

Concise Review

Genetic predisposition of cholesterol gallstone disease

Alejandro Hernández-Nazará;¹ Fátima Curiel-López;¹ Erika Martínez-López;¹ Zamira Hernández-Nazará;¹ Arturo Panduro¹

Abstract

Gallstone disease (GSD) is the result of the interaction between genetic and environmental factors and it is a major disease cause of surgery with high costs to health systems. Worldwide prevalence varies according to the ethnic population suggesting that high prevalence of GSD in certain ethnic groups is due to the presence of genetic factors implicated in different metabolic pathways. However, environmental factors play a determinant role in gene expression. This review summarizes the genes involved in biliary salt and cholesterol synthesis, lipids transport and the *Lith* genes. Future studies should be focused on the study of interactions between genetic and environmental factors which could be specific for each population.

Key words: Gallstone disease, cholesterol gallstone, polymorphisms, genetic susceptibility.

Introduction

Prevalence studies of GSD indicate considerable geographical and regional variations.^{1,2} The lowest GSD prevalence rate has been found in Asian and African populations with predominant pigment gallstones. However, the highest prevalence of GSD was found in European, Amerindian and Latin-American populations, occidentalized countries with predominant cholesterol gallstones. Pima Indians population in Chile has the most increased prevalence of GSD on 48%.¹ In Mexico, necropsy of serial studies shows an approximate prevalence of 14.3%

Address for Correspondence:

Manuscript received and accepted: 13 July and 31 August, 2006.

(16.2% in women and 5.6% in men).³ Also, Mexican-American women in an ultrasonographic study showed a higher prevalence of GSD on 27%.² Therefore, prevalence variability around the world could be explain by ethnic differences, familial aggregation and the presence of a genetic background. Moreover, significant evidence of major genetic determinants of sintomatic GSD on chromosome 1p has been found in Mexican Americans.⁴ One study showed that genetic effects accounted for 25%, shared environmental effects for 13%, and unique environmental effects for 62%.5 Moreover, the presence of risk factors such as obesity, insulin resistance syndrome (IRS) and a family history of GSD have been associated (Table I).^{1,6,7-10} The aquirement of a life style similar to developed countries, characterized by a hypercaloric diet, sedentary life and stress, mixed with a particulary genetic background can determine the development of GSD. This review summarizes the genes implicated in the principal metabolic pathways involved in the development of cholesterol gallstones, such as biliary salt and cholesterol synthesis, lipids transport and the Lith genes previously described in mice.

Physiopathology

The normal bile contains 70% of biliary salts, 22% phospholipids, 4% of cholesterol, 3% of proteins and 0.3% bilirrubin. Gallstones are classified according to physical composition (Table II). Cholesterol gallstones are more prevalent in western countries, so we will be focused on the different cholesterol metabolic pathways. Hepatic cholesterol can be derived from three sources: dietary, peripheral tissues and the liver de novo synthesis. In the liver, cholesterol can be converted into primary bile acids.11,12 There are four mechanisms in the formation of cholesterol gallstones: 1) bile supersaturated with cholesterol, 2) nucleation of cholesterol monohydrate with subsequent crystal retention and stone growth, 3) abnormal gallbladder motor function with delayed emptying and stasis, and 4) gastrointestinal hypomotility modulates the enterohepatic cycling of bile salts. Precipitation of cholesterol crystals from supersaturated bile is required for gallstone formation. Therefore, the precipitation risk is directly related with the concentration of cholesterol and inversely related with the concentrations of billiary salts and

¹ Department of Molecular Biology in Medicine, Old Civil Hospital of Guadalajara "Fray Antonio Alcalde" and Department of Molecular Biology and Genomic, University Center of Health Sciences (*CUCS*), University of Guadalajara. Guadalajara, Jalisco, Mexico. E-mail: apanduro@prodigy.net.mx.

Arturo Panduro MD, PhD.

Molecular Bidogy in Medicine, Civil Hospital of Guadalajara "Fray Antonio Alcalde", P.O. Box. 2-2500 Guadalajara, Jalisco, México 44280. Fax: 52(33)3614-7743. E-mail: apanduro@prodigy.net.mx.

