

Concise Review

Sorafenib, a systemic therapy for hepatocellular carcinoma

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Abstract

Hepatocellular carcinoma is a lethal disease that requires a multidisciplinary approach and management. Surgical therapy offers long-term survival; however, few patients are candidates. There has been no accepted systemic therapy for this disease until recently. This article briefly discusses the role of RAS/RAF/MEK/ERK signaling pathway in the pathogenesis of the disease and the promising role of sorafenib for advanced disease.

Key words: Liver cancer, hepatocellular carcinoma, HCV, HBV, sorafenib.

Hepatocellular carcinoma (HCC) is the most common primary liver cancer, with an estimated incidence of half a million new cases per year around the world. Its asymptomatic development, malignant progression, and the poor efficacy of current treatments entail a poor prognosis, with fewer than 5% of patients surviving five years after diagnosis. HCC is the third greatest cause of cancerrelated death in the world, and most of these deaths are registered in developing countries.²

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List abbreviations:

HCC, hepatocellular carcinoma; ERK, extracellular signal-regulated kinase; MEK, extracellular signal-regulated kinase; HBV, hepatitis B virus; HCV, hepatitis C virus; TKRs, tyrosine kinase receptors; GEFs, guanine nucleotide exchange factors; SOS protein, mammalian homologue of the Drosophila son of sevenless gene product; VEGF, vascular endothelial growth factor; PDGF, platelet-derived growth factor; EGF, epidermal growth factor; VEGFRs, VEGF receptors; HSP, heat shock protein.

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Intense research over the past 20 years has provided detailed information about the molecular mechanisms and signaling pathways involved in hepatocarcinogenesis. The RAS/RAF/MEK/ERK signaling pathway has an essential role in the regulation of normal hepatocyte proliferation. Defects in this signaling pathway are critical in HCC pathogenesis, making it an attractive target for chemotherapeutic agents. Sorafenib, an oral drug developed as a RAF inhibitor, is a promising agent for HCC therapy. Sorafenib is a multikinase inhibitor targeting the RAF/MEK/ERK pathway, with antiangiogenic effects.^{4,5} This review summarizes current knowledge of the RAS/RAF/MEK/ ERK signaling pathway and its implications in HCC pathogenesis, and focuses on the role of sorafenib in the therapy of HCC.

Epidemiology and risk factors

Liver-cancer-related death is a major health problem around the world. Despite being the sixth most common malignancy, HCC is highly lethal, representing the third greatest cause of death from malignancy worldwide, particularly in association with hepatitis B virus (HBV) infection in developing countries.³

The age-adjusted incidence of HCC has marked geographic variations, with the highest rates being observed in Asia and sub-Saharan Africa (25 cases per 100,000), and the lowest in North America and northern Europe.^{4,7} Unfortunately, several studies have shown a worldwide rise in HCC incidence in the last two decades, forecasting a devastating effect if health care policies are not intensified. Capocaccia et al.8 analyzed the database of the Surveillance Epidemiology and End Results program of the EUROCARE project and observed a fourfold increase in the incidence of HCC in southern Europe. In the United States, several studies have reported that the age-adjusted incidence of HCC has doubled over the last two decades,6-8 affecting Caucasian and Hispanic men particularly.9 At least 50% of the new cases in United States could be attributable to chronic Hepatitis C Virus (HCV) infection. 10 However, in almost 15%-50% of patients with HCC, there is no evidence of either viral hepatitis or heavy alcohol consumption, 11

Table I. Annual incidence of hepatocellular carcinoma related to the etiology of liver cirrhosis.

Author, year	No. Patients	Etiology of liver cirrhosis	Mean Follow-up*	HCC Annual Incidence**
Fattovich et al., 1997	361	HCV	60	1.4%
Chiaramonte et al., 1999	166	HCV	64.5	3.8%
Fattovich et al., 2002	136	HCV	66	2.5%
Solá et al., 2006	200	HCV	39	5.5%
	177	Alcoholic	39	1.7%
Fattovich et al., 2002	161	HBV	66	2.2%
Chiaramonte et al., 1999	66	HBV	64.5	1.7%
	27	HBV/HCV	64.5	7.6%
Ratziu et al., 2002	22	Cryptogenic, obesity-related	18	0.8%

^{*} Months

Abbreviations: HCC, hepatocellular carcinoma; HCV, hepatitis C virus; HBV, hepatitis B virus.

