



Note

A surprising finding: The curious case of a tongue lesion misdiagnosed as paracoccidioidomycosis

Bert Gerrits van den Ende^a, Anderson Messias Rodrigues^{b,c}, Rosane Christine Hahn^{d,e}, Ferry Hagen^{a,f,g,*}

^a Westerdijk Fungal Biodiversity Institute, Utrecht, The Netherlands

^b Laboratory of Emerging Fungal Pathogens, Department of Microbiology, Immunology, and Parasitology, Discipline of Cellular Biology, Federal University of São Paulo (UNIFESP), São Paulo, Brazil

^c Department of Medicine, Discipline of Infectious Diseases, Federal University of São Paulo (UNIFESP), São Paulo, Brazil

^d Laboratory of Mycology/Research, Faculty of Medicine, Federal University of Mato Grosso, Cuiabá, Mato Grosso, Brazil

^e Júlio Muller University Hospital – Eberh, Federal University of Mato Grosso, Cuiabá, Mato Grosso, Brazil

^f Institute for Biodiversity and Ecosystem Dynamics (IBED), University of Amsterdam, Amsterdam, The Netherlands

^g Department of Medical Microbiology, University Medical Center Utrecht, Utrecht, The Netherlands

ARTICLE INFO

Article history:

Received 10 August 2022

Accepted 23 November 2022

Available online 21 January 2023

Keywords:

Paracoccidioides

Nannizziopsis

Nanopore long-read sequencing

Mis-identification

ABSTRACT

Background: Paracoccidioidomycosis is an endemic mycosis caused by members of the *Paracoccidioides* genus. Brazil remains the focus area and, to a lesser extent, the disease has been reported from Argentina, Colombia and Venezuela.

Aims: A Venezuelan *Paracoccidioides brasiliensis* strain, isolated from a patient diagnosed with chronic multifocal paracoccidioidomycosis, was subjected to whole genome sequencing to provide more insight about *Paracoccidioides* outside the endemic focus area.

Methods: *P. brasiliensis* strain CBS 118890 was whole genome sequenced using nanopore; library preparation with the ‘native barcoding genomic DNA kit’ was followed by sequencing on Flongle and MinION flowcells. Batches of strain CBS 118890 were re-identified by sequencing the internal transcribed spacer (ITS) region, and final identification was made based on phylogenetic analysis.

Results: Surprisingly, the Venezuelan *P. brasiliensis* strain CBS 118890 turned out to be a *Nannizziopsis* species. The batches of this strain were ITS sequenced followed by phylogenetic analysis and resulted in the final identification of *Nannizziopsis arthrosporioides*.

Conclusions: *Nannizziopsis* infections are commonly seen in a wide variety of reptiles, but are particularly rare in human infections. This case underlines the need for molecular characterization of cases that clinically mimic paracoccidioidomycosis but that are serologically negative for *Paracoccidioides*.

© 2022 The Author(s). Published by Elsevier España, S.L.U. on behalf of Asociación Española de Micología. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Un hallazgo inesperado: el curioso caso de una lesión en la lengua diagnosticada erróneamente como paracoccidioidomycosis

RESUMEN

Antecedentes: La paracoccidioidomycosis es una micosis endémica causada por especies del género *Paracoccidioides*. Brasil sigue siendo el área con la mayor incidencia y, en menor medida, se ha informado de casos en Argentina, Colombia y Venezuela.

Objetivos: Una cepa venezolana de *Paracoccidioides brasiliensis*, obtenida de un paciente diagnosticado con paracoccidioidomycosis multifocal crónica, se sometió a secuenciación completa del genoma para obtener más información sobre *Paracoccidioides* fuera del área de foco endémico.

Palabras clave:

Paracoccidioides

Nannizziopsis

Secuenciación de nanoporos de lectura larga

Identificación errónea

* Corresponding author.

E-mail address: f.hagen@wi.knaw.nl (F. Hagen).

Métodos: Se secuenció el genoma completo de la cepa CBS 118890 de *P. brasiliensis* mediante la técnica de secuenciación de nanoporos; tras la preparación de la librería con el «native barcoding genomic DNA kit» se procedió a la secuenciación con el Flongle y MinION flowcells. Los lotes de la cepa CBS 118890 se volvieron a identificar mediante la secuenciación de la región del espaciador transcrito interno (ITS), y la identificación final se realizó en función del análisis filogenético.