Table I. Risk factors implicated in the development of cholesterol gallstones.

Hi	gh risk
1)	Female sex (estrogens)
2) 3)	Insulin Resistance (DM, IRS)
4)	Genetic factors
Mo	oderate risk
5)	Multiple pregnancies, contraceptives (estrogens)
6)	Bile salt loss (Chron's disease, ileum resection)
7)	Gallbladder dysmotility (vagotomy, octreotide, parenteral nutrition, rapid weight loss)
8)	Antihyperlipidemic drugs (clofibrate)

phospholipids.¹³ The pathogenesis of supersaturated bile is related to increased hepatic cholesterol synthesis and decreased bile acid formation.¹⁴ It is also known that cholesterol contributes to the hypomotility of the gallbladder.⁷

Genes involved in bile and cholesterol metabolism

Several genes implicated in the development of GSD have been described.¹⁵ We classified them according to their participation in different pathways such as biliary and cholesterol synthesis, cholesterol transport and *Lith* genes identified in mice (*Table III, Figures 1 and 2*) However, other groups of genes has been identified such as Mucin genes, genes related to gallbladder functions and genes related to inflammation, possibly induced by infection with *Helicobacter spp.*¹⁶

1. Cholesterol synthesis

1.1 Enzymes

1.1.1 3-Hydroxy Methyl-Glutaryl Coenzyme A Reductase (HMG-CoA):

It is the key enzyme in the *de novo* cholesterol synthesis pathway. Gene expression and the activity of the enzyme are regulated by intracellular cholesterol concentrations via cholesterol-derived oxysterols. In mice, cholesterol synthesis is reduced by decreasing the transcription of the genes encoding for HMG-CoAR. A relation seems to exist between HMG-CoAR expression/ activity and gallstone formation.¹¹ However, in a human study a similar activity of HMG-CoAR was found in patients with gallstones and gallstone-free patients even though the saturation of the gallbladder bile was higher in gallstone patients.¹²

1.1.2 Acyl-coenzyme A Cholesterol Acyltransferase (ACAT)

A cellular cholesterol sensor that esterifies the excess of intracellular cholesterol; then cholesteryl-es-

Table II. Classification of gallstones.

Type Pure	Main component Cholesterol	% Cholesterol > 90	% Frequency
			80-90
Mixed	Cholesterol, calcic salts, phospholipids	30-90	
Black pigments	Calcium bilirrubinate		
Brown pigments	Calcium phosphate and carbonate	< 30	< 20

ters are stored in cytosolic droplets or secreted into the circulation as part of lipoproteins.¹⁷ Hence, biliary cholesterol secretion is under control the activity of ACAT.² There are two genes, the Acat1 gene encodes a thiolase mitochondrial enzyme and the cytosolic acetoacetyl-CoA thiolase encoded by the Acat2 gene. Acat genes are expressed in liver and small intestine. Acat 2 esterifies the cholesterol in murine liver and Acat 1 is the main enzyme that catalyze cholesterol-esters in humans.^{18,19} Hepatic Acat deficiency in rodents²⁰ and humans²² can increase cholesterol availability for biliary secretion and the risk of GSD. Acat deficiency in the intestine diminishes cholesterol absorption resulting in the decreased biliary cholesterol output in liver and presumtive risk of GSD. Studies failed to find relationship between cholesterolosis and the amount of ACAT1 enzyme,²² suggesting the existence of others mechanisms on metabolism control of overall cholesterol.

1.2 Regulatory transcriptional factors

1.2.1 Sterol Regulatory Element Binding Proteins (SREBP)

SREBP1 and SREBP2 are basic helix-loop-helix leucine zipper transcription factors that regulate biosynthetic pathway of fatty acid (FA), cholesterol and LDL-R, by stimulating transcription of genes containing sterol-response-elements.23 Additionaly SREBPs regulate enzymes that generate NADPH and cytosolic acetyl-CoA, which are essential for lipogenesis.²⁴⁻²⁶ SREBP decrease transcription of the HMG-CoAR gene when high-cholesterol diet accumulates in the liver.²⁶ Three isoforms of SREBPs have been identified. SREBP1a and SREBP1c are transcribed from a single gene; SREBP2 is transcribed from a second gene. In liver and other organs, SREBP1c and SREBP2 are most highly expressed. SREBP1 is synthesized as a precursor; in sterol-depleted cells, the precursor is cleaved to generate a soluble fragment that translocates to the nucleus.²⁷ Sterols inhibit the cleavage of SREBP1. Proteolytic release requires a sterol-sensing protein (SCAP).28

