



Figure 1 Erythema on the wrist lesion after single-blind oral challenge with norfloxacin.

Our literature review revealed two reports of fixed drug eruption due to norfloxacin. Fernández-Rivas⁸ described a patient with several episodes of fixed drug eruption after ingestion of norfloxacin. Patch tests were negative with all the quinolones tested and the patient tolerated ciprofloxacin. In contrast with that case, our patient did not tolerate the other quinolones tested.

Rodríguez-Morales et al.⁹ reported a case of fixed drug eruption from quinolones (ciprofloxacin and norfloxacin) with a positive result in the patch test (ciprofloxacin) applied to the lesion. All patch tests were negative in our patient.

Controlled oral challenge seems to be the only way to determine the diagnosis and to demonstrate cross-reactivity, since the results of patch testing are inconclusive.⁵

In summary, we present a case of fixed drug eruption induced by norfloxacin, as confirmed by oral challenge and biopsy of the lesion.

Conflict of interest

The authors have no conflict of interest to declare.

Acknowledgement

This work was supported by grants from Spanish Health Ministry (FIS) network RIRAAF (RD07/0064).

References

1. Sánchez-Morillas L, Laguna-Martínez JJ, Reaño-Martos M, Gómez-Tembleque P, Rojo-Andrés E. Fixed drug eruption caused by amoxicillin-clavulanic acid. *Ann Allergy Asthma*. 2008;101:335.
2. Alonso MD, Martín JA, Quirce S, Dávila I, Lezaun A, Sánchez Cano M. Fixed eruption caused by ciprofloxacin with cross-sensitivity to norfloxacin. *Allergy*. 1993;48:296–7.
3. Sánchez-Morillas L, Rojas Pérez-Ezquerria P, Reaño-Martos M, Laguna Martínez JJ, Gómez-Tembleque Úbeda P. Systemic anaphylaxis caused by moxifloxacin. *Allergol Immunopathol*. 2010;38:226–7.
4. Ramsay B, Woodrow D, Cream JJ. An acantholytic bullous eruption after norfloxacin. *Br J Dermatol*. 1993;129:500.
5. Lozano Ayllón M, Gómez Martínez M, Mosquera MR, Laguna Martínez JJ, Orta Martiartu M, Fernández de Miguel C. Fixed eruption caused by ciprofloxacin without cross-sensitivity to norfloxacin. *Allergy*. 1993;50:598–9.
6. Dhar S, Sharma VK. Fixed drug eruption due to ciprofloxacin. *Br J Dermatol*. 1996;134:156–8.
7. Rojas Pérez-Ezquerria P, Sánchez-Morillas L, Santos Álvarez A, Gómez-Tembleque Úbeda P, Blanco Moratíel H, Laguna Martínez JJ. Fixed drug eruption caused by amoxicillin-clavulanic acid. *Contact Dermatitis*. 2010;63:294–6.
8. Fernández-Rivas M. Fixed drug eruption caused by norfloxacin. *Allergy*. 1997;52:477–8.
9. Rodríguez-Morales A, Alonso Llamazares A, Palacios Benito R, Martínez Cócera C. Fixed drug eruption from quinolones with a positive lesional patch test to ciprofloxacin. *Contact Dermatitis*. 2001;44:255.

L. Sánchez-Morillas^{a,*}, P. Rojas Pérez-Ezquerria^a, M.L. González Morales^b, C. Mayorga^c, R. González-Mendiola^a, J.J. Laguna Martínez^a

^a *Allergology Department, Hospital Central de la Cruz Roja, Madrid, Spain*

^b *Anatomy Pathology Department, Hospital Central de la Cruz Roja, Madrid, Spain*

^c *Allergology Department, Hospital Carlos Haya-Fundación IMABIS, Málaga, Spain*

*Corresponding author.