Resultados: Sorprendentemente, la cepa venezolana *P. brasiliensis* CBS 118890 resultó ser una especie de *Nannizziopsis*. Los lotes de esta cepa se secuenciaron mediante ITS seguido de un análisis filogenético y dieron como resultado la identificación de la especie *Nannizziopsis arthrosporioides*.

Conclusiones: Las infecciones por *Nannizziopsis* se observan comúnmente en una amplia variedad de reptiles, pero son particularmente raras en infecciones humanas. Este caso subraya la necesidad de la caracterización molecular de los casos que clínicamente reflejan paracoccidioidomicosis, pero que son serológicamente negativos para *Paracoccidioides*.

© 2022 El Autor(s). Publicado por Elsevier España, S.L.U. en nombre de Asociación Española de Micología. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

The order Onygenales harbours a large number and wide variety of human fungal pathogens, from dermatophytes causing superficial infections to the life-threatening pathogens that fall under the umbrella of endemic mycoses. The latter includes the genera *Blastomyces*, *Emergomyces*, *Histoplasma* and *Paracoccidioides* (family Ajellomycetaceae), and *Coccidioides* (family Onygenaceae).⁵ Of these fungi, *Paracoccidioides* is exclusively reported from Latin American countries, being represented by seven species: *Paracoccidioides americana*, *Paracoccidioides brasiliensis*, *Paracoccidioides restrepiensis* and *Paracoccidioides venezuelensis* (members of the *Paracoccidioides brasiliensis* species complex), *Paracoccidioides loboii*, *Paracoccidioides lutzii* and *Paracoccidioides ceti*.^{4,6,17,21} The proposed revision of the *Paracoccidioides* taxonomy was supported by genome sequencing studies.^{11,14,20}

In 2005, the CBS culture collection, hosted at the Westerdijk Fungal Biodiversity Institute (Utrecht, The Netherlands), received a *P. brasiliensis* strain from Venezuela that was obtained from a tongue lesion of a patient who was diagnosed with chronic multifocal paracoccidioidomycosis, for which itraconazole (200 mg per day) was administered. Unfortunately, given the time that had elapsed since the deposition of the strain, no further clinical information could be obtained. As *Paracoccidioides* strains from Venezuela were not yet commonly available with whole genome sequence data, we decided to perform long-read nanopore sequencing. For this purpose, the strain was subcultured into 10 ml malt extract broth and incubated for two weeks at 28 °C. Cells were harvested by centrifugation, followed by genomic DNA extraction as previously described.¹⁵ Nanopore library preparation was performed using the 'native barcoding genomic DNA kit' (EXP-NBD114 and SQK-LSK109; ONT, Oxford, UK), and the readings were generated with a pre-run – to check the quality of the prepared library – on a Flongle flow cell (FLO-FLG001; ONT), followed by a run onto a MinION flow cell (FLO-MIN106; ONT). Raw data were base called with high-accuracy settings and demultiplexed using Guppy v.6.0.1+652ffd179 (ONT). The FASTQ data from both runs were combined and subsequently subjected to *de novo* genome assembly using Flye v.2.9-b1768.⁸ The observed genome size was 24,246,093 bp, consisting of 6 contigs representing likely 5 chromosomes plus the mitochondrial genome. The lengths of these 5 chromosomes were 7,230,366 bp; 6,417,451 bp (=N50); 4,132,340 bp; 4,046,396 bp, and 2,395,022 bp. The circular mitochondrial genome was 24,518 bp in length. The coverage of the nuclear genome was 35X and that of the mitochondrial genome 218X. Data were deposited in NCBI Genome under the accession numbers BioProject PRJNA843904, BioSample SAMN28772366, Sequence Read Archive (SRA) SRR19450098 and SRR19450099, and the genome assembly CP098259–CP098264.

