

Coconut anaphylaxis: Case report and review



To the Editor,

The coconut (*Cocos nucifera*) is a monocotyledon plant belonging to the Aracaceae family. Although potentially severe, there are few allergic processes described that arise from ingestion of its fruit, although the majority of cases described to date have been anaphylactic reactions.^{1-7,10} The endosperm is the edible part of the fruit, and it is composed of 25% albumins and 75% globulins, the latter of which contains the proteins which have been implicated in allergic reactions, such as Coc n2, 7S globulin, and Coc n4, 11S globulin (known as cocosin). Under native conditions, molecular masses of the 7S and 11S globulins are 156 kDa and 326 kDa respectively, and the 7S globulin is composed of three protein subunits of 24 kDa, 22 kDa, and 16 kDa while the 11S globulin is composed of 55 kDa, 34 kDa, and 24 kDa subunits.⁸

Cross reactivity between coconuts, walnuts, hazelnuts and lentils has been described, and this is due to 7S and/or 11S globulins.^{1,4,6,7} Similarly, cases of monosensitization to coconut have been described.^{2,3,5}

Here we describe the case of a 12 and a half-year-old child with asthma and rhinitis caused by allergies which are being controlled with immunotherapy. The child is sensitized to *Dermatophagoides farinae* and *Dermatophagoides pteronyssinus*, *Alternaria* and dog and cat epitheliums, and has presented anaphylaxis from ingestion of crustaceans. At 12 years old, 10 min after ingesting a piece of coconut for the first time, the child presented an allergic reaction which lasted for an hour and consisted of spasmodic coughing, sneezing outbursts, rhinorrhea, vomiting, and difficulty in breathing with thoracic oppression and wheezing.

To investigate this coconut allergy, skin prick tests were performed with commercial extracts and the following mean wheal diameters were found: coconut, 14 mm, walnut and hazelnut, 5 mm, sesame, 6 mm and date, 3 mm with erythema. The histamine dihydrochloride (10 mg/ml) value detected was 7 mm.

Protein extracts, fresh coconut and coconut milk were prepared by homogenization in phosphate-buffered saline, dialization, and lyophilization. Serum specific immunoglobulin (Ig) E levels were measured using the enzyme allergosorbent technique (Specific IgE EIA kit HYTEC HYCOR Biomedical Ltd). Determination of serum specific IgE revealed the following values: fresh coconut, 0.6 kU_A/L, and coconut milk, 1 kU_A/L. Values detected against extracts from sesame, walnut and peanut were between 0.35 and 0 kU_A/L. Specific IgE against date extract was <0.35 kU_A/L.

Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) IgE immunoblotting revealed intense IgE binding bands at 27 kDa, 16 kDa, and a broad one between 20 and 25 kDa in both coconut extracts; some other faint bands were only detected in the fresh coconut extract at 80 kDa, 66 kDa, 39 kDa and 32/33 kDa (Fig. 1).

Finally, an open oral provocation test⁹ against walnut, hazelnut, sesame and date (foods whose tolerance were not known and to which sensitization was demonstrated in skin tests) was performed, and the result was negative. A provocation test against coconut was not performed as there was

the danger of an anaphylactic reaction and the family denied authorization to perform the test.

Nine cases of coconut allergy have been reported, all of which produce anaphylactic reactions,^{1-7,10} and four of which were in children.^{3,4,6,7} In four of the cases, the 11S globulin was identified as the principal agent responsible for the allergic process,^{1,2,4} in two of the cases it was the 7S globulin,⁶ and in one of the cases both proteins were implicated in the allergic reaction.⁷

The molecular masses of the most intense bands detected in this study coincide with the molecular masses described for the 7S globulin protein subunits, and this suggests that this protein is involved in the allergic process of this patient. A less intense band of approximately 80 kDa was also detected, and one with a similar molecular mass has been described in some cases of monosensitization to coconut.^{2,6}

In this case it was verified that the patient could tolerate ingestion of tree nuts (walnuts and hazelnuts), sesame seeds and dates, all of which had produced sensitization, and thus this is the second case described where there is an allergic reaction (anaphylactic) caused exclusively by ingestion of coconut in a child. The first case was of a 3-year-old child.³ The provocation results obtained, as well as low IgE specific values detected against walnut, sesame and date extracts lead us to believe that sensitization to these foods is due to cross reactive phenomena with coconut proteins, as has been shown in other cases.^{1,4,6,7}

In the last few years, the presence of molecules derived from coconuts has increased significantly in the food industry due to the use of coconut oil. Similarly, oil from oil palm, *Elaeis guineensis*, is also used in industry and the 7S globulin from the oil palm fruit is homologous to the 7S globulin from coconuts.⁸ Oils from both are also widely used in the cosmetic industry, where these oils can be found in 44–76%

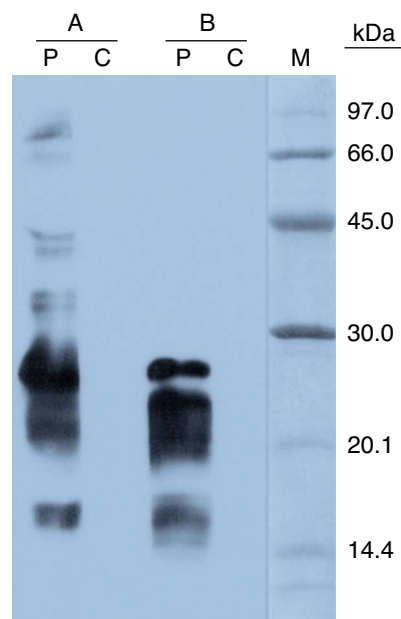


Figure 1 SDS-PAGE IgE immunoblotting result. (A) Fresh coconut extract. (B) Coconut milk extract. Lane P: patient serum. Lane C: control serum (pool of sera from non-atopic subject). Lane M: molecular mass marker.

