



REVISTA MÉDICA CLÍNICA LAS CONDES

<https://www.journals.elsevier.com/revista-medica-clinica-las-condes>

Miopatías congénitas

Congenital myopathies

Edoardo Malfatti MD PhD^{abc}✉

- ^a Service Neurologie Médicale, Centre de Référence Maladies Neuromusculaire Paris-Nord. CHU Raymond-Poincaré.
^b U1179 UVSQ-INSERM Handicap Neuromusculaire : Physiologie, Biothérapie et Pharmacologie appliquées, UFR des sciences de la santé Simone Veil, Université Versailles-Saint-Quentin-en-Yvelines. 104, boulevard Raymond Poincaré. Garches - France.
^c Unité de Morphologie Neuromusculaire, Institut de Myologie, Groupe Hospitalier Universitaire La Pitié-Salpêtrière; Paris, France.

INFORMACIÓN DEL ARTÍCULO

Historia del Artículo:

Recibido: 04 06 2018.

Aceptado: 10 10 2018.

Palabras clave:

Miopatías congénitas, miopatías nemalínicas, miopatías congénitas, miopatía congénita del núcleo central, miopatía corporal reducida, miopatías centronucleares.

Key words:

Congenital myopathies, nemaline myopathies, congenital myopathies with cores, core-rod myopathies, reducing body myopathy, cap myopathy, centronuclear myopathies.

RESUMEN

Las miopatías congénitas son un grupo de trastornos musculares esqueléticos hereditarios, clínica y genéticamente heterogéneos, definidos de acuerdo con los hallazgos histopatológicos observados en las biopsias musculares. Las lesiones histopatológicas más comunes son los agregados de proteínas, los cores y el aumento de las centralizaciones nucleares. En el campo de las miopatías congénitas musculares se han logrado avances en los últimos años, al definir nuevos trastornos. En particular, el desarrollo de las técnicas de secuenciación de nueva generación o secuenciación masiva (NGS) permitieron la identificación de alrededor de 20 nuevas formas de miopatías congénitas. Se han logrado algunos avances en el conocimiento de la alteración de la contractilidad muscular en el campo de las miopatías nemalínicas. Con la perspectiva de futuros ensayos clínicos, el alcanzar un diagnóstico genético e histopatológico y establecer las medidas de atención aceptadas internacionalmente, son fundamentales para reducir la carga de la enfermedad en los pacientes con miopatías congénitas.

ABSTRACT

Congenital myopathies are a group of primary hereditary, clinically and genetically heterogeneous skeletal muscle disorders, defined according to histopathologic lesions observed in muscle biopsies. The most common histopathologic lesions are protein aggregates, cores, and increased nuclear centralizations. The field of muscle congenital myopathies has met progress in the recent years by defining new disorders. In particular, next generation sequencing (NGS) techniques allowed the identification of around 20 novel forms of congenital myopathies. Some insights on the alteration of muscle contractility have been gained in the field of nemaline myopathies. In the perspective of future clinical trials, reaching out a confirmed genetic and histopathologic diagnosis and establish internationally accepted care measures are pivotal to reduce the disease burden of congenital myopathies patients.

INTRODUCTION

The core clinical features of congenital myopathies are early onset hypotonia, delayed motor milestones, muscular weakness, and respiratory muscle involvement¹. There is wide variability in severity. The most severe forms show profound muscle weakness or fetal akinesia, bulbar weakness with

deglutition problems and drooling, and respiratory involvement necessitating assistance. Patients can show dysmorphic features or skeletal deformities or arthrogryposis. Polyhydramnios or reduced fetal movements may be present during pregnancy. Miscarriages are often reported by patients' mother in cases of autosomal recessive inheritance.

✉ Autor para correspondencia

Correo electrónico: edoardo.malfatti@aphp.fr, edoardo.malfatti@gmail.com

e-ISSN: 2531-0186/ ISSN: 0716-8640

<https://doi.org/10.1016/j.rmclc.2018.10.003>

0716-8640/© 2018 Revista Médica Clínica Las Condes. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).



The classic phenotype associate neonatal hypotonia, gross motor delay, and proximal and axial weakness showing different degrees of severity. Recurrent respiratory infections are often reported. Patients may show skeletal deformities such as pectus excavatum, flat thorax and they develop scoliosis during growth spurt. At physical examination, patients have thin muscle bulk and reduced body weight. Facial and maxillary alterations including high-arched palate, tubular nose or low set ears are frequent and, in some conditions, there is a high-pitched or nasal voice. Ocular involvement including ptosis and ophthalmoplegia is present in specific congenital myopathies².

