



## Food Microbiology

# Zearalenone adsorption capacity of lactic acid bacteria isolated from pigs



María F. Vega<sup>a,\*</sup>, Susana N. Dieguez<sup>a,b,d</sup>, Belén Riccio<sup>b</sup>, Sandra Aranguren<sup>b</sup>, Antonio Giordano<sup>b</sup>, Laura Denzoin<sup>b</sup>, Alejandro L. Soraci<sup>b</sup>, María O. Tapia<sup>b</sup>, Romina Ross<sup>c</sup>, Ana Apás<sup>c</sup>, Silvia N. González<sup>c</sup>

<sup>a</sup> Departamento de Tecnología y Calidad de los Alimentos, Facultad de Ciencias Veterinarias, Universidad Nacional del Centro de la Provincia de Buenos Aires, Tandil, Buenos Aires, Argentina

<sup>b</sup> Laboratorio de Toxicología CIVETAN – CONICET, Facultad de Ciencias Veterinarias, Universidad Nacional del Centro de la Provincia de Buenos Aires, Tandil, Buenos Aires, Argentina

<sup>c</sup> Departamento de Salud Pública, Facultad de Bioquímica Química y Farmacia, Universidad Nacional de Tucumán, San Miguel de Tucumán, Tucumán, Argentina

<sup>d</sup> Comisión de Investigaciones Científicas de la Provincia de Buenos Aires CICPBA

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## ABSTRACT

The ability to adsorb zearalenone by five strain of lactic acid bacteria was evaluated: four strains of *Lactobacillus* spp. isolated from pig rectal swabs and one commercial strain (*Lactobacillus rhamnosus*). Several factors affecting the adsorption capacity were evaluated in order to improve the adsorption of the mycotoxin by bacteria. The stability of the zearalenone–bacteria complex was analyzed. In every case, bacterial adsorption capacity was higher than 40.0%. The strain showing the highest adsorption (68.2%) was selected for the following steps of this research. The adsorption percentages obtained after processing 6.5 and 7.5 mL MRS broth were 57.40% + 3.53 and 64.46% + 0.76, respectively. The stability of zearalenone–bacteria complex was evaluated by successively rinsing. In the first rinsing step 42.26% + 0.414 was still bound. In the second rinsing step 25.12% + 0.664 was still bound, whereas 15.82% + 0.675 remained in the pellet after the third rinse. Results obtained demonstrated that Lactic Acid Bacteria has capacity to adsorb zearalenone. Finally adsorption was increased using a higher volume of initial broth. These results could be used to design a new lyophilized powder for detoxification, using lactic acid bacteria as potential zearalenone adsorbents.

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\* Corresponding author.

E-mail: [mfvega@vet.unicen.edu.ar](mailto:mfvega@vet.unicen.edu.ar) (M.F. Vega).

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## Introduction

Zearalenone (ZEA) is a worldwide distributed mycotoxin as indicated by the International Agency for Research on Cancer (IARC).<sup>1–3</sup> Its toxicity and incidence was confirmed by recent reports.<sup>4,5</sup> This resorcylic lactone is produced mainly by *Fusarium graminearum* and is one of the most important toxins causing serious reproductive failures in pig production, due to its ability to couple 17- $\beta$ -estradiol receptors. This interference with cytosolic estrogen receptors of target cells makes it an endocrine disruptor. When the dangerousness of this mycotoxin is considered, two factors are important: its stability to heat, milling, storage and processing of feedstuffs,<sup>6</sup> and the severity of the intoxication, even if the ingestion takes place only for a few days as indicated by Anadón et al.<sup>7</sup>

Several physicochemical methods for detoxification have been widely used, but the tendency is to use biological methods that do not cause nutritional and palatability changes in feed.<sup>8</sup> There are several food additives for decontamination and/or detoxification of animal feedstuffs, such as the commercial adsorbent Mycosorb<sup>®</sup> (Alltech Inc.), in which the structure of yeast glucan layer is modified to increase its affinity and binding rate of mycotoxins above that of the native cell wall product.<sup>9</sup> In spite of its greater efficacy, the use of Mycosorb is limited because of its high price.

Interactions between *Fusarium* mycotoxins, ZEA, its derivative  $\alpha$ -zearalenol ( $\alpha$ -ZOL) and food-grade strains of *Lactobacillus* have been reported by several researchers.<sup>10–17</sup> El-Nezami et al.<sup>13</sup> demonstrated that, after co-incubation of ZEA and *Lactobacillus*, a considerable proportion (38.0%–46.0%) of these mycotoxins were recovered from bacterial pellets. Bacteria showed capacity to adsorb toxins and, as expected, results demonstrated that the binding depended on bacterial concentration. Co-incubation of ZEA and  $\alpha$ -ZOL with bacteria significantly affected the percentage of toxin bound, suggesting that these toxins may share the same binding site. Recently Tinyiro et al.<sup>18</sup> using some *Bacillus* strains, showed that the adsorption percentage was high (78.0% and 95.0%) and variation depended on the strain used. ZEA adsorption ability by *Planococcus* spp has also been demonstrated by Lu et al.<sup>19</sup>