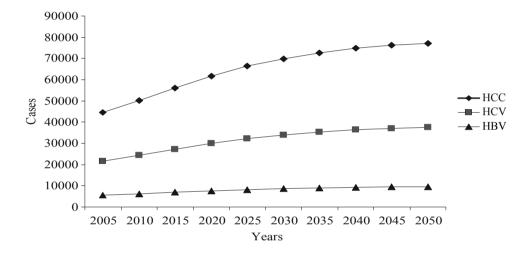


Figure 1. Trends in the prevalence of chronic liver disease in Mexico.^{19,20}

suggesting that such cases could be linked to nonalcoholic fatty liver disease and other etiologies of chronic liver disease.

A preexistent cirrhotic liver is a clinicopathological condition observed in 80%–90% of patients who develop HCC. ¹² Several studies have indicated that 1%–4% of all cirrhotic patients per year will developed HCC, ¹³ with differences according to the leading cause (table 1). ¹⁴⁻¹⁸ Liver cirrhosis has a critical impact on public health in Mexico, representing the third greatest cause of death in the general population, and predicted trends for the next five decades are not promising (figure 1). ^{19,20} The Mexican Association of Hepatology determined alcohol and HCV infection to be the main causes of liver cirrhosis in Mexico (39.5% vs 36.6%, respectively; p = 0.113), followed by cryptogenic cirrhosis. ²¹

Hepatocarcinogenesis

Hepatocarcinogenesis is a multistep process in which genetic abnormalities and epigenetic alterations accumulate, causing aberrant growth and malignant transformation of hepatocytes. Accumulation of such abnormalities leads to activation of mediators of cellular proliferation (proto-oncogenes and their mitogenic signaling pathways) resulting in neoplastic potential.

Hepatocellular carcinomas exhibit a high degree of genetic heterogeneity, and multiple molecular pathways may be involved in the pathogenesis of HCC. The best-characterized pathways are the heat shock protein (HSP)/ stress response signaling pathway, the Wnt pathway, and the MAPK pathway, and the associated involvement of growth factors and cytokines.²²⁻²⁴ This review focuses on the MAPK pathway (the RAF/RAS/MEK/ERK signaling pathway) and its major role in hepatocarcinogenesis.

The RAS/RAF/MAPK-ERK signaling pathway

The RAS/RAF/MAPK-ERK signaling pathway is an important mediator of tumor cell proliferation, differentiation and apoptosis. Studies have reported that MAPK expression is significantly higher in HCC compared to adjacent normal liver cells, 25,26 showing the critical role of this pathway in the pathogenesis of HCC (figure 2).

^{**} Data were recalculated from the original paper

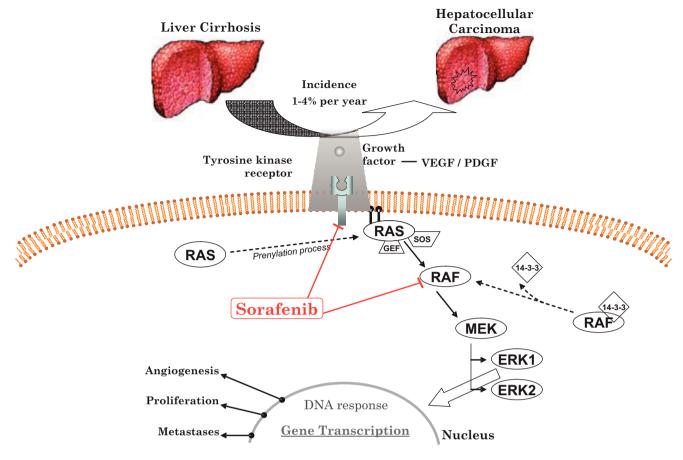


Figure 2. Role of the RAS/RAF/MEF/ERK pathway in hepatocarcinogenesis.

