



Review

Functional analysis of the MAPK pathways in fungi



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ABSTRACT

The Mitogen-Activated Protein Kinase (MAPK) signaling pathways constitute one of the most important and evolutionarily conserved mechanisms for the perception of extracellular information in all the eukaryotic organisms. The MAPK pathways are involved in the transfer to the cell of the information perceived from extracellular stimuli, with the final outcome of activation of different transcription factors that regulate gene expression in response to them. In all species of fungi, the MAPK pathways have important roles in their physiology and development; e.g. cell cycle control, mating, morphogenesis, response to different stresses, resistance to UV radiation and to temperature changes, cell wall assembly and integrity, degradation of cellular organelles, virulence, cell–cell signaling, fungus–plant interaction, and response to damage-associated molecular patterns (DAMPs).

Considering the importance of the phylogenetically conserved MAPK pathways in fungi, an updated review of the knowledge on them is discussed in this article. This information reveals their importance, their distribution in fungal species evolutionarily distant and with different lifestyles, their organization and function, and the interactions occurring between different MAPK pathways, and with other signaling pathways, for the regulation of the most complex cellular processes.

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Análisis funcional de las vías MAPK de los hongos

RESUMEN

Las vías de señalización de la proteína-cinasa activada por mitógenos (abreviadas como MAPK por sus siglas en inglés) son uno de los mecanismos más importantes y evolutivamente conservados para la percepción de información extracelular en organismos eucarióticos. Las vías MAPK están involucradas en la transferencia a la célula de la información recibida de estímulos extracelulares, que ofrecen como resultado final la activación de diferentes factores de transcripción que regulan la expresión de genes en respuesta a aquellos. En todas las especies de hongos, las vías MAPK tienen importantes funciones en su fisiología y desarrollo como, por ejemplo, el control del ciclo celular, el apareamiento, la morfogénesis, la respuesta a diferentes tipos de estrés, la resistencia a la luz UV y a los cambios de temperatura, la formación e integridad de la pared celular, la degradación de los orgánulos, la virulencia, la señalización célula-célula, la interacción hongo-planta y la respuesta a patrones moleculares asociados con el daño (abreviado como DAMP, por sus siglas en inglés).

Dada la importancia de las vías MAPK en hongos, en esta revisión se discute el conocimiento adquirido más recientemente sobre ellas. Esta información revela su importancia, su distribución en especies de hongos evolutivamente distantes y con estilos de vida diferentes, su organización y función, y las interacciones que ocurren entre diferentes vías MAPK, y entre estas y otras vías de señalización que regulan los procesos celulares más complejos.

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Mitogen-activated protein kinase (MAPK) pathways are one of the most important and evolutionarily conserved mechanisms of cellular signaling existing in eukaryotic organisms including animals, plants and fungi.^{23,79} The signal transduction processes in which MAP kinases are involved start with the sensing of environmental stimuli by receptors and proteins anchored to the cell membrane, such as the two-component signal transduction systems (TCS), receptor tyrosine kinases (RTKs) or multiprotein structures such as eisosomes, which in turn are attached to heterotrimeric or monomeric G proteins-coupled to receptors.^{3,23,39,50,99} These can interact with adaptor proteins, or directly activate a MAPKKK (MAP kinase kinase kinase) which in turn activates a MAPKK (MAP kinase kinase) by the phosphorylation of serine/threonine residues. This latter protein phosphorylates one or several MAPKs (MAP kinases) in serine/threonine/tyrosine residues, that finally give rise to the activation of transcription factors that induce or repress genes involved in the cellular adaptation or response to the sensed stimuli.^{6,32} In some of these pathways a scaffold (anchor) protein keeps associated with the different MAPKs.^{41,53,78}

Fungi are eukaryotic organisms with different lifestyles that possess protein kinases, including those organized in the form of MAPK systems, with high homology to kinases from animals, such as flies, worms, and humans.^{23,51} In fungi, the MAPK pathways are involved in different physiological and developmental processes, including cell cycle, mating, morphogenesis, sporulation, cell wall assembly and integrity, autophagy, pathogenesis, UV and heat-shock resistance, cell-cell signaling, fungus-fungus interactions, fungus-plant interactions (e.g. mycorrhiza), response to different forms of stress, response to damage-associated molecular patterns (DAMPs), etc.^{3,21,23,25,29,32,41,49,52,64,79}

Taking into consideration that some of these processes occur in higher eukaryotic organisms, involving also the action of MAPK modules, it may be concluded that fungi constitute excellent model organisms for the study and understanding of the mechanisms that operate in the signaling systems occurring in eukaryotic organisms in general. On these bases, in this review we analyze the functions of the MAPK pathways, the interactions between different MAPK pathways, and their interaction with other signaling pathways occurring in fungal species evolutionarily distant, and with different lifestyles.

An overview of the MAPK pathways in fungi

As described above, MAPK pathways are very important fungal systems involved in many physiological and developmental processes, stress response, virulence, interaction with other organisms, etc. They are generally conserved in all the species studied thus far and they have a very similar organization and functions^{3,21,23,25,29,32,41,49,52,64,79,93} (Fig. 1). The saprophytic yeast *Saccharomyces cerevisiae* and the human pathogen yeast *Candida albicans* gather the most information available about the fungal MAPK pathways. Indeed, *S. cerevisiae* was the first organism where genes related to sensing signals of mating were cloned.³ These genes were named *STE2* and *STE3* because their mutation caused sterility.⁶⁹ The MAPKs characterized in *S. cerevisiae* have homologues in many fungi belonging to different divisions, and with different lifestyles (e.g. see Table 1). In some of these fungi, MAPKs, and even the complete MAPK pathways, have been also characterized by molecular or biochemical studies, or predicted by bioinformatic analysis.

The existence of sensing proteins, such as Sln1, Sho1, Msb2, Opy2, Snf1, and even phytochrome (FphA), auxiliary proteins that interact with the external sensors Wsc, Gpr1, Ypd1, Ssk1, Cdc42, Bem4, and that subsequently activate the corresponding MAPK

pathways, have been described in different fungal species, such as *S. cerevisiae*, *C. albicans*, *Cryptococcus neoformans*, *Aspergillus nidulans*, *Fusarium graminearum*, *Fusarium oxysporum*, *Verticillium dahliae*, *Parastagonospora nodorum*, *Beuveria bassiana*, *Kluyveromyces lactis*, *Botrytis cinerea*, *Magnaporthe oryzae*, *Ustilago maydis*, etc.^{9,17,30,31,36,38,42,43,45,48,60,66,74,75,83,86,88–90,93}

These sensory proteins and auxiliary proteins form the two-component signal transduction (TCS) system, which together with G protein-coupled receptors (GPCRs), have been described in fungi to be involved in the signal transfer from the extracellular medium to the MAPK core, mainly under conditions of stress and virulence (see reviews in Rispail et al.⁷⁹; Velázquez-Zavala et al.⁸⁹; Ma and Li⁵⁰; Hagiwara et al.³²; Kou et al.⁴¹; Alvaro and Thorner³). These signal transduction mechanisms upstream of the MAPK core are discussed below.

The MAPK pathways not always regulate the same processes or induce the same cellular responses in the different fungal species. For example, in *C. albicans* the cell wall integrity (CWI) pathway is an important virulence factor,² but in contrast its homologue CWI pathway in the Basidiomycota phytopathogenic fungus *U. maydis* is only involved in sensing damage in the cell wall, forcing the cell to escape from the G2 phase of the cell cycle.¹³ This phenomenon occurs similarly in *S. cerevisiae*, with the difference that in the latter the homologue MAPK pathway induces a cell cycle arrest at the G2 phase when its cell wall is damaged.¹³

Mechanisms of transfer of the environmental signals through the MAPK pathway, and their connection with the downstream components

Signal transfer to MAPK pathways by the two-component signal transduction systems, and G-proteins coupled to receptors

The two-component signal transduction (TCS) system (originally described in bacteria, but now known to be present also in fungi and plants,^{32,50,73,79,86,90} and not in animals.⁵⁰), and G-proteins coupled receptors (GPCRs), are known to be the main mechanisms involved in receiving extracellular signals and in the further activation of MAPK or other pathways. In fungi, this system is involved in several processes including development, for example: osmotic and oxidative stress, cell and sexual cycle regulation, virulence, etc.^{32,50,74,90} In these organisms the TCS system is made by three components or signal transducers present in one or more copies: a histidine kinase (HK), a response regulator (RR), and a histidine-containing phospho-transmitter (HPT), that in turn phosphorylates the MAPKKK of the corresponding MAPK core.³²