Two fundamental rules are considered when bacterial strains are isolated in order to be administered to animals. The first is the host specificity, it is essential to achieve a good adaptation.<sup>20,21</sup> The second is the proximity of the ecosystem, it is vital that the microorganisms are isolated in the same place where it acts on the host.<sup>22,23</sup>

For all these reasons, the purpose of the present work was to study the ZEA adsorption capacity of lactic acid bacteria isolated from pig rectal swabs under *in vitro* conditions, in order to broaden scientific knowledge on feed additives based on the use of lactic acid bacteria as ZEA binders.

## Materials and methods

### Bacterial strains and culture conditions

*Lactobacillus rhamnosus* (R1) (Lyofast LR B Sacco) was purchased from Sacco SRL (Cadorago, Italy). This strain was used

as reference because of its demonstrated ability to adsorb ZEA.<sup>12,13,24</sup>

The tested *Lactobacillus* strains (L1, L2, L3, and L4) were isolated from 7 piglets kept on a single farm. The piglets, aged 21–60 days, were clinically healthy and no antibiotics had been administered during their lives. All samples were obtained within the same day. The strains were collected with a kit containing a swab and a transport medium (LAPTg semisolid agar without sugars). After being delivered to the laboratory, the swabs were broken off into peptone water, the dilutions were made and inoculated onto solid medium: LAPTg Agar,<sup>25–27</sup> then the plates were incubated in microaerophilia (candle jar system) at 37 °C for 48 h.

Strains of lactic acid bacteria (LAB): L1, L2, L3 and L4 were isolated at the “Laboratorio de Toxicología” at the “Universidad Nacional del Centro de la Provincia de Buenos Aires” Tandil, Buenos Aires, Argentina. LAB strains were kept at –20 °C in LAPTg broth (1.5% peptone, 1% tryptone, 1% glucose, 1% yeast extract, and 0.1% Tween 80), pH 6.8<sup>26</sup> containing 15% glycerol (v/v) until use.

### Reactivation process

The strains were defrosted at 37 °C for 15 min, then they were inoculated and incubated in LAPTg broth for 24 h at 37 °C in microaerophilia before genotypic an species identification. Before adsorption process two reactivation times were tested: 24 h ( $t_1$ ) and 72 h ( $t_2$ ).

### Genotypic identification

The strains isolated for this work L1, L2, L3 and L4, were characterized as lactobacilli by PCR, according to the methodology described by Dubernet et al.<sup>28</sup> Industrial strains provided by Sacco S.A.: *L. rhamnosus* (R1), *Lactobacillus delbrueckii* (R2), *Lactobacillus casei* (R3) and *Lactobacillus helveticus* (R4) were used only for genotypic identification as positive control. Identification was made by amplifying specific region of DNA. The amplification products were visualized by electrophoresis in agarose gels stained with ethidium bromide.<sup>29</sup> General primers for lactobacilli LbLMA1-rev (5'-CTC AAA ACT TTC AAA CAA AGT-3' and R16-1 5'-CTT GTA CAC ACC GCC CGT CA-3') were used.<sup>28</sup>

### Species identification

Genotypic identification was carried out by partial 16S rRNA gene sequencing. Genomic DNA was extracted according to Pospiech and Neumann.<sup>30</sup> Oligonucleotide primers (PLB16, 5-AGAGTTTGATCCTGGCTCAG-3, and MLB16, 5-GGCTGCTGGCAGTAGTTAG-3) were used to amplify the variable (V1) region of the 16S ribosomal RNA gene according to the protocol described by several authors.<sup>31–33</sup> PCR products were electrophoresed in 1% (wt/vol) agarose gels, stained and visualized as described above. Amplicons were excised from the gel and purified using a GFX PCR DNA gel band purification kit (GE Healthcare, UK). Purified PCR products were sequenced at CERELA-CONICET, Tucumán, Argentina, by using an ABI 3130 DNA sequencer (Applied Biosystems, Foster, CA). rRNA gene sequence alignments were performed using the multiple sequence alignment method<sup>34,35</sup> and

identification queries were fulfilled by a BLAST search<sup>36</sup> in GenBank (<http://www.ncbi.nlm.nih.gov/GenBank/>) and in the Ribosomal Database Project.<sup>37</sup>

#### Determination of Growth by Optical Density (OD<sub>560 nm</sub>)

The quantifying of the total number of cells for each strain (R1, L1, L2, L3, and L4) was made using a turbidimetric method. The OD<sub>560 nm</sub> of the bacterial suspension includes living cells, dead cells, and noncellular material that absorbs/scatters the light and indicates the total mass achieved. The wavelength of the spectrophotometer (UV-Vis Spectrophotometer Pharmacia LKB Biochrom Limited, Ultrospect III, Cambridge, England) was set at 560 nm. One milliliter of the sterile broth which was used to grow bacteria (LAPTg or MRS) was used to calibrate the equipment to zero absorbance and 1 mL of each sample was used to measure the OD<sub>560 nm</sub>.