Muscle weakness is mainly proximal and axial and patients report frequent falls, neck weakness or difficulties rising up from the supine position, necessitating rolling on one side. Facial weakness is recognizable with patients showing open mouth with an inverted V shape. Some congenital myopathies manifest with distal muscle involvement of legs and foot drop³. In the milder forms, difficulties in sport activities or effort intolerance can be the only manifesting signs, with patients developing muscle weakness later in life.

In general, patients tend to ameliorate during infancy and adolescence. However, certain forms are particularly aggressive leading to weakness progression and early adulthood gait loss (e.g. reducing body myopathy, centronuclear myopathies).

Some patients show prominent fatigability, effort intolerance or fluctuations that are due to the presence of secondary alterations of the neuromuscular junction⁴. Myalgias are frequent in patients with 'core myopathies' linked to *RYR1* gene mutations (see below).

Serum creatine kinase (sCK) level is normal or slightly elevated. Electrophysiological studies and electromyography show myopathic features.

Whole body muscle MRI is very useful in orientating the genetic screening. In fact, different profiles of distribution of muscle alterations have been found as constant features in particular entities⁵.

Muscle biopsy analysis with both light and electron microscopy techniques is a useful tool to confirm the diagnosis and orientate genetic analyses. However, these analyses must be performed in highly skilled and experienced centers in order to allow a good interpretation and avoid technical artifacts¹. Histopathologic features of each condition will be described in the sections relative to specific disorders.

Genetic pedigree and a careful family history are fundamental to establish the possible pattern of inheritance and start genetic analyses. More and more genes associated to congenital myopathies are described thank to exome/whole exome sequencing approaches⁶. For this reason, a constant and inte-

grated crosstalk between clinicians, histopathologists, and geneticists is necessary to provide the best accurate diagnosis for every patient. Nevertheless around 50 percent of patients do not show mutations in the already know genes. We expect to identify novel congenital myopathies in the next years.

A multidisciplinary approach is necessary for the follow up of congenital myopathy patients. This should include the intervention of pediatric neurologist, orthopedic, pulmonologist, cardiologist, nutritionist, and physical therapist. Clinician should refer the guidelines published by the International Standard of Care Committee for Congenital Myopathies' group⁷.

2. SPECIFIC ENTITIES

2.1 Nemaline myopathies

Nemaline (from the Greek word néma = rod) or rod myopathies (NM) probably represent the congenital myopathy most frequently encountered. They are characterized by the presence of small, rod-shaped inclusions inside the cytoplasm of the fiber⁸. Clinical spectrum is wide and comprises forms manifesting antenatally, or milder form with childhood onset weakness. NM are genetically heterogeneous and inheritance can be autosomal dominant (AD), de novo, or autosomal recessive. This heterogeneity is demonstrated by the fact that twelve genes:

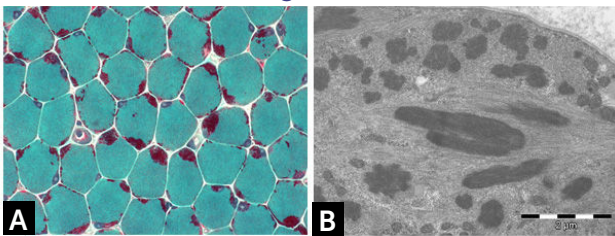
1. *ACTA1*, MIM#161800
2. *NEB*, MIM#256030
3. *TPM2*, MIM#609285
4. *TPM3*, MIM#609284
5. *TNNT1*, MIM#605355
6. *KBTBD13*, MIM#609273
7. *CFL2*, MIM#610687
8. *KLHL40* MIM#615340
9. *KLHL41* MIM#615731
10. *LMOD3* MIM#616112
11. *MYO18B* MIM# 607295
12. *MYPN* MIM#608517

Have been associated with these conditions⁸⁻⁹. The most frequently mutated gene is the nebuline gene, *NEB*, in around 50 percent of the genetically identified forms¹⁰. NM linked to actin gene, *ACTA1*, is the second most frequent form with more than 200 mutations reported¹¹. Cases with *ACTA1* de novo mutations seem to be particularly severe, while dominant forms show a classic nemaline myopathy phenotype.