Heterotrimeric G protein signaling occurs by its activation through a membrane G protein-coupled receptor (GPCRs). This occurs during the perception of an extracellular stimulus, in which the GPCR undergoes changes in its conformation, giving rise to the dissociation of the G proteins into a dimer, Gβ-Gγ, and a monomer, GTP-Gα. These components act downstream interacting with protein kinases which subsequently phosphorylate the MAPKKK of the MAPK core. After MAPK protein activation occurs, GTP bound to Gα is hydrolyzed, and re-association with the Gβ-Gγ heterodimer takes place.^{8,22,41,47,73,76}

Transfer of the signal received through the MAPK core

The extracellular signal perceived by receptors, and transmitted by the mechanisms described above is finally received by the MAPKKK protein. The activation of MAPK protein occurs by phosphorylation of specific amino acid residues. MAPKKK activates the MAPKK protein, and this protein in turn activates the MAPK protein, which finally activates transcription factors involved in the

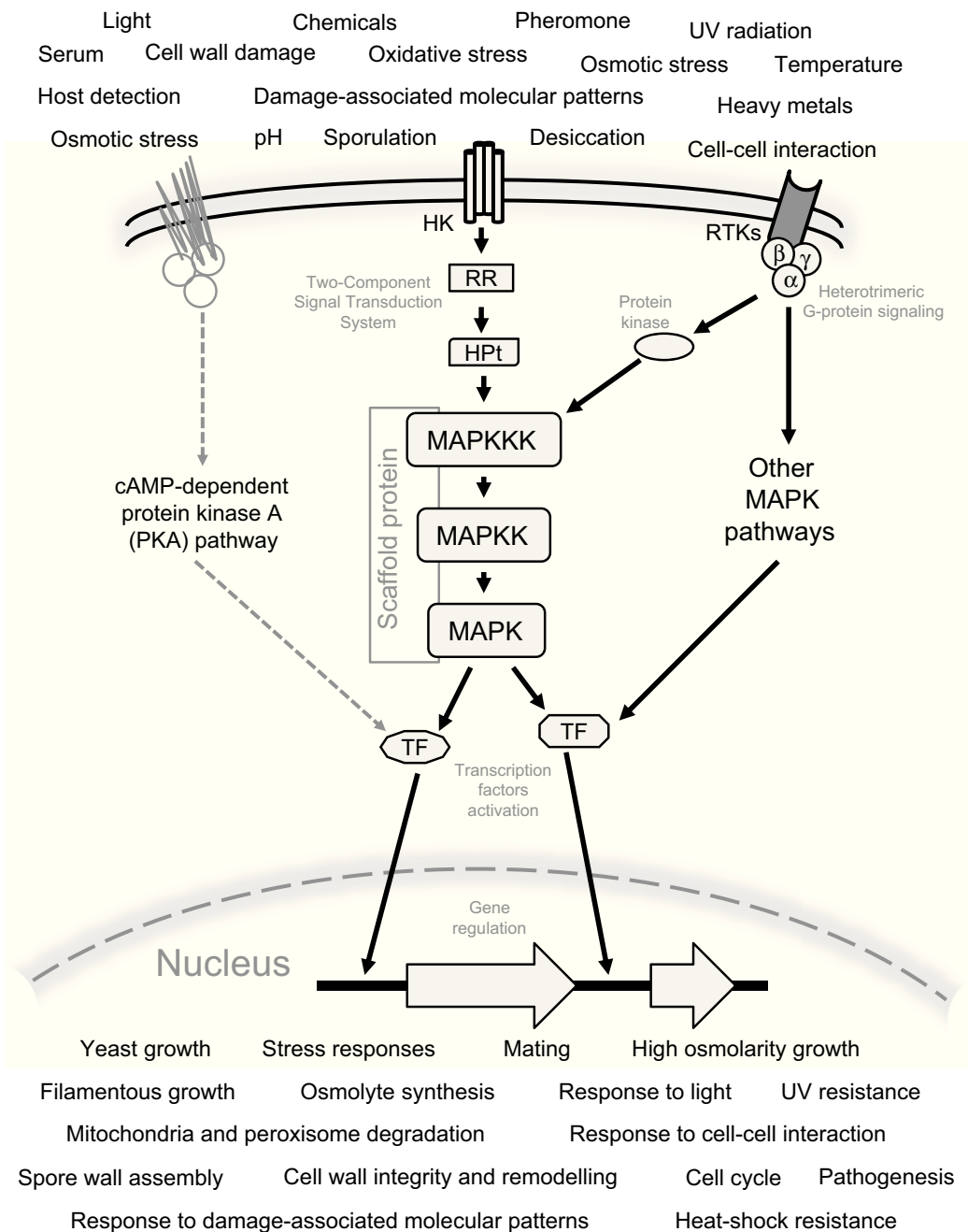


Fig. 1. General schematic representation of the signaling mechanism by the MAPK pathways in fungi. All possible sensed signals and cellular responses to these signals are included. Also the Two-Component Signal Transduction (TCS) system, G protein-coupled receptors, receptor tyrosine kinases (RTKs), and the interaction of MAPK with other signaling pathways, e.g. the cAMP-dependent protein kinase A (PKA) pathway are represented. The Scaffold protein is generally present in the MAPK pathway involved in mating. The following abbreviations are used: MAPKKK, MAP kinase kinase kinase; MAPKK, MAP kinase kinase; MAPK, MAP kinase; TF, transcription factors; HK, histidine kinase; RR, response regulator; HPt, histidine-containing phospho-transmitter.

transcriptional regulation of cellular response.^{6,32} MAPK proteins have the characteristic domain S.TKc (serine/threonine protein kinase), as well as ATP binding sites, and a phosphotransferase domain. It should be noticed that MAPKKKs also have SAM domains (sterile alpha motif domains involved in protein interaction and signal transduction), and Ras.bdg.2 domains (domains involved in its interaction with the Ras G proteins that allow the transfer of the perceived signals by trans-membrane receptors). These catalytic domains are generally conserved in the MAPKs of different fungal species, and their high homology is an evidence that they are involved in the same physiological phenomena and, accordingly, receive a similar name (Table 2). For example: Kss1 is involved

in filamentous growth; Fus3, in pheromone response and mating; Hog1, in osmotic and oxidative stress response; Slt2/Mpk1, in cell wall integrity; etc.^{2,3,9,32,41,50,73} Under this idea, Hog1, the principal component of the high-osmolarity glycerol (HOG) pathway is probably the most conserved MAPK protein in fungal species, and has high similarity mainly in the regions coding for their functional domains: STKc.Sty1.Hog1, catalytic domain; S.TKc, serine/threonine protein kinase; phosphotransferase; and ATP binding site (Fig. 2). Recently, in addition to the indispensable role of the HOG pathway in response to stress and osmosensing, an additional function was described in *A. nidulans*.⁹¹ Accordingly, it was described that this pathway was involved in the response to light

Table 1
MAPK components described in *Saccharomyces cerevisiae*, and their possible homologues in representative fungi of different phyla.