RAS is a cytosolic protein that, after a prenylation process, is located on the inner surface of the cellular membrane. This posttranslational processing anchors *RAS* protein to the cytoplasmic membrane, which is necessary for its biological activity. Misplaced *RAS* proteins are inactive, probably because they cannot recruit their target enzymes. Interestingly, a recent study suggested that prenylation is not necessary for endogenous *RAS* activation in normal cells.^{27,28}

Activation of RAS

Activation of the normal RAS signaling pathway is initiated by the interaction of several cytokines, hormones and extracellular growth factors with their tyrosine-kinases receptors (TKRs). As a result, ligand binding induces receptor dimerization and autophosphorylation, activating downstream intracellular signal cascades. First, there is recruitment of guanine nucleotide exchange factors (GEFs), such as RAS-GRF and SOS protein (mammalian homologue of the Drosophila son of sevenless gene product), to the inner surface of the cell membrane where RAS protein is also located after prenylation. RAS is a membrane-bound G protein. The biological activity of RAS is regulated through the

GDP/GTP cycle.²⁹ In the inactive state, *RAS* exists in the GDP-bound form. Because of TKR activation, GEF is recruited and located in the cell membrane, promoting the formation of the GTP-bound active state. In contrast, *RAS* becomes inactivated through hydrolysis of GTP by an intrinsic GTPase. Nevertheless, *in vitro* studies have demonstrated a low-activity level of this intrinsic GTPase; thus, effective hydrolysis of GTP is performed by several cytoplasmic GTPase-activating proteins, which rapidly induce the hydrolysis of GTP-bound RAS to the inactive GDP form.

Unregulated *RAS* pathway activity is observed in tumor cells, because of point mutations in the *RAS* gene family and the overexpression of TKRs and their ligands. The RAS genes encode four highly similar 21 kDa proteins, H-*RAS*, N-*RAS*, K-*RAS*4A and K-*RAS*4B. Point mutations in RAS genes are observed in approximately 20%–30% of all solid tumors, ²⁹ K-*RAS* being the most commonly affected. For example, K-*RAS* is mutated in up to 80% of pancreatic adenocarcinomas. ³⁰ N-*RAS* mutations are observed in 30% of cases of HCC; ³¹ in contrast, H-*RAS* mutations are rarely observed. These genetic derangements compromise the intrinsic GTP*ase* activity and the GAP-induced GTP*ase* activation of *RAS*, ³² with the loss of its ability to return to a quiescent state, lead-

ing to constitutive activation of RAS and subsequent stimulation of downstream effectors.

Overactivation of RTKs results from activating mutations or overexpression of growth factor ligands. Vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF) and epidermal growth factor (EGF) have important roles in dysregulated cell growth and metastases. VEGF and its receptors (VEGFRs), particularly VEGFR-2, are key molecules involved in endothelial cell proliferation, angiogenesis and vascular permeability. Elevated serum VEGF levels are associated with poor prognosis in patients with HCC. Turbermore, VEGF gene polymorphisms have been suggested as prognostic indicators for HCC. Experimental studies in mice have reported that approximately 70% of HCCs show high serum PDGFR-a levels. Second

Receptor overexpression and elevated ligand availability play an important role in the development of metastases in patients with HCC. As VEGFR and PDG-FR use the *RAS/RAF/MEK/ERK* pathway, its targeting as an anticancer therapy has been explored with great interest.

RAF is the best downstream effector of RAS

RAF, the best-characterized downstream effector of *RAS*, is a serine/threonine protein kinase positioned as the first signaling element of the MAPK pathway.^{39,40} The *RAF* gene family encodes three closely related cytosolic proteins: A*RAF*, B*RAF* and C*RAF* (also termed RAF-1). Whereas each isoform has a distinct expression profile in tissues, the three *RAF* species are found in normal liver cells and recent evidence has shown that they are over-expressed or mutated in HCC.^{41,42} These serine/threonine protein kinases share three conserved sequence regions, termed CR1, CR2 and CR3, that participates in the complex processes of *RAF* kinase activity regulation.