Cardiac involvement is not considered a core clinical feature of nemaline myopathies. However, *ACTA1* NM, the recently described forms linked to *MYO18B*, myosin 18B, coding for a non-conventional myosin, and *MYPN*, myopalladin, a Z-line protein, can associate both myopathy and cardiomyopathy¹²⁻⁹.

Histologically, all nemaline myopathies show the presence of fuchsinophilic protein inclusions visible with Gömöri trichrome staining (Fig.1A). The latter are often elongated, and vary in number and distribution inside the muscle fiber. In newborn muscle biopsies, rods identification can be particularly tricky. In these cases, electron microscopy is mandatory in order to disclose the presence of rods. With this technique they appear as electron dense bodies measuring 1–7 μm length and 0.3–2 μm width (Fig.1B). They present an orthogonally organized filamentous structure resembling to Z-line material. The exact origin and mechanisms of rod formation are largely unknown. Rods can be found inside myonuclei in *ACTA1* and *MYOPN* NM¹¹⁻⁹.

Figure 1.

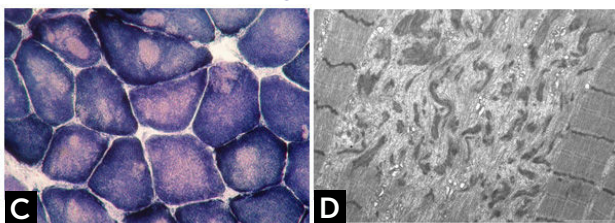


(A and B) Nemaline myopathy with mutations in *NEB* gene; (A) Clusters of rods at the periphery of fibers staining red at GT. (B) Electron micrograph showing nemaline bodies transversally and longitudinally oriented.

2. CONGENITAL MYOPATHIES WITH CORES

Core lesions are defined as well-defined rounded areas lacking oxidative activity (SDH, NADH etc.) corresponding to myofibrillar disorganization (Fig.1C). By electron microscopy a typical core correspond to wide areas of compacted and disorganized myofibrils, with Z-line streaming and absence of mitochondria, extending over numerous sarcomeres or almost along the full length of the fiber (Fig.1D)¹³. Cores can be central and single, single and peripheral, or multiple. Patients with core myopathies can present profound clinical severity with antennal manifestation or develop a proximal and mild muscle weakness in childhood or early adulthood. Inheritance is autosomal dominant or recessive and the majority of patients show mutation(s) of the *RYR1* gene, coding for the skeletal muscle ryanodin channel receptor, which is also associated with susceptibility to malignant hyperthermia and rhabdomyolysis¹⁴. AR *RYR1* mutated patients can show ocular involvement with ptosis and ophtalmoplegia. AD mutations in *MYH7* or *CCD18* genes have been also associated with 'core' myopathies^{15,16}.

Figure 1.



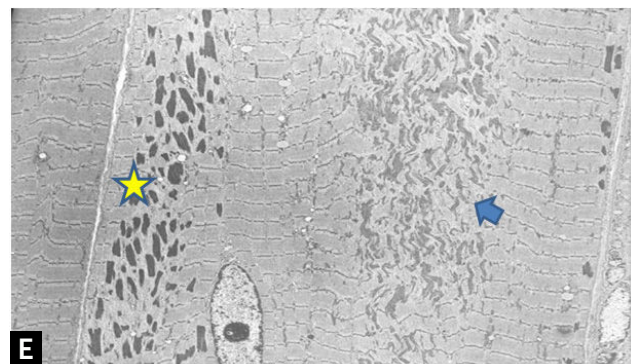
(C and D) Central core disease. (C) Centrally located cores evident as well-defined areas devoid of NADH-TR activity. (D) Electron micrograph of a core showing contraction of myofibrils and disruption of the Z-line.

The minicores are characterized by the presence of multiple foci of sarcomeric disorganization, with Z-line streaming, running over a few sarcomeres, even if occasionally they are longer with poorly-defined borders; mitochondria are absent from the altered areas¹⁷. Patients with minicores myopathy linked to AR mutations in selenoprotein N, *SEPN1* gene present with a constant clinical phenotype characterized by hypotonia and gross motor delay followed by prominent axial and diaphragmatic weakness associated with rigid spine, scoliosis, and respiratory failure¹⁸. Some patients showing ptosis, ophtalmoplegia and severe muscle weakness harbor AR mutations in the *RYR1* gene¹⁹⁻²⁰. Titin gene (*TTN*) mutations have been associated with congenital myopathy with contractures, possible cardiac involvement and cardiomyopathy and minicores in muscle biopsy²¹⁻²².