However, when processing the data, it became clear that the sequenced strain CBS 118890 was not *P. brasiliensis* but the distantly related species *Nannizziopsis arthrosporioides* (family Onygenaceae), known to behave pathogenic in reptiles.^{2,18} All available batches of this strain, including the original material stored as oil-preserved slant, were obtained from the CBS culture collection to rule out contamination or erroneous change of material. These strains were all subjected to sequencing the internal transcribed spacer (ITS) region of the ribosomal DNA using primers ITS1 and ITS4, as previously described.⁵ All batches resembled *N. arthrosporioides* when ITS sequences were identified against the NCBI nucleotide database (all ITS sequences identical, NCBI GenBank accession number ON725014). We extracted ITS data from the NCBI nucleotide database and performed phylogenetic analyses. Sequences were aligned using MAFFT v7.490, with the strategy setting set to 'FFT-NS-j'. The phylogenetic tree was built using IQ-tree v1.6.12 with the settings '-redo -nt AUTO -nm 1000 -m MFP+MERGE -abayes -alrt 1000 -bb 1000' and substitution model GTR+F+G4 was automatically calculated by IQ-tree on the basis of Akaike Information Criterion.^{7,12} In the phylogenetic tree depicted in Fig. 1, strain CBS 118890 clustered tightly together with the other *N. arthrosporioides* isolates, including the type-strain sequence. Unequivocally, strain CBS 118890 represents *N. arthrosporioides*, and the original identification as *P. brasiliensis* was erroneously made based on the clinical presentation. Culture characteristics were also atypical for *Paracoccidioides*, as can be observed by the colony macro- and micro-characteristics of the CBS 118890 strain in Fig. 2.

At present, there are twelve species recognized within *Nannizziopsis*: *N. arthrosporioides*, *Nannizziopsis barbatae*, *Nannizziopsis chlamydospora*, *Nannizziopsis crocodili*, *Nannizziopsis dermatitidis*, *Nannizziopsis draconii*, *Nannizziopsis guarroi*, *Nannizziopsis hominis*, *Nannizziopsis infrequens*, *Nannizziopsis obscura*, *Nannizziopsis pluriseptata*, and *Nannizziopsis vriesii*.^{18,19} The dreaded cause of fungal snake disease, *Ophidiomyces ophidiicola*, is closely related to the genus *Nannizziopsis*.⁹ *Nannizziopsis* infections are commonly seen in a wide variety of reptiles, but are particularly rare in human infections. A handful of human cases caused by *N. guarroi*, *N. hominis*, *N. infrequens*, *N. obscura*, and undetermined *Nannizziopsis* species have been reported.^{1,3,10,13,16,18,19,22} Retrospectively, we can add a human case of *N. arthrosporioides* infection to this list, an infection that was mimicked as paracoccidioidomycosis. So far, most *Nannizziopsis* cases in humans have been documented from people living in – or recently immigrated from – Western Africa. The presented case from Venezuela underlines the need for further molecular characterization in cases that clinically reflect paracoccidioidomycosis but that are serologically negative for *Paracoccidioides*, as they might be *Nannizziopsis* infections.