Protein	Ascomycota			Basidiomycota		Mucoromycota
	Name in <i>Saccharomyces cerevisiae</i>	<i>Candida albicans</i>	<i>Yarrowia lipolytica</i>	<i>Ustilago maydis</i>	<i>Cryptococcus neoformans</i>	<i>Rhizopus oryzae</i>
MAPKKK	Ste11	CAWG_04138 [59.6% (1.0 E-105)]	YALIOF13629g [56.1% (3.28 E-125)]	UM04258 [58.0% (2.54 E-102)]	Cryne_H99.1 6280 [57.0% (6.39 E-098)]	RO3G_14758 [51.4% (5.30 E-102)]
	Ssk2	CAWG_03313 [41.3% (0.0 E-000)]	YALIOA05247g [46.9% (0.00 E-000)]	UM01544 [47.3% (3.37 E-143)]	Cryne_H99.1 8685 [41.2% (5.68 E-132)]	RO3G_01661 [42.8% (2.72 E-152)]
	Ssk22	CAWG_03313 [40.9% (0.0 E-000)]	YALIOA05247g [46.8% (0.00 E-000)]	UM01544 [44.2% (3.24 E-137)]	Cryne_H99.1 8685 [43.5% (7.29 E-107)]	RO3G_01661 [45.2% (4.14 E-143)]
	Bck1	CAWG_04138 [44.9% (4.0 E-65)]	YALIOF08855g [53.3% (3.01 E-139)]	UM01662 [61.0% (1.25 E-099)]	Cryne_H99.1 11167 [54.6% (1.64 E-072)]	RO3G_07651 [53.7% (9.61 E-073)]
MAPKK	Ste7	CAWG_01727 [39.3% (5.0 E-61)]	YALIOB15906g [46.2% (3.97 E-033)]	UM01514 [55.9% (1.39 E-057)]	Cryne_H99.1 1550 [56.7% (2.32 E-051)]	RO3G_01555 [57.1% (1.94 E-056)]
	Pbs2	CAWG_02916 [64.0% (8.0 E-144)]	YALIOB15906g [69.3% (3.46 E-135)]	UM06342 [63.7% (2.31 E-096)]	Cryne_H99.1 676 [59.6% (1.95 E-101)]	RO3G_04837 [63.3% (9.04 E-111)]
	Mkk1	CAWG_04324 [50.5% (1.0 E-82)]	YALIOB13178g [66.4% (1.16 E-120)]	UM04864 [61.0% (1.41 E-084)]	Cryne_H99.1 9585 [51.0% (3.06 E-068)]	RO3G_02519 [48.2% (7.44 E-051)]
	Mkk2	CAWG_04324 [50.5% (6.0 E-84)]	YALIOB13178g [62.2% (1.80 E-119)]	UM04864 [58.0% (3.75 E-086)]	Cryne_H99.1 9585 [52.6% (3.46 E-071)]	RO3G_02519 [47.0% (1.32 E-048)]
MAPK	Fus3	CAWG_03179 [59.0% (5.0 E-124)]	YALIOE23496g [63.3% (1.12 E-145)]	UM03305 [60.1% (1.58 E-141)]	Cryne_H99.1 2279 [56.0% (1.80 E-124)]	RO3G_16338 [62.2% (1.92 E-144)]
	Kss1	CAWG_03179 [57.1% (4.0 E-123)]	YALIOE23496g [63.6% (1.11 E-099)]	UM03305 [67.8% (2.89 E-124)]	Cryne_H99.1 2261 [66.8% (1.03 E-120)]	RO3G_16338 [64.3% (1.66 E-102)]
	Hog1	CAWG_04100 [79.0% (0.0 E-000)]	YALIOE25135g [85.0% (0.00 E-000)]	UM02357 [82.0% (0.00 E-000)]	Cryne_H99.1 11389 [81.0% (0.00 E-000)]	RO3G_12796 [81.7% (0.00 E-000)]
	Slt2/Mpk1	CAWG_01373 [54.7% (5.0 E-153)]	YALIOB02816g [72.9% (0.00 E-000)]	UM00421 [70.0% (3.17 E-134)]	Cryne_H99.1 4067 [60.4% (8.75 E-100)]	RO3G_05692 [66.5% (3.76 E-143)]
	Smk1	CAWG_00052 [53.9% (6.0 E-105)]	YALIOB02816g [48.7% (9.21 E-098)]	UM00421 [49.5% (2.57 E-076)]	Cryne_H99.1 4067 [45.7% (6.61 E-087)]	RO3G_05692 [50.2% (1.15 E-087)]

sensed by phytochrome FphA, and its interaction with protein phosphotransferase YdpA.

Transcription factors downstream of MAPK pathways

The last part in the transduction of extracellular signals by the MAPK pathways is the activation by phosphorylation of downstream master transcription factors, which in turn regulate the transcription of other transcription factors and other genes involved in the response to the extracellular signal sensed. Interestingly, the transcription factors are one of the most important points of interaction and interconnection between different MAPK pathways, as well as in other signaling pathways. An excellent example of all the aspects described above is the *U. maydis* transcription factor Prf1. This transcription factor regulates two fundamental processes in this fungus, mating and the pathogenic process, and it can be activated by both the cAMP-dependent protein kinase A (PKA) pathway (during mating) and MAPK PMM pathway (during the pathogenic process) (revised by Brefort et al.⁸).

Throughout the study of MAPK and other pathways in fungi, different transcription factors involved in transcriptional regulation of cellular responses have been described. For example, as previously mentioned, Yap2 is the point of interconnection between *S. cerevisiae* CWI and HOG pathways⁶³; in *A. nidulans*, AtfA is involved in the response to stress³⁷; in *U. maydis*, the role

of PacC/Rim101 as a crossover point between CWI, HOG, PMM and Pal/Rim pathways during response to pH and dimorphism have been suggested.^{24,58,59} Also in *C. albicans*, the transcription factors Cph1, Efg1 and Tec1 activated by the Cek1 MAPK pathway, regulate filamentous growth and virulence.^{9,96}

Likewise, the downstream activation by MAPK pathways of the transcription factors Sst2, Bni1, Far1, Ste12, Sko1, Rck1, Rck2, Msn2, Msn4, Hot1, Smp1, Rlm1, Mcm1, Msg5, Sdp1, Pir3, Mbp1, Swi6, Swi4, Fks2, Ppz1, Ppz2, Smp1, Far1, Flo8, Sfl1, Sbf1, NapA, Pap1, etc., has been described as important or indispensable for many important physiological processes in several and different fungi.^{32,79,96}

Finally, it is important to mention that also an epigenetic regulation has been suggested for some of the transcription factors described here, and therefore the cellular processes that they regulate.^{55,73}

Interaction between MAPK pathways, or with other signaling pathways

Several MAPK pathways may be involved in more than one cellular process

In *C. albicans* the already mentioned HOG pathway, described to be involved in the adaptation to high osmolarity stress, interacts with CWI pathway, and they together are involved in

Table 2
MAPK pathways described in phylogenetically distant fungi and with different lifestyles.

Phylum	Fungus	Lifestyle	MAPK pathways described	Response to	Involved in	
Ascomycota	<i>Candida albicans</i>	Animal pathogen	Cek1	Cell wall damage	Cell wall construction, filamentous growth	
			Hog1	Osmotic stress, oxidative stress, UV radiation, serum, heavy metals, chemical compounds	Adaptation to stress, UV resistance, heat-shock resistance	
			Mkc1	Cell wall damage	Cell wall integrity and remodeling, virulence	
	<i>Saccharomyces cerevisiae</i>	Saprophytic	Cek2	Stress	Growth	
			Fus3	Pheromone response	Mating, cell cycle	
			Kss1	Morphological switch, starvation	Pseudomycelium formation	
			Smk1 Hog1	Sporulation Osmotic stress and oxidative stress	Spore wall assembly High osmolarity growth, mitochondria degradation	
	<i>Yarrowia lipolytica</i>	Saprophytic	Fus3	Slr2/Mpk1	Cell wall stress, nutrition, temperature	Cell wall integrity and remodeling, mitochondria and peroxisomes degradation
				Fus3	Pheromone response	Mating, cell cycle, filamentous growth
			Stl2	Cell wall stress	Cell wall remodeling, cell cycle	
Hog1			High osmolarity	Osmolyte synthesis, cell cycle		
Kss1			Starvation	Filamentous growth		
Basidiomycota	<i>Ustilago maydis</i>	Plant pathogen	PMM	Pheromone response, plant invasion	Pathogenesis, mating, morphogenesis, cell cycle	
			CWI	Cell wall stress	Cell wall integrity, cell cycle	
	<i>Cryptococcus neoformans</i>	Animal pathogen	Hog1	Osmotic stress	High osmolarity growth	
			Cpk1	Pheromone response	Mating	
Mucoromycota	<i>Rhizopus oryzae</i>	Animal and plant pathogen	Hog1	Desiccation, osmotic shock	Stress responses	
			Mpk1	Cell wall perturbation	Cell wall integrity	
			Fus3	Pheromone response	Mating	
			Kss1	Starvation	Filamentous invasion	
			Hog1	High osmolarity	High osmolarity growth	
			Mpk1	Hypotonic shock	Cell wall integrity	

The corresponding citation appears in the text.

morphogenesis, integrity of the cell wall, growth, response to stress, and virulence^{35,77,80}; it was revised by Brown et al.⁹ These interactions were confirmed by the mutation of the upstream components (*SHO1*, *SLN1*, *YDP1*, *SSK1*, *MSB2*, *OPY2*) of these two pathways (HOG or CWI). The mutant strains were avirulent in mice and *Galleria mellonella* (the greater wax moth or honeycomb moth³⁵); a different behavior and susceptible phenotype to different types of stress was also shown.⁸⁸ The same behavior was also found in the Ascomycota fungus *B. bassiana*, a fungal pathogen of insects.⁹⁰ An important evidence of interaction between MAPK pathways is the TCS system, since in general the sensors and proteins belonging to this system can interact with more than one MAPK pathway, as demonstrated in different fungi, such as *S. cerevisiae*, *C. albicans*, *B. bassiana*, *V. dahliae*, *F. oxysporum*, *B. cinerea*, *K. lactis*, *Aspergillus fumigatus*, etc.,^{17,33,45,74,75,86,88–90} revised in Ma and Li,⁵⁰ and Higawara et al.³³

In fungi, the interaction among proteins belonging to different MAPK pathways, apart from the interaction of different whole MAPK pathways, may take place. Thus, the *S. cerevisiae* MAPK Ste7 interacts generally with Fus3 in the mating reaction of the yeast, but also with Kss1 for pseudomycelium formation. Similarly, the MAPK Ste11 interacts with Ste7 for mating, but also with Pbs2 during osmotic stress.⁹⁶ The same cross interactions take place in other fungi, e.g. *C. albicans*, *U. maydis*, and *Rhizopus oryzae*.⁷⁹ Also in *S. cerevisiae* the interaction between CWI and HOG pathways has been suggested by the existence of the same transcription factor in both pathways, Yap2.⁶³ In *A. nidulans*, the MAPK SakA and the transcription factor AtfA are components of multiple signaling pathways involved in response to stress and in development (e.g.

mating, DNA damage response, mRNA stability, protein synthesis, cell cycle regulation, and mitochondrial function).³⁷ Besides, in *Fusarium verticillioides* the MAPK pathway FvBCK1 involved in growth and development is also involved in cell wall biogenesis and in the response to osmotic and oxidative stresses,⁹⁴ suggesting the interconnection of this MAPK pathway (growth and development) with other MAPK pathways present in this fungus (CWI and HOG).