2.3 Core-rod myopathy

Core-rod myopathy shows the association of nemaline bodies with well-defined cores within separate muscle fiber regions (Fig.1E). Numerous core-rod myopathies are associated with autosomal dominant (AD) or autosomal recessive (AR) mutations in the gene encoding the skeletal muscle ryanodine receptor (*RYR1*)²³⁻²⁴. In contrast, only few clinically heterogeneous patients with core-rod myopathy linked to nebulin (*NEB*) gene mutation have been reported. Clinical features can correspond to typical congenital phenotype or consist of prominent distal weakness with bilateral foot-drop³.

Figure 1.



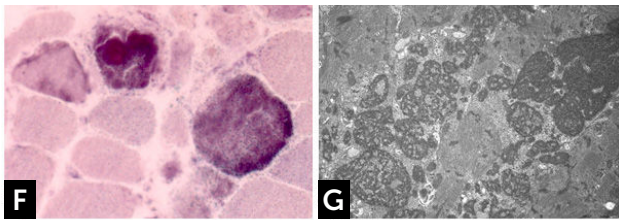
(E) Core-rod myopathy. Electron micrographs showing a fiber harboring both rods (indicated by a yellow star) and a core lesion (indicated by a blue arrow).

2.4 Reducing body myopathy

Reducing bodies were described around 40 years ago as cytoplasmic inclusions that reduce nitro-blue tetrazolium (NBT) and thus stain strongly with the menadione-NBT reaction²⁵. Pioneer studies speculated that reducing activity of the bodies is likely to be due to protein sulfhydryl groups. More recently, it has been demonstrated that RBs correspond to protein aggregates whose major constituent is FHL1²⁶. Mutations in *FHL1* gene (OMIM 300163) have been associated with reducing body myopathy (*RBM*), and other disorders²⁶.

The clinical phenotype varies from severe, congenital or early childhood onset disorders manifested with delayed motor milestones, prominent muscle weakness and rapid loss of ambulation, rigid spine, to milder, less progressive conditions manifested later in life²⁷. RBs are observed as bright pink cytoplasmic inclusions on H&E and red on the modified Gömöri trichrome stain. They strongly react for menadione-NTB with and without alpha-glycerolphosphate, (Fig.1F) but they are devoid of oxidative, especially SDH and COX, and ATPase activities. The number and the size of RBs can vary among the samples and some fibers are completely replaced by them. Cytoplasmic bodies are seen as collections of red granules in the vicinity of reducing bodies. By electron microscopy, RBs correspond to large inclusions composed by osmiophilic granular material (Fig. 1G). Immuno-electron microscopy studies showed that FHL1 is indeed found inside the RB and confirmed the expected FHL1 localization in skeletal muscle adjacent to the Z-line/band I²⁸.

Figure 1.



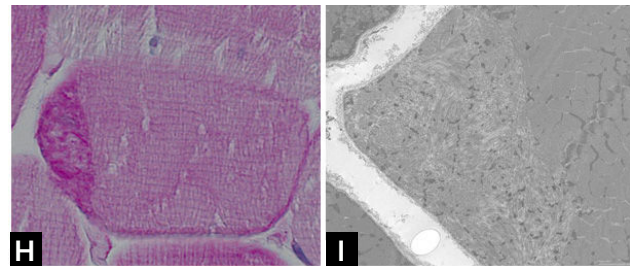
(F and G) Reducing body myopathy (RBM) due to *FHL1* mutations. (F) RBs stain blue strongly reducing NBT with menadione reaction without α -glycerolphosphate. (G) Electron microscopy. RB material corresponds to coarse tubulofilaments.

2.5 Cap myopathy

Cap myopathy is a rare congenital myopathy, characterized by the presence of peripherally-placed, well-delimited structures resembling a “cap”²⁹. Clinical features are variable with forms presenting like a typical congenital myopathies and others showing prominent respiratory involvement and marked maxillofacial deformations¹. Caps appear green and granular with Gömöri trichrome, stain strongly with PAS (Fig. 1H), and NADH, but they are pale with SDH. ATPases reactions do not stain the cap structures.

