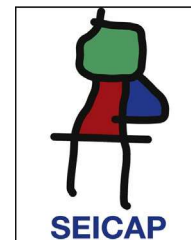




Allergologia et immunopathologia

Sociedad Española de Inmunología Clínica,
Alergología y Asma Pediátrica

www.elsevier.es/ai



ORIGINAL ARTICLE

Methacholine challenge test by wheezing and oxygen saturation in preschool children with asthma



S. Caussade^{a,*}, J.A. Castro-Rodriguez^a, S. Contreras^a, R. Bugueño^b, R. Ramirez^b,
O. Padilla^c, H. Einisman^a, N. Holmgren^a

^a Department of Pediatrics, School of Medicine, Pontificia Universidad Católica de Chile, Santiago, Chile

^b School of Medicine, Pontificia Universidad Católica de Chile, Santiago, Chile

^c Department of Public Health, School of Medicine, Pontificia Universidad Católica de Chile, Santiago, Chile

Received 15 October 2013; accepted 25 February 2014

Available online 16 June 2014

KEYWORDS

Methacholine;
Wheeze;
SpO₂;
Asthma;
Preschool

Abstract Methacholine challenge test (MCT) performed with spirometry is a commonly used test to evaluate bronchial hyperreactivity (BHR) in children. However, preschoolers do not usually collaborate.

Objectives: To assess the usefulness of MCT through clinical evaluation (wheezing auscultation and decreased pulse arterial oxygen saturation [SpO₂]) in recurrent wheezing preschoolers with asthma, in comparison to healthy controls.

Methods: We performed the MCT (modified Cockcroft method) on healthy and on asthmatic preschoolers. The end point was determined by the presence of wheezing in the chest and/or tracheal auscultation (PCw) and/or a decrease in SpO₂ of ≥ 5 from the baseline value (PCSpO₂). Maximal methacholine concentration was 8 mg/ml.

Results: The study population comprised 65 children: 32 healthy and 33 asthmatic children. There were no differences in demographic characteristics between the groups. The median methacholine doses for PCw and for PCSpO₂ were significantly lower among asthmatic than healthy children: 0.5 mg/ml (0.25–0.5 mg/ml) vs. 2 mg/ml (1–4 mg/ml), respectively, $p < 0.001$; and 0.25 mg/ml (0.25–0.5 mg/ml) and 2 mg/ml (0.5–4 mg/ml), respectively, $p < 0.001$. The best cut-off point of PCw was observed at a methacholine concentration of 0.5 mg/ml (AUC = 0.72 [95% CI = 0.66–0.77]), its sensitivity was 91%, specificity 43%, PPV 16% and NPV 98%. For PCSpO₂ the best cut-off point was a methacholine concentration of 1 mg/ml (AUC = 0.85 [95% CI 0.81–0.89]), with sensitivity of 80%, specificity 74%, PPV 49%, and NPV 92%. There were no adverse reactions.

Conclusion: MCT using clinical parameters such as wheezing auscultation and SpO₂ measurement could be a useful and safe test to confirm BHR among preschoolers.

© 2013 SEICAP. Published by Elsevier España, S.L.U. All rights reserved.

* Corresponding author.

E-mail addresses: solangecaussade@gmail.com, mcaussa@med.puc.cl (S. Caussade).

Introduction

Asthma is the most prevalent chronic disease among children and in the vast majority of cases it starts at preschool age.^{1,2} However, it continues to be a difficult disorder to diagnose in preschoolers. This is partly because clinical symptoms of asthma are variable and non-specific, given that other wheezing disorders exist.^{3,4} Diagnosis and management of asthma in recurrent wheezing preschoolers are still primarily based on subjective clinical features and findings from medical examinations (atopic manifestations, parental asthma, or response to controller therapy [e.g. inhaled corticosteroids]).^{3,4}

On the other hand, bronchial hyperreactivity (BHR) is a traditional hallmark of asthma. Its presence is a good predictor of severity, morbidity, and decline of lung function among asthmatic children.^{5,6} Due to its high sensitivity, methacholine challenge test (MCT) performed with spirometry is one of the most common tests for measuring BHR in schoolchildren and adolescents.⁷ Nevertheless, since preschoolers usually collaborate poorly in performing acceptable and reproducible serial spirometry manoeuvres, methacholine challenge tests with measurements that require little cooperation, e.g. impulse oscillometry, interrupter technique, and simpler clinical methods such as wheezing auscultation and pulse oximetry saturation [SpO₂], have been developed.^{8–12}

The aim of this study was to assess the usefulness and safety of a clinical method for measuring methacholine bronchial hyperresponsiveness through wheezing auscultation, decreased pulse arterial oxygen saturation [SpO₂] and respiratory rate [RR] in recurrently wheezing preschoolers with asthma and healthy controls. The second aim was to establish methacholine concentration at which a significant airway response occurs.