More than one MAPK pathway is involved in a single phenomenon

The HOG pathway (Hog1) is associated with the CWI pathway (Mkc1) (described to be involved in the regulation of glucan and chitin synthesis^{71,72}) and the filamentous growth pathway (Cek1) (involved in the regulation of genes encoding protein-O-mannosyltransferases¹²), to maintain the integrity of the cell wall of *C. albicans*.⁹ Under the same concept, in *S. cerevisiae* it has been described the requirement of two MAPK pathways for the process of autophagy: CWI (Stl2) and HOG (Hog1) pathways, involved in the degradation of mitochondria, although the *STL2* gene is also independently involved in the degradation of peroxisomes.⁵² Again in *S. cerevisiae*, the CWI, HOG and filamentous growth pathways are together involved in the response to stress.²⁷

Interaction of MAPK pathways with other signaling pathways

In addition to these interactions among the MAPK pathways, the interaction of MAPK pathways with other signal transduction pathways has been also described in fungi. In *U. maydis*, it has

Table 3
Interaction of the MAPK pathways with other signaling pathways in different fungi.

Fungus	Signaling pathways involved	Phenomenon	Reference
<i>Saccharomyces cerevisiae</i>	Filamentous growth (MAPK), PAL/RIM, cyclin-dependent kinases, TOR, mitochondrial retrograde (RTG) pathway	Morphogenesis	17
	CWI (MAPK), PKA	Response to stress	19
	CWI (MAPK), PAL/RIM	Cell wall assembly	14
	Filamentous growth (MAPK), PKA	Morphogenesis	5
<i>Saccharomyces pombe</i>	Pheromone response (MAPK), PKA	Mating	5
<i>Candida albicans</i>	Filamentous growth (MAPK), PKA	Dimorphism and mating	7,94
<i>Yarrowia lipolytica</i>	Filamentous growth (MAPK), PKA	Dimorphism	15,16
<i>Neurospora crassa</i>	Pheromone response (MAPK), PKA	Fruiting bodies development	74
<i>Aspergillus fumigatus</i>	HOG (MAPK), calcineurin signaling	Pathogenesis and virulence	20
<i>Colletotrichum orbiculare</i>	Mating (MAPK), filamentous growth (MAPK), PKA	Conidiation, pathogenesis and virulence	33
	Mating (MAPK), filamentous growth (MAPK), PKA	Morphogenesis, appressorium formation	40
<i>Coniothyrium minitans</i>	CWI (MAPK), nox complex signal	Conidiation and pathogenesis	89
<i>Ustilago maydis</i>	PMM (MAPK), CWI (MAPK), HOG (MAPK), PAL/RIM	Cell wall biogenesis, response to stress, response to pH, dimorphism	23,56,57
	PMM (MAPK), PKA	Mating, dimorphism, pathogenesis and virulence	8,52
<i>Cryptococcus neoformans</i>	Pheromone response (MAPK), PKA	Mating, pathogenesis and virulence	5
<i>Hypsizygus marmoreus</i>	MAPK, PKA, blue light signaling	Early stages of fruiting bodies development	93

been revealed. In *C. albicans* this phenomenon is controlled by the Cek1 pathway,⁹ in *U. maydis* by the PMM pathway both *in vitro*⁵⁴ and *in vivo* conditions,^{4,61} and in *Y. lipolytica* by the Kss1 pathway.¹⁵ Similarly, the transition from yeast to the pseudomycelial morphologies in *S. cerevisiae* (that properly does not grow in a filamentous form), involves the Kss1 MAPK pathway.²⁶ Recently, in addition, the participation of a number of different signaling pathways – the filamentous growth MAPK, PKA, PAL/RIM, TOR (targets of rapamycin) and RTG (mitochondrial retrograde), as well as the cyclin-dependent kinases – were found by means of a genetic screen to be involved in the pseudomycelium formation of *S. cerevisiae*.¹⁷

As mentioned above, the Cek1 MAPK pathway in *C. albicans*, a homologue of the *S. cerevisiae* Kss1 MAPK pathway that is involved in pseudomycelium formation, is necessary for the filamentous growth. Mutations in the components of the core Cek1 pathway (Ste11, Hst7, Cek1), adaptor proteins (St20, Ras1, Cdc42), or transcription factors (Cph1, Efg1, Tec1) controlled by this pathway, affect or suppress the mycelial growth of *C. albicans*, and significantly attenuate or suppress its virulence.^{9,96} A similarly phenomenon occurs when the gene *HOG1* is deleted in this fungus,³⁵ demonstrating the requirement of the filamentous growth and HOG pathways during the distinctive and pathogenic processes of *C. albicans*. In addition, mutation of the TCS system (upstream interactive proteins) in this pathway affects the *C. albicans* polarized growth under nutrient limitation conditions.⁷⁵

As already pointed out, fungal MAPK pathways can interact with other signal transduction pathways during the morphogenetic processes (Table 3). For example, in *C. albicans* the MAPK (Cek1) and PKA pathways operate in synergy during the yeast-to-mycelium transition (dimorphism),⁹⁶ whereas in *Y. lipolytica*^{15,16} and *U. maydis*⁵⁴ MAPK is required for the mycelial growth, and PKA for growing in the yeast form. That is to say, in these latter fungi these pathways are functionally antagonistic.

As an example of the regulation of dimorphism by the MAPK pathway in fungi, Fig. 3A shows an important number of genes regulated by the MAPK PMM pathway during the dimorphic transition from yeast to mycelium in *U. maydis*.^{58,59} This study represents the first analysis where gene regulation of the whole eukaryotic

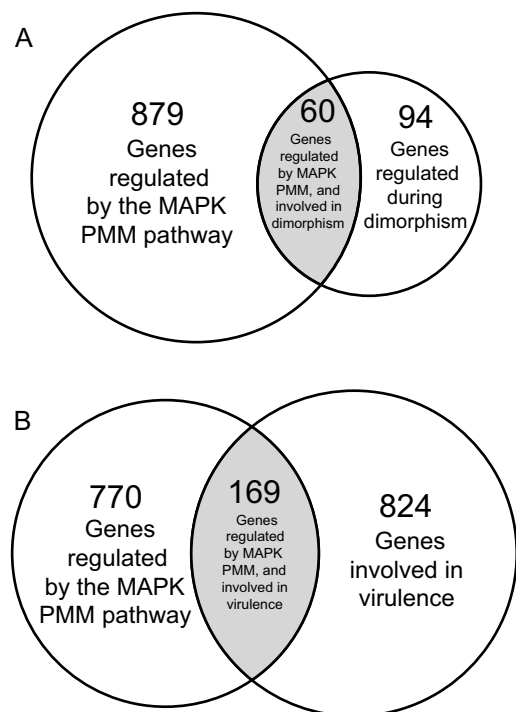


Fig. 3. Venn diagram showing the number of genes regulated by the MAPK (PMM) in *Ustilago madis* dimorphism and virulence. (A) Venn diagram showing 60 genes regulated by the MAPK pathway during the *U. maydis* dimorphism. (B) Venn diagram showing 169 genes regulated by the MAPK pathway involved the *U. maydis* pathogenic process.

genome by a MAPK pathway was done.⁵⁸ In *U. maydis*, approximately 14% (equivalent to 939 genes) of its genome was found to be regulated by the MAPK pathway. Among the genes differentially regulated, there were genes encoding proteins involved in cell cycle regulation [cyclin-dependent kinases (cdk), cell-division cycle (CDC), Hos4-subunit of the Set3 complex, DIP1, etc.]; transcription factors [PacC (response to pH changes), AtfA (response to different

types of stress), white collar 1 (response to light), etc.]; cellular transport and secretion (transport of substances, metals, effectors, etc.); signal transduction mechanism [Sok1-protein kinase; Pbs2-tyrosine protein kinase, GTPases, Crk1 (MAPK protein), etc.]; synthesis of cell wall [chitin deacetylase, glucan synthase (Kre6), chitinases, chitin synthases, proteins involved in *N*-glycosylation and *O*-mannosylation (Rot1)]; and finally in differentiation processes [proteins involved in organization of the actin cytoskeleton, cell polarity (Kin7a-motor protein), vesicle trafficking; actin polymerization (Ysc84-protein), etc.]. Also genes involved in pathogenesis and virulence were also differentially expressed (see below).