Table III. Candidate genes implicated in the development of	GSD.
---	------

		Gene/Protein	Locus	Function/alteration	*
	Enzymes	3HMGCoA ACAT1	5q13.3-q14 11q22.3-q23.1	Key enzyme in the de novo cholesterol synthesis pathway.	А
~		ACAT2	6q25.3-q26	Esterifies intracellular cholesterol excess.	А
Synthesia	Transcriptional Factors	SREBP SHP	17p11.2 1p36	Controls fatty acid and cholesterol homeostasis. Regulates the expression of genes involved in transportation and bile acids synthesis	A C
terol		LXR	12q13.3	Regulate the metabolism of several important lipids.	С
holes		ГЛК	12q	and expression by regulating target genes.	С
0		LRH-1	1q32.1	Serves as a tissue-specific competence factor for bile acid synthesis.	С
		ΡΡΑRγ	3p25	Some genes involved in fatty acid metabolism are activated by PPAR.	С
	Transmembranal	MDR 3	217q22	Facilitates the phosphatidylcholine transport to the bile canalicule	Δ
		ABCA1	9q22-q31	Acts as a cholesterol efflux pump in the cellular lipid removal pathway	A
		ABCG5,	2p21	Crucial in the hepatobiliary and intestinal	
		ABCG8	10.01.01	cholesterol excretion and uptake.	A
		SR-BI	12q24.31	Influences the absorption of cholesterol.	A
±.		IFABP	4q28–4q31	Transports non esterified fatty acids.	C
odsu	Intracellular	NCP-1	18q11-q12	Critical for intestinal cholesterol absorption.	С
Tra		SCP2	1p32	of abalasterol/lipid	C
<u>5</u>		OSBP	11a12.1	Implicated in cholesterol synthesis and trafficking in the liver	C
ster		CAV1	7031 1-31 2	Plays an important role in the regulation of intracellular	C
ole		CATVI	/451.1 51.2	cholesterol homeostasis and it may modulate biliary secretion.	А
G		LFABP	2p11	Regulates lipid cholesterol transport between membranes	
			-	and is required for cholesterol synthesis and metabolism.	С
	Plasmatic	Apo A1	11q23	In the bile acts as an anti-nucleation agent in the formation	
				of gallstones.	С
		CETP	16q21	Facilitates the exchange of neutral lipids among	G
		Ano P	2224	the plasma lipoproteins.	C
		Apo E	2p24 19a13 2	Acts as a ligand for LDL and quilomicrons to their	C
		Apo E	1)415.2	receptors mediating the plasmatic response to	
				dietary cholesterol consumption.	С
sis		CYP7A1	8a11-a12	Catalyzes the first reaction in the catabolism of cholesterol	
liar alt these		0117/11	~q++ q+=	and is the major regulator in the synthesis of biliary	
Bi S Syni				acids in the liver.	А
e p		Lith1	2	A candidate gene for <i>Lith_L</i> is the hile salt export nump	
Litl		Lith 2	19	BsepI, whereas Lith-2 was associated with Mrp2.	А
0				· · · · · · · · · · · · · · · · · · ·	

* A = Associated genes, C = Candidate genes

1.2.2 Small Heterodimeric Partner (SHP)

SHP heterodimerizes with several nuclear receptors and it is expressed in the liver and intestine. It has been demonstrated that activation of FXR (see ahead), by natural and synthetic agonists, increases SHP levels, which in turn reduces SREBP-1c expression²⁹⁻³¹ mechanism proposed to explain the reduction of CYP7A1 expression by bile acids, which also invoked SHP as a mediator.³² SHP is also a potent repressor of LRH1 and its target genes. Together, SHP and LRH1 are important factors in the regulation of gene expression involved in transport and synthesis of bile acids, thereby influencing the formation of supersaturated cholesterol.²⁹