#### Pellets adsorption process

In first place a stock ZEA solution was prepared by dissolving 25 mg ZEA (99.5% purity, Sigma, St. Louis, USA) in 125 mL chloroform (HPLC grade, Biopack, Argentina) to obtain a final concentration of 200 µg/mL. This solution was kept at -18 °C until its use. 95 µL of the stock ZEA solution were evaporated to dryness under an N<sub>2</sub> current at 60 °C. The dry residue was resuspended in 50 µL of methanol (HPLC grade, Sintorgan, Argentina) and 1500 µL of a 0.9% NaCl (Sigma, St. Louis, USA) (SP1) or 0.9% CaCl<sub>2</sub> (Sigma, St. Louis, USA) (SP2) to give a ZEA concentration of 12.3 µg/mL in both solutions.

In second place the adsorption pellets were prepared. LAB were activated and grown in LAPTg medium. Cultures were incubated in microaerophilia (candle jar system) for 72 h at 37 °C. OD<sub>560 nm</sub> was measured, then bacteria were harvested by centrifugation (Sigma Laborzentrifugen 1K15, St. Louis, United States) at 5900 × g, 4 °C, 15 min.

Finally bacterial pellets were rinsed twice with NaCl 0.9% solution and then resuspended in 1.5 mL of SP1 solution or SP2 solution accordingly and incubated for 1 hour at 37 °C, with gentle stirring every 30 min as indicated by Bueno.<sup>11</sup>

After incubation, the solutions were centrifuged (5900 × g, 4 °C, 15 min), then the supernatant was filtered through a 0.22 µm nylon membrane and analyzed by HPLC–UV.<sup>11,38</sup>

As a control to demonstrate that the adsorption process recovers all the mycotoxin reference strain (R1) was used. ZEA in the supernatant and in the pellet (extracted with methanol) were quantified, these two concentrations were added and compared with the initial mycotoxin.

#### ZEA determination by HPLC

The HPLC system was equipped with a Gilson 151 UV-Vis detector, and Gilson 712 software (Gilson, Inc., Middleton, USA). Separation was achieved by HPLC, using a C18 stationary phase, 250 mm × 4.6 mm i.d., 5 µm column. The mobile phase consisted of acetonitrile (HPLC grade, Sintorgan, Villa Marteli, Argentina):water (ultra purified, Elga) mixture (55:45) working in isocratic mode at 1.2 mL/min. The column was maintained at 30 °C. The injection volume was 20 µL and the

chromatographic run time was 20 min. Detection was carried out at 236 nm.<sup>39</sup> To quantify ZEA using external standard calibration seven concentrations 0.63, 1.25, 2.5, 5, 10, 15 and 20 µg/mL were prepared with standards (Sigma St. Louis, USA), the responses were found to be linear (coefficient of correlation >0.998). Recorded retention times for β-ZOL, α-ZOL and ZEA were 6.70 min; 9.24 min; and 17.19 min respectively.

#### Adsorption percentage (ADS%) quantification

ADS% was quantified by comparing mycotoxins peak areas in the supernatant obtained after the adsorption process to peak areas for SP1 or SP2 solutions without bacterial addition, representing positive controls. The ADS% of ZEA was calculated according to Eq. (1).<sup>38,40</sup>

$$ADS\% = 100 - \frac{AUC_2 * 100}{AUC_1} \quad (1)$$

Being:

ADS%: adsorption percentage.

AUC<sub>2</sub>: peak area of the mycotoxin in the supernatant after the adsorption process.

AUC<sub>1</sub>: peak area of the mycotoxin in the positive control (SP1 or SP2).

#### Strain selection

Two parameters were used for LAB strain selection: The OD<sub>560 nm</sub> achieved after the reactivation process, used as indicator of mass production, and ADS%. We used both variables following Eq. (2) because we consider that they have the same importance for strain selection.

$$X = \frac{OD_{560 nm} * 0.5 + ADS\% * 0.5}{0.5 + 0.5} \quad (2)$$

Being:

X = mean of OD<sub>560 nm</sub> and ADS%.

OD<sub>560 nm</sub> = optical density.

ADS% = adsorption percentage.

#### Optimization of adsorption process conditions

With the aim of increasing adsorption percentage, using the selected strain, four variables of the technique developed by Bueno<sup>11</sup> were evaluated.

