



# Allergologia et immunopathologia

Sociedad Española de Inmunología Clínica,  
Alergología y Asma Pediátrica

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## REVIEW

# Change in gut microbiota for eczema: Implications for novel therapeutic strategies



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Received 7 April 2017; accepted 19 May 2017

Available online 24 December 2017

## KEYWORDS

Gut microbiota;  
Inflammation;  
Eczema;  
Probiotics treatment

**Abstract** Eczema is one of the most common inflammatory diseases, often constituting a life-long burden for afflicted individuals. The complex interaction of host genetic and multiple environmental factors contribute to its pathogenesis. A relationship between maladjustment of gut microbiota and eczema has been brought into the light of day in most previous studies. In eczema preclinical models, specific intestinal microbial species have been demonstrated to prohibit or dwindle immune responsiveness, indicating that these strains among commensal gut bacteria may exert either a moribific or phylactic function in eczema progression. As such, oral probiotics can serve as a medicinal approach for eczema therapy. Given that relative scientific work is still at the early stage, only limited data are available in the field. New sequencing techniques have been fortunately performed to gain access to an extended research on the relationship between gut bacterial flora and human diseases. In the current review, we identified the role of intestinal microbiota in the development of eczema and how specific bacterial strains adjust the immune responsiveness in the midst of disease progression. Probiotics as an applicable treatment for eczema were evaluated in other threads as well.

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## Introduction

Eczema, also known as atopic dermatitis, is a common childhood condition characterised by the inflammation of skin with intense itching.<sup>1</sup> The prevalence of eczema has been increasing worldwide during the past decades, par-

ticularly in industrialised nations and amongst children.<sup>2–4</sup> Even though research studies on genetic predisposition to eczema have implicated genes (e.g. *IL4*, *IL4R*, *IL13*, *CMA1*, *SPINK5*, *FLG*, *IL-6*),<sup>5–12</sup> the aetiology of systemic inflammation still requires further investigation. The gut is a crucial immune organ besides its function in metabolism and in the body it includes the biggest lymphoid tissue mass.<sup>13</sup> The gut is a habitat to vast and various species of microbes.<sup>13,14</sup> Crucial signals derived from the gut microbiota contribute to the development of host immunity. Hence, Gut microbes

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are very important to maintain human health and disease. Recent studies in experimental subjects have demonstrated that maladjustment of the intestinal microbiota is related to the development of eczema.<sup>15,16</sup> Changes in terms of the abundance of certain intestinal microbial species have been revealed to prohibit or dwindle immune responsiveness in eczema experimental subjects, they may be biomarkers in eczema prevention and therapy. In spite of being a relatively novel area of research, evidence hitherto indicates that the intestinal microbiota may serve as fecund targets for prevention or control of eczema which is primarily characterised by innate and adaptive immune dysbiosis. In the current study, we reviewed recent series to further investigate changes in microbiota composition which trigger eczema disease, and the efficacy of probiotics/prebiotic in the therapy for eczema is also assessed based on the previous studies.

## Eczema pathogenesis and the role of gut microbiota within it

### Eczema pathogenesis

Eczema is a chronic form of childhood disorder that is characterised by remitting and relapsing cutaneous symptoms. These symptoms include itching and dryness, flaking, blistering, oozing, and bleeding.<sup>17</sup> Eczema is often the first manifestation of atopy in infants who will develop asthma or allergic rhinitis later in childhood.<sup>18,19</sup> Different dendritic cells subtypes, such as Langerhans cells (LC), inflammatory dendritic epidermal cells (IDEC) and plasmacytoid dendritic cells (PDC), play a key role in eczema and impact on the mechanisms underlying eczema, such as the recruitment of inflammatory cells, T-cell priming, and cytokine and chemokine release. Lesional skin of eczema patients harbours significant numbers of LC; IDEC and PDC expressing the high-affinity receptor for immunoglobulin E (IgE).<sup>20–22</sup> An enhanced T helper 2 (Th2) immune response, reflected by an increased frequency of allergen-specific T cells producing interleukin (IL)-4, -5 and -13, and a decrease in interferon (IFN)- $\gamma$ -producing T-cells.<sup>23,24</sup> IL-4 can be involved in IgE isotype switching, while IL-5 can attract eosinophils and prolong their survival, which may result in the peripheral blood eosinophilia and increased IgE serum levels in eczema patients.<sup>25</sup> Moreover, there is a preferential apoptosis of circulating Th1 cells in eczema, which may also contribute to Th2 predominance in eczema patients.<sup>26</sup> Interestingly, eczema patients have significantly increased numbers of circulating regulatory T cells that exhibit normal immunosuppressive activities in vitro.<sup>27</sup> Studies on genetic predisposition to eczema have implicated eczema-related genes such as (e.g. *IL4*, *IL4R*, *IL13*, *CMA1*, *SPINK5*, *FLG*, *IL-6*),<sup>5–12</sup> environmental factors have also been shown to contribute to the disease pathogenesis. There are also other possible mechanisms that aberrant barrier functions in gut mucosa lead to greater antigen transfer across the mucosal barrier and the routes of transport are altered, thereby evoking aberrant immune responses and release of pro-inflammatory cytokines with further impairment of the barrier functions.<sup>28</sup> Such increased inflammation would lead to further increases in intestinal permeability and in

a vicious circle of increasing allergenic responses, and a more permanent dysregulation of the immune responses to ubiquitous antigens in genetically susceptible individuals.<sup>29</sup> The alteration of gut microbiota has an important impact on the peripheral and central immune system. Many unclear mechanisms still exist.

### The role of gut microbiota in eczema

Recently, it has been indicated that microbial triggers have been implicated in eczema.<sup>30</sup> The vast majority of these studies suggested that subjects with eczema exhibit alterations in the relative abundance of “beneficial” and potentially “harmful” bacteria compared to healthy subjects (Table 1). There is convincing evidence from experimental subjects suggesting that kind of factors (e.g., diet, level of physical activity, during pregnancy, and with the use of broad-spectrum antibiotics) are related to eczema disease via affecting gut microbiota. The existence of a link between eczema and the gut microbiota was indicated based on studies in patients with eczema. Infants with eczema harboured significantly lower gut microbial diversity when compared to healthy controls, suggesting that alteration of the entire intestinal microbiota and the lack of exposure to certain bacterial targets may be contributing factors that amounted to their diseased state.<sup>31,32</sup> In keeping with the observation, by comparing with healthy subjects, gut microbial diversity significantly decreased in infants with IgE-associated eczema, the diversity of the bacterial phylum *Bacteroidetes* and phylum *Proteobacteria* also significantly reduced, the level of the phylum *Proteobacteria* significantly decreased.<sup>33</sup> *Proteobacteria* comprises gram-negative bacteria, typically with endotoxin lipopolysaccharides (LPS) incorporated into the cell wall. Endotoxin can induce a TH1 response through the innate immune system by enhancing IL-12 production from monocytes and dendritic cells,<sup>34</sup> and low exposure to endotoxin has been associated with an increased risk of atopic eczema.<sup>35</sup> In addition, a strong endotoxin exposure might down regulate atopy-promoting Th2 responses.<sup>33</sup> Similarly, Nylund et al.,<sup>36</sup> also found that infants with eczema appeared to have a significant decrease in the abundance of *Bacteroidetes* compared to healthy infants. Therefore, these increasing studies suggest that changes in the composition of gut microbiota play a significant role in induction and furthering the progression of eczema. The abundance of *Ruminococcaceae* was significantly lower at one week of age in infants with IgE-associated eczema than controls.<sup>37</sup> Meanwhile, the abundance of *Ruminococcus* was significantly negatively associated with TLR2-induced IL-6 and TNF- $\alpha$ . The abundance of the phylum *Proteobacteria* and the family *Enterobacteriaceae* significantly decreased in infants with IgE-associated eczema compared to controls. The abundance of *Proteobacteria* was significantly inversely related with TLR4-induced TNF- $\alpha$ . The abundance of *Enterobacteriaceae* was significantly negatively associated with TLR4-induced TNF- $\alpha$  and IL-6. At one year,  $\alpha$ -diversity of *Actinobacteria* was significantly lower in infants with IgE-associated eczema compared with controls. *Ruminococcaceae* belonging to *Firmicute* have been associated with the maintenance of gut health. There is emerging interest in the role of