1.2.3 Liver X Receptor (LXR)

Two members are known in this subfamily: LXR (NR1H3) and LXR (NR1H2). Both have a different expression pattern and form heterodimers with another nuclear receptor, the retinoic X receptor (RXR). Whereas LXR (NR1H3) is ubiquitously expressed, LXR (NR1H2) is most highly expressed in



Figure 1. Key proteins diagram of cholesterol and salt bile metabolism. Cholesterol from diet (D) and new synthesis (N) •. Cholesteryl esters ■, Acid Bile ! , Cholesterol synthesis: 3 Hidroxy Methyl Glutaryl CoA Reductase (HMGCoAR) Acyl Coenzyme A Cholesterol Acyltransferase (ACAT). Regulatory Transcriptional Factors: Sterol Regulatory Element Binding Proteins (SREBPs), Liver X Receptor (LXR), Farsenoid X Receptor (FXR), Liver Receptor Homologue (LRH1), Small Heterodimeric Partner (SHP), Peroxisome Proliferator-Activated Receptor Gamma (PPARy).Transmembranal transport of cholesterol and bile constituents 🛥 : Multi- Drug Resistance Transporter 3 (MDR3), ATP-Binding Cassette 1 (ABCA1), ATP-Binding Cassette, Subfamily G, Member 5 and 8 (ABCG5/8), Scavenger Receptor Class B Type I (SR-BI). Intracellular trafficking of cholesterol: Niemann-Pick C1 Like 1 (NPC1L1), Sterol Carrier Protein 2 (SCP2), Oxysterol-Binding Protein 1(OSBP1), Caveolin 1 (CAV1), Liver Faty Acid Binding Protein (L-FABP). Plasmatic Transport of cholesterol: Apolipoprotein A1 (ApoA1), Cholesterol Esters Transfer Protein (CETP), Apoliprotein B (ApoB), Apolipoprotein E (ApoE) and Bile salt biosynthesis: Cholesterol 7a Hidroxilasa (CYP7A1).

the liver and intestinal tract which are the tissues most involved in cholesterol metabolism.³⁰ The expression of the LXR target genes are lowered via SHP-dependent mechanism.³²

1.2.4 Farsenoid X Receptor (FXR)

FXR forms part of the RXR that binds with high affinity to bile acid.³³ Bile acids are able to activate FXR which regulates the target genes in bile acid uptake, synthesis, transport and cholesterol metabolism. Identified target genes are CYP7A1.^{34,35} ApoC-II,³⁶ CYP8B1, basolateral sodium taurocholate cotransporter protein, hepatic canalicular bile salt transporter, ileal bile acid binding protein, SHP and phospholipid transfer protein.^{29,30} There is evidence that the production of certain apolipoproteins



Figure 2. The term Monogenic, Associated and Candidate genes were used to classify the genes implicated in the development of GSD. *Monogenic*. Already unknown and is a unique gene that promotes by his own the development of GSD. *Associated genes*. Participate on regulation of the principal metabolic pathways. Alteration of several of these genes could favor the development of GSD. *Candidate genes*. Genes indirectly involved in several metabolic pathways which could represents a risk factor for GSD.

is regulated by the bile acid–activated nuclear receptor FXR.^{36,37}

1.2.5 Liver Receptor Homologue (LRH1)

Also known as NR5A2, it belongs to the NR5A or the Ftz-F1 subfamily of nuclear receptors and is expressed in liver, pancreas, intestine, and ovary.³⁸ LHR-1 is an orphan nuclear receptor that binds as a monomer. LRH1 serves as a tissue-specific competence factor for bile acid synthesis. LRH1 target genes include: α -fetoprotein, SHP, CETP, CYP7A1 and CYP8B1.²⁹

1.2.6 Peroxisome Proliferator-Activated Receptors (PPARs)

Is a nuclear receptor activated by hypolipidemic compounds like fibrates, but FA have been found to be the natural ligands. Genes involved in FA metabolism and β -oxidation have been found to be activated by PPAR. Specific PPAR γ target gene is peroxisomal acyl-CoA oxidase.¹² PPAR γ and molecules like PPAR γ coactivator-1 are involved in inflammatory gallbladder³⁹ and cholesterol formation.⁴⁰

