



Note

## Cutaneous infection by *Phaeoacremonium parasiticum*



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### ABSTRACT

**Background:** *Phaeoacremonium parasiticum* is considered a rare infectious agent that is part of a heterogeneous group of fungi causing phaeohyphomycosis. This organism is capable of producing subcutaneous infections, eumycetomas, osteomyelitis, arthritis, myositis and also disseminated diseases, such as fungemia and endocarditis.

**Case report:** We describe a case of cutaneous infection by *P. parasiticum* in a kidney transplant patient. The identification of this microorganism was performed by microbiological and histopathological studies and confirmed with the sequence of the gene encoding β-tubulin and a real time panfungal PCR targeting 18S ribosomal RNA gene. The microorganism was correctly identified by phenotypic and molecular methods. The patient was treated with oral antifungal therapy and a debulking surgery and evolved without any complication.

**Conclusions:** The diagnosis of this infection is difficult and usually affects kidney transplant patients, but the reasons of this association are still unknown.

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## Infección cutánea por *Phaeoacremonium parasiticum*

### RESUMEN

**Palabras clave:**

Feohomicosis

Infección cutánea

Trasplante renal

**Antecedentes:** *Phaeoacremonium parasiticum* es considerado un agente infeccioso poco común que forma parte de un grupo heterogéneo de hongos causantes de feohomicosis. Este microorganismo es capaz de producir infección cutánea, eumicetoma, osteomielitis, artritis, miositis e incluso enfermedad diseminada como fungemia y endocarditis.

**Caso clínico:** Se describe un caso de infección cutánea por *P. parasiticum* en un paciente trasplantado renal. Para la identificación del microorganismo se realizaron pruebas microbiológicas e histopatológicas, y se confirmó la identificación con la secuenciación del gen de la β-tubulina y una PCR a tiempo real para la detección del gen 18S rRNA. El microorganismo fue identificado correctamente por métodos fenotípicos y moleculares. El paciente recibió tratamiento con antifúngicos orales y citorreducción quirúrgica, y evolucionó sin ninguna complicación.

**Conclusiones:** El diagnóstico de esta infección es difícil y se presenta habitualmente en pacientes trasplantados renales. Sin embargo, la asociación de esta infección con este tipo de pacientes no ha sido aún explicada.

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*Phaeoacremonium* species have a wide distribution in the environment. It was thought that they only produced disease in plants,<sup>2,8</sup> but in 1974 the first case of cutaneous infection in a kidney transplant patient was described. *Phaeoacremonium parasiticum*, previously known as *Phialophora parasitica*, is considered a rare infectious agent although in the last years new cases have been described.<sup>2,7,12,13</sup> *P. parasiticum* is part of a heterogeneous group of fungi causing phaeohyphomycosis, a disease that includes a broad spectrum of infections caused by fungi that produce septate hyphae with melanin in the tissue.<sup>1,10</sup> This organism produces subcutaneous infections, eumycetomas, osteomyelitis, arthritis, myositis and also disseminated diseases, such as fungemia and endocarditis.<sup>17</sup> We describe a case of cutaneous infection caused by *P. parasiticum* in an immunosuppressed kidney transplant patient.

### Case report

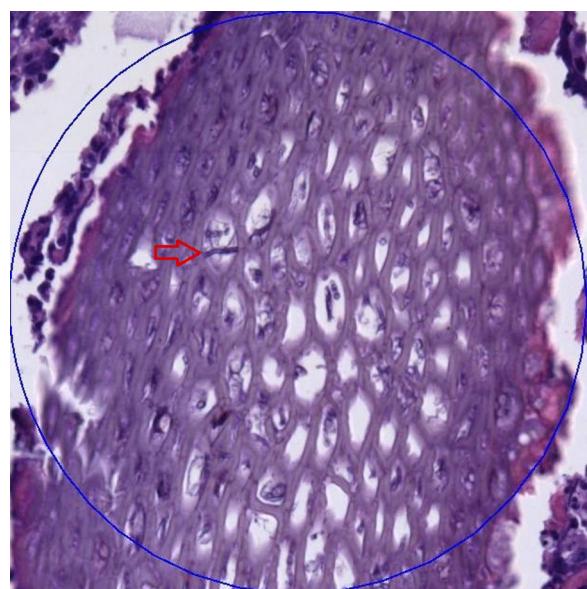
A 77 year-old man went to a local hospital with an injury in his right hand caused with plants of his garden. The wound was cleaned and a daily antiseptic cleaning was prescribed. However, after several weeks the wound did not improve. He came back to the same centre, and at the time of admission the patient explained that he was under immunosuppressive treatment due to a kidney transplant in 2015.