**Table 1** Changes in microbiota composition associated with eczema and potential therapeutic strategies.

Models	Disease	Implicated microbiota	New therapeutic strategies	Implicated microbiota	Reference
Infants	Eczema	<i>Proteobacteria</i> ↓	NO	NO	33,37
Infants	Eczema	<i>Bacteroidetes</i> ↓	NO	NO	36
Infants	Eczema	<i>Ruminococcaceae</i> ↓, <i>Enterobacteriaceae</i> ↓	NO	NO	37
Infants	Eczema	<i>Bifidobacterium</i> ↓, <i>Staphylococcus</i> ↑	NO	NO	43
Infants	Eczema	<i>Bifidobacterium</i> ↓, <i>Clostridium</i> ↓, lactic-acid-producing bacteria (LAB)↑, <i>Enterococci</i> ↑	NO	NO	45
Infants	Eczema	<i>Clostridium</i> clusters IV and XIVa↑	NO	NO	36
Infants	Eczema	<i>Lactobacilli/Enterococci</i> ↑	NO	NO	48
Infants	Eczema	<i>Enterococcus</i> ↑, <i>Klebsiella</i> ↑, <i>Shigella</i> ↑, <i>Bifidobacterium</i> ↓	NO	NO	17
Infants	Eczema	<i>Campylobacter</i> ↑, <i>Roseburia</i> ↓	NO	NO	53
Infants	Eczema	<i>Escherichia coli</i> ↑	NO	NO	56,57
Infants	Eczema	<i>Clostridium difficile</i> ↑	NO	NO	57,58
Infants	Eczema	<i>Bifidobacterium</i> <i>catenulatum</i> ↑, <i>B. breve</i> ↓	NO	NO	67
Infants	Eczema	<i>Staphylococcus aureus</i> ↓	NO	NO	73
Infants	Eczema	NO	Probiotic ( <i>Lactobacillus</i> F19)	NO	79
Infants	Eczema	NO	Probiotic ( <i>L. rhamnosus</i> HN001)	NO	80,81,82
Infants	Eczema	NO	Probiotic ( <i>L. fermentum</i> VRI-033PCC)	NO	84
Infants	Eczema	NO	Probiotic ( <i>L. reuteri</i> )	NO	85
Infants	Eczema	NO	Probiotic ( <i>L. rhamnosus</i> GG)	NO	49,50,51,90,91,92,95
Infants	Eczema	NO	Probiotic ( <i>B. lactis</i> Bb-12)	NO	50
Infants	Eczema	NO	Probiotic ( <i>B. bifidum</i> )	NO	98
Infants	Eczema	NO	Mixed probiotics ( <i>L.</i> <i>rhamnosus</i> LPR and <i>B. longum</i> BL999, or <i>L. paracasei</i> ST11 and <i>B. longum</i> BL999)	NO	100
Infants	Eczema	NO	Mixed probiotics ( <i>B. bifidum</i> , <i>B. lactis</i> , and <i>L. lactis</i> )	NO	101
Infants	Eczema	NO	Mixed probiotics ( <i>L.</i> <i>rhamnosus</i> GG, <i>L. rhamnosus</i> LC705, <i>B. breve</i> Bb99, and <i>Propionibacterium</i> <i>freudenreichii</i> ssp. <i>shermanii</i> JS)	NO	102

No refers to no test or no research.

*Ruminococcus* colonisation in infancy.<sup>38</sup> A possible health benefit is the production of ruminococcins, such as ruminococcin A, which is a bacteriocin that can inhibit the development of *Clostridium* species.<sup>39</sup> Thus, reduced abundance of potentially immunomodulatory gut bacteria is associated with exaggerated inflammatory cytokine responses to TLR-ligands and subsequent development of IgE-associated eczema. Abrahamsson et al.<sup>33</sup> reported the diversity of the bacterial genus *Bacteroides* to be significantly reduced. *Bacteroides* species have also been demonstrated to have anti-inflammatory properties. *Bacteroides* can break down complex plant polysaccharides<sup>40</sup>

and their abundance has been associated with increased short-chain fatty acid concentrations in the infant gut after introduction of the first solid foods.<sup>41</sup> Furthermore, *B. fragilis* polysaccharide has been shown in a mice model to direct the cellular and physical maturation of the developing immune system via its ability to direct the development of CD4+ T cells, thus inducing the differentiation of Th1 lineage and correction of the Th1/Th2 imbalance.<sup>42</sup> All in all, the *Bacteroides* have significance in the development and maintenance of gut and balanced mucosal immunity. In accordance with the observation, Watanabe et al.,<sup>43</sup> demonstrated eczematous subjects had significantly

lower counts of *Bifidobacterium* than healthy subjects, and the frequency of occurrence of *Staphylococcus* was significantly higher in eczematous subjects than in healthy subjects. *Bifidobacterium* can stimulate the production of Th1-type cytokines and leads to Th1-dominant immunity.<sup>44</sup> Therefore, the amount of *Bifidobacterium* in the intestine might be related to the onset of eczema in the host. Mah et al.,<sup>45</sup> revealed toddlers suffering from eczema harboured significantly lower abundance of *Bifidobacterium* and *Clostridium*, but significantly higher counts of total lactic-acid-producing bacteria (LAB) and *enterococci* compared to controls. Similarly, another study shown children with eczema had increased abundance of the *Clostridium* clusters IV and XIVa,<sup>36</sup> which are typically abundant in adults. Thus, children with eczema prematurely changed towards gut microbiota of adult-type. Gut microbiota of infant-type may enhance the normal mucosal barrier function by affecting the maturation of the gut epithelium and immune functions and reducing intestinal inflammation.<sup>46,47</sup> Kirjavainen et al.<sup>48</sup> testified infants with eczema in the highly sensitised group (HSG) had significantly greater numbers of *lactobacilli/enterococci* than those in the sensitised group (SG). *Lactobacilli* are probiotic bacteria, which can manipulate immune function, adhere to epithelial cells of the intestinal mucosa and colonise on the surface of gastrointestinal mucosa to form a bacterial membrane barrier to protect epithelial cells of the intestinal mucosa against injury from a variety of pathogenic microorganisms. Meanwhile, a number of other studies have also found that *lactobacilli* are associated with beneficial effects in the management of atopic eczema.<sup>49–51</sup> A possible explanation is that allergic sensitisation promotes the growth of *lactobacilli* and/or *enterococci* by causing changes in the gut ecology. In addition, Some species of *enterococci* have virulence factors that can compromise the gut barrier and in theory could thereby affect atopic sensitisation.<sup>52</sup> Hong et al.,<sup>17</sup> found that *Bifidobacterium* was present at significantly higher abundance in non-eczema infants compared to those with eczema, while *Enterococcus*, *Klebsiella* and *Shigella* were present at significantly higher abundances in eczema infants during early stages of infancy. *Bifidobacterium* was less diverse in eczema infants than the non-eczema group. For example, at one month of age, the *B. angulatum*, the *B. adolescentis*, *B. dentium*, the *B. catenulatum* group, the *B. bifidum* group and the *B. longum* group were only detected in non-eczema infants. At three months of age, the *B. dentium* and the *B. bifidum* group were only detected in non-eczema infants. At 12 months of age, the relative abundances of the *B. longum* group was significantly higher in non-eczema infants than eczema infants. Recently, one study found *Campylobacter* was significantly more abundant in infants with eczema and non-allergic controls, while *Roseburia* was significantly less abundant in participants with eczema than in controls.<sup>53</sup> *Campylobacter* can disrupt the intestinal epithelial barrier, which allowed the translocation of non-invasive bacteria such as *Escherichia coli*<sup>54</sup> and primed the intestine for inflammatory responses in susceptible infants. *Roseburia* is a butyrate-producing bacterium inhibiting histone deacetylase activity, leading to hyperacetylation of histones and thus suppression of nuclear factor-kappa B activation, and reinforcing the colonic defence barrier by enhancing the production of