Electron microscopy confirms the presence of well-delimited structures containing fragments of sarcomeres, cellular debris, and amorphous material (Fig.1I). Mutations have been identified in the tropomyosin-2 (*TPM2*), in the tropomyosin-3 (*TPM3*), and more recently in the alpha-actin gene (*ACTA1*)². The association of caps and nemaline bodies in the same patient have been described by us in *TPM3*-related cap myopathy²⁹. We recently described a cohort of patients sharing a slowly progressive congenital cap myopathy with facial involvement and asymmetric weakness, and homozygous truncating mutations in *MYPN* gene, encoding myopalladin³⁰.

Figure 1.



(H and I) Cap disease; (H) Cap structure appearing as well-demarcated polar structures with intense PAS staining. (I) Electron micrograph of a Cap showing amorphous material with dispersed Z-line fragments.

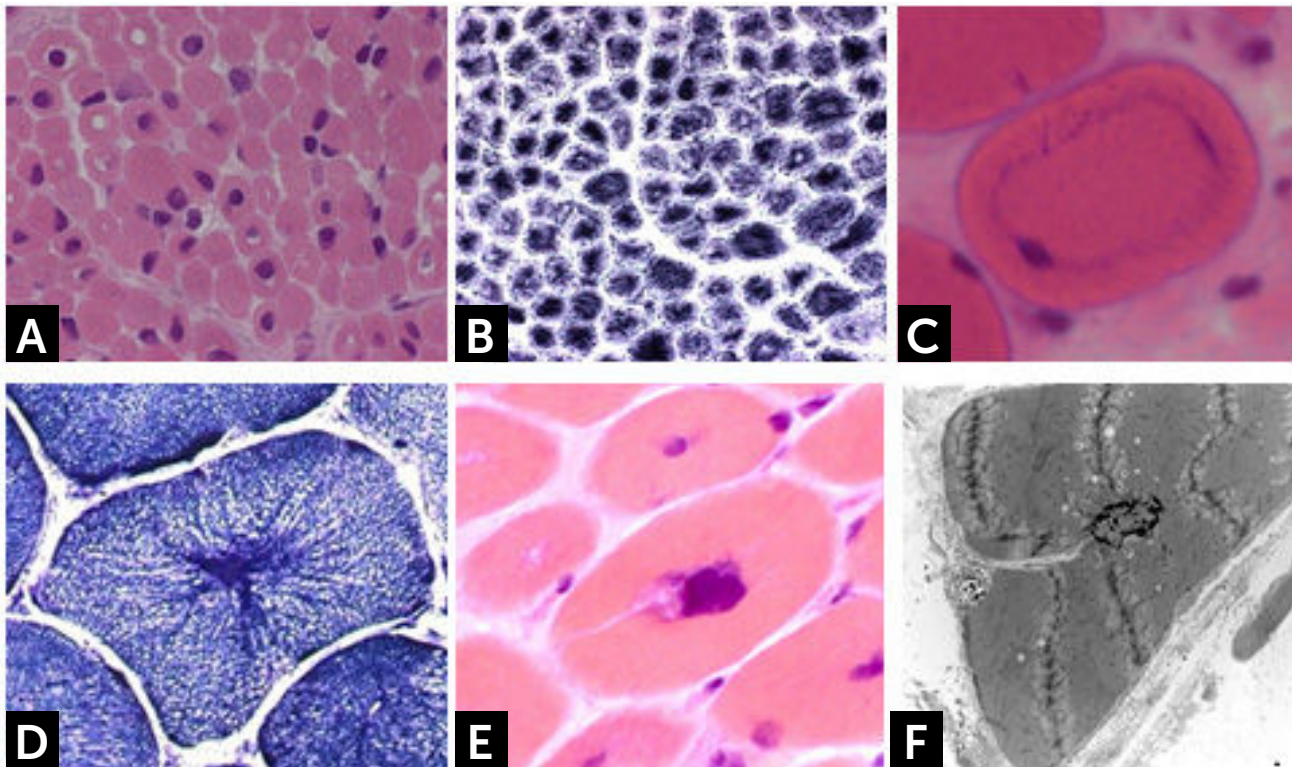
2.6 Centronuclear myopathies

Centronuclear congenital myopathies are genetic conditions characterized by the presence of a high incidence of centrally or internally placed nuclei in rows in muscle fibers³¹. Four forms have identified: (1) a severe/lethal X-linked recessive form, also named myotubular myopathy with mutation in the *MTM1* gene; (2) a clinically variable sporadic or AD form associated with *DNM2* gene mutation, and (3) a moderate or severe AR or AD form related to *BIN1* gene mutations. (4). Recently AR mutations in *SPEG* gene have been found in three patients presenting severe clinical form of congenital myopathy with dilated cardiomyopathy³².