- (i) Culture media used in the reactivation process: bacterial growth was compared in two culture media: MRS and LAPTg. First bacterial growth was determined by OD<sub>560 nm</sub> and cells were observed by scanning electron microscopy (SEM) (JEOL 35 CF from JEOL Ltd., Tokyo, Japan). Finally bacteria were cultured until OD<sub>560 nm</sub> = 1 was reached. This value was standardized for every strain. The adsorbent pellets were obtained for both culture media and the ADS% were calculated.
- (ii) Contact solution: the effects of solutions SP1 and SP2 on adsorption percentage were compared.
- (iii) Initial volume of culture media: the effects of an initial volume of 7.5 mL and 6.5 mL were compared.

(iv) Viable and non-viable strain: the effects of sterilizing (120 °C, 15 min) and non-sterilizing strain were compared.

### Evaluation of the selected method

Using data from 3 samples on 4 separate days of assay the selected method was also evaluated. The precision was expressed by the relative standard deviation (RSD). Note that the RSD is called as same as the coefficient of variation (CV) in statistics. Within day precision (repeatability) was calculated by the mean coefficient of variation (CV) which was required to be less than 15% in accordance with the Guidance for Industry – Bioanalytical Method Validation prepared by the Biopharmaceutics Coordinating Committee in the Center for Drug Evaluation and Research (CDER) in cooperation with the Center for Veterinary Medicine (CVM) at the Food and Drug Administration.<sup>41</sup> Between days precision (intermediate precision) was expressed as between days coefficient of variation (CV<sub>bd</sub>), which was calculated using Eq. (3).<sup>42</sup>

$$CV_{bd} = \frac{SD_{bd} * 100}{\mu} \quad (3)$$

Being:

$\mu$  = average media.

$SD_{bd}$  = between days standard deviation (calculated as the square root of between days variance).

Between days variance is obtained after subtracting the contribution of within day variability, using the following Eq. (4)

$$SD_{bd}^2 = SD_{(\mu)}^2 + \frac{(n-1)}{n} * SD_{wd}^2 \quad (4)$$

Being:

$SD^2(\mu)$  = variance of every day mean.

$n$  = number of observations per day.

$SD_{wd}^2$  = average within day variance.

$$SD_{wd}^2 = \frac{\sum_{d=1}^D \sum_{r=1}^n (x_{dr} - \bar{x}_d)^2}{D * (n-1)} \quad (5)$$

where:

$D$  = total number of days.

$n$  = total number of replicates per day.

$x_{dr}$  = result for replicate  $r$  on day  $d$ .

$\bar{x}_d$  = average of all replicates on day  $d$ .

$$SD_{\mu}^2 = \frac{\sum_{d=1}^D (\bar{x}_d - \bar{\bar{x}})^2}{(D-1)} \quad (6)$$

where:

$D$  = total number of days.

$\bar{x}_d$  = average of all replicates on day  $d$ .

$\bar{\bar{x}}$  = average of all results.

### Scanning electron microscopy (SEM)

The pellets obtained after the reactivation process and after the adsorption process were fixed with a modifier Karnofsky's solution (1.6% glutaraldehyde + 2.6% paraformaldehyde

in 0.1 M sodium phosphate buffer (pH 7.2)). After 24 h at 4 °C, they were dehydrated by successive passages in solutions (ethyl alcohol-acetone) with greater graduation of ethyl alcohol each time (30%, 50%, 70%, 90% y 100%), the lapse for each passage was 10 min. After drying, the samples were glued on stubs using carbon tape and coated with gold (ion sputter JFC-1100). Bacteria were analyzed using a JEOL 35CF scanning electron microscope.

### Stability of the ZEA-selected *Lactocobacillus* complex

Using the strain with the highest ADS%, ZEA dissociation was study by successive rinses. In first place, the selected adsorption process was carried out, and then the pellet obtained was suspended in 1.0 mL of purified water and maintained at room temperature for 10 min. Then, the suspension obtained was centrifuged at 5900 ×  $g$ , 4 °C, 15 min, and finally ZEA concentration was analyzed in the supernatant. This step was repeated up to three times. At the end, ZEA remaining bound was recovered with 1.0 mL of methanol. ZEA quantification in all supernatants was performed by HPLC as indicated in ZEA determination. ZEA bound to the bacterial pellet was calculated according to Eq. (7).

$$\ln(ZEA\ bound_{\mu M}) = \ln\left(\frac{\text{Initial ZEA} * \%ADS}{100}\right) \quad (7)$$

Being:

%ADS = adsorption percentage.

Initial ZEA =  $\mu$ M of Initial ZEA in the positive control SP1 = 38.64  $\mu$ M.

### Biotransformation of ZEA

The metabolization of ZEA into  $\alpha$ -ZOL and  $\beta$ -ZOL was also investigated because ZEA concentration could be reduce as a consequence of biotransformation as indicated by Boswald et al.<sup>43</sup> and not because of ZEA-bacteria complex formation. HPLC analysis of a mixed standard solution containing ZEA and both metabolites was used to identify the presence or absence of each compound.