Methods

This study was carried out at the Pontificia Universidad Catolica de Chile, Santiago, Chile. We prospectively enrolled preschool children related to university employees, outpatients of the paediatric clinic, and children recruited from kindergartens in Santiago. The children were classified as asthmatics or healthy (control group). Asthmatics were included in the study if they had had three or more wheezing episodes in the last 12 months and if they had shown a clinical response to bronchodilator and to controller drugs (e.g. inhaled corticosteroids [ICS] or leukotriene inhibitors).¹³ They were classified by their severity, according to guidelines, as having intermittent, mild or moderate persistent asthma.¹⁴ None of the children had a personal history of prematurity, neonatal lung disease, pneumonia, lung resection, central airway obstructive disease or other cardiopulmonary chronic diseases.

The MCT was performed in our paediatric lung function laboratory during the summer of 2008–2010. Neither asthmatic nor healthy children had had any upper or lower respiratory symptoms for at least three weeks prior to the study, nor active rhinitis symptoms. Asthmatics had been free of controller drugs ICS and leukotriene inhibitors for at least one month prior to the study, anti-histamines for at

least one week, and short acting bronchodilators for at least 8 h. Children were seated with one parent in a non-stressful environment. The MCT was performed using the 2-min tidal breathing method developed by Cockcroft et al.^{12,15} with doubling doses of methacholine solutions from 0.06 to 8 mg/ml dissolved in saline. The children were previously nebulised with saline to establish control values. We used a nebuliser and a facemask (Pari Star[®], Midlothian, USA). Sixty seconds after the end of each nebulisation, two independent observers (SC and RB or RR) simultaneously determined the respiratory rate for 1 min, observed SpO₂ in the monitor, and auscultated the presence of wheezing (over the trachea, and upper front and lower back of the thorax), asking the child to breathe deeper than tidal volume. SpO₂ was monitored with a pulse oximeter (Masimo Rad 9[®], Masimo Corporation, Irvine, CA, USA). Nebulisations were carried out every 5 min until a maximum of 8 mg/ml of methacholine; nebulisers were calibrated following ATS recommendations,¹⁵ with an output of 0.13 ml/min \pm 10%.

The endpoint of MCT was set with the concentration of methacholine that determined one or more of the following events: presence of wheezing at auscultation (PC_w), and/or decrease of ≥ 5 from control SpO₂ for at least 10 s (PC_{SpO₂}) and/or increase in RR $\geq 50\%$ from control RR (PC_{RR}). If more than one event was present, i.e. presence of PC_w and PC_{SpO₂}, we called it PC_{w-SpO₂}. Once the test was completed, children were nebulised with 0.25 mg of ipratropium bromide + 0.5 mg fenoterol bromide (Berodual[®], Boehringer Ingelheim, Rhein, Germany) with oxygen flow of 6 l/min for 10 min. The MCT was stopped and considered a failure if the child was uncooperative (cried, hyperventilated, spoke during nebulisation or removed the facemask) or if adverse effects appeared (tearing, nasal symptoms, headache) or if parents requested the test to be stopped.

All parents signed informed consent forms to authorise the participation of their children in the study. The Ethics Committee of the Medical Research Center of the School of Medicine of the Pontificia Universidad Catolica de Chile approved the study (CE #0016/08).