- 2. Cholesterol transport
- 2.1 Transmembranal transport of cholesterol
- 2.1.1 Multidrug Resistance Transporter 3 (MDR3) MDRs are P-glycoproteins and were discovered as large cell membrane proteins overproduced in cancer cells resistant to a diverse set of hydrophobic

drugs.⁴¹ MDR3 gene is separated from the MDR1 gene by 34 kb. Both human genes are transcribed in the same direction, MDR3 being located downstream from MDR1.⁴² MDR3 codes for a phosphatidylcholine transporter protein in the membrane of the hepatocyte that facilitates the phosphatidylcholine transport to the bile canalicule. Certain authors have reported cases of acute pancreatitis caused by microlithiasis due to mutations in the MDR3 gene, resulting in a phosphatidylcholine deficiency associated with a rapid crystallization of cholesterol.⁴³

2.1.2 ATP-Binding Cassette 1 (ABCA1)

Is a cholesterol and phospholipid efflux pump mediated by ApoA1, essential for HDL formation and controls the first step of reverse cholesterol transport.14,44 Overexpression in transgenic Abca1 mice increases plasma HDL-cholesterol levels, hepatic delivery of HDL cholesteryl-esters and biliary cholesterol concentrations.⁴⁵ Tangier disease is caused by mutations in the ABCA1; these patients have a massive tissue deposition of sterols with near to zero plasma levels of HDL.46 In gallbladder epithelial cells, ABCA147 is regulated by LXR and RXR48 and modulates biliary cholesterol concentrations and its excretion from the body.14 Retinoids and others ABCA1 regulators offer a novel class of agents for treating elevated cholesterol or prevention of GSD in rodents. Aramchol is a FA-bile acid conjugate that induces ABCA1-dependent cholesterol efflux without affecting transcriptional control.49

2.1.3 ATP-Binding Cassette, Subfamily G, Member 5 and 8 (ABCG5/8)

The second set of ABC-half transporters implicated to have a role in the physiological pathways by which dietary cholesterol, as well as non-cholesterol sterols, traffics in human body.^{50,51} The two genes are tandemly grouped.⁵² The ABCG5/8 couple is crucial for hepatobiliary and intestinal cholesterol excretion, expressed in enterocytes and the canalicular membrane.51,53 Liver and intestine maintain sterol balance with respect to noncholesterol sterols.54 In the liver, they are the main player in the secretion of cholesterol and sterols into the bile.55 Mutations of these genes increases dramatically the plasma and hepatic cholesterol levels in response to changes in dietary cholesterol content and cause a rare human disorder, sitosterolemia.^{13,52,56} This disorder has been identified in Mexican-American patients with high concentrations of sterols in plasma tissues⁵² indicating high risk for GSD.57

2.1.4 Scavenger Receptor Class B Type I (SR-BI) This protein is a multifunctional receptor able to bind to anionic phospholipids, native and oxidized LDL and apoptotic cells. It has affinity for HDLs and mediates the selective uptake of cholesterol-esters. It is expressed in intestine and it has been suggested that it contributes to the entrance of cholesterol in the body.⁵⁸ Polymorphisms studied are associated with variation in plasma concentrations of fasting triglyceride.⁵⁸ It has been suggested a possible mechanism involved in the absorption of cholesterol in gallbladder synergized by the union of ApoA1 also present in the bile.⁵⁹

- 2.1.5 Intestinal Fatty Acid Binding Protein (IFABP)
 - The FABP2 gene belongs to a family expressed in a tissue-specific manner.⁶⁰ Protein IFABP is expressed only in epithelial cells of the small intestine,⁶¹ transporting non esterified FA from the plasma membrane, through the aqueous cytosol, to the endoplasmic reticulum (ER),⁶² influencing lipid absorption and plasma levels of lipids.⁶³⁻⁶⁵ FABPs are also hypothesized to serve as cytosolic FA carriers to transport FAs among cellular organelles where FAs have various functions. As intracellular transporters, FABPs deliver regulatory lipids to the nucleus of the cell where the lipids can influence PPAR mediated gene expression.⁶⁶
- 2.2 Intracelullar transport of cholesterol
- 2.2.1 Niemann Pick Type C-1 (NCP1)