In other fungi, such as *C. albicans*⁷⁰ and *Y. lipolytica*,⁶⁷ genes involved in similar processes have been also identified by transcriptomic analysis during their dimorphic transition, corroborating the importance of the MAPK pathway in the differentiation processes of these fungi.

MAPK pathways in the development of fruiting bodies

Another morphogenetic phenomenon that occurs in some fungi is the formation of fruiting bodies, complex structures involved in sexual reproduction. Interestingly, the role of a MAPK pathway in developing ascocarps of *N. crassa* has been described, as well as its possible interaction with the PKA pathway under the same mechanisms described above.⁷⁶ It is known that during the formation of these structures, the corresponding extracellular stimuli are transferred through heterotrimeric G or Ras proteins, and then to the MAPK and PKA pathways, which regulate gene expression leading to the formation of ascocarps.⁷⁶ Accordingly, a recent study described the importance of MAPK, PKA and blue light signaling pathways in the early stages of the formation of fruiting bodies (mycelial knot, mycelial pigmentation, and primordium) in the edible Basidiomycota species *H. marmoreus*.⁹⁵ Similarly, in the Shiitake fungus *Lentinula edodes*, the importance of the MAPK pathway, also in early stages of the fruiting bodies development, has been described.⁸⁴ In the Ascomycota fungi *Cordyceps militaris*,⁹⁷ *Sordaria macrospora*,⁸⁵ and *Phaeosphaeria nodorum*³⁸ the role of MAPK pathway in fruiting bodies formation has been suggested, particularly the HOG pathway in the latter fungus.³⁸ Moreover, in the Basidiomycota fungus *Pleurotus eryngii*, differentially expressed genes involved in MAPK signaling, particularly in the cell wall biosynthesis (MAPK CWI), were identified during fruiting body formation.²⁵ In *U. maydis*, despite its taxonomic classification, the ability to form basidiocarps under controlled conditions has been described.¹¹ Interestingly, mutant strains in genes encoding MAPK core proteins were unable to form basidiocarps (León-Ramírez et al., in preparation). This observation confirms the fundamental role of MAPK pathways during basidiocarps formation in fungal species. Nevertheless, despite these studies, there is still scant information about how the MAPK pathways regulate the phenomenon of fruiting body formation in fungi.

MAPK pathways involved in fungal virulence

In all plant or animal pathogenic fungi thus far studied, MAPK pathways have been identified to be involved in their respective virulent processes. For example, the role of MAPK pathways during pathogenesis has been described for the plant pathogens *U. maydis*,⁸ *Alternaria alternata*,¹⁸ *B. cinerea*,^{45,82} *F. graminearum*,^{30,31} *F. oxysporum*,⁷⁴ *C. orbiculare*,³⁴ *Magnaporthe grisea*,^{10,41} *V. dahliae*,⁸⁶ *Cochliobolus heterostrophus*,⁴⁶ *P. nodorum*,³⁸ *Fusarium verticillioides*,⁹⁴ *C. minitans*,⁹¹ inter alia, and for the human pathogens *C. albicans*,^{9,35,77,80} *C. neoformans*,³⁶ and *A. fumigatus*.^{20,87,92} Also a similar role in the pathogenesis was disclosed in the MAPK pathway of the insect pathogen *B. bassiana*.⁹⁰

In other types of interactions between fungi and plants, the signaling by the MAPK pathways has been described as necessary; e.g. in the Ascomycota fungus *Epichloë festucae*, a MAPK plays a crucial role during its symbiotic interaction with the plant *Lolium perenne*.⁷ A similar requirement occurs in the mycorrhizal relationship between *Rhizophagus irregularis* and its soy bean host.⁴⁹

Regarding the molecular mechanism of the MAPK signaling pathways in virulence, deeper analyses have been performed with *C. albicans*, probably the most important fungal pathogen in humans. This fungus has the ability to colonize different parts of the human body changing its morphology from yeast to an invasive mycelium.⁴⁰ When *C. albicans* infects and colonizes a human being, the fungus is faced with a hostile environment caused by the defense system of the host. Under these conditions the HOG, the filamentous growth, and CWI MAPK pathways play a key role, firstly by sensing the hostile environmental conditions, and secondly orchestrating the alterations in the genetic machinery of the fungus that leads to a physiological alteration associated to virulence such as change in cell morphology, changes in cell wall composition and structure, ability to respond to several stresses, secretion of different proteins including hydrolytic enzymes, etc.^{9,35,80,98} Of the four known *C. albicans* MAPK pathways, CWI (Mkc1), HOG (Hog1), filamentous growth (Cek1), and growth (Cek2) (see Table 2), CWI and HOG are involved in the response to the stress imposed by the immune system of the host during the infection process, including the activity of the phagocytes.^{2,35,77,80} It is therefore not surprising that inactivation of the Hog1 and Mkc1 pathways reduces the virulence of *C. albicans* in mice,^{1,2,9} and that mutant strains in these genes cannot bind to, or infect, the intestinal mucosa, also being susceptible to bile salts.⁷⁷ It has been described the role of the sensor protein Msb2, a glycoprotein of the Cek1 pathway involved in cell wall integrity and filamentous growth. Msb2 is secreted to the medium and prevents the inflammatory response caused by the antimicrobial peptides (AMPs) produced by the host.⁸³ Mbs2 is part of the TCS system, and the *C. albicans* mutant strains in the genes of the TCS system are susceptible to different stresses.⁸⁸ Similarly, genes encoding proteins of this system are essential at the early stages of the pathogenesis processes developed by other fungi: *U. maydis*,⁴³ *V. dahliae*,⁸⁶ *B. bassiana*,⁹⁰ *B. cinerea*,⁴⁵ *F. oxysporum*,⁷⁴ *M. oryzae*,^{41,60} and several species of *Aspergillus*.³²

Interactions of MAPK pathways with other signaling pathways during the pathogenic processes of fungi have been described. For example, in *A. fumigatus*, the HOG and calcineurin signaling pathways are required for its virulence.²⁰ Similarly, in *C. minitans*, a fungal species used as a biocontrol agent, the Nox complex signal along with CWI (Stl2) pathway regulate its pathogenicity and conidiation,⁹¹ and in *U. maydis* two important phenomena, mating and virulence, are regulated by the MAPK PMM and PKA pathways (revised by Brefort et al.⁸). *U. maydis* is the causal agent of *Zea mays* smut disease, and under controlled conditions can infect different plants.^{44,56,57,65} Apparently in this fungus, and in contrast to *C. albicans*, only the MAPK PMM pathway is involved in virulence⁸ through the MAPKs Kpp2⁶⁸ and Crk1.²⁸ Therefore when the genes encoding proteins of the PMM pathway are deleted, the virulence of the fungus over the maize is reduced or eliminated.^{61,62} In Fig. 3B we show the number of genes putatively regulated by the MAPK PMM pathway and required for pathogenic processes or the acting of some molecules in *U. maydis*: degradative proteins of plant cell wall, effectors, virulence factors, or regulating transcription factors are some of them. Other genes implied in virulence and identified by its deletion or by bioinformatic analysis are also shown. The regulation of these genes by the MAPK PAM pathway confirms the key role in the signal transduction processes occurring during the pre-penetration and pathogenic process of this fungus, similarly to what occurs in many other pathogenic fungi.

Conclusions and perspectives

It is evident that MAPK pathways are signal transduction and cellular communication mechanisms highly conserved in all fungi, similarly to what occurs in higher eukaryotic organisms. Accordingly, MAPK pathways are involved in the most essential physiological and development processes occurring in fungal species, and their mutation leads to aberrant phenotypes, severe damages under different growth conditions, alterations in development and differentiation, and decrease or loss of virulence in the pathogenic species. The data accumulated in the study of fungal species analyzed thus far evidence that, practically, with only subtle variations, the same MAPK pathways exist in all the analyzed species, although their number may be variable. It may be concluded also that, although they act very similarly and work by the same mechanism, Receptor – Two-Component Signal Transduction system (TCS) – MAPK core – Transcription factors – Gene regulation (Fig. 1), the regulated processes are not always the same. It is also important to recall that during practically all important physiological processes in fungi, there is an active interconnection between MAPK pathways and also with other signaling pathways, especially with the PKA. In many cases, this interaction of the MAPK pathways with other pathways occurs upstream of the pathways involved, for instance in the sensory and auxiliary proteins.