mucins and antimicrobial peptides, as well as the expression of tight junction proteins.<sup>55</sup> Disorder of the intestinal microflora might play a role in the onset of eczema and the aggravation of eczema.

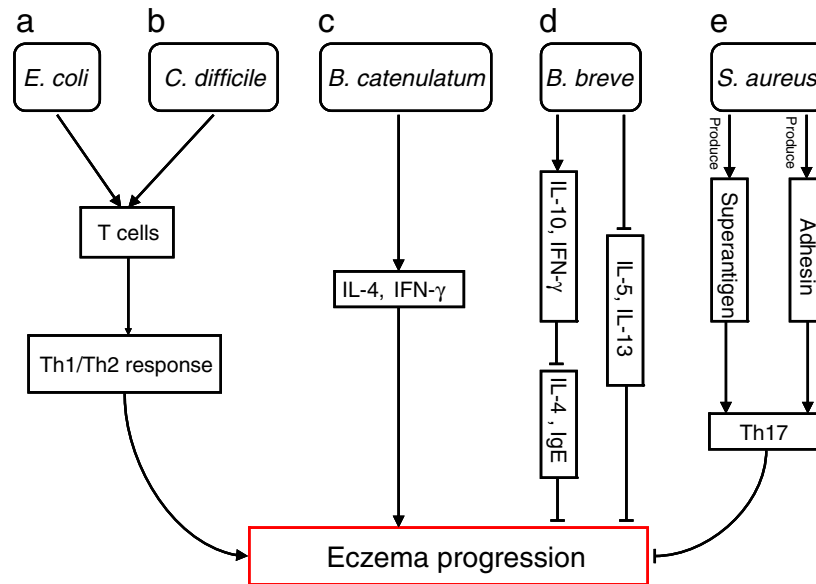
### Possible bacterial species involved

The differences in the gut microbiota composition precede the manifestation of eczema. Numerous studies have extensively investigated how specific species are involved in progressions of eczema (Fig. 1)

*Escherichia coli* was significantly higher in infants who were going to develop atopic eczema compared with healthy controls.<sup>56</sup> Previously, one study reported the number of *E. coli* to be positively correlated with total serum IgE levels in infants with eczema, indicating that the presence of *E. coli* is associated with the extent of atopic sensitisation.<sup>48</sup> *E coli* (for example, due to LPS) may evoke an inflammatory response in the gut leading to increased allergen uptake and thereby greater atopic sensitisation in infants with atopic eczema.

*E. coli*<sup>57</sup> and *Clostridium difficile*<sup>57,58</sup> were significantly positive associated with a risk of developing eczema. The presence of *E. coli* and *C. difficile* could induce a decrease of other beneficial bacteria, which could result in reduced induction of Treg cells by these beneficial bacteria leading to immune dysregulation. In the absence of optimal levels of immune regulation, an individual may develop a Th1 (such as Crohn's disease or autoimmunity) or Th2 (such as atopic diseases) mediated inflammatory disorder. Secondly, *E. coli* and *C. difficile* could have a direct effect on the production of cytokines by antigen-presenting cells, thereby affecting the differentiation of T cells.<sup>59</sup> Another hypothesis is that *E. coli* and/or *C. difficile* increase the intestinal permeability (for instance by the production of toxins). This increased permeability of the intestinal barrier could facilitate the penetration of innocuous antigens and subsequent sensitisation.<sup>60</sup> Indeed it has been shown that *C. difficile* toxins A and B compromise the intestinal cell barrier.<sup>61,62</sup> Furthermore, increased intestinal permeability has been described in patients with food allergies, eczema and asthma compared with healthy subjects.<sup>63–66</sup>

*B. catenulatum* and *B. breve* colonisation can influence the development of eczema in infants.<sup>67</sup> Namely, *B. catenulatum* was associated with a higher risk of developing eczema. However, *B. breve* was associated with a lower risk of eczema. The mechanisms through which these *bifidobacteria* exert their effects are unknown. The immune effects of exposure to microorganisms are both species and strain dependent, for example the capacity to induce FoxP3 Treg cells<sup>68</sup> and stimulate the production of different pro- and anti-inflammatory cytokines.<sup>68,69</sup> *B. catenulatum* is reported to be a strong inducer of IL-4 and IFN- $\gamma$  production.<sup>70</sup> *B. breve* induces the production of regulatory and Th1 cytokines. Inoue et al.,<sup>71</sup> found *B. breve* M-16V treatment could suppress the production of Th2 cytokine (IL-4) and IgE by inducing the secretion of regulatory (IL-10) and Th1 (IFN- $\gamma$ ) cytokines in a murine model of allergic inflammation. Similarly, synbiotic (*B. breve* M-16V and prebiotics) supplement could reduce allergen-induced Th2 (IL-4, IL-5 and IL-13)



**Figure 1** Bacterial species involved in eczema progression. a: *E. coli* may mediate a Th1/Th2 response by affecting T cells; b: *C. difficile* may mediate a Th1/Th2 response by affecting T cells; c: *B. catenulatum* may induce IL-4 and IFN- $\gamma$  production; d: *B. breve* may suppress the production of Th2 cytokine (IL-4) and IgE by inducing the secretion of regulatory (IL-10) and Th1 (IFN- $\gamma$ ) cytokines, and reduce allergen-induced Th2 (IL-5 and IL-13) responses; e: *S. aureus* may elicit a Th17 response. Abbreviations: Th1: T helper 1; Th2: T helper 2; IL: Interleukin; IFN- $\gamma$ : Interferon- $\gamma$ ; IgE: immunoglobulin E; Th17: T helper 17.

responses<sup>72</sup> in asthmatic adults. Taken together, the development of infant eczema can be influenced by modulation of specific *bifidobacteria* patterns in early life, depending on their immunomodulatory properties.