To activate *RAF* kinase, GTP-bound *RAS* directly interacts with *RAF*, promoting its recruitment to the cell membrane, an essential step for its activation. ^{43,44} The *RAS* effector domain binds to *RAF* via CR1. Once located in the membrane, *RAF* requires several modifications to become active. ⁴⁵ Nevertheless, *in vitro* studies have revealed that such an interaction is insufficient to stimulate *RAF* kinase activity, suggesting that others cofactors are necessary for *RAF* activation *in vivo*. ^{46,47}

Cytosolic *CRAF* exists as a complex formed with the dimeric cofactor 14-3-3, a highly conserved chaperonin protein that binds at phosphorylated serine residues S259 and S621, inactivating the protein. *CRAF* interaction with *RAS* displaces the cofactor 14-3-3, exposing the serine residues to desphosphorylation by protein phosphatase 2A or other phosphatases. In addition, the activation of *CRAF* also requires phosphorylation of other serine and tyrosine residues, particularly S338 and Y341.⁴⁸ In *BRAF*, the final phosphorylation of S338 and Y341 is omitted because S445, a homologous site on *BRAF*, is constitutively phos-

phorylated; thus, BRAF is immediately activated after interaction with GTP-bound RAS. This property confers to BRAF a higher kinase activity level, making it the strongest activator of downstream MEK pathway.

Once activated, *RAF* phosphorylates *MAPKKs* (*MEK*1 and *MEK*2) at residues S218 and S222. All *RAF* isoforms are able to activate *MEK*1; however, only B*RAF* and C*RAF* activate *MEK*2. ⁴² In turn, downstream effectors of *MAPKKs*, *ERK*1 and *ERK*2, are activated by phosphorylation at residues T183 and Y185, with further activation of the nuclear transcription factors Elk-1, *fos*, *jun*, AP-1, *myc* and nuclear factor-kB (NF-kB), which regulate gene expression associated with cell proliferation, differentiation, angiogenesis or apoptosis. ⁴⁹

Constitutive activation of *RAF* and *RAS* are indistinguishable in their potential to induce malignant transformation. *RAF* protein is mutated in approximately 7% of all malignancies because of point mutations, deletions, amplification and rearrangements of RAF. Tannapfel *et al.* recently reported that *BRAF* mutations are rare in HCC.⁵⁰ Hwang *et al.* have shown that the *CRAF* gene is upregulated in 40% of cirrhotic livers and 50% of HCCs; as a consequence, *CRAF* protein is overexpressed in 91.2% and 100% of these tumors, respectively.⁵¹

Epidemiological data have provided strong evidence about the role of chronic HCV infection and cirrhosis in the development of HCC.⁵² In addition, experimental models have shown the development of HCC in transgenic mice expressing the HCV core gene.⁵³ However, the precise mechanisms involved are unclear. An *in vitro* study showed that HCV core protein activates the MEK/ERKs signaling pathway in mammalian epithelial cells, with constitutive RAF-1 activity. HCV core protein has also been shown to bind the 14-3-3 protein both *in vivo* and *in vitro*, suggesting that such an interaction with this chaperonin exposes the serine residues S259 and S621 to desphosphorylation, a key step in RAF activation.⁵⁴

Sorafenib: a promising therapy

Sorafenib, a bi-aryl urea, was initially recognized as a CRAF inhibitor.⁴ Further studies in different cell lines and xenograft models have demonstrated that sorafenib is a potent multikinase inhibitor, including wild-type and mutant BRAF, VEGFR2, VEGFR3, PDGFR-• • FLT3, Ret and c-Kit, and has antiangiogenic effects.⁵⁵⁻⁵⁷

The direct effects of sorafenib have been evaluated *in vitro* in two distinct HCC cell lines. Sorafenib inhibited cell proliferation and induced cell apoptosis in both cell lines in a dose-dependent manner.⁵⁷ In addition, sorafenib inhibited MEK and ERK phosphorylation. In the same study, the *in vivo* effects of sorafenib were evaluated in a xenograft model, in which sorafenib produced significant and dose-dependent tumor growth inhibition of 49% and 78%, respectively. Sorafenib produced durable

partial tumor regression in 50% of the mice, indicating direct effects on tumor cell proliferation/survival *in vivo*.⁵⁷