In the culture from the sample taken from the lesion, a filamentous fungi, identified as *Exophiala dermatitidis* by microscopic identification, grew. The patient started an antifungal treatment with voriconazole, but after four days the patient stopped the treatment due to of intolerance.

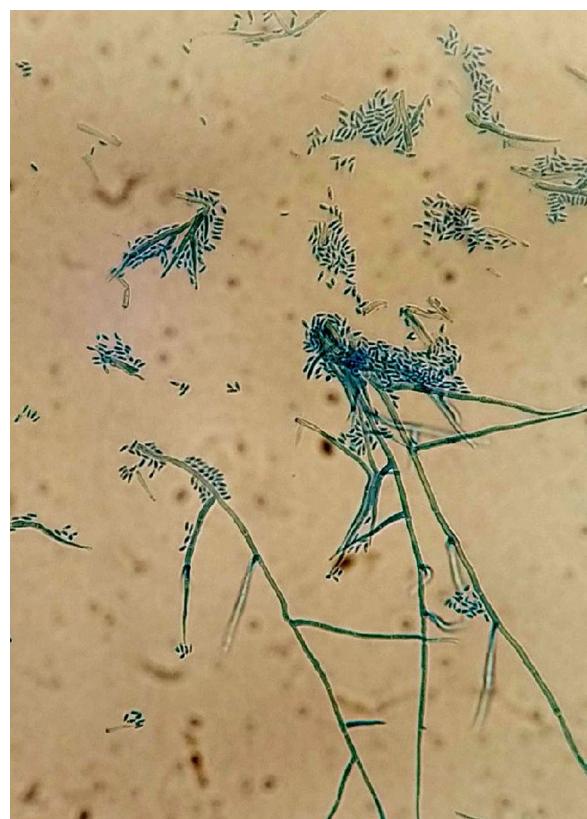
After 10 months, time in which the lesion was still present, the patient went to the dermatology department of our hospital. Physical examination revealed a nodular lesion in the right hand without any systemic symptoms. Oral treatment with itraconazole was started and a debulking surgery was performed. During the surgery, samples for culture and histopathological study were taken. The histopathological examination of the tissue showed aggregates of neutrophils forming microabscesses, a moderate lymphoid infiltrate and numerous granulomas. In the centre of one granuloma, a material of vegetal appearance with a fungal structure consistent with non-septate hyphae was observed (Fig. 1).

Samples were plated on potato dextrose agar and incubated at 30 °C. After 7 days of incubation, colonies of fluffy appearance with irregular borders and olive-grey colour were observed. A direct microscopic examination of the culture with lactophenol cotton blue staining showed hyaline hyphae, thin-walled phialides tapering towards the tip with small funnel-shaped collarettes and hyaline conidia in balls. This morphology is consistent with *Phaeoacremonium* genus (Fig. 2). The identification of the isolate was confirmed by the analysis of the sequence of the gene encoding β-tubulin and a real time panfungal PCR targeting 18S ribosomal RNA gene. DNA extraction was performed as described by Turenne<sup>15</sup> with some changes.<sup>16</sup> For the extraction, vortexing with glass beads to improve the lysis of fungal cells wall was performed. Amplification was performed in an automated PCR-System Smart-cycler (Cepheid, USA) with cycles of 95 °C for 120 s, 45 cycles at 95 °C for 10 s, 52 °C for 30 s and 72 °C for 10 s, using Syber-green to detect the amplified products (Sensifast SYBR Hi-Rox Kit, Bioline, UK). A melting curve analysis was performed in both meth-

The PCR products of both targets were sequenced (BigDye, Applied Biosystem) and the sequences obtained were compared with those available on GenBank using a BLAST search. *P. parasiticum* species identification was obtained; the sequences obtained



**Fig. 1.** Haematoxylin-eosin stain (40× magnification) showing a material of vegetal appearance with a fungal structure of non-septate hyphae in the centre of one the granulomas.



**Fig. 2.** Lactophenol cotton blue stain (40× magnification) of a 7 day-culture of the sample. Hyaline hyphae, thin-walled phialides with small funnel-shaped collarettes, and hyaline conidia in balls are shown.

with the two amplification assays matched two sequences with accession numbers KU375504 and KX268647, with 100% and 99% of similarity respectively.