E-mail address: lsanchezmorillas@hotmail.com (L. Sánchez-Morillas).

doi:10.1016/j.aller.2011.10.004

Interleukin (IL)-22 serum level in hypersensitivity pneumonitis (HP) in a mushroom worker

To the Editor,

Hypersensitivity pneumonitis (HP) is a pulmonary disease with symptoms of dyspnoea and cough resulting from the inhalation of an antigen to which the patient has been previously sensitised.¹

HP is a disease that occurs as a consequence of exposure to organic dust. Initially, it was associated with farming (mouldy grain or hay handling), hence the term farmer's lung.² With time, a large variety of environmental settings and antigens have been described.³

HP caused by inhalation of mushroom spores has increased, and HP has been described as an occupational hazard of mushroom plant workers.⁴

Edible oyster mushrooms, *Pleurotus species*, are cultivated all over the world.⁵

The indoor cultivation of *Pleurotus ostreatus* regularly led to allergic symptoms in workers.⁶

IL-22 is a pro-inflammatory cytokine belonging to the IL-10 family and represents an important effector molecule of activated lymphocyte T helper (Th)22, Th1, Th17 cells.⁷

IL-22 increases the innate immunity of tissue cells, protects tissues from damage and enhances their regeneration. It exerts pro-inflammatory effects, is present in systemic and local inflammation, and exerts an important role in the antimicrobial defence.⁸

Given the fact that IL-22 prominently increases the expression of a range of antimicrobially acting proteins in various epithelia suggests a role for this cytokine in the innate immune defence, especially against extracellular bacteria. Moreover, further mechanisms used by IL-22 to promote the defence against intracellular bacteria, fungi, viruses and parasites are currently under exploration.⁹

Simonian et al. showed that a subset of $\gamma\delta$ T cells represents the predominant source of Th17 cytokine IL-22 in a murine model of HP. Preventing expression of IL-22 they accelerated lung fibrosis. Direct blockage of IL-22 also enhanced collagen deposition in the lung, whereas administration of recombinant IL-22 inhibited lung fibrosis. These data revealed a protective pathway that involves the inhibition of $\gamma\delta$ T cells by regulatory IL-22-secreting $\gamma\delta$ T cells.¹⁰

Therefore we evaluated IL-22 serum levels in a patient suffering from HP and then we compared them with seven healthy subjects of the same sex and similar age (mean \pm SD: 9.45 ± 6.14 pg/ml).

IL-22 serum concentrations were measured by a quantitative enzyme immunoassay technique. The assay was performed using a commercially available kit (R&D Systems Europe Abingdon, UK).

A 23-year-old man reported recurrent episodes of chest tightness, cough, wheezing symptoms accompanied by rhinoconjunctivitis, fever (37.5–38.5°C), and fatigue; these symptoms occurred in the evenings after work for about a year.

The patient reported to work continuously for about three years in a greenhouse, where he cultivated edible mushrooms (*P. ostreatus*); moreover he stated that his contact, even if occasionally, had begun about ten years before.

We carried out the common blood tests; the differential blood count showed a slight increase in white blood cells ($11,660\text{mm}^3$) and decrease in haematocrit (37.5%), the C-reactive protein (CRP) and the fibrinogen were over the range (14.60mg/dl and 410g/dl respectively) while serum iron was below the standard (25mcg/dl), the specific IgE were positive for *Dermatophagoides farinae* (18.90 KU/l), *Dermatophagoides pteronyssinus* (21.00 KU/l), *Cynodon dactylon* (2.33 KU/l), *Fetuca elaitor* (6.46 KU/l), *Cladosporium herbarum* (6.21 KU/l), and *Parietaria judaica* (1.13 KU/l).

Positive specific IgE could explain the respiratory symptoms but not the fever, so we performed a chest X-ray that showed striated patchy opacities; proceeding in the diagnostic work we performed a high resolution computed tomography (HRCT) lung scanning that showed in the expiratory phase a marked and uneven alteration in density in both lungs for the coexistence of higher density and hyperlucency areas (part of the mosaic attenuation). These areas