Two open-label, uncontrolled, phase I trials evaluated sorafenib in 86 patients with solid tumors refractory to standard treatment, including one patient with HCC.58,59 Overall, sorafenib was safe and well tolerated at doses of 400 mg b.i.d. Even when the majority of patients experienced at least one adverse event, toxicities were mostly mild to moderate. Two phase I clinical trials demonstrated efficacy in the treatment of patients with advanced HCC using combination regimens with other anticancer agents such as doxorubicin. 60,61 Patients received continuous oral sorafenib 400 mg b.i.d. in four-week cycles (median number of treatment cycles was four, range 1-19). Three patients had partial responses (duration ranged from 12 to 14.5 months), eight had minor responses, 46 had stable disease (• •16 weeks) and 48 had progressive disease (imaging assessment), with a median overall survival of 9.2 months. In addition, relatively infrequent dose-limiting toxicities were observed in this study, including fatigue (9.5%), diarrhea (8%) and hand and foot skin reactions (5.1%), with grade 3 toxicities the most common.61

Results from the international double-blind placebocontrolled SHARP trial were presented during the Annual Meeting of the American Society of Clinical Oncology.62 After stratification for portal vein and/or extrahepatic invasion and ECOG status, the researchers randomized 602 patients with Child-Pugh A cirrhosis and HCC to receive either placebo (n = 303) or sorafenib 400 mg b.i.d. (n = 299) from March 2005 to April 2006. Intention-totreat analysis revealed that sorafenib-treated patients lived 46.3 weeks as compared to 34.4 weeks in the placebo group (p = 0.00058). In addition, time-to-progression was significantly longer in the sorafenib-treated group (24 vs 12.3 weeks; p = .000007). No complete response was observed during the study period and few partial responses were observed (7/299 in the sorafenib-treated group vs 2/303 in the placebo-treated group). The study was stopped in February 2007 because of results favoring sorafenib were found in the second planned interim analysis of October 2006.62 Based on these final data, sorafenib seems to have a role as a disease stabilizer rather than as a cure for HCC.

Conclusions

Molecule-targeted therapies for cancer are promising, particularly those directed to malignancies with a currently poor prognosis, such as HCC. The available literature shows for the first time that systemic therapy with sorafenib prolongs survival in HCC patients. We anticipate great interest in the publication of the full report of the SHARP trial and its impact on the scientific community worldwide.