Antifungal susceptibility testing was performed on Sensititre YeastOne panel (Thermo scientific diagnostic systems, UK) following EUCAST guide.<sup>4</sup> The minimal inhibitory concentration (MIC)

values obtained were the following: voriconazole 0.06 µg/ml, itraconazole 0.12 µg/ml, posaconazole 0.06 µg/ml and amphotericin B 8 µg/ml. The isolate was considered sensitive to azoles and had a decreased sensitivity to amphotericin B. The patient continued with oral itraconazole and progressed favourably.

*P. parasiticum* is an infrequent microorganism that causes infection especially in transplant patients receiving immunosuppressive treatment.<sup>6,8,11,13,14</sup> There are few reports of infections caused by this microorganism, but some reviews suggest an association with kidney transplantation. Up to 36% of infections occurred in kidney transplant patients,<sup>1,3,10</sup> as our case, and in some cases an invasive infection was observed.<sup>16</sup> However, the cause of this association is not yet clear. Patients with several immunocompromising haematological diseases, stem cell transplantation, or rheumatoid arthritis treated with infliximab have also been recognized as a risk group for infections by *P. parasiticum*.<sup>8,11</sup> Although a disseminated infection is rare it can be fatal in some cases. No death related to localized infection has been reported.<sup>11</sup>

Traumatic implantation (i.e. injuries caused by plants) is the main infective route, taking into account that species of *Phaeoacremonium* are ubiquitous in nature.<sup>11</sup> After the trauma, dermis is the mainly affected tissue and, in some cases, the subsequent lesion relapses.<sup>11</sup> The aetiology of phaeohyphomycosis involves a broad spectrum of microorganisms such as *Acrophialophora*, *Alternaria*, *Cladosporium*, or *Exophiala*, among other genera, included *Phaeoacremonium*, being the latter a very rare cause.<sup>1,3</sup> Due to the similar morphology of these microorganisms it is challenging to obtain a reliable identification only under a microscope.<sup>2,6,9,12</sup> Therefore, the sequencing of certain regions of the β-tubulin gene and 18S ribosomal RNA gene is recommended to reach a definitive diagnosis.<sup>2,6</sup> The performance of only microscopic identification may lead to misdiagnoses as happened in our case at the very beginning, when the fungus was identified as *E. dermatitidis*. The combination of molecular and morphological diagnosis is the most appropriate way to achieve a reliable identification.<sup>6</sup>

Antifungal susceptibility testing for *Phaeoacremonium* is not standardized yet, thus caution must be taken when interpreting the results. Although a specific treatment against this microorganism is still unknown, it is believed that azoles can give optimum results. A study of the *in vitro* activity of several antifungals against *P. parasiticum* showed a MIC range of voriconazole 0.125–2 µg/ml, mean 0.55 µg/ml, amphotericin B range 1–16 µg/ml, mean 3.08 µg/ml, and itraconazole range 0.25–32 µg/ml, mean 6.17 µg/ml.<sup>8</sup> Some reports describe a lower activity of itraconazole.<sup>1,3,5,8</sup> However, in our study the strain seemed to be sensitive to itraconazole, and this antifungal was used as part of the patient treatment.

In conclusion, the diagnosis of this infection is difficult. However, there is a need to continue investigating, especially on kidney transplant patients. The combined treatment of surgery and antifungal therapy seems to be the best option to treat this infection.