### Statistical analysis

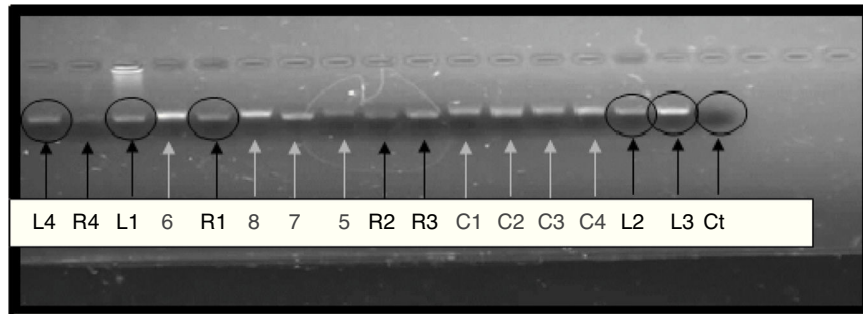
All samples were analyzed in triplicate. Results were expressed as mean  $\pm$  standard deviation (SD). Data were statistically evaluated using the InfoStat program, 2010. (InfoStat group FCA, Universidad Nacional de Córdoba, Argentina). A two-tailed Student's t-test for paired samples ( $p < 0.05$ ) was used to determine significant differences between treatments.

## Results and discussion

### Genotypic identification

Results from the PCR molecular technique are shown in Fig. 1. All strains assayed and the reference strains *L. rhamnosus* (R1), *L. delbrueckii* (R2), *L. casei* (R3) and *L. helveticus* (R4) were positive (amplified) with general primers for *Lactocobacillus*. In the negative control (Ct), as is expected, no amplification was observed.





**Fig. 1 – PCR amplification products for *Lactobacilli* isolated from pig rectal swaps (L1–L2–L3–L4), and isolated from other sources (5–6–7–8–C1–C2–C3–C4) and reference strains (R1–R2–R3–R4) using general primers: LbLMA1-rev y R16-1. Ct, control (without any bacterial strain).**

### Species identification

The comparative analysis of the sequences obtained from L4 strain with those described in various databases, showed a high identity with strains belonging to Phylum “Firmicutes” Class “Bacilli” Order “Lactobacillales” Family “Lactobacillaceae” Gender “Lactobacillus” and within this genus to species “plantarum” and “pentosus”. Given the phylogenetic proximity of these species, additional studies were performed to differentiate between them. L4 strain did not ferment glycerol and D-xylose, which are typically fermented by *L. pentosus* but not by *L. plantarum*.<sup>44,45</sup> These results indicate that the selected strain (L4) was a *L. plantarum*.

### Adsorption process

When time  $t_2$  (72h) was used in the reactivation process the reference strain R1 gave an adsorption value of  $79.8\% \pm 6.8$ . This result was similar to the results obtained by El-Nezami<sup>12</sup> when nonviable bacteria were used (80%). With this reactivation time it was probable that waste and dead cells began to accumulate and it caused some changes in the adsorption process. In the next assay reactivation time  $t_1$  (24 h) was used. This change gave an adsorption value of  $30.5\% \pm 5.4$ , it was close than values reported for viable cells.<sup>13</sup>

For R1 strain, the sum of the ZEA percentages, in the supernatant and in the methanol used to extract mycotoxin in the pellet was 97.73%. This value was close to 100% mycotoxin which indicated that the method achieved a high mycotoxin extraction.

### Strain selection

The strains L1, L2, L3 and L4 showed adsorption percentages between 46% and 64% (Table 1), these were higher than those showed by Bueno<sup>11</sup> and similar to those described by El-Nezami.<sup>13</sup> The second parameter analyzed was  $OD_{560\text{nm}}$ , as Bueno<sup>11</sup> indicated in order to achieve a good bacteria production, the value should be 1 or higher. Only in one case (L1 strain) the value was lower. According to our results, L4 strain achieved the highest average (X) between  $OD_{560\text{nm}}$  and ADS%, so it was decided to use this strain in subsequent research. Several genera of bacteria have been studied to evaluate their

**Table 1 – Results of the parameters used for strain selection [ $OD_{560\text{nm}}$ , % ADS and average (X) of both values]. Reactivation time used 24 h.**

Strain	Adsorption parameters		
	$OD_{560\text{nm}}$	% ADS	(X)average
L4	$1.062 \pm 0.085^a$	$64.2 \pm 1.5^c$	$32.618^c$
L3	$1.361 \pm 0.118^b$	$47.6 \pm 1.5^b$	$24.501^b$
L2	$1.426 \pm 0.143^b$	$49.1 \pm 0.0^b$	$25.278^b$
L1	$0.905 \pm 0.014^a$	$46.5 \pm 5.8^b$	$23.698^b$
<i>L. rhamnosus</i> (R1)	$1.129 \pm 0.019^a$	$30.5 \pm 5.4^a$	$15.825^a$

Results were represented as mean  $\pm$  SD. Different superscript letters in the same column indicate statistical significant differences ( $p < 0.05$ ).

zearalenone adsorption ability. Bueno<sup>11</sup> reported adsorption levels ranging from 0 to 36.44% for Bifidobacteria, 1.74 to 33.69% for Lactobacilli, and 0 to 59.09% for Actinomycetes. El-Nezami<sup>13</sup> reported the ZEA adsorption percentages of two food grade strains (*L. rhamnosus* GG and *L. rhamnosus* LC705). Furthermore, Lu et al<sup>19</sup> studied ZEA adsorption by *Planococcus spp.*, achieving 21.82% adsorption for viable bacteria and 42.82% for inactivated bacteria.