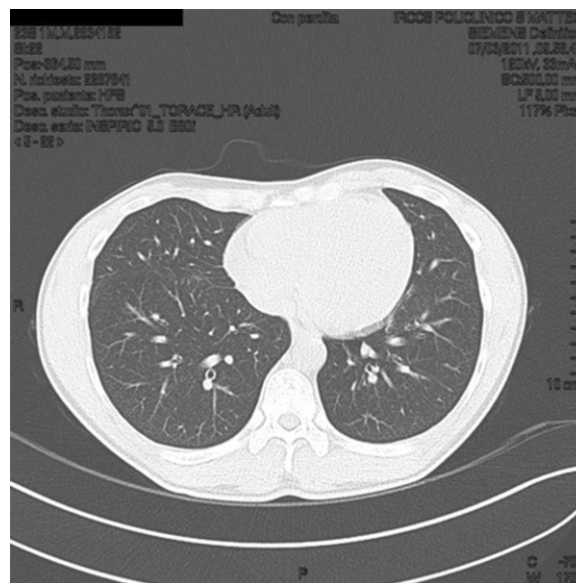


Figure 1 High resolution computed tomography (HRCT) lung at T0 shows in the expiratory phase a marked and uneven alteration in density of both lungs for the coexistence of higher density and hyperlucency areas (part of the mosaic attenuation). These areas represent pulmonary expiratory air trapping secondary to obstruction of the small airways.

represent pulmonary expiratory air trapping secondary to obstruction of the small airways (Fig. 1).

Bronchoalveolar lavage (BAL) fluid demonstrated an increase of total cells ($2.3 \times 10^6/\text{ml}$), lymphocytes (55% of the total cells), and eosinophils (1%). CD4/CD8 ratio of lymphocyte surface markers was 0.4.

The results of BAL (marked lymphocytosis, CD4+/CD8+ ratio lower than one) confirmed the clinical suspicion of hypersensitivity pneumonitis.

So, after obtaining written informed consent, a 10-ml blood sample was collected from the antecubital vein, to perform IL-22 serum levels assay at T0 (Table 1).

The IL-22 serum levels were lower than the minimum of seven controls (2.460 pg/ml vs. 4.270 pg/ml).

Table 1 IL-22 serum levels in seven healthy subjects (Controls) and in patient suffering from hypersensitivity pneumonitis (HP) at T0 and at T1 (three months after removal from the workplace).

Control no.	IL-22 serum levels (pg/ml)
1	11.516
2	5.886
3	5.371
4	12.049
5	5.577
6	21.458
7	4.270
Patient	IL-22 serum levels (pg/ml)
T0	2.460
T1	0.868

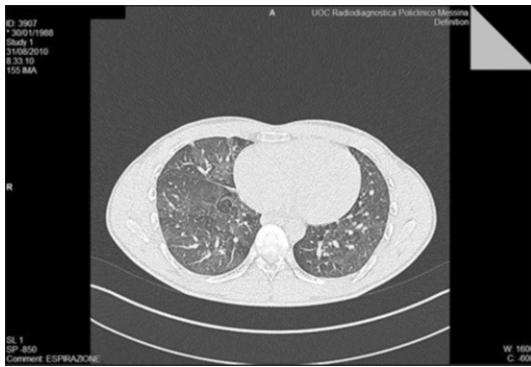


Figure 2 High resolution computed tomography (HRCT) lung at T1 shows some images of the mantle micronodular lung parenchyma, which do not increase in size than the previous examination. At the current appreciation we do not identify aspects frosted glass, or inter- and intralobular septal thickening, or clumps of inflammatory nature. We do not appreciate honeycombing, bronchiectasis or no bronchiolectasis traction. The density of lung parenchyma in breath is regular. It indicates only moderate diffuse abnormalities of the bronchial walls, which appear minimally thickened. This finding has not progressed from the previous examination.

We removed the patient from his workplace and after three months we repeated the blood tests: the white blood cells, haematocrit, CRP, fibrinogen and serum iron were normalised, the HRCT scanning showed some images of the mantle micronodular lung parenchyma, no increase in size with respect to previous examination. No frosted glass aspect was observed, or inter- and intralobular septal thickening, or clumps of inflammatory nature. Nor was there detectable honeycombing, bronchiectasis or bronchiolectasis traction. The density of lung parenchyma during breathing was regular. This reveals only moderate diffuse abnormalities of the bronchial walls, which appear minimally thickened. This finding has not progressed from the previous examination (Fig. 2).