## References

1. Alayeto J, Fabregó A, Puig Verdie A, Sorli Redo ML, Horcajada Gallego JP, Portillo Bordonabe ME. Feohifomicosis subcutánea causada por *Phaeoacremonium parasiticum*. Rev Iberoam Micol [revista electrónica]. 2015;32:265–8. Available from: <https://doi.org/10.1016/j.riam> [accessed 03.12.16].
2. Aroca A, Raposo R, Lunello P. A biomarker for the identification of four *Phaeoacremonium* species using the β-tubulin gene as the target sequence. Appl Microbiol Biotechnol [revista electrónica]. 2008;80:1131–40.
3. Baddley JW, Mostert L, Summerbell RC, Moser SA. *Phaeoacremonium parasiticum* infections confirmed by β-tubulin sequence analysis of case isolates. J Clin Microbiol [revista electrónica]. 2006;44:2207–11. Available from: <https://doi.org/10.1128/JCM> [accessed 03.12.16].
4. Chairman JL, Arendrup MC, Arikan S, Barchiesi F, Bille J, Schmalreck A, et al. EUCAST método de dilución en caldo para la determinación de las concentraciones mínimas inhibitorias de antifúngicos para hongos filamentosos formadores de conidias; 2016.
5. Chowdhary A, Meis JF, Guarro J, Hoog GS, De Kathuria S, Arendrup MC, et al. ESCMID and ECMM joint clinical guidelines for the diagnosis and management of systemic phaeohyphomycosis: diseases caused by black fungi [revista electrónica]; 2014.
6. Colombi MA, Alanio A, Denis B, Melica G, Garcia-Hermoso D, Levy B, et al. Dual invasive infection with *Phaeoacremonium parasiticum* and *Paraconiothyrium cyclothyrioides* in a renal transplant recipient: case report and comprehensive review of the literature of *Phaeoacremonium* phaeohyphomycosis. J Clin Microbiol [revista electrónica]. 2015;53:2084–94. Available from: <https://doi.org/10.1128/JCM> [accessed 15.12.16].
7. Crous PW, Gams W, Wingfield MJ, van Wyk PS. *Phaeoacremonium* gen. nov. associated with wilt and decline diseases of woody hosts and human infections. Mycologia [revista electrónica]. 1996;88:786–96.
8. El-Herte RL, Schouweiler KE, Farah RS, Arbulu R, Diekema D, Wanat KA, et al. *Phaeoacremonium parasiticum* phaeohyphomycosis in a patient with systemic lupus erythematosus treated successfully with surgical debridement and voriconazole: a case report and review of the literature. ID Cases [revista electrónica]. 2014;1:84–8. Available from: <https://doi.org/10.1016/j.idcr> [accessed 15.12.16].
9. Ellis DT. *Phaeoacremonium parasiticum*. Mycology Online [revista electrónica]; 2017;1–2. Available from: <https://www.mycology.adelaide.edu.au/description/hyphomycetes/phaeoacremonium> [accessed 15.12.16].
10. Marques SA, Camargo RM, Summerbell RC, De Hoog GS, Ishioka P, Chambô-Cordaro LM, et al. Subcutaneous phaeohyphomycosis caused by *Phaeoacremonium parasiticum* in a renal transplant patient. Med Mycol [revista electrónica]. 2006;44:671–6.
11. Mazzurco JD, Ramirez J, Fivenson DP. Phaeohyphomycosis caused by *Phaeoacremonium* species in a patient taking infliximab. J Am Acad Dermatol [revista electrónica]. 2012;66:333–5. Available from: <https://doi.org/10.1016/j.jaad> [accessed 20.12.16].
12. Mostert L, Groenewald JZ, Summerbell RC, Robert V, Sutton DA, Padhye AA, et al. Species of *Phaeoacremonium* associated with infections in humans and environmental reservoirs in infected woody plants. J Clin Microbiol [revista electrónica]. 2005;43:1752–67. Available from: <https://doi.org/10.1128/JCM.43.4.1752-1767> [accessed 03.12.16].
13. Mulcahy H, Chew FS. *Phaeoacremonium parasiticum* myositis: a case report with imaging findings. Radiol Case Rep [revista electrónica]. 2011;6:1–6. Available from: <https://doi.org/10.2484/rccr.v6i2> [accessed 03.12.16].
14. Shah SK, Parto P, Lombard GA, James MA, Beckles DL, Lick S, et al. Probable *Phaeoacremonium parasiticum* as a cause of cavitary native lung nodules after single lung transplantation. Transplant Infect Dis [revista electrónica]. 2013;15:9–13. Available from: <https://doi.org/10.1111/tid> [accessed 20.12.16].
15. Turenne CY, Sanche SE, Hoban DJ, Karlowsky JA, Kabani AM. Rapid identification of fungi by using the ITS2 genetic region and an automated fluorescent capillary electrophoresis system. J Clin Microbiol. 1999;37:1846–51.
16. Villanueva M, Bonany P, García R, Redondo E, Puig J. Utilidad de la PCR panfúngica en el diagnóstico de infecciones por hongos. Científicas. 2017;5:8–11.
17. Mathur VPB, Kumar MS. Phaeohyphomycosis of subcutaneous tissue. Indian J Med Microbiol [revista electrónica]. 1974;27:66–9. Available from: <https://www.ijmm.org> [accessed 11.12.16].