Statistical analysis

To calculate the sample size, we used a previous study¹⁶ in which we found a median of PC_w among asthmatic preschoolers of 0.25 mg/ml (0.06–4 mg/ml) and 1 mg/ml (0.5–8 mg/ml) among healthy children. Therefore, to find significant differences between the two groups we needed at least 10 children in each group (with 80% power and 95% of significance level). We used the Student *t*-test for comparative analysis of the general characteristics of the groups. PC_w and PC_{SpO₂} were not normally distributed. Their results were expressed as median and interquartile range. We also obtained their logarithmic values (not shown). Mann–Whitney and/or Kruskal–Wallis test was used to compare the median of methacholine concentrations between the groups. A ROC curve analysis was performed to determine the cut-off points of wheezing auscultation and SpO₂ fall. We calculated the AUC (area under curve). Sensitivity, specificity, and positive (PPV) and negative predictive values (NPV) with their 95% CI for PC_w and PC_{SpO₂} were additionally calculated. Two-tailed *p* values of ≤ 0.05 were considered significant.

Table 1 Demographic and clinical characteristics of healthy and asthmatic children.

	Healthy <i>n</i> = 32	Asthmatic <i>n</i> = 33	<i>p</i>
Age (months)	54.7 ± 10.2	53.6 ± 9.5	0.88
Males (<i>n</i>)	12	20	0.35
Height (cm)	106.5 ± 6.7	106.6 ± 6.4	0.9
BMI	16.2 ± 0.99	17.1 ± 2.3	0.059
Basal SpO ₂ (%)	98.4 ± 1.1	98 ± 1.3	0.9
Basal respiratory rate (/mn)	22 ± 3.8	20 ± 3.2	0.11
Basal cardiac rate (/mn)	104 ± 9	96 ± 5	0.04
Smoker in the household (<i>n</i>)	19	14	0.17
Parent asthma (<i>n</i>)	1	11	<0.01
Allergic rhinitis (<i>n</i>)	0	21	<0.0001
Allergic dermatitis (<i>n</i>)	1	4	0.36

Numbers are expressed as *n* or mean ± SD.

SPSS v 15.0 statistical software package (IBM, Armonk, NY, USA) was used for the analysis.

Results

We recruited 35 healthy and 37 asthmatic preschoolers. Seven preschoolers (three healthy) were excluded due to lack of collaboration to perform the MCT. There were no significant differences between healthy and asthmatic children in terms of age, gender, body mass index (BMI) and tobacco consumption at home. Basal SpO₂ and RR were also similar between both groups, but cardiac rate was significantly higher in healthy children (Table 1). Asthmatic children had higher prevalence of dermatitis and rhinitis and parental asthma.

Among asthmatics, 17/33 (52%) had intermittent symptoms, 14/33 (42%) had mild persistent symptoms and 2/33 (6%) had moderate persistent symptoms. For analysis, these last two children were included in the "mild persistent group". Among asthmatics, the average onset age of wheezing was eight months and the average number of annual wheezing episodes was three.

The agreement between the two independent observers was 94% (34/36) for wheezing and 100% for SpO₂. However, the measurement of RR (performed for 1 min) was difficult and there was a high degree of discrepancy (42%) between the observers. Therefore RR was not considered for analysis. Considering the minimal doses to obtain one of the two clinical parameters (PC_w or PC_{SpO₂}) as positive, there was no difference between asthmatics with intermittent and persistent symptoms: 0.38 mg/ml (0.16–0.89 mg/ml) and 0.54 mg/ml (0.26–1.12 mg/ml) respectively for PC_w and 0.59 mg/ml (0.06–5.78 mg/ml) and 0.25 mg/ml (0.14–1.46 mg/ml) respectively for PC_{SpO₂}. Consequently, we joined the two sub-groups of asthmatics and compared it to the healthy control group.

The maximal methacholine dose for asthmatic and healthy children was 8 mg/ml and the lowest SpO₂ was 90%. There were two children in the asthmatic group and four in the healthy group who did not show any positive test (Table 2). PC_w did not show differences by age nor gender in both asthmatic and control group. The same observation was found for PC_{SpO₂}.

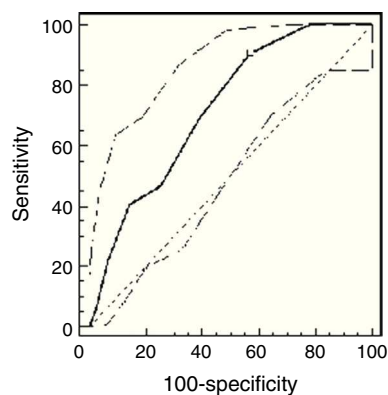
Table 2 Clinical signs at the endpoint of MCT in healthy and asthmatic children.^a

Clinical sign	Healthy <i>n</i> = 32	Asthmatics <i>n</i> = 33
Wheezing alone	14 (44%)	17 (52%)
Decreased SpO ₂ alone	12 (38%)	11 (33%)
Wheezing + decreased SpO ₂	2 (6%)	3 (9%)
Non-responders	4 (12%)	2 (6%)

^a There were no differences between the groups.