References

- El-Serag HB. Epidemiology of hepatocellular carcinoma. Clin Liver Dis 2001; 5: 87-107, vi.
- Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. CA Cancer J Clin 2005; 55: 74-108.
- Huynh H, Nguyen TT, Chow KH, Tan PH, Soo KC, Tran E. Overexpression of the mitogen-activated protein kinase (MAPK) kinase (MEK)-MAPK in hepatocellular carcinoma: its role in tumor progression and apoptosis. *BMC Gastroenterol* 2003; 3: 19.
- Wilhelm S, Chien DS. BAY 43-9006: preclinical data. Curr Pharm Des 2002; 8: 2255-7.
- Wilhelm SM, Carter C, Tang L, Wilkie D, McNabola A, Rong H, Chen C, et al. BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. *Cancer Res* 2004; 64: 7099-109.
- Bosch FX, Ribes J, Diaz M, Cleries R. Primary liver cancer: worldwide incidence and trends. Gastroenterology 2004; 127: S5-S16.
- El-Serag HB, Mason AC, Key C. Trends in survival of patients with hepatocellular carcinoma between 1977 and 1996 in the United States. *Hepatology* 2001; 33: 62-5.
- Capocaccia R, Sant M, Berrino F, Simonetti A, Santi V, Trevisani F. Hepatocellular carcinoma: trends of incidence and survival in Europe and the United States at the end of the 20th century. Am J Gastroenterol 2007; 102: 1661-70; quiz 0, 71.
- El-Serag HB. Hepatocellular carcinoma: recent trends in the United States. Gastroenterology 2004; 127: S27-34.
- El-Serag HB. Epidemiology of hepatocellular carcinoma in USA. Hepatol Res 2007; 37 Suppl 2: S88-94.
- El-Serag HB, Lau M, Eschbach K, Davila J, Goodwin J. Epidemiology of hepatocellular carcinoma in Hispanics in the United States. Arch Intern Med 2007; 167: 1983-9.
- Fattovich G, Stroffolini T, Zagni I, Donato F. Hepatocellular carcinoma in cirrhosis: incidence and risk factors. *Gastroenterol-*0gy 2004: 127: S35-50.
- Bailey MA, Brunt EM. Hepatocellular carcinoma: predisposing conditions and precursor lesions. *Gastroenterol Clin North Am* 2002: 31: 641-62.
- Fattovich G, Giustina G, Degos F, Tremolada F, Diodati G, Almasio P, Nevens F, et al. Morbidity and mortality in compensated cirrhosis type C: a retrospective follow-up study of 384 patients. *Gastroenterology* 1997; 112: 463-72.
- Chiaramonte M, Stroffolini T, Vian A, Stazi MA, Floreani A, Lorenzoni U, Lobello S, et al. Rate of incidence of hepatocellular carcinoma in patients with compensated viral cirrhosis. *Cancer* 1999; 85: 2132-7.
- Fattovich G, Pantalena M, Zagni I, Realdi G, Schalm SW, Christensen E. Effect of hepatitis B and C virus infections on the natural history of compensated cirrhosis: a cohort study of 297 patients. Am J Gastroenterol 2002; 97: 2886-95.
- Ratziu V, Bonyhay L, Di Martino V, Charlotte F, Cavallaro L, Sayegh-Tainturier MH, Giral P, et al. Survival, liver failure, and hepatocellular carcinoma in obesity-related cryptogenic cirrhosis. *Hepatology* 2002; 35: 1485-93.
- Sola R, Alvarez MA, Balleste B, Montoliu S, Rivera M, Miquel M, Cirera I, et al. Probability of liver cancer and survival in HCVrelated or alcoholic-decompensated cirrhosis. A study of 377 patients. *Liver Int* 2006; 26: 62-72.
- National System of Health Information. [Bases de datos en formato de cubo dinámico]. http://sinais.salud.gob.mx/.
- Mendez-Sanchez N, Villa AR, Chavez-Tapia NC, Ponciano-Rodriguez G, Almeda-Valdes P, Gonzalez D, Uribe M. Trends in liver disease prevalence in Mexico from 2005 to 2050 through mortality data. *Ann Hepatol* 2005; 4: 52-5.
- Mendez-Sanchez N, Aguilar-Ramirez JR, Reyes A, Dehesa M, Juorez A, Castneda B, Sanchez-Avila F, et al. Etiology of liver cirrhosis in Mexico. *Ann Hepatol* 2004; 3: 30-3.