*Staphylococcus aureus* can produce a variety of T-cell-activating enterotoxins, called superantigens.<sup>73</sup> Gut colonisation by *S. aureus* strains carrying a certain combination of superantigen and adhesin genes was negatively associated with subsequent development of atopic eczema.<sup>73</sup> Such strains may provide stimulation and promote maturation of the infantile immune system. Thus, *S. aureus* could have a protective effect through the broad immune stimulation afforded by this bacterium. Mucosal colonisation by *S. aureus* might elicit a Th17 response<sup>74</sup> strengthening the skin and mucosal barriers. This may, in turn, reduce the risk of sensitisation and entering into the atopic march. To speculate, exposure of the newborn infant to *S. aureus* superantigens with limited pathogenic potential, such as SELM, might provide a method to prevent the development of an atopic phenotype.<sup>73</sup>

### Probiotics/prebiotics therapy

Probiotics are widely used in medical application to prevent or treat many diseases, such as diarrhoea, obesity, in particular immune disorders like eczema, allergy and eczema. A great number of studies have testified that modulating gut microbiota may be an effective strategy to cure and maintain eczema. The therapeutic effects of probiotics on eczema have also been confirmed in experimental subjects.

*Lactobacilli*, not only can enhance release of anti-inflammatory factors,<sup>75-77</sup> but also appears to protect against the invasion of pathogenic bacteria.<sup>78</sup> In a

double-blind, placebo-controlled randomised intervention trial, infants with a high risk for eczema were fed cereals with or without *Lactobacillus* F19.<sup>79</sup> The risk of eczema significantly decreased in the probiotic groups compared placebo groups. Meanwhile, the interferon- $\gamma$  (IFN- $\gamma$ )/interleukin 4 (IL4) mRNA ratio (the Th1/Th2 ratio) was significantly higher in the probiotic than the placebo group. These results suggested that *Lactobacillus* F19 may have effect on the T cell-mediated immune response. Another study randomly assigned women to take *L. rhamnosus* HN001, or placebo daily at gestation week 35 until six months of breastfeeding, and their infants were with high risk for eczema randomly assigned to receive the same treatment from birth to two years.<sup>80,81</sup> Infants receiving *L. rhamnosus* had a significantly reduced risk of eczema. Marlow et al.,<sup>82</sup> found that HN001 interacted with Toll-like receptor (TLR) that resulted in a significantly reduced risk of eczema. The main role of TLRs is as pattern recognition receptors, such that TLRs are expressed on T cells and involved in maintaining the balance between Th1 and Th2 immune responses. In addition, Gill et al.<sup>83</sup> found mice fed HN001 had significantly higher IFN- $\gamma$  levels than controls. HN001 may have a protective effect of HN001 against eczema by influencing cytokine production. More recently, children aged 6-18 months with moderate or severe eczema treated with *L. fermentum* VRI-033PCC,<sup>84</sup> showed a significant reduction in eczema scores compared to placebo. Similarly, the mothers received *L. reuteri* ATCC 55730 daily from gestational week 36 until delivery. Their babies then continued with the same product from birth until 12 months of age and were followed up for another year.<sup>85</sup> The *L. reuteri* group had significantly less IgE-associated eczema. However, skin prick test reactivity was also significantly less common in the treated than in the placebo group. *L. reuteri* prevents

TNF- $\alpha$ -induced IL-8 expression in murine epithelial cells,<sup>86</sup> diminishes inflammatory bowel disease in murine models,<sup>87</sup> and induces human IL-10 producing regulatory T cells by modulating dendritic cell function in vitro.<sup>88</sup> Another mode of action of *L. reuteri* could be an indirect effect through an influence on the composition of the intestinal microbiota, as *L. reuteri* strains produce the antimicrobial metabolite reuterin and inhibit pathogenic bacteria, without inhibiting normal bacterial residents of the gastrointestinal tract in vitro.<sup>89</sup> Keeping with the observation, infants with atopic eczema were fed an extensively hydrolysed whey formula with or without *Lactobacillus* GG.<sup>49</sup> *Lactobacillus* GG supplement show a significant improvement of atopic eczema after one month's intervention concomitant with a significant reduction in the concentrations of concentration of faecal  $\alpha_1$ -antitrypsin tumour necrosis factor- $\alpha$  (TNF- $\alpha$ ). Another study showed that probiotics *L. rhamnosus* GG administered pre- and postnatally for six months to children at high risk of atopic disease reduced the risk of developing atopic eczema later in infancy and childhood compared with that in infants receiving placebo.<sup>51,90,91</sup> Similarly, *L. rhamnosus* GG was given prenatally to mothers who had at least one first-degree relative with atopic eczema for four weeks before expected delivery and to their children, postnatally, for three months.<sup>92</sup> *L. rhamnosus* GG significantly reduced the risk of developing atopic eczema. Probiotic administration to the pregnant and lactating mother increased the amount of anti-inflammatory cytokine transforming growth factor- $\beta$ 2 (TGF- $\beta$ 2) in the mother's milk, which was suggested to increase its immune-protective potential and to be associated with a reduction in the risk of atopic eczema during the first two years of life. Transforming growth factor  $\beta$ 2 (TGF- $\beta$ 2) is considered a key immunoregulatory factor in promoting IgA production and induction of oral tolerance.<sup>93,94</sup> Viljanen et al.<sup>95</sup> found that in infants with IgE-associated atopic eczema-dermatitis syndrome (AEDS), treatment with LGG induced significantly higher C-reactive protein levels than in the placebo group, concomitantly, IL-6 levels significantly increased after treatment with LGG. Isolauri et al. also reported that oral administration of *Lactobacillus* GG or *Bifidobacterium lactis* Bb-12<sup>50</sup> in infants manifested atopic eczema significantly reduce the extent, severity and subjective symptoms of atopic eczema, in parallel with a significant reduction in the concentration of soluble CD4 (sCD4) in serum and eosinophilic protein X (EPX) in urine. Reduction of soluble CD4 is a marker of T-cell activation. Soluble CD4 has been found to be elevated in several diseases associated with chronic inflammation,<sup>96</sup> while urinary EPX has been shown to reflect the activity of allergic inflammation in childhood asthma.<sup>97</sup>

The possible influences of probiotic *Bifidobacterium bifidum* on infants with eczema was evaluated. *B. bifidum* supplementation significantly reduces the Scoring Atopic Dermatitis (SCORAD) index of infants with eczema as compared with prior to treatment and the controls.<sup>98</sup> Further study found that *B. bifidum* supplementation significantly enhances the levels of *B. bifidum* in the intestine. *B. bifidum* colonise in the intestinal tract and resist exogenous pathogens, so as to enhance the immune status of the body. *B. bifidum* also have a number of other roles, including improving the barrier function of the intestinal immune

system, regulating the immune response and reducing the production of inflammatory cytokines and the inflammatory response.<sup>99</sup>