Methacholine dose for PC_w was significantly lower among the asthmatic than healthy children: 0.5 mg/ml (0.25–0.5 mg/ml) vs. 2 mg/ml (1–4 mg/ml), respectively, *p* < 0.001. Similarly, the methacholine dose for PC_{SpO₂} was lower among the asthmatics than the controls: 0.25 mg/ml (0.25–0.5 mg/ml) and 2 mg/ml (0.5–4 mg/ml), respectively, *p* < 0.001. Only four asthmatic children and two healthy children had a positive PC_w and PC_{SpO₂} test (Table 2), so we could not draw any valid conclusion combining both variables.

Using the ROC analysis, the best cut-off point of PC_w was observed at a methacholine concentration of 0.5 mg/ml (AUC = 0.72 [95% CI = 0.66–0.77]). Its sensitivity was 91% [95% CI = 75–98%], specificity 43% [37–49%], PPV 16% and NPV 98% (Fig. 1). For PC_{SpO₂} the best cut-off point was

**Figure 1** ROC curve PC_w: cut-off 0.5 mg/ml methacholine concentration. Dashed lines represent 95% confidence interval.

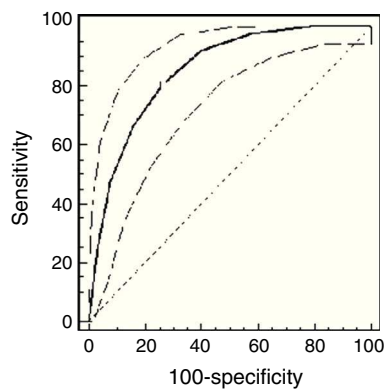


Figure 2 ROC curve PC_{SpO2}: cut-off 1 mg/ml methacholine concentration. Dashed lines represent 95% confidence interval.

at a methacholine concentration of 1 mg/ml (AUC = 0.85 [95% CI 0.81–0.89]), sensitivity 80% [70–88%], specificity 74% [68–79%], PPV 49%, and NPV 92% (Fig. 2).

In the asthmatic group, there were 21 children with allergic rhinitis and 12 without rhinitis. However, there were no differences in PC_w median between those with rhinitis and those without rhinitis: 0.25 mg/ml (0.25–0.5 mg/ml) and 0.5 mg/ml (0.5–2 mg/ml), respectively, $p = 0.37$.

There were no adverse reactions in any children.

Discussion

In the present study, we found that preschool children with intermittent and mild persistent asthma had a significantly lower methacholine dose for PC_w and PC_{SpO2} than healthy controls. The best cut-off point of PC_w was observed at a methacholine concentration of 0.5 mg/ml and of PC_{SpO2} at 1 mg/ml, both with good sensitivity and NPV. We report the term PC_w as an analogy to PC₂₀, as has been previously proposed.⁷ Besides, we added the PC_{SpO2} also as an analogy to PC₂₀. This study shows that measuring methacholine induced BHR using PC_w and PC_{SpO2} among preschool children with intermittent and mild persistent asthma is a useful, simple and low risk method.

Wheezes are produced by the fluttering of the airways walls and fluid together, and are induced by a critical air-flow velocity.¹⁷ It must be considered that the conditions for hearing wheezing may change from breath to breath depending on the intensity of inhalation or exhalation or the presence of airway secretions.¹⁸ Optimal evaluation of wheezing is now available through devices that record and analyse breathing sounds, thus improving sensitivity and objectivity.^{19,20} In this study we asked children to breathe deeper than tidal volume to ensure a better breath sound auscultation. To avoid having a biased observation, both observers auscultated simultaneously, and they agree in 94%. We evaluated children 1 min after nebulisation was complete. We did not hear breath sounds during nebulisation, as Godfrey et al. did.²¹ They assessed BHR to adenosine 5'-monophosphate in asthmatic preschoolers with acoustic analysis of both lungs and found that wheezing would start during nebulisation in 31% of children and that the site, timing and characteristics of the first wheeze showed high variability. Moreover, in a study in preschool children, Bentur

et al.²² found that obtaining the standardised FEV₁ (Forced Expiratory Volume in one second) fall of 20% by spirometry (PC_{20-FEV1}) frequently preceded wheezing auscultation. This suggested that PC_{20-FEV1} determinations must be safer than the method used in our study. However, we did not have any adverse reactions and no child showed a fall of SpO₂ below 90%.