- Blanc JF, Lalanne C, Plomion C, Schmitter JM, Bathany K, Gion JM, Bioulac-Sage P, et al. Proteomic analysis of differentially expressed proteins in hepatocellular carcinoma developed in patients with chronic viral hepatitis C. *Proteomics* 2005; 5: 3778-89.
- Lee HC, Kim M, Wands JR. Wnt/Frizzled signaling in hepatocellular carcinoma. Front Biosci 2006; 11: 1901-15.
- Yoon S, Seger R. The extracellular signal-regulated kinase: multiple substrates regulate diverse cellular functions. *Growth Factors* 2006; 24: 21-44.
- Schmidt CM, McKillop IH, Cahill PA, Sitzmann JV. Increased MAPK expression and activity in primary human hepatocellular carcinoma. *Biochem Biophys Res Commun* 1997; 236: 54-8.
- Downward J. Targeting RAS signalling pathways in cancer therapy. Nat Rev Cancer 2003; 3: 11-22.
- 27. Khwaja A, Dockrell ME, Hendry BM, Sharpe CC. Prenylation is not necessary for endogenous Ras activation in non-malignant cells. *J Cell Biochem* 2006; 97: 412-22.
- Khosravi-Far R, Campbell S, Rossman KL, Der CJ. Increasing complexity of Ras signal transduction: involvement of Rho family proteins. Adv Cancer Res 1998; 72: 57-107.
- Bos JL. ras oncogenes in human cancer: a review. Cancer Res 1989; 49: 4682-9.
- 30. Almoguera C, Shibata D, Forrester K, Martin J, Arnheim N, Perucho M. Most human carcinomas of the exocrine pancreas contain mutant c-K-ras genes. *Cell* 1988; 53: 549-54.
- 31. Gu JR, Hu LF, Cheng YC, Wan DF. Oncogenes in human primary hepatic cancer. *J Cell Physiol* Suppl 1986; 4: 13-20.
- Trahey M, McCormick F. A cytoplasmic protein stimulates normal N-ras p21 GTPase, but does not affect oncogenic mutants. Science 1987; 238: 542-5.
- 33. Zwick E, Bange J, Ullrich A. Receptor tyrosine kinases as targets for anticancer drugs. *Trends Mol Med* 2002; 8: 17-23.
- Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. *Nat Med* 2003; 9: 669-76.
- Poon RT, Ng IO, Lau C, Zhu LX, Yu WC, Lo CM, Fan ST, et al. Serum vascular endothelial growth factor predicts venous invasion in hepatocellular carcinoma: a prospective study. *Ann Surg* 2001; 233: 227-35.
- Poon RT, Ho JW, Tong CS, Lau C, Ng IO, Fan ST. Prognostic significance of serum vascular endothelial growth factor and endostatin in patients with hepatocellular carcinoma. *Br J Surg* 2004; 91: 1354-60.
- 37. Kong SY, Park JW, Lee JA, Park JE, Park KW, Hong EK, Kim CM. Association between vascular endothelial growth factor gene polymorphisms and survival in hepatocellular carcinoma patients. *Hepatology* 2007; 46: 446-55.
- Stock P, Monga D, Tan X, Micsenyi A, Loizos N, Monga SP. Platelet-derived growth factor receptor-alpha: a novel therapeutic target in human hepatocellular cancer. *Mol Cancer Ther* 2007; 6: 1932-41.
- 39. Marshall CJ. Ras effectors. Curr Opin Cell Biol 1996; 8: 197-204.
- Beeram M, Patnaik A, Rowinsky EK. Raf: a strategic target for therapeutic development against cancer. J Clin Oncol 2005; 23: 6771-90
- Leevers SJ, Paterson HF, Marshall CJ. Requirement for Ras in Raf activation is overcome by targeting Raf to the plasma membrane. *Nature* 1994; 369: 411-4.
- 42. Sridhar SS, Hedley D, Siu LL. Raf kinase as a target for anticancer therapeutics. *Mol Cancer Ther* 2005; 4: 677-85.
- 43. Hu CD, Kariya K, Tamada M, Akasaka K, Shirouzu M, Yokoyama S, Kataoka T. Cysteine-rich region of Raf-1 interacts with activator domain of post-translationally modified Ha-Ras. *J Biol Chem* 1995; 270: 30274-7.
- Chong H, Vikis HG, Guan KL. Mechanisms of regulating the Raf kinase family. Cell Signal 2003; 15: 463-9.