A mixture of probiotics has also been used for the treatment of eczema. Rautava et al.,<sup>100</sup> completed a study in which mothers with allergic disease and atopic sensitisation were randomly assigned to receive (1) *Lactobacillus rhamnosus* LPR and *Bifidobacterium longum* BL999 (LPR+BL999), (2) *L. paracasei* ST11 and *B. longum* BL999 (ST11+BL999), or (3) placebo, beginning two months before delivery and during the first two months of breast-feeding. The infants were followed until the age of 24 months. Probiotic supplementation significantly reduced the risk of developing eczema. Keep with the observation, a mixture of probiotics (*Bifidobacterium bifidum*, *Bifidobacterium lactis*, and *Lactococcus lactis*) was prenatally administered to mothers of high-risk children (i.e. positive family history of allergic disease) and to their offspring for the first 12 months of life.<sup>101</sup> Parental-reported eczema during the first three months of life was significantly lower in the intervention group compared with placebo. In addition, the intervention group was significantly more frequently colonised with higher numbers of *Lactococcus lactis*. Furthermore, at age three months, in vitro production of IL-5 was significantly decreased in the probiotic-group compared with the placebo-group. Similarly, pregnant women carrying high risk children to use the probiotic mixture (*Lactobacillus rhamnosus* GG (ATCC 53103); *L. rhamnosus* LC705 (DSM 7061); *Bifidobacterium breve* Bb99 (DSM 13692); and *Propionibacterium freudenreichii* ssp. *shermanii* JS (DSM 7076)) or a placebo for two to four weeks before delivery, their infants received the same probiotics plus galacto-oligosaccharides or a placebo for six months.<sup>102</sup> Probiotic treatment significantly reduced eczema and atopic eczema. The abundance of *Lactobacilli* and *bifidobacteria* was significantly higher in the guts of supplemented infants. Interestingly, in infants with eczema, the same combination of probiotic bacteria as used in this study induced an significant increase in plasma IL-10 levels.<sup>95</sup> Primary prevention of eczema by perinatal administration of probiotic bacteria indeed involves modulation of the early colonisation of the intestinal microbiota, which may result in modulating the development and maturation of the infants immune system. Gut microbiota contact directly with extensions of dendritic cells, which orchestrate the mucosal immune homeostasis. Commensal bacteria stimulate the innate immune system and contribute to the generation of regulatory lymphocytes, which, through IL-10 and TGF- $\beta$ , establish and maintain mucosal immune tolerance.<sup>103</sup> Modulation of the immune response via interaction with intestinal dendritic cells with subsequent effects on T-cell differentiation and induction of regulatory T cells has been suggested.<sup>88</sup> Furthermore, recognition of commensal bacteria by TLRs on intestinal epithelial cells and cells of the mucosal immune system is essential for intestinal (immune) homeostasis.<sup>104,105</sup> Probiotic signalling through TLRs may contribute to maintaining mucosal and intestinal homeostasis and thereby preventing eczema. Above all, probiotics intervention might be a potential effective approach in the treatment of eczema via restoring gut microbiota. Therapies that may most efficiently bring the disease under control are still being sought.

## Concluding remarks

The human intestinal hosts possess trillions of microorganisms, also collectively known as the bacterial flora. An increasing number of studies are progressively unravelling the fascinating interaction between hosts and microbiota.<sup>106–108</sup> More and more evidence indicates that intestinal microbiota exert a critical function in keeping healthy and the presence of eczema. Its mechanism, however, remains elusive despite promising results from previous studies. The specific strains among commensal gut bacteria may exert either a phylactic or moribific effect into eczema progression. Additionally, previous series completed in both animal and human models demonstrated that an effective strategy of preventing and managing eczema might target on gut microbiota. Probiotics/prebiotic can confer health by modulating the composition of gut microbiota and restoring the physiological bacterial flora. Many studies have provided a compelling rationale for exploring the oral probiotics administered as adjunctive therapies to eczema. That being said, limited data are available in this area amid, and the relevant scientific work is still at the early stage. A lot of studies need to be carried out in the future, as below. Initially, a host provides a large and complex environment for gut bacterial flora. It is as yet unclear whether the alteration of intestinal microbiota contributes to the development of eczema or its presence reflects the primary cause of changes in gut microbiota. There is a need for a profound understanding of the interactive mechanism between intestinal microbiota and host. Of note, a great number of studies need to be completed in diverse populations or mammal models and various types of food. As such, it is singularly critical to offer an entirely novel approach to cure diseases for sound health through monitoring and controlling gut microbiota. Compared with developing novel drugs for inflammation, it might cost less to seek novel approaches, such as monitoring and manipulating human gut microbiota if required using probiotics and/or prebiotics (non-digestible food additives triggering the development and/or activity of bacteria). The opportunity exists to develop updated probiotics in accordance with the interaction between specific microbiota and eczema. In addition, it is plausible to develop probiotics from intestinal microbiota in healthy groups, for instance, faecal transplant as a therapeutic strategy has whetted more appetite. Similarly, it is significant to determine how prebiotics and/or probiotics change the constitution of intestinal microbiota (reconstructing bacterial flora) and how relative it is to eczema. Up to now, only a fraction of human studies have reported the alteration of gut microbiota in pre- and post-probiotics and/or prebiotics treatment among eczema patients. There is also a need for construction of “*Pan-microbiome*” studies which may play an essential role in a further understanding of eczema aetiopathogenesis. Most importantly, recent findings were validated in studies of the microbiota at diverse stages of eczema (and in differing at-risk populations). Given few relevant researches in the area, more studies should be carried out to elucidate the interaction between specific microbiota and eczema, and a wealth of well-controlled clinical studies on gut microbiota are required to make sure safety in patients. Synergistic efforts in vivo and in vitro are also to be asked for so as to advance our knowledge in the area, which

is promising for its existing potential in biomarker reorganisation and novel therapeutic targets. All in all, we have opened up a completely novel access to the understanding and therapy of eczema and more research is urgently needed in this emerging field.

## Ethical disclosures

**Confidentiality of data.** The authors declare that no patient data appears in this article.

**Right to privacy and informed consent.** The authors declare that no patient data appears in this article.

**Protection of human subjects and animals in research.** The authors declare that no experiments were performed on humans or animals for this investigation.

## Funding

None.

## Conflict of interest

None declared.

## Acknowledgements

We thank Yue Cai and Wei Pan for critical review and important intellectual contributions to the final version of the manuscript.

## References

1. Barnetson RSC, Rogers M. Childhood atopic eczema. *BMJ*. 2002;324:1376–9.
2. Beasley R. Worldwide variation in prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and atopic eczema: ISAAC. *Lancet*. 1998;351:1225–32.
3. Asher MI, Montefort S, Björkstén B, Lai CK, Strachan DP, Weiland SK, et al. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phases One and Three repeat multicountry cross-sectional surveys. *Lancet*. 2006;368:733–43.
4. Williams H, Stewart A, Mutius EV, Cookson W, Anderson HR. Is eczema really on the increase worldwide? *J Allergy Clin Immunol*. 2008;121:947–54.
5. He JQ, Chan-Yeung M, Becker AB, Dimich-Ward H, Ferguson AC, Manfreda J, et al. Genetic variants of the IL13 and IL4 genes and atopic diseases in at-risk children. *Genes Immun*. 2003;4:385–9.
6. Kawashima T, Noguchi E, Arinami T, Yamakawa-Kobayashi K, Nakagawa H, Otsuka F, et al. Linkage and association of an interleukin 4 gene polymorphism with atopic dermatitis in Japanese families. *J Med Genet*. 1998;35:502–4.
7. Liu X, Nickel R, Beyer K, Wahn U, Ehrlich E, Freidhoff LR, et al. An IL13 coding region variant is associated with a high total serum IgE level and atopic dermatitis in the German Multicenter Atopy Study (MAS-90). *J Allergy Clin Immunol*. 2000;106:167–70.