This gene codes for transmembranal protein Niemann-Pick C1 Like 1 (NPC1L1), localized in jejunal enterocytes that is critical for intestinal phytosterols and cholesterol absorption containing a sterol sensing domain homologous to the domains found in HMG-CoAR.⁶⁷ NPC1L1 is also a peripheral cholesterol transporter for the energy-dependent vesicular trafficking process of endocytosed lipoprotein cholesterol.⁶⁸⁻⁷⁰ Hepatic NPC1L1 is an important factor that regulates biliary cholesterol secretion in mice, because its inactivation produces an impaired biliary cholesterol secretion in cholesterol-fed mice.⁷¹

2.2.2 Sterol Carrier Protein 2 (SCP2)

This protein plays an important role in intracellular trafficking of cholesterol/lipids⁷²⁻⁷⁴ specially to mitochondria.⁷⁵ Different authors localize the SCP2 in peroxisomes and others in the cytosol.⁷⁶ SCP2 contains both a thiolase domain and a sterol carrier-protein domain and is the key enzyme in β -oxidation of bile acid intermediates. SCP2 is necessary for the rapid transport of newly synthesized cholesterol into bile as well as the conversion of free cholesterol into cholesterol-esters.^{76,77} Increased hepatic SCP2 expression correlated with biliary cholesterol hypersecretion^{78,79} in human patients⁸⁰ and development of GSD in mice.⁸¹

2.2.3 Oxysterol-Binding Protein (OSBP)

A citosolyc protein involve in cholesterol synthesis and trafficking in the liver. OSBP transports sterols from lysosomes to the nucleus, where sterol downregulates genes like LDL-R, HMG-CoAR, HMG-CoAS. OSBP regulates cellular transport of cholesterol, sphingomielyn, oxysterol and sterol by secretory vesicles and control of signalling cascades. OSBP acts as a cholesterol-binding scaffolding protein coordinating the activity of phosphatases to control the extracellular signal-regulated kinase-signaling pathway.⁸²⁻⁸⁴

2.2.4 Caveolin (CAV 1)

CAV1 is highly expressed in intrahepatic basolateral and canalicular membranes. CAV1 mediates endocytosis by caveolae, which are plasma membrane invaginations that are highly enriched in cholesterol and sphingomyelin.85,86 CAV1 is implicated in cell signaling, transcytosis and in regulation of intracellular cholesterol transport.⁸⁷ In hepatocytes, the SR-BI has been shown to be associated with CAV1 indicating their role in cholesterol uptake. Lipid droplets are potential target organelles for caveolar endocytosis demonstrating a role for CAV1 in the maintenance of free cholesterol levels in adipocytes.⁸⁸ The transcription of Cav1 is under the positive control of SREBP pathway, increasing when intracellular cholesterol levels are high.⁸⁹ PPARy overexpression cans upregulate Cav1 expression in macrophages. CAV1 facilitates the transport of cholesterol from the ER to the plasma membrane and it may modulate biliary secretion.⁹⁰

- 2.2.5 Liver Fatty Acid Binding Protein (L-FABP)
 - L-FABP is an abundant cytoplasm component of the hepatocyte that regulates lipid cholesterol transport between membranes. It is required for cholesterol synthesis and metabolism^{13,91} and responsible for the diffusional mechanism of FA transfer to membranes.⁹² L-FABP expression is regulated by PPARγ.⁹³ Polymorphism T94/T94 exhibit higher ApoB levels whereas carriers of the A94 allele seem to be protected against high ApoB levels when consuming a saturated fat diet.⁹⁴