SpO₂ measurement is an indirect sign of bronchoconstriction, reflecting hypoxaemia caused by ventilation/perfusion alteration due to smooth muscle contraction, vasodilatation or both.²³ We must consider that baseline SpO₂ values correspond to the upper part of the oxyhaemoglobin saturation curve. So small changes in SpO₂ could be associated with large drops in arterial oxygenation.²⁴ Some researchers had used SpO₂ < 90–91% or a 3–5 decrease as the end-point.^{8–10,22} A fall in SpO₂ has been reported to have a good correlation with transcutaneous oxygen pressure and respiratory resistance in MCT carried out in asthmatic infants and preschoolers.^{24,25} Although the measurement of SpO₂ is simpler and more widely available, we found that it could be used to assess bronchial hyperresponsiveness, and if wheezing is not present we could consider a concentration of 1 mg/ml for PC_{SpO2}. This parameter could be used for safety and BHR diagnosis in younger children.

Our study shows a lack of coincidence between decreased SpO₂ (PC_{SpO2}) and the presence of wheezing (PC_w); both parameters were positive only in four asthmatic and two healthy preschoolers. This issue was reported by different authors with opposing results. Yong et al.¹⁰ obtained wheezing in 78%, decreased SpO₂ in 10% and both variables in only 12% of recurrently wheezing <4-year-old children. In a group of 146 young asthmatics, Springer et al.⁷ found the presence of wheezing alone in 6.8%, decreased SpO₂ in 5.5% and both in 13.6%. Koh et al.¹¹ found wheezing alone in 37%, had decreased SpO₂ alone in 37%, and both parameters in 26% preschool asthmatic children. Kivastik et al.⁸ stopped the test due to wheezing in 27% of healthy, coughing and wheezer children, in 33% because of decreased SpO₂, and in 35% due to both variables. Our discordance could be explained by the effect of bronchoconstriction on breathing patterns, with minute ventilation and respiratory rate variations.²⁶

The differences in PC_w and PC_{SpO2} values between healthy and asthmatic preschool children in our study are remarkable, with a fourfold value in the last group. We found PC_w median of 2 mg/ml and 0.5 mg/ml and for PC_{SpO2} 2 mg/ml and 0.25 mg/ml in healthy and asthmatic children, respectively. Our results were expressed as median because they were not normally distributed. Other researchers reported their results as geometric mean. De Mir et al.²⁷ studied 16 healthy and 63 asthmatic children aged from six months to four years old; they found higher values than ours in both groups, considering wheezing and/or SpO₂ decrease as end-points: 13.3 ± 5.02 mg/ml in healthy children and 5.8 ± 3.9 mg/ml in asthmatic children. Kivastik⁸ found a geometric mean of PC_w of 2.88 mg/ml in nine healthy children and 1.28 mg/ml in 25 asthmatic children. In these studies they had a low number of healthy children, inhaled corticosteroids were permitted for use as usual in children with recurrent wheezing, and they used a different methodology to calculate PC of the total group. These facts could explain the differences from our results.

Regarding the severity of asthma, we did not find significant differences in PC_w or PC_{SpO_2} , similar to the findings of Wang et al.²⁵ In contrast, Avital et al.²⁸ reported a close relationship between the degree of clinical BHR and the severity of symptoms using the tracheal auscultation technique. Perhaps our results can be explained by the presence of a low asthma severity in our patients. It is also known that age correlates with an increase in methacholine-induced BHR in asthmatic children,²⁹ but in our study we only found a similar trend, without statistical differences.