It seems necessary to point out that, although large advances on the study of MAPK pathways in fungi have been recently uncovered, more information on some phenomena or processes that occur in these organisms is necessary. For example, the nature of different receptors, as well as the interactions occurring between the different signaling pathways present in fungi are still unknown. Similarly, the role of the MAPK pathways in the formation of fruiting bodies in fungi is still a poorly studied matter, and no information is available on the signaling pathways that are regulating this process in coordination with the MAPK pathways. Also necessary in this aspect is some information on the nature of the genes regulated in several fungal processes.

Regarding the pathogenic processes, it is important to have more information on whether the MAPK pathways regulate the synthesis of virulence factors, effector proteins, etc., involved in the pathogenic processes per se, and of the systems used by fungi to avoid the host defense mechanisms, especially in fungi with biotrophic or hemibiotrophic life styles. It would be important also to increase the knowledge on how MAPK pathways regulate fungus symbiotic relationships with other organisms: fungus–plant (e.g. mycorrhizae, orchids), fungus–algae (e.g. lichens), and fungus–insects (e.g. *Ambrosia* beetles, ant gardens), etc. Finally, it is necessary to study in depth the understanding of the epigenetic regulation of the physiological and developmental processes where MAPK pathways are involved.

Despite these unfilled aspects it is obvious that our knowledge on the roles played by MAPK pathways in fungal cells has widened very rapidly in the most recent years, and that the knowledge gathered on them makes clear their importance in all the living processes of the members of the Phylum Fungi.

Conflict of interest

The authors declare that they have no conflicts of interest.

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References

- Alonso-Monge R, Navarro-García F, Román E, Negrodo AI, Eisman B, Nombela C, et al. The Hog1 mitogen-activated protein kinase is essential in oxidative stress response and chlamydospore formation in *Candida albicans*. *Eukaryot Cell*. 2003;2:351–61.
- Alonso-Monge R, Román E, Arana DM, Pla J, Nombela C. Fungi sensing environmental stress. *Clin Microbiol Infect*. 2009;1:17–9.
- Alvaro CG, Thorner J. Heterotrimeric G protein coupled receptor signaling in yeast mating pheromone response. *J Biol Chem*. 2016;291:7788–95.
- Andrews DL, Egan JD, Mayorga ME, Gold SE. The *Ustilago maydis* *ubc4* and *ubc5* genes encode members of a MAPK kinase cascade required for filamentous growth. *Mol Plant Microbe Interact*. 2000;13:781–6.
- Bahn YS, Xue C, Idnurm A, Rutherford JC, Heitman J, Cardenas ME. Sensing the environment: lessons from fungi. *Nat Rev Microbiol*. 2007;5:57–69.
- Banuet F. Signaling in the yeasts: an informational cascade with links to the filamentous fungi. *Microbiol Mol Biol Rev*. 1998;62:249–74.
- Becker Y, Eaton CJ, Brasell E, May KJ, Becker M, Hassing B, et al. The fungal cell-wall integrity MAPK cascade is crucial for hyphal network formation and maintenance of restrictive growth of *Epichloë festucae* in symbiosis with *Lolium perenne*. *Mol Plant Microbe Interact*. 2015;28:69–85.
- Brefort T, Doehlemann G, Mendoza-Mendoza A, Reissmann S, Djamei A, Kahmann R. *Ustilago maydis* as pathogen. *Annu Rev Phytopathol*. 2009;47:423–45.
- Brown AJ, Budge S, Kaloriti D, Tillmann A, Jacobsen MD, Yin Z, et al. Stress adaptation in a pathogenic fungus. *J Exp Biol*. 2014;217:144–55.
- Bruno KS, Tenjo F, Li L, Hamer Je Xu JR. Cellular localization and role of kinase activity of PMK1 in *Magnaporthe grisea*. *Eukaryot Cell*. 2004;3:1525–32.
- Cabrera-Ponce JL, León-Ramírez CG, Verver-Vargas A, Palma-Tirado L, Ruiz-Herrera J. Metamorphosis of the Basidiomycota *Ustilago maydis*: transformation of yeast-like cell into basidiocarps. *Fungal Genet Biol*. 2012;49:765–71.
- Cantero PD, Ernst JF. Damage to the glycosylated activates PMT-directed O-mannosylation via the Msb2-Cek1 pathway in *Candida albicans*. *Mol Microbiol*. 2011;80:715–25.
- Carbó N, Pérez-Martín J. Activation of the cell wall integrity pathway promotes escape from G2 in the fungus *Ustilago maydis*. *PLoS Genet*. 2010;6:e1001009.
- Castrejón F, Gómez A, Sanz M, Duran A, Roncero C. The RIM101 pathway contributes to yeast cell wall assembly and its function becomes essential in the absence of mitogen-activated protein kinase Slt2p. *Eukaryot Cell*. 2006;5:507–17.
- Cervantes-Chávez JA, Ruiz-Herrera J. STE11 disruption reveals the central role of a MAPK pathway in dimorphism and mating in *Yarrowia lipolytica*. *FEMS Yeast Res*. 2006;6:801–15.
- Cervantes-Chávez JA, Ruiz-Herrera J. The regulatory subunit of protein kinase A promotes hyphal growth and plays an essential role in *Yarrowia lipolytica*. *FEMS Yeast Res*. 2007;7:929–40.
- Chavel CA, Caccamise LM, Li B, Cullen PJ. Global regulation of a differentiation MAPK pathway in yeast. *Genetics*. 2014;198:1309–28.
- Chung KR. Mitogen-activated protein kinase signaling pathways of the tangerine pathotype of *Alternaria alternata*. *MAP Kinase*. 2013;2:16–23.
- Cohen A, Kupiec M, Weisman R. Glucose activates TORC2-Gad8 protein via positive regulation of the cAMP/cAMP-dependent protein kinase A (PKA) pathway and negative regulation of the Pmk1 protein-mitogen-activated protein kinase pathway. *J Biol Chem*. 2014;289:21727–37.
- De Castro PA, Chen C, De Almeida RS, Freitas FZ, Bertolini MC, Morais ER, et al. ChIP-seq reveals a role for CrzA in the *Aspergillus fumigatus* high-osmolarity glycerol response (HOG) signaling pathway. *Mol Microbiol*. 2014;94:655–74.
- Dettmann A, Heilig Y, Valerius O, Ludwing S, Seiler S. Fungal communication requires the MAK-2 pathway elements STE-20 and RAS-2, the NRC-1 adapter STE-50 and the MAP Kinase scaffold HAM-5. *PLoS Genet*. 2014;10:e1004762. <http://dx.doi.org/10.1371/journal.pgen.1004762>
- Dohlman HG, Thorner JW. Regulation of G protein-initiated signal transduction in yeast: paradigms and principles. *Annu Rev Biochem*. 2001;70:703–54.
- Dohlman HG. Thematic minireview series: complexities of cellular signaling revealed by simple model organisms. *J Biol Chem*. 2016;291:7786–7.
- Fonseca-García C, León-Ramírez CG, Ruiz-Herrera J. The regulation of different metabolic pathways through the Pal/Rim pathway in *Ustilago maydis*. *FEMS Yeast Res*. 2012;12:547–56.
- Fu YP, Liang Y, Dai YT, Yang CT, Duan MZ, Zhang Z, et al. De novo sequencing and transcriptome analysis of *Pleurotus eryngii* subsp. *tuoliensis* (Bailiungu) mycelia in response to cold stimulation. *Molecules*. 2016;21, pii: E560. doi: 10.3390/molecules21050560.
- Gancedo M. Control of pseudohyphae formation in *Saccharomyces cerevisiae*. *FEMS Microbiol Rev*. 2001;25:107–23.
- García R, Botet J, Rodríguez-Peña JM, Bermejo C, Ribas JC, Revuelta JL, et al. Genomic profiling of fungal cell wall-interfering compounds: identification of a common gene signature. *BMC Genomics*. 2015;16:683. <http://dx.doi.org/10.1186/s12864-015-1879-4>
- Garrido E, Voss U, Müller P, Castillo-Lliva S, Kahmann R, Pérez-Martín J. The induction of sexual development and virulence in the smut fungus *Ustilago maydis* depends on Crk1, a novel MAPK protein. *Genes Dev*. 2004;18:3117–30.
- Gruber S, Zeilinger S. The transcription factor Ste12 mediates the regulatory role of the TMK1 MAP kinase in mycoparasitism and vegetative hyphal fusion in the filamentous fungus *Trichoderma atroviride*. *PLoS One*. 2014;9:e111636. <http://dx.doi.org/10.1371/journal.pone.0111636>

