwer CD. Microvascular dysfunction as a link between obesity, insulin resistance and hypertension. *Diabetes Res Clin Pract*. 2014;103:382–7.
 41. Muniyappa R, Yavuz S. Metabolic actions of angiotensin II and insulin: a microvascular endothelial balancing act. *Mol Cell Endocrinol*. 2013;378:59–69.
 42. Brillante DG, O’Sullivan AJ, Howes LG. Arterial stiffness in insulin resistance: the role of nitric oxide and angiotensin II receptors. *Vasc Health Risk Manag*. 2009;5:73–8.
 43. Aroor AR, Demarco VG, Jia G, Sun Z, Nistala R, Meininger GA, et al. The role of tissue Renin–Angiotensin–aldosterone system in the development of endothelial dysfunction and arterial stiffness. *Front Endocrinol (Lausanne)*. 2013;4:161.
 44. Nóbrega N, Araújo NF, Reis D, Facine LM, Miranda CAS, Mota GC, et al. Hydrogen peroxide and nitric oxide induce anticontractile effect of perivascular adipose tissue via renin angiotensin system activation. *Nitric Oxide*. 2019;84:50–9.
 45. Traupe T, Lang M, Goettsch W, Munter K, Morawietz H, Vetter W, et al. Obesity increases prostanoid-mediated vasoconstriction and vascular thromboxane receptor gene expression. *J Hypertens*. 2002;20:2239–45.
 46. Hu ZW, Kerb R, Shi XY, Wei-Lavery T, Hoffman BB. Angiotensin II increases expression of cyclooxygenase-2: implications for the function of vascular smooth muscle cells. *J Pharmacol Exp Ther*. 2002;303:563–73.
 47. Kamo T, Akazawa H, Komuro I. Pleiotropic effects of angiotensin II receptor signaling in cardiovascular homeostasis and aging. *Int Heart J*. 2015;56:249–54.
 48. de Queiroz D, Ramos-Alves F, Santos-Rocha J, Duarte G, Xavier F. Losartan reverses COX-2-dependent vascular dysfunction in offspring of hyperglycaemic rats. *Life Sci*. 2017;184:71–80.
 49. Ishida K, Matsumoto T, Taguchi K, Kamata K, Kobayashi T. Mechanisms underlying altered extracellular nucleotide-induced contractions in mesenteric arteries from rats in later-stage type 2 diabetes: effect of ANG II type 1 receptor antagonism. *Am J Physiol Heart Circ Physiol*. 2011;301:H1850–61.
 50. Matsumoto T, Ishida K, Nakayama N, Taguchi K, Kobayashi T, Kamata K. Mechanisms underlying the losartan treatment-induced improvement in the endothelial dysfunction seen in mesenteric arteries from type 2 diabetic rats. *Pharmacol Res*. 2010;62:271–81.
 51. Wanderer S, Mrosek J, Gessler F, Seifert V, Konczalla J. Vasomodulatory effects of the angiotensin II type 1 receptor antagonist losartan on experimentally induced cerebral vasospasm after subarachnoid haemorrhage. *Acta Neurochirurgica (Wien)*, 160(2):277–284.
 52. Zou MH, Shi C, Cohen RA. High glucose via peroxynitrite causes tyrosine nitration and inactivation of prostacyclin synthase that is associated with thromboxane/prostaglandin H(2) receptor-mediated apoptosis and adhesion molecule expression in cultured human aortic endothelial cells. *Diabetes*. 2002;51:198–203.
 53. Cosentino F, Eto M, De Paolis P, van der Loo B, Ullrich V, Kouroedov A, et al. High glucose causes upregulation of cyclooxygenase-2 and alters prostanoid profile in human endothelial cells: role of protein kinase c and reactive oxygen species. *Circulation*. 2003;107:1017–23.
 54. Guo Z, Su W, Allen S, Pang H, Daugherty A, Smart E, et al. Cox-2 up-regulation and vascular smooth muscle contractile hyperreactivity in spontaneous diabetic db/db mice. *Cardiovasc Res*. 2005;67:723–35.
 55. Bayorh M, Ganafa A, Socci R, Eatman D, Silvestrov N, Abukhalaf I. Effect of losartan on oxidative stress-induced hypertension in Sprague-Dawley rats. *Am J Hypertens*. 2003;16 Pt 1:387–92.