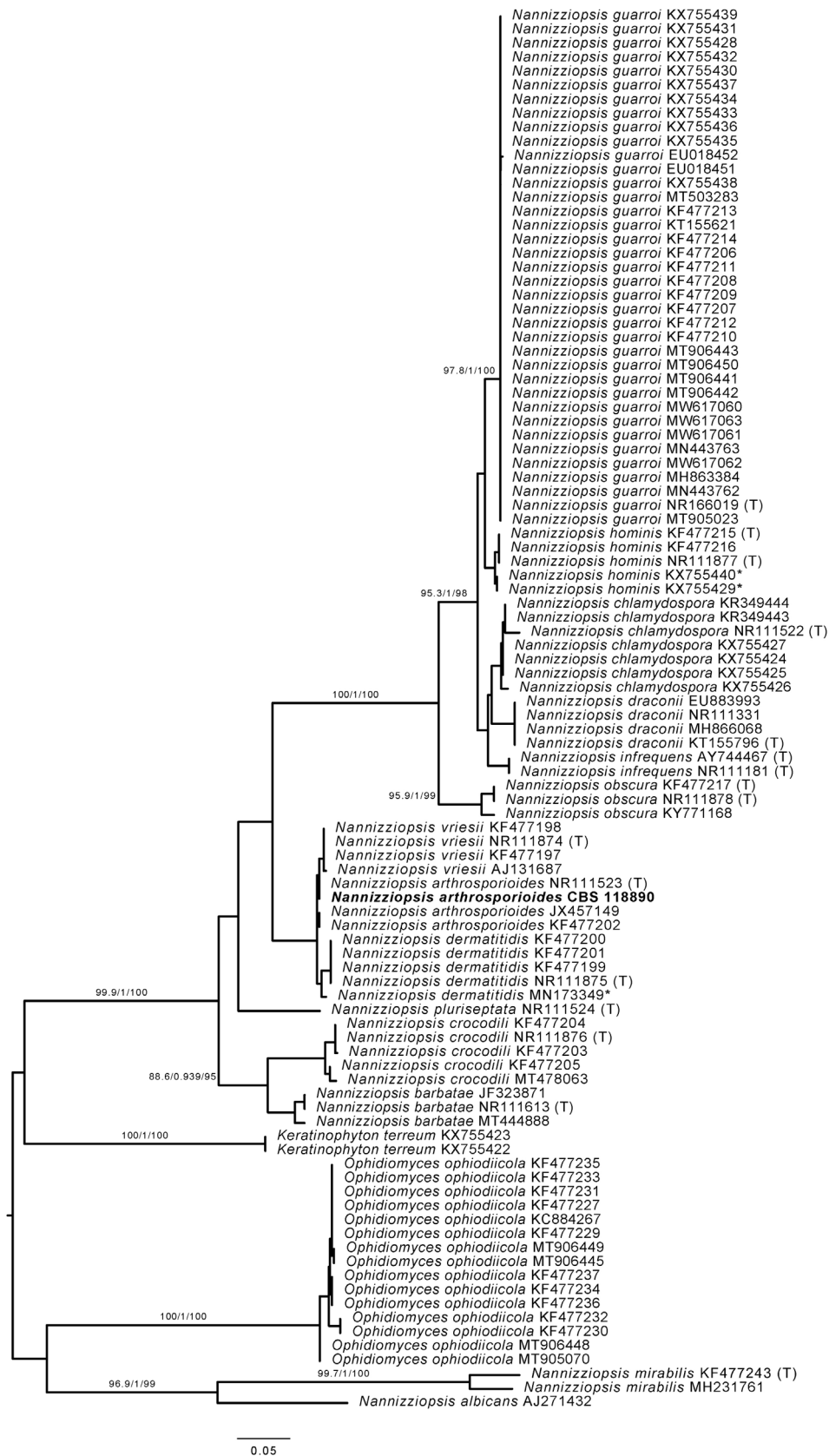


Fig. 1. Phylogenetic analysis of *Nannizziopsis* strains. * = Sequences KX755429 and KX755440 were originally deposited as *Nannizziopsis guarroi* but cluster together with *Nannizziopsis hominis*. Sequence MN173349 was originally identified by Chen and colleagues (2021) as *Nannizziopsis arthrosporioides* but is now placed within the group of *Nannizziopsis dermatitidis* strains.² (T) = Sequence originated from type-strain of specified species. The scale bar represents the nucleotide substitution ratio. Numbers next to major branches and values >85/0.85/85 represent the support of the likelihood ratio test, Bayesian interference, and bootstrapping, respectively.¹¹ *Nannizziopsis arthrosporioides* strain CBS 118890 is highlighted in bold.