30. Gu Q, Chen Y, Liu Y, Zhang C, Ma Z. The transmembrane protein FgSho1 regulates fungal development and pathogenicity via the MAPK module Ste50-Ste11-Ste7 in *Fusarium graminearum*. *New Phytol.* 2015;206:315–28.
31. Gu Q, Zhang C, Liu X, Ma Z. A transcription factor FgSte12 is required for pathogenicity in *Fusarium graminearum*. *Mol Plant Pathol.* 2015;16:1–13.
32. Hagiwara D, Sakamoto K, Abe K, Gomi K. Signaling pathways for stress responses and adaptation in *Aspergillus* species: stress biology in the post-genomic era. *Biosci Biotechnol Biochem.* 2016;80:1667–80.
33. Hagiwara D, Suzuki S, Kamei K, Gono T, Kawamoto S. The role of AtfA and HOG MAPK pathways in stress tolerance in conidia of *Aspergillus fumigatus*. *Fungal Genet Biol.* 2014;73:138–49.
34. Harata K, Kubo Y. RasGTPase activating protein Colra1 is involved in infection-related morphogenesis by regulating cAMP and MAPK signaling pathways through CoRas2 in *Colletotrichum orbiculare*. *PLoS One.* 2014;9:e109045, <http://dx.doi.org/10.1371/journal.pone.0109045>
35. Herrero-de-Dios C, Alonso-Monge R, Pla J. The lack of upstream elements of the Cek1 and Hog1 mediated pathways leads to a synthetic lethal phenotype upon osmotic stress in *Candida albicans*. *Fungal Genet Biol.* 2014;69:31–42.
36. Idnurm A, Bahn YS, Nielsen K, Lin X, Fraser JA, Heitman J. Deciphering the model pathogenic fungus *Cryptococcus neoformans*. *Nat Rev Microbiol.* 2005;3:753–64.
37. Jaimes-Arroyo R, Lara-Rojas F, Bayram Ö, Valerius O, Braus GH, Aguirre J. The *SrkA* kinase is part of the *SakA* Mitogen-Activated Protein Kinase interactome and regulates stress response and development in *Aspergillus nidulans*. *Eukaryot Cell.* 2015;14:495–510.
38. John E, Lopez-Ruiz F, Rybak K, Mousley CJ, Oliver RP, Tan KC. Dissecting the role of histidine kinase and HOG1 mitogen-activated protein kinase signalling in stress tolerance and pathogenicity of *Parastagonospora nordorum* on wheat. *Microbiology.* 2016;162:1023–36.
39. Kabeche R, Madrid M, Casado J, Moseley JB. Eisosomes regulate phosphatidylinositol 4,5-bisphosphate (PI(4,5)P₂) cortical clusters and Mitogen-Activated Protein (MAP) Kinase signaling upon osmotic stress. *J Biol Chem.* 2015;290:25960–73.
40. Kim S, Kim E, Shin DS, Kang H, Oh KB. Evaluation of morphogenic regulatory activity of farnesic acid and its derivatives against *Candida albicans* dimorphism. *Bioorg Med Chem Lett.* 2002;12:895–8.
41. Kou Y, Naqvi NI. Surface sensing and signaling networks in plant pathogenic fungi. *Semin Cell Dev Biol.* 2016;57:84–92.
42. Lam MH, Snider J, Rehal M, Wong V, Aboualizadeh F, Drecun L, et al. A comprehensive membrane interactome mapping of Sho1p reveals Fps1p as a novel key player in the regulation of the HOG pathway in *S. cerevisiae*. *J Mol Biol.* 2015;427:2088–103.
43. Lanver D, Mendoza-Mendoza A, Brachmann A, Kahmann R. Sho1 and Msb2-related proteins regulate appressorium development in the smut fungus *Ustilago maydis*. *Plant Cell.* 2010;22:2085–101.
44. León-Ramírez CG, Cabrera-Ponce JL, Martínez-Espinoza AD, Herrera-Estrella L, Méndez-Morán L, Reynaga-Peña CG, et al. Infection of alternative host plant species by *Ustilago maydis*. *New Phytol.* 2004;164:337–46.
45. Leroch M, Mueller N, Hinsenkamp I, Hahn M. The signalling mucin Msb2 regulates surface sensing and host penetration via BMP1 MAP kinase signalling in *Botrytis cinerea*. *Mol Plant Pathol.* 2015;16:787–98.
46. Lev S, Sharon A, Hadar R, Ma H, Horwitz BA. A mitogen-activated kinase of the corn leaf pathogen *Cochliobolus heterostrophus* is involved in conidiation, appressorium formation, and pathogenicity: diverse roles for mitogen-activated protein kinase homologs in foliar pathogens. *Proc Natl Acad Sci U S A.* 1999;96:13542–7.
47. Li L, Wright SJ, Krystofova S, Park G, Borkovich KA. Heterotrimeric G protein signaling in filamentous fungi. *Annu Rev Microbiol.* 2007;61:423–52.
48. Liu W, Zhou X, Li G, Li L, Kong L, Wang C. Multiple plant surface signals are sensed by different mechanisms in the rice blast fungus for appressorium formation. *PLoS Pathog.* 2011;7:e1001261, <http://dx.doi.org/10.1371/journal.ppat.1001261>
49. Liu Z, Li Y, Ma L, Wei H, Zhang J, He X, et al. Coordinated regulation of arbuscular mycorrhizal fungi and soybean MAPK pathway genes improved mycorrhizal soybean drought tolerance. *Mol Plant Microbe Interact.* 2015;28:408–19.
50. Ma D, Li R. Current understanding of HOG-MAPK pathway in *Aspergillus fumigatus*. *Mycopathologia.* 2013;175:13–23.
51. Manning G. Genomic overview of protein kinases. *WormBook.* 2005;13:1–19.
52. Mao K, Wang K, Zhao M, Xu T, Klionsky DJ. Two MAPK signaling pathways are required for mitophagy in *Saccharomyces cerevisiae*. *J Cell Biol.* 2011;193:755–67.
53. Marcus S, Polverino A, Barr M, Wigler M. Complexes between STE5 and components of the pheromone-responsive mitogen-activated protein kinase module. *Proc Natl Acad Sci U S A.* 1994;91:7762–6.
54. Martínez-Espinoza AD, Ruiz-Herrera J, León-Ramírez CG, Gold SE. MAP kinase and cAMP signaling pathways modulate the pH-induced yeast-to-mycelium dimorphic transition in the corn smut fungus *Ustilago maydis*. *Curr Microbiol.* 2004;49:274–81.
55. Martínez-Soto D, González-Prieto JM, Ruiz-Herrera J. Transcriptomic analysis of the *GCN5* gene reveals mechanisms of the epigenetic regulation of virulence and morphogenesis in *Ustilago maydis*. *FEMS Yeast Res.* 2015;15:fov055.
56. Martínez-Soto D, Pérez-García FE, Ruiz-Herrera J. Arabidopsis infection by haploid or diploid strains of *Ustilago maydis* reveals its capacity as a necrotrophic or biotrophic phytopathogen. *Fungal Genom Biol.* 2016;6:133, <http://dx.doi.org/10.4172/2165-8056.1000133>
57. Martínez-Soto D, Robledo-Briones AM, Estrada-Luna AA, Ruiz-Herrera J. Transcriptomic analysis of *Ustilago maydis* infecting *Arabidopsis* reveals important aspects of the fungus pathogenic mechanisms. *Plant Signal Behav.* 2013;8:e25059.
58. Martínez-Soto D, Ruiz-Herrera J. Regulation of the expression of the whole genome of *Ustilago maydis* by a MAPK pathway. *Arch Microbiol.* 2015;197:575–88.
59. Martínez-Soto D, Ruiz-Herrera J. Transcriptomic analysis of the dimorphic transition of *Ustilago maydis* induced in vitro by a change in pH. *Fungal Genet Biol.* 2013;5(58–59):116–25.
60. Martín-Urdiroz M, Osés-Ruiz M, Ryder LS, Talbot NJ. Investigating the biology of plant infection by the rice blast fungus *Magnaporthe oryzae*. *Fungal Genet Biol.* 2016;90:61–8.
61. Mayorga ME, Gold SE. A MAP kinase encoded by the *ubc3* gene of *Ustilago maydis* is required for filamentous growth and full virulence. *Mol Microbiol.* 1999;34:485–97.
62. Mayorga ME, Gold SE. The *ubc2* gene of *Ustilago maydis* encodes a putative novel adaptor protein required for filamentous growth, pheromone response and virulence. *Mol Microbiol.* 2001;41:1365–79.
63. Mazzola D, Pimentel C, Caetano S, Amaral C, Menezes R, Santos CN, et al. Inhibition of Yap2 activity by MAPKAP kinase Rck1 affects yeast tolerance to cadmium. *FEBS Lett.* 2015;589:2841–9.
64. Medina-Castellanos E, Esquivel-Naranjo EU, Heil M, Herrera-Estrella A. Extracellular ATP activates MAPK and ROS signaling during injury response in the fungus *Trichoderma atroviride*. *Front Plant Sci.* 2014;5:659, <http://dx.doi.org/10.3389/fpls.2014.00659>
65. Méndez-Morán L, Reynaga-Peña CG, Springer PS, Ruiz-Herrera J. *Ustilago maydis* infection of the nonnatural host *Arabidopsis thaliana*. *Phytopathology.* 2005;95:480–8.
66. Mizuno T, Masuda Y, Ire K. The *Saccharomyces cerevisiae* AMPK, Snf1, negatively regulates the Hog1 MAPK pathway in ER stress response. *PLoS Genet.* 2015;11:e1005491, <http://dx.doi.org/10.1371/journal.pgen.1005491>
67. Morales-Vargas AT, Domínguez A, Ruiz-Herrera J. Identification of dimorphism-involved genes of *Yarrowia lipolytica* by means of microarray analysis. *Res Microbiol.* 2012;163:378–87.
68. Müller P, Aichinger C, Feldbrügge M, Kahmann R. The MAP kinase kpp2 regulates mating and pathogenic development in *Ustilago maydis*. *Mol Microbiol.* 1999;34:1007–17.
69. Nakayama N, Miyajima A, Arai K. Nucleotide sequences of STE2 and STE3, cell type specific sterile genes from *Saccharomyces cerevisiae*. *EMBO J.* 1985;4:2643–8.
70. Nantel A, Dignard D, Bachewich C, Harcus D, Marcil A, Bouin AP, et al. Transcription profiling of *Candida albicans* cells undergoing yeast-to-hyphal transition. *Mol Biol Cell.* 2002;13:3452–65.
71. Navarro-García F, Alonso-Monge R, Rico H, Pla J, Sentandreu R, Nombela C. A role for the MAP kinase gene MKC1 in cell wall construction and morphological transition in *Candida albicans*. *Microbiology.* 1998;144:411–24.
72. Navarro-García F, Sánchez M, Pla J, Nombela C. Functional characterization of the MKC1 gene of *Candida albicans*, which encodes a mitogen-activated protein kinase homolog related to cell integrity. *Mol Cell Biol.* 1995;15:2197–206.
73. Ortiz-Urquiza A, Keyhani NO. Stress response signaling and virulence: insights from entomopathogenic fungi. *Curr Genet.* 2015;61:239–49.
74. Perez-Nadales E, Di Pietro A. The transmembrane protein Sho1 cooperates with the mucian Msb2 to regulate invasive growth and plant infection in *Fusarium oxysporum*. *Mol Plant Pathol.* 2015;16:593–603.
75. Pitoniak A, Chavel CA, Chow J, Smith J, Camara D, Karunanithi S, et al. Cdc42p-interacting protein Bem4p regulates the filamentous-growth mitogen-activated protein kinase pathway. *Mol Cell Biol.* 2015;35:417–36.
76. Pöggeler S, Nowrousian M, Kück U. Fruiting-body development in Ascomycetes. In: Kues F, editor. *The mycota. Growth, differentiation and sexuality*. Berlin Heidelberg: Springer-Verlag; 2006. p. 325–55.
77. Prieto D, Román E, Correia I, Pla J. The HOG pathway is critical for the colonization of the mouse gastrointestinal tract by *Candida albicans*. *PLoS One.* 2014;27:e87128, <http://dx.doi.org/10.1371/journal.pone.0087128>
78. Printen JA, Sprague GF Jr. Protein-protein interactions in the yeast pheromone response pathway: Ste5p interacts with all members of the MAP kinase cascade. *Genetics.* 1994;138:609–19.
79. Rispail N, Soanes DM, Ant C, Czajkowski R, Grünler A, Hugue R, et al. Comparative genomics of MAPK kinase and calcium-calmodulin signalling components in plant and human pathogenic fungi. *Fungal Genet Biol.* 2009;46:287–98.
80. Román E, Alonso-Monge R, Miranda A, Pla J. The Mkk2 MAPKK regulates cell wall biogenesis in cooperation with Cek1-pathway in *Candida albicans*. *PLoS One.* 2015;10:e0133476, <http://dx.doi.org/10.1371/journal.pone.0133476>
81. Ruiz-Herrera J, editor. *Dimorphic fungi: their importance as models for differentiation and fungal pathogenesis*. Bentham e Books; 2012.
82. Schamber A, Leroch M, Diwo J, Mendgen K, Hahn M. The role of mitogen activated protein (MAP) kinase signalling components and the Ste12 transcription factor in germination and pathogenicity of *Botrytis cinerea*. *Mol Plant Pathol.* 2010;11:105–19.
83. Szafranski-Schneider E, Swidrigall M, Cottier F, Tielker D, Román E, Pla J, et al. Msb2 shedding protects *Candida albicans* against antimicrobial peptides. *PLoS Pathog.* 2012;8:e1002501.
84. Szeto CY, Leung GS, Kwan HS. Le.MAPK and its interacting partner, Le.DRMIP, in fruiting body development in *Lentinula edodes*. *Gene.* 2007;393:87–93.
85. Teichert I, Steffens EK, Schanab N, Fränzel B, Krisp C, Wolters DA, et al. PRO40 is a scaffold protein of the cell wall integrity pathway, linking the MAP kinase module to the upstream activator protein kinase C. *PLoS Genet.* 2014;10:e1004582, <http://dx.doi.org/10.1371/journal.pgen.1004582>