8. Mao XQ, Shirakawa T, Yoshikawa T, Yoshikawa K, Kawai M, Sasaki S, et al. Association between genetic variants of mast-cell chymase and eczema. *Lancet*. 1996;348:581–3.
9. Kato A, Fukai K, Oiso N, Hosomi N, Murakami T, Ishii M. Association of SPINK5 gene polymorphisms with atopic dermatitis in the Japanese population. *Br J Dermatol*. 2003;148:665–9.
10. Palmer CNA, Irvine AD, Terronkwiatkowski A, Zhao Y, Liao H, Lee SP, et al. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. *Nat Genet*. 2006;38:441–6.
11. Brown SJ, Irvine AD. Atopic eczema and the filaggrin story. *Semin Cutan Med Surg*. 2008;27:128–37.
12. Quaranta M, Knapp B, Garzorz N, Mattii M, Pullabhatla V, Pennino D, et al. Intra-individual genome expression analysis reveals a specific molecular signature of psoriasis and eczema. *Sci Transl Med*. 2014;6:244ra290.
13. Salminen S, Bouley C, Boutron-Ruault MC, Cummings JH, Franck A, Gibson GR, et al. Functional food science and gastrointestinal physiology and function. *Br J Nutr*. 1998;80:S147–71.
14. Guarner F, Malagelada JR. Gut flora in health and disease. *Lancet*. 2003;361:512–9.
15. Chan CWH, Wong RS, Law PTW, Wong CL, Tsui SKW, Tang WPY, et al. Environmental factors associated with altered gut microbiota in children with eczema: a systematic review. *Int J Mol Sci*. 2016;17.
16. Bridgman SL, Kozyrskyj AL, Scott JA, Becker AB, Azad MB. Gut microbiota and allergic disease in children. *Ann Allergy Asthma Immunol*. 2016;116:99–105.
17. Hong PY, Lee BW, Aw M, Shek LPC, Yap GC, Chua KY, et al. Comparative analysis of fecal microbiota in infants with and without eczema. *PLoS One*. 2010;5:e9964.
18. Beck LA, Leung DYM. Allergen sensitization through the skin induces systemic allergic responses. *J Allergy Clin Immunol*. 2000;106:258–63.
19. Hulst AEVD, Klip H, Brand PLP. Risk of developing asthma in young children with atopic eczema: a systematic review. *J Allergy Clin Immunol*. 2007;120:565–9.
20. Beiber T, de la Salle H, Wollenberg A, Hakimi J, Chizzonite R, Ring J, et al. Human epidermal Langerhans cells express the high affinity receptor for immunoglobulin E (Fc epsilon RI). *J Exp Med*. 1992;175:1285.
21. Wollenberg A, Kraft S, Hanau D, Bieber T. Immunomorphological and ultrastructural characterization of Langerhans cells and a novel, inflammatory dendritic epidermal cell (IDEC) population in lesional skin of atopic eczema. *J Invest Dermatol*. 1996;106:446–53.
22. Novak N, Allam JP, Hagemann T, Jenneck C, Laffer S, Valenta R, et al. Characterization of Fc epsilon RI-bearing CD123 dendritic cell antigen-2 plasmacytoid dendritic cells in atopic dermatitis. *J Allergy Clin Immunol*. 2004;114:364–70.
23. Leung DY, Boguniewicz M, Howell MD, Nomura I, Hamid QA. New insights into atopic dermatitis. *J Clin Invest*. 2004;113:651–7.
24. van Reijnsen FC, Bruijnzeelkoomen CA, Kalthoff FS, Maggi E, Romagnani S, Westland JK, et al. Skin-derived aeroallergen-specific T-cell clones of Th2 phenotype in patients with atopic dermatitis. *J Allergy Clin Immunol*. 1992;90:184–93.
25. Leberman DA, Coffman RL. Interleukin 4 causes isotype switching to IgE in T cell-stimulated clonal B cell cultures. *J Exp Med*. 1988;168:853–62.
26. Akdis M, Trautmann A, Klunker S, Daigle I, Küçüksezer UC, Deglmann W, et al. T helper (Th) 2 predominance in atopic diseases is due to preferential apoptosis of circulating memory/effector Th1 cells. *FASEB J*. 2003;17:1026–35.
27. Ou LS, Goleva EC, Leung DY. T regulatory cells in atopic dermatitis and subversion of their activity by superantigens. *J Allergy Clin Immunol*. 2004;113:756–63.
28. Isolauri E. Dietary modification of atopic disease: use of probiotics in the prevention of atopic dermatitis. *Curr Allergy Asthma Rep*. 2004;4:270–5.
29. Isolauri E, Kalliomäki M, Laitinen K, Salminen S. Modulation of the maturing gut barrier and microbiota: a novel target in allergic disease. *Curr Pharm Des*. 2008;14:1368–75.
30. West CE. Gut microbiota and allergic disease: new findings. *Curr Opin Clin Nutr Metab Care*. 2014;17:261–6.
31. Forno E, Onderdonk AB, Mccracken J. Diversity of the gut microbiota and eczema in early life. *Clin Mol Allergy*. 2008;6:1–9.
32. Ismail IH, Oppedisano F, Joseph SJ, Boyle RJ, Licciardi PV, Robins-Browne RM, et al. Reduced gut microbial diversity in early life is associated with later development of eczema but not atopy in high-risk infants. *Pediatr Allergy Immunol*. 2012;23:674–81.
33. Abrahamsson TR, Jakobsson HE, Andersson AF, Björkstén B, Engstrand L, Jenmalm MC. Low diversity of the gut microbiota in infants with atopic eczema. *J Allergy Clin Immunol*. 2011;129:434–40.
34. Doreswamy V, Peden DB. Modulation of asthma by endotoxin. *Clin Exp Allergy*. 2011;41:9–19.
35. Gehring U, Bolte G, Borte M, Bischof W, Fahlbusch B, Wichmann HE, et al. Exposure to endotoxin decreases the risk of atopic eczema in infancy: a cohort study. *J Allergy Clin Immunol*. 2001;108:847–54.
36. Nylund L, Satokari R, Nikkilä J. Microarray analysis reveals marked intestinal microbiota aberrancy in infants having eczema compared to healthy children in at-risk for atopic disease. *BMC Microbiol*. 2013;13:1–11.
37. West CE, Rydén P, Lundin D, Engstrand L, Tulic MK, Prescott SL. Gut microbiome and innate immune response patterns in IgE-associated eczema. *Clin Exp Allergy*. 2015;45:1419–29.
38. Coppa GV, Gabrielli O, Zampini L, Galeazzi T, Ficcadenti A, Padella L, et al. Oligosaccharides in 4 different milk groups, *Bifidobacteria*, and *Ruminococcus obeum*. *J Pediatr Gastroenterol Nutr*. 2011;53:80–7.
39. Dabard J, Bridonneau C, Phillippe C, Anglade P, Molle D, et al. *Ruminococcus*, a new lantibiotic produced by a *Ruminococcus gnavus* strain isolated from human feces. *Appl Environ Microbiol*. 2001;67:4111–8.
40. Xu J, Bjursell MK, Himrod J, Deng S, Carmichael LK, Chiang HC, et al. A genomic view of the human-*Bacteroides thetaiotaomicron* symbiosis. *Science*. 2003;299:2074–6.
41. Koenig JE, Spor A, Scalfone N, Fricker AD, Stombaugh J, Knight R, et al. Succession of microbial consortia in the developing infant gut microbiome. *Proc Natl Acad Sci U S A*. 2011;108:4578–85.
42. Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell*. 2005;122:107–18.
43. Watanabe S, Narisawa Y, Arase S, Okamoto H, Ikenaga T, Tajiri Y, et al. Differences in fecal microflora between patients with atopic dermatitis and healthy control subjects. *J Allergy Clin Immunol*. 2003;111:587–91.
44. Arkwright PD, David TJ. Intradermal administration of a killed *Mycobacterium vaccae* suspension (SRL 172) is associated with improvement in atopic dermatitis in children with moderate-to-severe disease. *J Allergy Clin Immunol*. 2001;107:531–4.
45. Mah KW, Björkstén B, Lee BW, van Bever HP, Shek LP, Tan TN, et al. Distinct pattern of commensal gut microbiota in toddlers with eczema. *Int Arch Allergy Immunol*. 2006;140:157–63.