Furthermore, the patient reported the disappearance of symptoms after only seven days off work.

We repeated the IL-22 assay at T1, the level was still lower than the minimum control (0.868 pg/ml vs. 4.270 pg/ml).

IL-22 has emerged as an important cytokine in mucosal immunity. The mechanisms by which IL-22 protects epithelial structures in the lung from damage induced by numerous environmental insults that are under intense investigation. Emerging data suggest that IL-22 may provide mucosal protection by inducing antimicrobial peptides from epithelial cells in the lung as well as maintaining epithelial integrity either by preventing injury or accelerating epithelial repair after a variety of insults.⁸

Although the IL-22 is reported to play a role in HP, the studies are limited to murine models; in our opinion this is the first report of involvement of this IL in a human patient suffering from HP.

We can hypothesise that the low IL-22 serum levels found in our patient at T0 were related to the patient's inability to

respond to injury and could explain the patient's susceptibility to developing HP. The decrease in IL-22 serum levels, after three-month suspension from work in the greenhouse was, in our opinion, related to removal from the antigenic stimulus.

IL-22 could thus become a useful marker for understanding in advance who is at risk of HP, but other studies and larger series are needed.

Acknowledgement

We would like to thank Ms. A. Donato for the editing of the text.

References

1. Lacasse Y, Selman M, Costabel U, Dalphin JC, Ando M, Morell R, et al. Clinical prediction rule for the diagnosis of active hypersensitivity pneumonitis (HP): the HP study. *Am J Respir Crit Care Med.* 2003;168:952–8.
2. Campbell JM. Acute symptoms following work with hay. *Br Med J.* 1932;2:1143–4.
3. Cormier Y, Schuyler M. Hypersensitivity pneumonitis and organic dust toxic syndromes. In: *Asthma and the workplace.* New York: Marcel Dekker; 2006.
4. Saikai T, Tanaka H, Fuji M, Sugawara H, Takeya I, Tsunematsu K, et al. Hypersensitivity pneumonitis induced by the spore of *Pleurotus Eryngii* (Eringi). *Intern Med.* 2002;41:571–3.
5. Farr DF. Mushroom industry: diversification with additional species in the United States. *Mycologia.* 1983;75:351–60.
6. Cox A, Folgering HTM, Van Griensven LJLD. Extrinsic allergic alveolitis caused by spores of the oyster mushroom *Pleurotus ostreatus*. *Eur Respir J.* 1988;1:466–8.
7. Wolk K, Witte E, Witte K, Warszawska K, Sabat R. Biology of interleukin-22. *Semin Immunopathol.* 2010;32:17–31.
8. Aujla SJ, Kolls JK. IL-22: a critical mediator in mucosal host defense. *J Mol Med.* 2009;87:451–4.
9. Witte E, Witte K, Warszawska K, Sabat R, Wolk K. Interleukin-22: a cytokine produced by T, NK and NKT cell subsets, with importance in the innate immune defense and tissue protection. *Cytokine Growth Factor Rev.* 2010;21:365–79.
10. Simonian PL, Wehrmann F, Roark CL, Born WK, O'Brien RL, Fontenot AP. $\gamma\delta$ T cells protect against lung fibrosis via IL-22. *J Exp Med.* 2010;207:2239–53.

Salvatore Saitta^{a,*}, Alfio Proietto^b, Giovanna Spataro^c, Sebastiano Gangemi^{a,d}

^a School and Unit of Allergy and Clinical Immunology, Department of Human Pathology, University of Messina, Italy

^b Unit of Pneumology, Department of Experimental Medicine and Pharmacology, University of Messina, Italy

^c Department of Social and Environmental Medicine, University of Messina, Italy

^d Institute of Biomedicine and Molecular Immunology-National Research Council, Palermo, Italy

*Corresponding author.

E-mail address: saittasalvatore@tiscalinet.it (S. Saitta).

doi:10.1016/j.aller.2012.01.001