This study has some limitations. First, we only recruited intermittent and mild persistent asthmatic children, so these results cannot be applied in more severe asthmatic categories. Second, since this study was not longitudinal, we cannot determine if the presence of BHR among preschool children with intermittent and mild persistent asthma could be a marker of illness persistence later in life.^{5,6} Although sensitivity and NPV have high values, specificity and PPV are limited especially for PC_w . Thus, BHR could be present in a child without asthma. Then MCT by this method could be useful to rule out asthma, like in older children and adults.¹⁵ Third, considering the prolonged time of the MCT, it was refused by seven children (9.7%); we should have considered a shorter test to increase its level of acceptance, as other researchers did.^{8,27} Fourth, the combination PC_w and PC_{SpO_2} often lacked coincidence, and this could be the best reliable parameter to evaluate clinical bronchial responsiveness to methacholine; we could not carry out analysis because of the fair number of cases in this situation. We know that FEV1 measurement is the best and safest parameter for detecting airway obstruction, nevertheless the aim of our study was to find alternative measures for patients who fail to perform spirometry. However, a strength in this study is the fact that a significant sample size was achieved and even exceeded. We used a safe method and followed ATS guidelines^{12,15} in performing the MCT (being critical for comparing results in clinical practice). Neither adverse reactions nor severe decreases in SpO2 were observed. Finally, we confirmed that checking SpO2 during MCT should be mandatory, given the fact that a decrease suggests bronchial obstruction in children without detectable wheezing.

Conclusion

Although wheeze auscultation and SpO2 cannot completely replace lung function tests to evaluate BHR, this clinical method confirms that BHR is present in almost all preschool patients with three or more annual wheezing episodes. It also provides additional information about the normal response to methacholine in healthy children. This could be a safe, useful and tolerable method to use in young children who are uncooperative with spirometry, or when laboratories lack the equipment necessary to evaluate methacholine induced bronchoconstriction in this age group.

Ethical disclosures

Protection of human subjects and animals in research. The authors declare that the procedures followed were in accordance with the regulations of the responsible Clinical

Research Ethics Committee and in accordance with those of the World Medical Association and the Helsinki Declaration.

Confidentiality of data. The authors declare that they have followed the protocols of their work centre on the publication of patient data and that all the patients included in the study have received sufficient information and have given their informed consent in writing to participate in this study.

Right to privacy and informed consent. Right to privacy and informed consent. The authors have obtained the informed consent of the patients and/or subjects mentioned in the article. The author for correspondence is in possession of this document.

Conflict of interest

The authors have no conflict of interest to declare.

Acknowledgments

The authors thank the children, their parents and the kindergarten teachers.

References

1. Yunginger JW, Reed CE, O'Connell EJ, et al. A community-based study of the epidemiology of asthma. Incidence rates, 1964–1983. *Am Rev Respir Dis.* 1992;146:888–94.
2. Asher MI, Montefort S, Bjorksten B, Lai CKW, Strachan DP, Weiland SK, et al., the ISAAC Phase Three Study Group. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC phases one and three repeat multicountry cross-sectional surveys. *Lancet.* 2006;368:733–43.
3. Bacharier LB, Guilbert TW. Diagnosis and management of early asthma in preschool-aged children. *J Allergy Clin Immunol.* 2012;130:287–96.
4. Bush A. Diagnosis of asthma in children under five. *Prim Care Respir J.* 2007;16:7–15.
5. Weiss ST, Van Natta ML, Zeiger RS. Relationship between increased airway responsiveness and asthma severity in the childhood asthma management program. *Am J Respir Crit Care Med.* 2000;162:50–6.
6. Sears MR, Greene JM, Willan AR, Wiecek EM, Taylor DR, Flannery EM, et al. A longitudinal, population-based, cohort study of childhood asthma followed to adulthood. *N Engl J Med.* 2003;349:1414–22.
7. Cockcroft D. Direct challenge tests. Airway hyperresponsiveness in asthma: its measurement and clinical significance. *Chest.* 2010;138 2 Suppl.:185–245.
8. Kivastik J, Gibson AM, Primhak RA. Methacholine challenge in pre-school children. Which outcome measure? *Respir Med.* 2007;101:2555–60.
9. Springer C, Godfrey S, Picard E, Uwyyed K, Totshild M, Hananya S, et al. Efficacy and safety of methacholine bronchial challenge performed by auscultation in young asthmatic children. *Am J Respir Crit Care Med.* 2000;162:857–60.
10. Yong SC, Smith CM, Wach R, Kurian M, Primhak RA. Methacholine challenge in preschool children: methacholine-induced wheeze versus transcutaneous oximetry. *Eur Respir J.* 1999;14:1175–8.
11. Koh YY, Kang H, Yoo Y, Kim DK, Yu J, Kim CK. Wheeze detection as a measure of bronchial challenge in young children with