Results were represented as mean  $\pm$  SD. Different superscript letters in the same column indicate statistical significant differences ( $p < 0.05$ ).

### Evaluation of the adsorption process conditions

For the selected *Lactobacillus* strain L4 the change of various parameters in order to improve the adsorption process was evaluated.

Culture media comparison, L4 showed significantly higher  $OD_{560\text{nm}}$  in MRS ( $1.707 \pm 0.16$ ) than in LAPTg ( $1.101 \pm 0.15$ ) these differences were statistically significant ( $p < 0.05$ ). With respect to adsorption percentages, using the same  $OD_{560\text{nm}}$  (1) higher results were also obtained using MRS media ( $57.38\% \pm 4.68$ ) than LAPTg ( $49.33\% \pm 4.10$ ), but these difference were not statistically significant ( $p > 0.05$ ). Taking into account a future industrial use and observing that MRS broth achieved the higher production it was selected as culture media.

In addition, scanning electron microscopy revealed few differences comparing bacterial morphology during incubation in both broths (Fig. 2A and B), bacillus seemed longer and thinner when cultured in LAPTg than in MRS (marked in

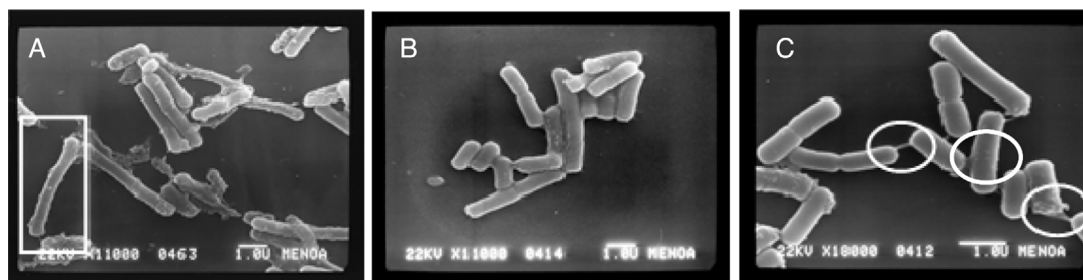


Fig. 2 – Scanning electron micrographs of L4 strain in (A) LAPTg media, (B) MRS media, (C) after the adsorption process.

Fig. 2A with a square), that should be studied in future investigations. These changes would indicate different morphology of the cellular wall and could explain different adsorption percentage achieved when this different culture media were used in the reactivation process prior ZEA adsorption. Furthermore, after the adsorption process, L4 presents several intercellular bonds (marked in Fig. 2C with circles), this being an interesting strain property that should be studied in future investigations (Fig. 2C).

A higher %ADS was obtained after incubation with SP1 than with SP2 solution, ( $57.28\% \pm 1.85$  and  $49.33\% \pm 6.11$ , respectively). These differences were not statistically significant ( $p > 0.05$ ), which indicates that a change of contact solution does not produce a significant change in the adsorption. For this reason, the technique was not changed and SP1 was maintained as contact solution.

Adsorption percentages obtained after processing 6.5 and 7.5 mL MRS broth were  $57.40\% \pm 3.53$  and  $64.46\% \pm 0.76$ , respectively. These differences were statistically significant ( $p < 0.05$ ). Therefore the use of 7.5 mL of media was selected to carry out the adsorption process.

After sterilizing culture media, the ADS% was  $82.4\% \pm 3.3$  and  $73.5\% \pm 1.0$  for viable strain ADS%. These differences were statistically significant ( $p < 0.05$ ). These findings could be considered in order to decide if non-viable bacteria may be used as mycotoxins adsorbent with higher ADS% or viable bacteria with lower ADS% but with possible probiotic effect at the same time. These findings confirm the results of other researches.<sup>13</sup>

Table 2 shows the ADS% obtained by processing three daily samples for four different days, under identical conditions. Relative standard deviations (%RSD) were calculated for each day, these values 1.81 to 7.23 were similar with that obtained by Kolk-Clauw et al.,<sup>39</sup> using the same technology for determination of ZEA (HPLC UV), and they never exceed the value of 15% that was our goal proposal. Repeatability ( $S_r$ ), also known as within-run precision was calculated, under identical conditions successive measurements obtained were close in a value of 6.00. Using the overall mean obtained (70.10) and the relative standard deviation maximum allowed (15%), the maximum standard deviation claimed was 10.52. This value was less than SD achieved in each day and less than the repeatability obtained.