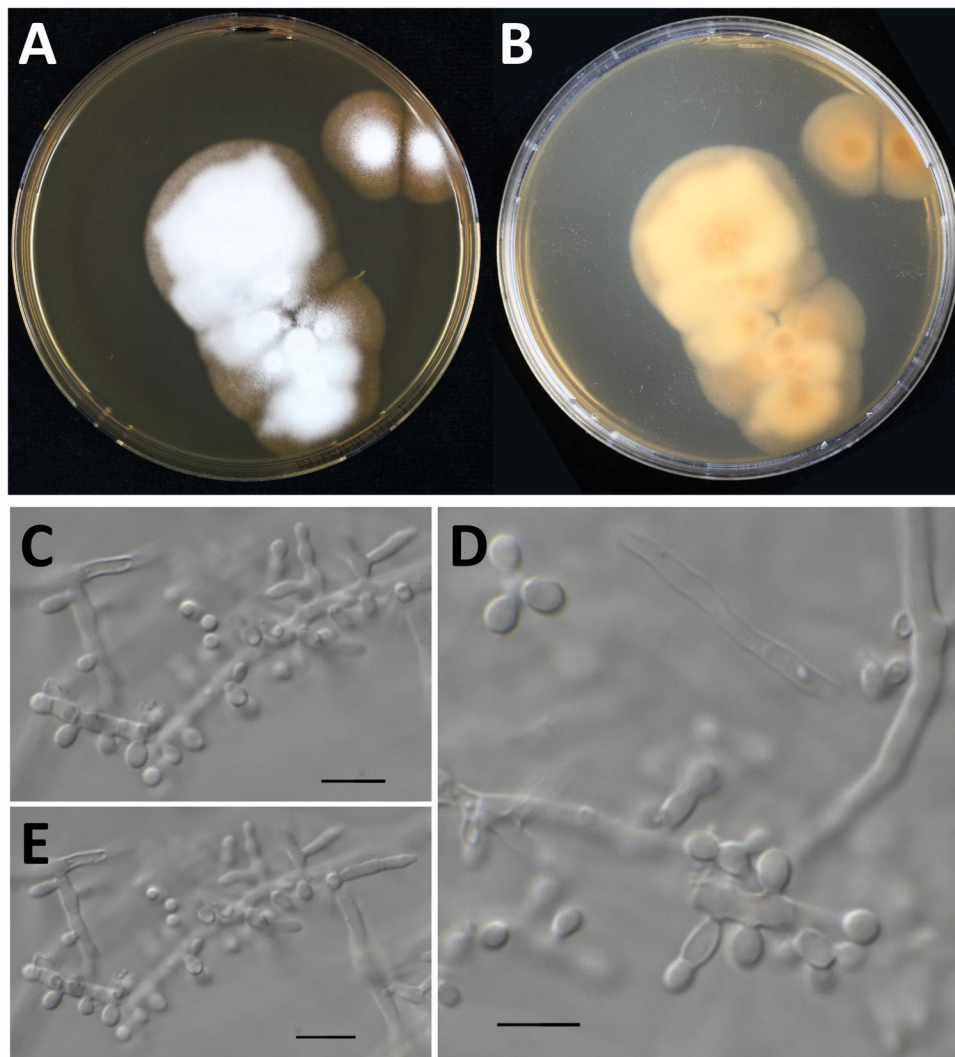


Fig. 2. Culture characteristics of *Nannizziopsis arthrosporioides* CBS 118890. Panels A and B: Culture onto malt extract agar incubated for two-weeks at 35 °C, colony is about 5 cm in diameter, velvety to powdery (panel A, obverse; panel B, reverse). Panels C–E: Microscopy of a two-week old *N. arthrosporioides* culture showing aleuriospores and subglobose spores, scale bar = 10 μ m.

References

1. Baggott A, McGann H, Barton R, Ratner J. Disseminated *Nannizziopsis obscura* infection in a renal transplant patient – the first reported case. *Med Mycol Case Rep.* 2017;17:20–4. <http://dx.doi.org/10.1016/j.mmcr.2017.06.002>.
2. Chen YH, Chi MJ, Sun PL, Yu PH, Liu CH, Cano-Lira JF, et al. Histopathology, molecular identification and antifungal susceptibility testing of *Nannizziopsis arthrosporioides* from a captive Cuban Rock Iguana (*Cyclura nubila*). *Mycopathologia.* 2020;185:1005–12. <http://dx.doi.org/10.1007/s11046-020-00481-6>.
3. Garcia-Hermoso D, Hamane S, Fekkar A, Jabet A, Denis B, Siguier M, et al. Invasive infections with *Nannizziopsis obscura* species complex in 9 patients from West Africa, France, 2004–2020. *Emerg Infect Dis.* 2020;26:2022–30. <http://dx.doi.org/10.3201/eid2609.200276>.
4. Hahn RC, Hagen F, Mendes R, Burger E, Nery AF, Siqueira N, et al. Paracoccidioidomycosis: current status and future trends. *Clin Microbiol Rev.* 2022;35:e0023321. <http://dx.doi.org/10.1128/cmr.00233-21>.
5. Irinyi L, Serena C, Garcia-Hermoso D, Arabatzis M, Desnos-Ollivier M, Vu D, et al. International Society of Human and Animal Mycology (ISHAM)-ITS reference DNA barcoding database – the quality controlled standard tool for routine identification of human and animal pathogenic fungi. *Med Mycol.* 2015;53:313–37. <http://dx.doi.org/10.1093/mmy/myv008>.
6. Kandemir H, Dukik K, de Melo Teixeira M, Stielow JB, Zohra Delma F, Al-Hatmi AMS, et al. Phylogenetic and ecological reevaluation of the order *Onygenales*. *Fungal Div.* 2022. <http://dx.doi.org/10.1007/s13225-022-00506-z>.
7. Katoh K, Rozewicki J, Yamada KD. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Brief Bioinform.* 2019;20:1160–6. <http://dx.doi.org/10.1093/bib/bbx108>.
8. Kolmogorov M, Yuan J, Lin Y, Pevzner PA. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol.* 2019;37:540–6. <http://dx.doi.org/10.1038/s41587-019-0072-8>.
9. Lorch JM, Knowles S, Lankton JS, Michell K, Edwards JL, Kapfer JM, et al. Snake fungal disease: an emerging threat to wild snakes. *Philos Trans R Soc Lond B Biol Sci.* 2016;371:20150457. <http://dx.doi.org/10.1098/rstb.2015.0457>.
10. Mascitti H, Sivadon-Tardy V, Bougnoux ME, Duran C, Tordjman M, Colombier MA, et al. Arthritis caused by *Nannizziopsis obscura*, France. *Emerg Infect Dis.* 2022;28:1929–31. <http://dx.doi.org/10.3201/eid2809.220375>.
11. Mavengere H, Mattox K, Teixeira MM, Sepúlveda VE, Gomez OM, Hernandez O, et al. *Paracoccidioides* genomes reflect high levels of species divergence and little interspecific gene flow. *mBio.* 2020;11:e01999–2020. <http://dx.doi.org/10.1128/mBio.01999-20>.
12. Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, von Haeseler A, et al. IQ-TREE 2 new models and efficient methods for phylogenetic inference in the genomic era. *Mol Biol Evol.* 2020;37:1530–4. <http://dx.doi.org/10.1093/molbev/msaa015>.
13. Most ZM, Lieu T, Filkins L, Nicolaiades R, Rakheja D, Gelfand A, et al. Disseminated *Nannizziopsis* infection in an adolescent with a STAT1 mutation. *Open Forum Infect Dis.* 2020;7:ofaa390. <http://dx.doi.org/10.1093/ofid/ofaa390>.
14. Muñoz JF, Farrer RA, Desjardins CA, Gallo JE, Sykes S, Sakthikumar S, et al. Genome diversity, recombination, and virulence across the major lineages of *Paracoccidioides*. *mSphere.* 2016;1:e00213–6. <http://dx.doi.org/10.1128/mSphere.00213-16>.
15. Navarro-Muñoz JC, de Jong AW, Gerrits van den Ende B, Haas PJ, Then ER, Mohd Tap R, et al. The high-quality complete genome sequence of the opportunistic fungal pathogen *Candida vulturna* CBS 14366T. *Mycopathologia.* 2019;184:731–4. <http://dx.doi.org/10.1007/s11046-019-00404-0>.