- cough-variant asthma and with classic asthma. *Acta Paediatr.* 2007;96:1223–7.
12. An official American Thoracic Society/European Respiratory Society Statement: Pulmonary function testing in preschool children. *Am J Respir Crit Care Med.* 2007;175:1304–45.
 13. Castro-Rodriguez JA, Rodriguez-Martinez CE, Custovic A. Infantile and preschool asthma. *Eur Respir Monogr.* 2012;56:10–21.
 14. Global Initiative for Asthma. Global strategy for asthma management and prevention updated; 2011. http://www.ginasthma.org/uploads/users/files/GINA_Report2011_May4.pdf
 15. Guidelines for Methacholine and Exercise Challenge Testing—1999. The Official Statement of the American Thoracic Society. *Am J Respir Crit Care Med.* 2000;161:309–29.
 16. Paz FE, Vega LE, Sanchez I, Bertrand PJ, Caussade S. PC wheezing in preschool children with asthma. In: *ATS 2008 Toronto international conference.* 2008. p. A708 [Poster Board K57].
 17. Meslier N, Charbonneau G, Racineux JL. Wheezes *Eur Respir J.* 1995;8:1942–8.
 18. Pasterkamp H. Acoustic markers of airway responses during inhalation challenge in children. *Pediatr Pulmonol.* 2004;26 Suppl.:175–6.
 19. Godfrey S, Uwytyed K, Springer C, Avital A. Is clinical wheezing reliable as the endpoint for bronchial challenges in preschool children. *Pediatr Pulmonol.* 2004;37:193–200.
 20. Sanchez I, Alvarez C, Claveria C, Lisboa C. Acoustic analysis of respiratory sounds during methacholine challenge in preschool children. *Rev Med Chile.* 2001;129:1271–8.
 21. Godfrey S, Cohen L, Avital A, Springer C. Timing and nature of wheezing at the endpoint of a bronchial challenge in preschool children. *Pediatr Pulmonol.* 2005;39:262–7.
 22. Bentur L, Beck R, Elias N, Barak A, Efrati O, Yahav Y, et al. Methacholine bronchial provocation measured by spirometry versus wheeze detection in preschool children. *BMC Pediatr.* 2005;5:19–25.
 23. Cockcroft D, Hurst T, Marciniuk D, Cotton D, Laframboise K, Nagpal A, et al. Routine pulse oximetry during methacholine challenges is unnecessary for safety. *Chest.* 2000;118:1378–2138.
 24. Prendiville A, Maxwell DL, Rose A, Silverman M. Histamine-induced airway obstruction in infancy: changes in oxygenation. *Pediatr Pulmonol.* 1988;4:164–8.
 25. Wang J, Mochizuki H, Muramatsu R, Arakawa H, Tokuyama K, Morikawa A. Evaluation of bronchial hyperresponsiveness by monitoring of transcutaneous oxygen tension and arterial oxygen saturation during methacholine challenge in asthmatic children. *J Asthma.* 2006;43:145–9.
 26. Steward IC, Parker A, Catterall JR, Douglas NJ, Flenley DC. Effect on bronchial challenge on breathing patterns and arterial oxygenation in stable asthma. *Chest.* 1989;95:65–70.
 27. De Mir Messa I, Moreno Galdó A, Cobos Barroso N, Gartner S, Martin de Vicente C, Rovira Amigo S, et al. Bronchial hyperresponsiveness to methacholine in children under 4 years with recurrent bronchitis. *Arch Bronconeumol.* 2010;46:621–7.
 28. Avital A, Bar-Yishay E, Springer C, Godfrey S. Bronchial provocation tests in young children using tracheal auscultation. *Am Respir Dis.* 1991;144:36–829.
 29. Le Souef P, Sears M, Sherrill D. The effect of size and age of subject on airway responsiveness in children. *Am J Respir Crit Care Med.* 1995;152:576–9.