Muscle biopsies of boys presenting the X-linked *MTM1* form show the presence of centrally located nuclei (Fig.2A), and a subsarcolemmal pale halo evident with oxidative enzymes (Fig.2B). Female carriers of one *MTM1* mutation may show mild muscle weakness and present a histopathologic findings consisting of basophilic loops found in the muscle fibers cytoplasm (Fig.2C). These structures resemble a necklace and show increased oxidative activity and PAS staining³¹.

Dynamin-2 (*DNM2*) gene, encodes for a large GTPase implicated in endocytosis and membrane trafficking. *DNM2* mutations are associated both to autosomal dominant (AD) forms of CNM (mild or late-childhood and adult onset forms) and sporadic cases (severe early childhood type). Muscle biopsies show a high percentage of nuclear internalizations located especially in small hypotrophic fibers. Numerous fibers show the “radiated sarcoplasmic strands” (RSS fibers), that correspond to a particular radiated disposition of the sarcoplasmic reticulum giving to the fiber the aspect of a bicycle wheel (Fig. 2D). Mutations in the amphiphysin 2 (*BIN1*) gene can be found both at AR state in severe neonatal or early-childhood type or at AD³³⁻³⁴ in the adult-onset progressive myopathy. Clusters of centrally located nuclei are found in a homogeneous population of rounded atrophic type 1 fibers. Connective tissue augmentation and fibro-adipose replacement without necrosis and regeneration are also seen. The presence of clus-

Figure 2.



(A). H&E staining of frozen muscle section from a patient presenting myotubular myopathy (*MTM1*). Presence of rounded muscle fibers with centrally located nuclei, resembling myotubes. (B) NADH reaction showing clear peripheral halo. (C) Necklace fiber in a biopsy from a female *MTM1* symptomatic carrier. (D) Particular radiated disposition of the sarcoplasmic reticulum as the "spokes of a wheel" in a *DNM2* mutated patient muscle. (E) Cluster of myonuclei and membrane invagination in a muscle biopsy from a patient with AR *BIN1* mutations. (F) Electron microscopy shows a centralized nucleus with membrane invagination.

ters of centrally located nuclei (Fig. 2E) surrounded by abundant amorphous material containing autophagic vacuoles and sarcolemmal membrane alterations consisting of invaginations, (Fig. 2F), vacuolization, and triads alterations are observed by electron microscopy (personal observation)³⁴.

3.7 Congenital fiber-type disproportion (CFTD)

CFTD is defined by the presence of smaller type 1 fibers (at least 12%) compared to type 2 fibers. Such change has been regarded for a long time as unspecific due to its presence in numerous congenital myopathies. Patients usually present with neonatal hypotonia variably associated with contractures. More recently, mutations in *TPM3*, *TPM2*, *RYR1*, *ACTA1*, *SEPN1*, and *MYH7* genes have been found in congenital myopathy patients showing CFTD without any other histopathological lesion².

3. DISEASES MECHANISMS

The mechanisms leading to muscle fiber degeneration and muscle weakness in congenital myopathies are largely

unknown. Regarding the group of nemaline myopathies, all the known defective protein in these conditions are sarcomeric structural protein such as nebulin, actin, tropomyosins or myopalladin, thin filaments Kelch associated proteins, or the non-conventional myosin 18 B protein, found in the proximity of the Z line. All this proteins belong to the sarcomere, the constitutive myofibrillar unit necessary for muscle contraction. We can compare the sarcomere as a highly organized structure resembling the pentagram of a music partition. Thus, it is possible that a structural sarcomeric modification provoked by a genetic alteration could lead to its disarray. In the case of nemaline myopathies the sarcomeric derangement could lead to nemaline bodies accumulation.