- 45. Traverse S, Cohen P, Paterson H, Marshall C, Rapp U, Grand RJ. Specific association of activated MAP kinase (Raf) with the plasma membranes of ras-transformed retinal cells. *Oncogene* 1993; 8: 3175-81.
- 46. Zhang XF, Settleman J, Kyriakis JM, Takeuchi-Suzuki E, Elledge SJ, Marshall MS, Bruder JT, et al. Normal and oncogenic p21ras proteins bind to the amino-terminal regulatory domain of c-Raf-1. *Nature* 1993; 364: 308-13.
- Kubicek M, Pacher M, Abraham D, Podar K, Eulitz M, Baccarini M. Dephosphorylation of Ser-259 regulates Raf-1 membrane association. *J Biol Chem* 2002; 277: 7913-9.
- Fabian JR, Daar IO, Morrison DK. Critical tyrosine residues regulate the enzymatic and biological activity of Raf-1 kinase. *Mol Cell Biol* 1993; 13: 7170-9.
- 49. Garnett MJ, Marais R. Guilty as charged: B-RAF is a human oncogene. *Cancer Cell* 2004; 6: 313-9.
- Tannapfel A, Sommerer F, Benicke M, Katalinic A, Uhlmann D, Witzigmann H, Hauss J, et al. Mutations of the BRAF gene in cholangiocarcinoma but not in hepatocellular carcinoma. *Gut* 2003; 52: 706-12.
- 51. Hwang YH, Choi JY, Kim S, Chung ES, Kim T, Koh SS, Lee B, et al. Over-expression of c-raf-1 proto-oncogene in liver cirrhosis and hepatocellular carcinoma. *Hepatol Res* 2004; 29: 113-21.
- Shimotohno K, Watashi K, Tsuchihara K, Fukuda K, Marusawa H, Hijikata M. Hepatitis C virus and its roles in cell proliferation. *J Gastroenterol* 2002; 37 Suppl 13: 50-4.
- Moriya K, Fujie H, Shintani Y, Yotsuyanagi H, Tsutsumi T, Ishibashi K, Matsuura Y, et al. The core protein of hepatitis C virus induces hepatocellular carcinoma in transgenic mice. *Nat Med* 1998; 4: 1065-7.
- 54. Giambartolomei S, Covone F, Levrero M, Balsano C. Sustained activation of the Raf/MEK/Erk pathway in response to EGF in stable cell lines expressing the Hepatitis C Virus (HCV) core protein. *Oncogene* 2001; 20: 2606-10.
- 55. Ahmad T, Eisen T. Kinase inhibition with BAY 43-9006 in renal cell carcinoma. *Clin Cancer Res* 2004; 10: 6388S-92S.
- 56. Clark JW, Eder JP, Ryan D, Lathia C, Lenz HJ. Safety and pharmacokinetics of the dual action Raf kinase and vascular endothelial growth factor receptor inhibitor, BAY 43-9006, in patients with advanced, refractory solid tumors. *Clin Cancer Res* 2005; 11: 5472-80.
- 57. Liu L, Cao Y, Chen C, Zhang X, McNabola A, Wilkie D, Wilhelm S, et al. Sorafenib blocks the RAF/MEK/ERK pathway, inhibits tumor angiogenesis, and induces tumor cell apoptosis in hepatocellular carcinoma model PLC/PRF/5. Cancer Res 2006; 66: 11851-8.
- 58. Awada A, Hendlisz A, Gil T, Bartholomeus S, Mano M, de Valeriola D, Strumberg D, et al. Phase I safety and pharmacokinetics of BAY 43-9006 administered for 21 days on/7 days off in patients with advanced, refractory solid tumours. *Br J Cancer* 2005; 92: 1855-61.
- 59. Moore M, Hirte HW, Siu L, Oza A, Hotte SJ, Petrenciuc O, Cihon F, et al. Phase I study to determine the safety and pharmacokinetics of the novel Raf kinase and VEGFR inhibitor BAY 43-9006, administered for 28 days on/7 days off in patients with advanced, refractory solid tumors. *Ann Oncol* 2005; 16: 1688-94.
- 60. Richly H, Kupsch P, Passage K, Grubert M, Hilger RA, Voigtmann R, Schwartz B, et al. Results of a phase I trial of BAY 43-9006 in combination with doxorubicin in patients with primary hepatic cancer. *Int J Clin Pharmacol Ther* 2004; 42: 650-1.
- Abou-Alfa GK, Schwartz L, Ricci S, Amadori D, Santoro A, Figer A, De Greve J, et al. Phase II study of sorafenib in patients with advanced hepatocellular carcinoma. *J Clin Oncol* 2006; 24: 4293-300.
- 62. ASCO 2007: plenary top 5. Oncology 2007; 21: 896.