86. Tian L, Xu J, Zhou L, Gou W. VdMsb regulates virulence and microscle-rotia production in the fungal plant pathogen *Verticillium dahliae*. *Gene*. 2014;550:238–44.
87. Valiante V, Macheleidt J, Föge M, Brakhage AA. The *Aspergillus fumigatus* cell wall integrity signaling pathway: drug target, compensatory pathways, and virulence. *Front Microbiol*. 2015;6:325.
88. Vallejo MC, Mayinger P. Delayed turnover of unphosphorylated Ssk1 during carbon stress activates the yeast Hog1 Map Kinase pathway. *PLoS One*. 2015;10:e0137199, <http://dx.doi.org/10.1371/journal.pone.0137199>
89. Velázquez-Zavala N, Rodríguez-Gonzalez M, Navarro-Olmos R, Ongay-Larios L, Kawasaki L, Torres-Quiroz F, et al. Ineffective phosphorylation of Mitogen-Activated Protein Kinase Hog1p in response to high osmotic stress in the yeast *Kluyveromyces lactis*. *Eukaryot Cell*. 2015;14:922–30.
90. Wang ZL, Li F, Li C, Feng MG. Bbssk1, a response regulator required for conidiation, multi-stress tolerance, and virulence of *Beauveria bassiana*. *Appl Microbiol Biotechnol*. 2014;98:5607–18.
91. Wei W, Zhu W, Cheng J, Xie J, Jiang D, Li G, et al. Nox complex signal and MAPK cascade pathway are cross-linked and essential for pathogenicity and conidiation of mycoparasite *Coniothyrium minitans*. *Sci Rep*. 2016;6:24325, <http://dx.doi.org/10.1038/srep24325>
92. Winkelströter LK, Bom VL, de Castro PA, Ramalho LN, Goldman MH, Brown NA, et al. High osmolarity glycerol response PtcB phosphatase is important for *Aspergillus fumigatus* virulence. *Mol Microbiol*. 2015;96:42–54.
93. Yu Z, Armant O, Fischer R. Fungi use the SakA (HogA) pathway for phytochrome-dependent light signalling. *Nat Microbiol*. 2016;1:16019, <http://dx.doi.org/10.1038/nmicrobiol.2016.19>
94. Zhang C, Wang J, Tao H, Dang X, Wang Y, Chen M, et al. FvBck1, a component of cell wall integrity MAP kinase pathway, is required for virulence and oxidative stress response in sugar cane Pokkah Boeng pathogen. *Front Microbiol*. 2015;6:1096, <http://dx.doi.org/10.3389/fmicb.2015.01096>
95. Zhang J, Ren A, Chen H, Zhao M, Shi L, Chen M, et al. Transcriptome analysis and its application in identifying genes associated with fruiting body development in basidiomycete *Hypsizygus marmoreus*. *PLoS One*. 2015;10:e0123025, <http://dx.doi.org/10.1371/journal.pone.0123025>
96. Zhao X, Mehrabi R, Xu JR. Mitogen-activated protein kinase pathways and fungal pathogenesis. *Eukaryot Cell*. 2007;6:1701–14.
97. Zheng P, Xia Y, Xiao G, Xiong C, Hu X, Zhang S, et al. Genome sequence of the insect pathogenic fungus *Cordyceps militaris*, a valued traditional Chinese medicine. *Genome Biol*. 2011;12:R116.
98. Zhu W, Filler SG. Interactions of *Candida albicans* with epithelial cells. *Cell Microbiol*. 2010;12:273–82.
99. Zwick E, Bange J, Ullrich A. Receptor tyrosine kinase signalling as target for cancer intervention strategies. *Endocr Relat Cancer*. 2001;8:161–73.