Fine morphological studied conducted by us on muscle samples from patients with NEB mutations showed a positive correlation between clinical severity and myofibrillar smallness and disruption associated with rods deposition in severe/lethal cases¹⁰. Other studies measuring single fibers contractility in muscle biopsies from patients with different nemaline myopa-

thies confirmed that the contractile alteration is related to the shortness of thin filaments³⁴.

In the case of core myopathies linked to *RYR1* gene mutations, the possible pathogenic mechanisms could be related to calcium mishandling/toxicity provoked by altered *RYR1* channel function¹⁴. However, the relation between calcium mishandling and core lesion formation remains to be clarified.

In the centronuclear myopathy group, a mechanism of altered development of skeletal muscle fibers have been hypothesized in the myotubular myopathy, while *DNM2* and *amphysisin 2* seem to be involved in the intracellular trafficking³¹.

4. CONCLUSIONS

In this article, we reviewed main clinical, morphological and genetic features of the main congenital myopathies. It is possible that other congenital myopathies forms will be discovered in future years with the application of more advanced sequencing approaches such as RNA sequencing. The future challenge for clinicians, researchers and patients will be the development of therapeutic approach. With this in mind, a deep clinical, morphological, and genetic phenotyping are and remain essential steps for the understanding of this group of myopathies.

Declaration of interest:

The author has nothing to disclose

REFERENCES

- Malfatti E, Romero NB. Diseases of the skeletal muscle. *Handb Clin Neurol*. 2017;145:429-451.
- Romero NB, Clarke NF. Congenital myopathies. *Handb Clin Neurol*. 2013;113:1321-36.
- Malfatti E, Monges S, Lehtokari VL, et al. . Bilateral foot-drop as predominant symptom in nebulin (*NEB*) gene related "core-rod" congenital myopathy. *Eur J Med Genet* Oct;58(10):556-61.
- Natera-de Benito D, Nascimento A, Abicht A, et al. *KLHL40*-related nemaline myopathy with a sustained, positive response to treatment with acetylcholinesterase inhibitors. *J Neurol*. 2016 Mar;263(3):517-23.
- Quijano-Roy S, Carlier RY. Muscle magnetic resonance imaging: a new diagnostic tool with promising avenues in therapeutic trials. *Neuropediatrics*. 2014 Oct;45(5):273-4
- Kaplan JC, Hamroun D. The 2016 version of the gene table of monogenic neuromuscular disorders (nuclear genome). *Neuromuscul Disord*. 2016 Apr-May;26(4-5):330.
- Wang CH, Dowling JJ, North K et al. Consensus statement on standard of care for congenital myopathies. *J Child Neurol*. 2012 March; 27 (3): 363-382.
- Malfatti E, Romero NB. Nemaline myopathies: State of the art. *Rev Neurol (Paris)*. 2016 Oct;172(10):614-619.
- Miyatake S, Mitsuhashi S, Hayashi YK, et al. Biallelic Mutations in *MYPN*, Encoding Myopalladin, Are Associated with Childhood-Onset, Slowly Progressive Nemaline Myopathy. *Am J Hum Genet*. 2017 Jan 05;100(1):169-78.
- Malfatti E, Lehtokari VL, Böhm J et al . Muscle histopathology in nebulin-related nemaline myopathy: ultrastructural findings correlated to disease severity and genotype. *Acta Neuropathol Commun*. 2014 Apr 12;2:44.
- Nowak KJ, Ravenscroft G, Laing NG. Skeletal muscle alpha-actin diseases (actinopathies): pathology and mechanisms. *Acta Neuropathol* 2013 Jan;125(1):19-32.
- Malfatti E, Böhm J, Lacène E et al. A premature stop codon in *MYO18B* is associated with severe nemaline myopathy with cardiomyopathy. 2015. *J Neuromuscular Diseases* 2015. 2(3) 2015; p.219-227
- Fardeau M. 1982. Congenital myopathies . Pp. 161-203 in F.L. Mastaglia and S.J. Walton, eds. *Skeletal muscle pathology*. Churchill Livingstone, London.
- Maggi L, Scoto M, Cirak S, et al. Congenital myopathies--clinical features and frequency of individual subtypes diagnosed over a 5-year period in the United Kingdom. *Neuromuscular disorders* 2013 Mar;23(3):195-205
- Romero NB, Xie T, Malfatti E et al. Autosomal dominant eccentric core disease caused by a heterozygous mutation in the *MYH7* gene. *J Neurol Neurosurg Psychiatry* 2014 85(10):1149-52.
- Majczenko K, Davidson AE, Camelo-Piragua S, et al Dominant mutation of *CCDC78* in a unique congenital myopathy with prominent internal nuclei and atypical cores. *Am J Hum Genet*. Aug 2012 10;91(2):365-71.
- Jungbluth H. Multi-minicores disease. *Orphanet J Rare Dis*. 2007; 2:31.
- Ferreiro A, Quijano-Roy S, Pichereau C et al. Mutations of the selenoprotein *N* gene, which is implicated in rigid spine muscular dystrophy, cause the classical phenotype of multiminiore disease: reassessing the nosology of early-onset myopathies. *Am J Hum Genet* 2002a 71:739-749.
- Ferreiro A, Monnier N, Romero NB et al. A recessive form of central core disease, transiently presenting multi-minicores disease, is associated with a homozygous mutation in the ryanodine receptor type 1 gene. *Ann Neurol* 2002b 51: 750-759.
- Jungbluth H, Zhou H, Hartley L et al. Miniore myopathy with ophthalmoplegia caused by mutations in the ryanodine receptor type 1 gene. *Neurology* 2005 65: 1930-1935.
- Carmignac V, Salih MA, Quijano-Roy S et al. C-terminal titin deletions cause a novel early onset myopathy with fatal cardiomyopathy. *Ann*