46. Maynard CL, Elson CO, Hatton RD, Weaver CT. Reciprocal interactions of the intestinal microbiota and immune system. *Nature*. 2012;489:231–41.
47. Rosenfeldt V, Benfeldt E, Valerius NH, Pærregaard A, Michaelsen KF. Effect of probiotics on gastrointestinal symptoms and small intestinal permeability in children with atopic dermatitis. *J Pediatr*. 2004;145:612–6.
48. Kirjavainen PV. Aberrant composition of gut microbiota of allergic infants: a target of bifidobacterial therapy at weaning? *Gut*. 2002;51:51–5.
49. Majamaa H, Isolauri E. Probiotics: a novel approach in the management of food allergy. *J Allergy Clin Immunol*. 1997;99:179–85.
50. Isolauri E, Arvola T, Sütas Y, Moilanen E, Salminen S. Probiotics in the management of atopic eczema. *Clin Exp Allergy*. 2000;30:1604–10.
51. Kalliomäki M, Salminen S, Arvilommi H, Kero P, Koskinen P, Isolauri E. Probiotics in primary prevention of atopic disease: a randomised placebo-controlled trial. *Ambulatory Child Health*. 2001;7:334–5.
52. Franz CM, Holzapfel WH, Stiles ME. Enterococci at the crossroads of food safety? *Int J Food Microbiol*. 1999;47:1–24.
53. Tang MF, Sy HY, Kwok JSL, Tam WH, Hon KL, Tung CKC, et al. Eczema susceptibility and composition of faecal microbiota at 4 weeks of age: a pilot study in Chinese infants. *Br J Dermatol*. 2016.
54. Lamb-Rosteski JM, Kalischuk LD, Inglis GD, Buret AG. Epidermal growth factor inhibits *Campylobacter jejuni*-induced claudin-4 disruption, loss of epithelial barrier function, and *Escherichia coli* translocation. *Infect Immun*. 2008;76:3390–8.
55. Wang HB, Wang PY, Wang X, Wan YL, Liu YC. Butyrate enhances intestinal epithelial barrier function via up-regulation of tight junction protein Claudin-1 transcription. *Dig Dis Sci*. 2012;57:3126–35.
56. Penders J, Stobberingh EE, Thijs C, Adams H, Vink C, Van RR, et al. Molecular fingerprinting of the intestinal microbiota of infants in whom atopic eczema was or was not developing. *Clin Exp Allergy*. 2006;36:1602–8.
57. Penders J, Thijs C, Pa VDB, Kummeling I, Sniijders B, Stelma F, et al. Gut microbiota composition and development of atopic manifestations in infancy: the KOALA Birth Cohort Study. *Gut*. 2007;56:661–7.
58. van Nimwegen FA, Penders J, Stobberingh EE, Postma DS, Koppelman GH, Kerkhof M, et al. Mode and place of delivery, gastrointestinal microbiota, and their influence on asthma and atopy. *J Allergy Clin Immunol*. 2011;128:948–55, e943.
59. Karlsson H, Hessel C, Rudin A. Innate immune responses of human neonatal cells to bacteria from the normal gastrointestinal flora. *Infect Immun*. 2002;70:6688–96.
60. Linneberg A, Østergaard C, Tvede M, Andersen LP, Nielsen NH, Madsen F, et al. IgG antibodies against microorganisms and atopic disease in Danish adults: the Copenhagen Allergy Study. *J Allergy Clin Immunol*. 2003;111:847–53.
61. Pothoulakis C. Effects of *Clostridium difficile* toxins on epithelial cell barrier. *Ann N Y Acad Sci*. 2000;915:347–56.
62. Feltis BA, Wiesner SM, Kim AS, Erlandsen SL, Lyerly DL, Wilkins TD, et al. *Clostridium difficile* toxins A and B can alter epithelial permeability and promote bacterial paracellular migration through HT-29 enterocytes. *Shock*. 2000;14:629–34.
63. Majamaa H, Isolauri E. Evaluation of the gut mucosal barrier: evidence for increased antigen transfer in children with atopic eczema. *J Allergy Clin Immunol*. 1996;97:985–90.
64. Benard A, Desreumeaux P, Huglo D, Hoorelbeke A, Tonnel AB, Wallaert B. Increased intestinal permeability in bronchial asthma. *J Allergy Clin Immunol*. 1996;97:1173–8.
65. Caffarelli C, Cavagni G, Menzies IS, Bertolini P, Atherton DJ. Elimination diet and intestinal permeability in atopic eczema: a preliminary study. *Clin Exp Allergy*. 1993;23:28–31.
66. Pike MG, Heddle RJ, Boulton P, Turner MW, Atherton DJ. Increased intestinal permeability in atopic eczema. *J Invest Dermatol*. 1986;86:101–4.
67. Ismail IH, Boyle RJ, Licciardi PV, Oppedisano F, Lahtinen S, Robins-Browne RM, et al. Early gut colonisation by *Bifidobacterium breve* and *B. catenulatum* differentially modulates eczema risk in children at high-risk of developing allergic disease. *Pediatr Allergy Immunol*. 2016;27:838–46.
68. Roock SD, Elk MV, Dijk MEAV, Timmerman HM, Rijkers GT, Prakken BJ, et al. Lactic acid bacteria differ in their ability to induce functional regulatory T cells in humans. *Clin Exp Allergy*. 2010;40:103–10.
69. Lopez P, Gueimonde M, Margolles A, Suarez A. Distinct *Bifidobacterium* strains drive different immune responses in vitro. *Int J Food Microbiol*. 2010;138:157–65.
70. Pozo-Rubio T, Mujico JR, Marcos A, Puertollano E, Nadal I, Sanz Y, et al. Immunostimulatory effect of faecal *Bifidobacterium* species of breast-fed and formula-fed infants in a peripheral blood mononuclear cell/Caco-2 co-culture system. *Br J Nutr*. 2011;106:1216–23.
71. Inoue Y, Iwabuchi N, Xiao JZ, Yaeshima T, Iwatsuki K. Suppressive effects of *bifidobacterium breve* strain M-16V on T-helper type 2 immune responses in a murine model. *Biol Pharm Bull*. 2009;32:760–3.
72. Ma VDP, Lutter R, Smids BS, Weersink EJ, Js VZD. Synbiotics reduce allergen-induced T-helper 2 response and improve peak expiratory flow in allergic asthmatics. *Allergy*. 2011;66:39–47.
73. Nowrouzian FL, Lina G, Hodille E, Lindberg E, Hesselmar B, Saalman R, et al. Superantigens and adhesins of infant gut commensal *Staphylococcus aureus* strains and association with subsequent development of atopic eczema. *Br J Dermatol*. 2017;176:439–45.
74. Islander U, Andersson A, Lindberg E, Adlerberth I, Wold AE, Rudin A. Superantigenic *Staphylococcus aureus* stimulates production of interleukin-17 from memory but not naive T cells. *Infect Immun*. 2010;78:381–6.
75. Brisbin JT, Gong J, Parvizi P, Sharif S. Effects of lactobacilli on cytokine expression by chicken spleen and cecal tonsil cells. *Clin Vaccine Immunol*. 2010;17:1337–43.
76. Ou CC, Lin SL, Tsai JJ, Lin MY. Heat-killed lactic acid bacteria enhance immunomodulatory potential by skewing the immune response toward Th1 polarization. *J Food Sci*. 2011;76:M260–7.
77. Sierra S, Lara-Villoslada F, Sempere L, Olivares M, Boza J, Xaus J. Intestinal and immunological effects of daily oral administration of *Lactobacillus salivarius* CECT5713 to healthy adults. *Anaerobe*. 2010;16:195–200.
78. Lee JH, Valeriano VD, Shin YR, Chae JP, Kim GB, Ham JS, et al. Genome sequence of *Lactobacillus mucosae* LM1, isolated from piglet feces. *J Bacteriol*. 2012;194:4766.
79. West CE, Hammarström ML, Hernell O. Probiotics during weaning reduce the incidence of eczema. *Pediatr Allergy Immunol*. 2009;20:430–7.
80. Wickens K, Black P, Stanley TV, Mitchell E, Barthow C, Fitzharris P, et al. A protective effect of *Lactobacillus rhamnosus* HN001 against eczema in the first 2 years of life persists to age 4 years. *Clin Exp Allergy*. 2012;42:1071–9.
81. Wickens K, Black PN, Stanley TV, Mitchell E, Fitzharris P, Tannock GW, et al. A differential effect of 2 probiotics in the prevention of eczema and atopy: a double-blind, randomized, placebo-controlled trial. *J Allergy Clin Immunol*. 2008;122:788–94.
82. Marlow G. Differential effects of two probiotics on the risks of eczema and atopy associated with single nucleotide polymorphisms to Toll-like receptors. *Pediatr Allergy Immunol*. 2015;26:262–71.
83. Gill HS, Rutherford KJ, Prasad J, Gopal PK. Enhancement of natural and acquired immunity by *Lactobacillus rhamnosus*