16. Nourrisson C, Vidal-Roux M, Cayot S, Jacomet C, Bothorel C, Ledoux-Pilon A, et al. Invasive infections caused by *Nannizziopsis* spp. molds in immunocompromised patients. *Emerg Infect Dis.* 2018;24:549–52, <http://dx.doi.org/10.3201/eid2403.170772>.
17. Roberto TN, de Carvalho JA, Beale MA, Hagen F, Fisher MC, Hahn RC, et al. Exploring genetic diversity, population structure, and phylogeography in *Paracoccidioides* species using AFLP markers. *Stud Mycol.* 2021;100:100131, <http://dx.doi.org/10.1016/j.simyco.2021.100131>.
18. Sigler L, Hambleton S, Paré JA. Molecular characterization of reptile pathogens currently known as members of the *Chrysosporium* anamorph of *Nannizziopsis vriesii* complex and relationship with some human-associated isolates. *J Clin Microbiol.* 2013;51:3338–57, <http://dx.doi.org/10.1128/JCM.01465-13>.
19. Stchigel AM, Sutton DA, Cano-Lira JF, Cabañes FJ, Abarca L, Tintelnot K, et al. Phylogeny of *Chrysosporia* infecting reptiles: proposal of the new family *Nannizziopsiaceae* and five new species. *Persoonia.* 2013;31:86–100, <http://dx.doi.org/10.3767/003158513X669698>.
20. Teixeira MM, Cattana ME, Matute DR, Muñoz JF, Arechavala A, Isbell K, et al. Genomic diversity of the human pathogen *Paracoccidioides* across the South American continent. *Fungal Genet Biol.* 2020;140:103395, <http://dx.doi.org/10.1016/j.fgb.2020.103395>.
21. Vilela R, Huebner M, Vilela C, Vilela G, Pettersen B, Oliveira C, et al. The taxonomy of two uncultivated fungal mammalian pathogens is revealed through phylogeny and population genetic analyses. *Sci Rep.* 2021;11:18119, <http://dx.doi.org/10.1038/s41598-021-97429-7>.
22. Zhao Y, Nozdrin M, Dalla Pria A, Bracchi M. *Nannizziopsis* immune reconstitution inflammatory syndrome in a patient with HIV: first reported case. *Eur J Case Rep Intern Med.* 2021;8:003021, <http://dx.doi.org/10.12890/2021.003021>.