Results were represented as mean  $\pm$  SD. Different superscript letters in the same row indicate significant differences ( $p < 0.05$ ).

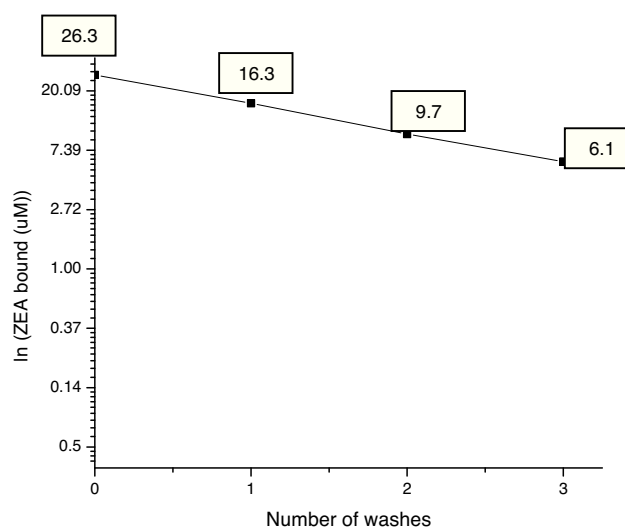


Fig. 3 – ZEA remaining from ZEA-L4 complex after successive rinsing.

Within day precision (repeatability) and between day precision were calculated and as required both values were less than 15%.

#### Stability of the ZEA-bacteria complex

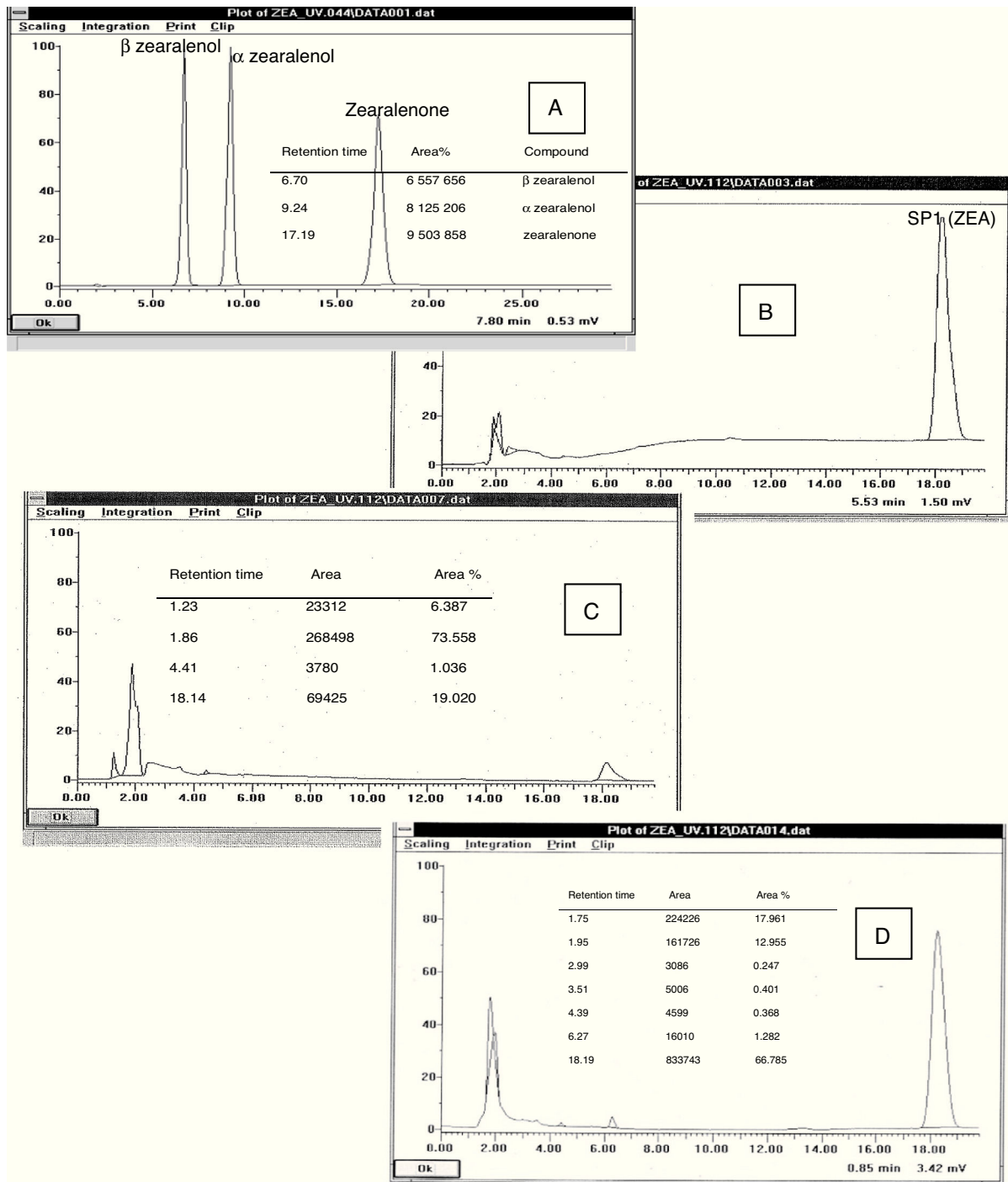
Using the improved process with an initial ZEA concentration of  $38.7 \mu\text{M}$ , the initial adsorption percentage was  $68.20\% \pm 1.74$  (ZEA bound:  $26.3 \mu\text{M}$ ). In the first rinsing step  $42.26\% \pm 0.414$  was still bound to the pellet (ZEA bound:  $16.3 \mu\text{M}$ ). In the second rinsing step,  $25.12\% \pm 0.664$  was still bound to the pellet (ZEA bound:  $9.7 \mu\text{M}$ ), whereas  $15.82\% \pm 0.675$  remained in the pellet (ZEA bound  $6.1 \mu\text{M}$ ) after the third rinse.