- (HN001) *Lactobacillus acidophilus* (HN017) and *Bifidobacterium lactis* (HN019). *Br J Nutr*. 2000;83:167–76.
84. Weston S, Halbert A, Richmond P, Prescott SL. Effects of probiotics on atopic dermatitis: a randomised controlled trial. *Arch Dis Child*. 2005;90:892–7.
  85. Abrahamsson TR, Jakobsson T, Böttcher MF, Fredrikson M, Jenmalm MC, Björkstén B, et al. Probiotics in prevention of IgE-associated eczema: a double-blind, randomized, placebo-controlled trial. *J Allergy Clin Immunol*. 2007;119:S237.
  86. Ma D, Forsythe P, Bienenstock J. Live *Lactobacillus rhamnosus* [corrected] is essential for the inhibitory effect on tumor necrosis factor alpha-induced interleukin-8 expression. *Infect Immun*. 2004;72:5308–14.
  87. Peña JA, Rogers AB, Ge Z, Ng V, Li SY, Fox JG, et al. Probiotic *Lactobacillus* spp. diminish helicobacter hepaticus-induced inflammatory bowel disease in interleukin-10-deficient mice. *Infect Immun*. 2005;73:912–20.
  88. Smits HH, Engering A, Van dKD, de Jong EC, Schipper K, van Capel TM, et al. Selective probiotic bacteria induce IL-10-producing regulatory T cells in vitro by modulating dendritic cell function through dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin. *J Allergy Clin Immunol*. 2005;115:1260–7.
  89. Jacobsen CN, Nielsen VR, Hayford AE, Møller PL, Michaelsen KF, Pærregaard A, et al. Screening of probiotic activities of forty-seven strains of *Lactobacillus* spp. by in vitro techniques and evaluation of the colonization ability of five selected strains in humans. *Appl Environ Microbiol*. 1999;65:4949–56.
  90. Kalliomäki M, Salminen S, Poussa T, Arvilommi H, Isolauri E. Probiotics and prevention of atopic disease: 4-year follow-up of a randomised placebo-controlled trial. *Lancet*. 2003;361:1869–71.
  91. Kalliomäki M, Salminen S, Poussa T, Isolauri E. Probiotics during the first 7 years of life: a cumulative risk reduction of eczema in a randomized, placebo-controlled trial. *J Allergy Clin Immunol*. 2007;119:1019–21.
  92. Rautava S, Kalliomäki M, Isolauri E. Probiotics during pregnancy and breastfeeding might confer immunomodulatory protection against atopic disease in the infant. *J Allergy Clin Immunol*. 2002;109:119–21.
  93. Stavnezer J. Regulation of antibody production and class switching by TGF-beta. *J Immunol*. 1995;155:1647–51.
  94. Petitprez, Khalife, Fontaine, Lafitte, Capron, Grzych. Cytokine mRNA expression in lymphoid organs associated with the expression of IgA response in the rat. *Scand J Immunol*. 1999;49:14–20.
  95. Viljanen ME, Haahtela T, Korpela R, Kuitunen M, Sarnesto A, Vaarala O, et al. Induction of inflammation as a possible mechanism of probiotic effect in atopic eczema-dermatitis syndrome. *J Allergy Clin Immunol*. 2005;115:1254–9.
  96. Goto H, Gidlund M. Soluble CD4: a link between specific immune mechanisms and non-specific inflammatory responses? *Scand J Immunol*. 1996;43:690–2.
  97. Koller DY, Halmerbauer G, Frischer T, Roithner B. Assessment of eosinophil granule proteins in various body fluids: is there a relation to clinical variables in childhood asthma? *Clin Exp Allergy*. 1999;29:786–93.
  98. Lin RJ, Qiu LH, Guan RZ, Hu SJ, Liu YY, Wang GJ. Protective effect of probiotics in the treatment of infantile eczema. *Exp Ther Med*. 2015;9:1593–6.
  99. Ozdemir O. Any benefits of probiotics in allergic disorders? *Allergy Asthma Proc*. 2010;31:103–11.
  100. Rautava S, Kainonen E, Salminen S, Isolauri E. Maternal probiotic supplementation during pregnancy and breast-feeding reduces the risk of eczema in the infant. *J Allergy Clin Immunol*. 2012;130:816–7.
  101. Niers L, Martín R, Rijkers G, Sengers F, Timmerman H, Van UN, et al. The effects of selected probiotic strains on the development of eczema (the PandA study). *Allergy*. 2009;64:1349–58.
  102. Kukkonen K, Savilahti E, Haahtela T, Juntunen-Backman K, Korpela R, Poussa T, et al. Probiotics and prebiotic galactooligosaccharides in the prevention of allergic diseases: a randomized, double-blind, placebo-controlled trial. *J Allergy Clin Immunol*. 2007;119:192–8.
  103. Kelly D, Conway S, Aminov R. Commensal gut bacteria: mechanisms of immune modulation. *Trends Immunol*. 2005;26:326–33.
  104. Artis D. Epithelial-cell recognition of commensal bacteria and maintenance of immune homeostasis in the gut. *Nat Rev Immunol*. 2008;8:411–20.
  105. Rakoffnahoum S, Paglino J, Eslamivarzaneh F, Edberg S, Medzhitov R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell*. 2004;118:229–41.
  106. Kang YB, Cai Y, Zhang H. Gut microbiota and allergy/asthma: from pathogenesis to new therapeutic strategies. *Allergol Immunopathol*. 2017;45:305–9.
  107. Kang Y, Zhang X, Cai Y, Su J, Kong X. Gut microbiota and metabolic disease: from pathogenesis to new therapeutic strategies. *Rev Med Microbiol*. 2016;27:141–52.
  108. Kang Y, Cai Y, Zhang X, Kong X, Su J. Altered gut microbiota in RA: implications for treatment. *Z Rheumatol*. 2017;76:451–7.