of skin products (protective creams, soaps, body lotions, shampoos, etc.).¹¹

It has been suggested that sensitization to proteins from certain foods such as coconuts could be due to topical exposure through the skin. Whether it is through this route or the oral route by ingestion of foods containing products derived from coconuts, it is very probable that exposure to coconut proteins is now occurring from very early ages and thus the risk of sensitization to this fruit could facilitate an increase in allergic reactions to coconuts in the coming years.

In summary, we present a case of anaphylaxis in a child due to ingestion of coconut, and it is very probable that 7S globulin is the principal protein implicated in this allergic process.

Ethical disclosures

Protection of human and animal subjects. The authors declare that no experiments were performed on humans or animals for this article.

Data confidentiality. The authors declare that no patient details appear in this article.

Privacy rights and informed consent. The authors declare that no patient details appear in this article.

Conflicts of interest

We declare that we do not have any financial or personal relationship with regard to the submitted publication.

References

1. Teuber SS, Rich Peterson W. Systemic allergic reaction to coconut (*Cocos nucifera*) in 2 subjects with hypersensitivity to tree nut and demonstration of cross-reactivity to legumin-like seed storage proteins: new coconut and walnut food allergens. *J Allergy Clin Immunol.* 1999;103:1180–5.
2. Rosado A, Fernandez Rivas M, Gonzalez Mancebo E, León F, Campos C, Tejedor MA. Anaphylaxis to coconut. *Allergy.* 2002;57:182–3.

3. Tella R, Gaig P, Lombardero M, Paniagua MJ, Garcia-Ortega P, Richart C. A case of coconut allergy. *Allergy.* 2003;58:825–6.
4. Nguyen SA, More DR, Wishman BA, Hagan LL. Cross-reactivity between coconut and hazelnut proteins in a patient with coconut anaphylaxis. *Ann Allergy Asthma Immunol.* 2004;92:281–4.
5. Martin E, Tornero P, De Barrio M, Perez CI, Beitia JM, Baeza ML. Monosensitization to coconut. *J Allergy Clin Immunol.* 2004;113 2 Suppl:S315.
6. Benito C, Gonzalez Mancebo E, Alonso Diaz de Durana MD, Tolon RM, Fernandez Rivas M. Identification of a 7S globulin as a novel coconut allergen. *Ann Allergy Asthma Immunol.* 2007;98: 580–4.
7. Manso L, Pastor C, Perez-Gordo M, Cases B, Sastre J, Cuesta-Herranz J. Cross-reactivity between coconut and lentil related to a 7S globulin and an 11S globulin. *Allergy.* 2010;65: 1487–8.
8. Garcia RN, Arocena RV, Laurena AC, Tecson-Mendoza EM. 11S and 7S globulins of coconut (*Cocos nucifera* L.): purification and characterization. *J Agric Food Chem.* 2005;53:1734–9.
9. Sicherer SH. Oral food challenges for diagnosis and management of food allergies. *UpToDate.* 2013. March.
10. Stutius LM, Sheehan WJ, Rangsithienchai P, Bharmanee A, Scott JE, Young MC, et al. Characterizing the relationship between sesame, coconut, and nut allergy in children. *Pediatr Allergy Immunol.* 2010;21:1114–8.
11. Newhall KK, Amoroso LS, Sinacore JM, Pongracic JA. Presence of common food allergens in commercially available pediatric skin care products. *J Allergy Clin Immunol.* 2004;113: S235.

A. Michavila Gomez^{a,*}, M. Amat Bou^b, M.V. Gonzalez Cortés^b, L. Segura Navas^b, M.A. Moreno Palanques^a, B. Bartolomé^c

^a *Pediatric Allergy Unit, Castellón General Hospital, Spain*

^b *Pediatric Service, Castellón General Hospital, Spain*

^c *Research and Development Department, Bial-Aristegui, Bilbao, Spain*

* Corresponding author.

E-mail address: amichavila@gmail.com

(A. Michavila Gomez).

<http://dx.doi.org/10.1016/j.aller.2013.09.004>

Audit of the use of intravenous immunoglobulin for antibody deficiencies in a Clinical Immunology Unit



To the Editor,

Intravenous immunoglobulin preparations (IVIG) are indicated as the treatment of choice in a number of primary and secondary antibody deficiencies.^{1–3} There are many options for IgG replacement, and the choice is an individual one based on many factors.⁴ The present audit was developed to examine the current hospital practice of IVIG use for

replacement therapy of antibody deficiencies in a Clinical Immunology Unit.

A two-month (February–March 2013) prospective audit of all IVIG use for replacement therapy of antibody deficiencies in two Immunology Day Hospitals was conducted at the Clinical Immunology Unit in the Gregorio Marañón Hospital in Madrid. Patients receiving subcutaneous immunoglobulin were not included in this study. Medical files and dispensing records were prospectively examined. A questionnaire was used. Solid organ transplantation recipients receiving IVIG therapy because of IgG hypogammaglobulinaemia and severe infections were not included in this audit as they were receiving replacement IVIG therapy in other facilities of the hospital.