Fig. 3 shows  $\ln(\text{ZEA bound})$  vs number of washes. Dissociation of ZEA was relatively linear. The dissociation constant, which could be estimated from the slope, was higher than that obtained by El-Nezami et al.<sup>12</sup> These results could probably be due to the use of different bacteria strains than those used by El-Nezami et al.,<sup>12</sup> specifically in terms of their differences in the cell wall. Some compounds of the cell wall are really important in the adsorption process as indicated by Yiannikouris et al.<sup>46,47</sup>

**Table 2 – Adsorption percentage (%ADS) of L4 after the adsorption process for three samples in four different days. Estimation of repeatability (within day precision) and intermediate precision (between day precision).**

	Day 1	Day 2	Day 3	Day 4	Mean	SD <sup>2</sup> ( $\mu$ )	SD <sup>2</sup> <sub>wd</sub>	SD <sup>2</sup> <sub>bd</sub>
% ADS	64.46	65.30	75.61	75.01	70.095 ( $\mu$ )	31.409	36.042	55.437
SD	1.17 <sup>a</sup>	1.35 <sup>a</sup>	5.47 <sup>a</sup>	1.73 <sup>a</sup>				
%RSD (CV)	1.81	2.06	7.23	2.31				

Results were represented as mean  $\pm$  SD. Different superscript letters in the same row indicate significant differences ( $p < 0.05$ ). Within day precision (repeatability) and between day precision were calculated and as required both values were less than 15%. Within-day precision: 3.36%. Between-day precision: 10.62%.



**Fig. 4 – Chromatographic profiles. (A) Chromatographic profile of standards:  $\alpha$  zearalenol,  $\beta$  zearalenol and zearalenona. (B) Chromatographic profile of ZEA solution (SP1). (C) Chromatographic profile of L4 supernatant after co-incubating with ZEA (1 h, 37 °C). (D) Chromatographic profile of ZEA from L4 pellet after solubilization with methanol.**

### Zearalenone determination by HPLC chromatography

Results from a mixed standard solution of ZEA,  $\alpha$  and  $\beta$  zearalenol are showed in Fig. 4 A. Results from SP1 solution without contact with adsorbent are showed in Fig. 4 B, peaks corresponding to  $\alpha$  and  $\beta$  zearalenol were not observed. Results from the supernatant of L4 strain after incubation (37 °C, 1 h) with the mycotoxin are showed in Fig. 4C, peaks corresponding to  $\alpha$  and  $\beta$  zearalenol were not observed. Results from L4 pellet after resuspension with methanol are showed in Fig. 4D. Peak corresponding to  $\beta$  zearalenol with a small Area % was observed, peak corresponding to  $\alpha$  zearalenol was not observed, it was important to demonstrated that probably this *Lactobacillus* did not metabolize the mycotoxin as indicated by El-Nezami et al.<sup>13</sup>. The sum of ZEA in supernatant and in pellet was almost the initial ZEA added in SP1 solution, it showed that the initial mycotoxin in (SP1) solution was almost completely recovered.

### Conclusions

This work demonstrated that a *Lactobacillus plantarum* (L4 strain) isolated from pigs has great potential as adsorbent of the mycotoxin zearalenone. The results showed the best conditions for the adsorption process. The initial culture medium used for bacterial reactivation, the viability of the cells and the amount of bacterial mass measured by OD<sub>560nm</sub> were strongly involved in the percentage of adsorption of *L. plantarum* to ZEA. The studies showed high stability of the complex bacteria-mycotoxin and the absence of metabolites of the toxin. This study is the first step in the development of a new ZEA adsorbent based on a lyophilized powder of a locally isolated strain and other components that help to improve reproductive health.

### Conflicts of interest

The authors declare no conflicts of interest.

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