- Neurol 2007 61:340-351.
22. Chauveau C, Bonnemann CG, Julien C et al. Recessive TTN truncating mutations define novel forms of core myopathy with heart disease. *Hum Mol Genet* 2013 23(4):980-91.
 23. Monnier N, Romero NB, Lemale J et al. An autosomal dominant congenital myopathy with cores and rods is associated with neomutation in RYR1 gene encoding the skeletal muscle ryanodine receptor. *Hum Mol Genet* 2000 9: 2599-2608.
 24. Hernandez-Lain A, Husson I, Monnier N et al. De novo RYR1 heterozygous mutations (I4898T) causing lethal core-rod myopathy in twins. *EJMG* 2011 54: 29-33.
 25. Brooke MH and Neville HE. Reducing body myopathy. *Neurology* 1972 ; 22: 829-40.
 26. Schessl J, Zou Y, McGrath MJ, et al. Proteomic identification of FHL1 as the protein mutated in human reducing body myopathy. *J Clin Invest* 2008 ; 118: 904-12.
 27. Schessl J, Taratuto AL, Sewry C, et al. Clinical, histological and genetic characterization of reducing body myopathy caused by mutations in FHL1. *Brain* 2009 ; 132:452-64.
 28. Malfatti E, Olivé M, Taratuto AL, et al. (2013). Skeletal muscle biopsy analysis in reducing body myopathy and other FHL1-related disorders. *J Neuropathol Exp Neurol.* 2013 Sep;72(9):833-45.
 29. Malfatti E, Schaeffer U, Chapon F, Yang Y, Eymard B, Xu R, Laporte J, Romero NB.
Combined cap disease and nemaline myopathy in the same patient caused by an autosomal dominant mutation in the TPM3 gene. Neuromuscul Disord. 2013 Dec;23(12):992-7.
 30. Lornage X, Malfatti E, Chéraud C, et al. Recessive MYPN mutations cause cap myopathy with occasional nemaline rods. *Ann Neurol.* 2017 Mar;81(3):467-473.
 31. Romero NB Centronuclear myopathies: a widening concept. *Neuromuscul Disord.* 2010 20(4):223-8.
 32. Agrawal PB, Pierson CR, Joshi M et al. SPEG interacts with myotubularin, and its deficiency causes centronuclear myopathy with dilated cardiomyopathy. *Am J Hum Genet* 2014 7;95(2):218-26.
 33. Nicot AS, Toussaint A, Tosch V et al. Mutations in amphiphysin 2 (BIN1) disrupt interaction with dynamin 2 and coause autosomal recessive centronuclear myopathy. *Nat Gene* 2007. 33:1134-1139
 33. Böhm J, Biancalana V, Malfatti E, et al. Adult-onset autosomal dominant centronuclear myopathy due to BIN1 mutations. *Brain* 2014 137(Pt 12):3160-70.
 34. Winter JM, Joureau B, Lee EJ et al. Mutation-specific effects on thin filament length in thin filament myopathy. *Ann Neurol.* 2016 Jun;79(6):959-69.