2.3 Plasmatic transport of cholesterol

2.3.1 Apolipoprotein AI (ApoAI)

Apo AI is the main apolipoprotein on HDL. ApoAI is a cofactor for Lecitin Cholesterol Acyl-Transferase (LCAT), which is responsible for the formation of most cholesteryl-esters in plasma. ApoAI is related to CAV1 and both are involved in the regulation of intracellular cholesterol trafficking for the assembly of cellular lipids to ApoAI-HDL.⁹⁵ ApoAI promotes efflux of cholesterol from cells. ApoA1 knockout mice have low plasma HDL-cholesterol levels and their rate of hepatic cholesterol synthesis is 50% lower than wild-type mice. ApoA1 and ApoA2 are secreted in to bile,⁹⁶ and bile acids influence expression of ApoA1.⁹⁷ In contrast, ApoA1 overexpressing mice have been reported to have a 2-fold increase in biliary output of bile acid and cholesterol. In humans, it has been observed that ApoAI removes certain lipids from the bile and acts as an anti-nucleation agent in the formation of gall-stones.⁹⁸

2.3.2 Cholesterol Esters Transfer Protein (CETP)

This protein facilitates the exchange of neutral lipids among the plasma lipoproteins and induces a transfer of cholesterol-esters of the HDL toward the lipoproteins rich in tryglicerydes (TG) in exchange for TG. An enzimatic deficiency causes hyperalphalipoproteinemia and the G to A sustitution in the intron 14 splice donor is a common mutation. Therefore, a high activity of CETP could decrease levels of HDL-cholesterol and high levels of TG, a lipid pattern that increases the risk to develop GSD.⁹⁹

2.3.3 Apolipoprotein B (ApoB)

ApoB is the main protein of chylomicrons and LDL. There are two main forms: ApoB48 and ApoB100. The first is synthesized exclusively by the gut and the second by the liver. ApoB acts as a ligand for the LDL-R mediated by endocytosis. ApoB has been associated with familial hypobetalipoproteinemia, an autosomal dominant disorder of lipid metabolism characterized by extremely low plasma levels of ApoB, as well as low levels of LDL and total cholesterol.¹⁰⁰ The XbaI polymorphism is associated with differences in plasma LDL-cholesterol levels and contributes relatively in the development of GSD in certain populations.¹⁰¹

2.3.4 Apolipoprotein E (ApoE)

It is a major protein component of VLDL and minor of HDL. ApoE acts as a ligand for LDL and quilomicrons to their receptors mediating the plasmatic response to the dietary cholesterol. In familial type III hyperlipoproteinemia, there is impaired clearance of chylomicron remnants and VLDL, increased plasma cholesterol and TG due to a defect in ApoE.¹⁰² ApoE has three allelic variants: E2, E3 and E4. Carriers of E2 allele present low concentrations of total cholesterol and LDL-cholesterol in plasma, while E4 allele carriers present higher levels of LDL and total cholesterol that causes differences in the affinity to ligand-receptor binding. Carriers of E4 allele have high risk of GSD because it increases the lipoprotein uptake and consequently the hepatic and biliary concentration of cholesterol.^{103,104} In contrast, E2 allele provides a protection against GSD.⁹⁸

- 3. Bile salt biosynthesis
- 3.1 Cholesterol 7α Hidroxilasa (CYP7A1)

The first reaction in the catabolism of cholesterol is mediated by the enzyme CYP7A1 that produces biliary acids. LRH1 and RXR/FXR heterodimers regulate CYP7A1 expression and bile acid synthesis.⁴⁸ A deletion in these gene results in the loss of the function of the enzyme increasing the levels of LDL that predispose to premature GSD due to the inability for cholesterol solubilization in the biliary salts. Several studies suggest that a substitution of A for C in the position -204 of the promoter of the gene CYP7A1 has been associated with variations in the concentrations of LDL-cholesterol. In men, the C variant has been associated with an increased rate of total cholesterol/HDL index and higher levels of LDL-cholesterol in plasma.¹⁰⁵

4. Lith genes

Genetic era in GSD research began with the detection of the first cholesterol gallstone genes (Lith) by quantitative trait Loci (QTL) mapping in crosses between gallstone-susceptible (C57L/J) and gallstone-resistant (AKR) inbred strains of mice.¹⁰⁶ QTL analysis localizes additional unknown gallstone genes. It provides the genetic basis for the orthologous human. Lith genes might encode lipid transporters in the canalicular membrane that could transfer lipid molecules into hepatic bile or regulators of cholesterol metabolism. The major murine cholesterol gallstone QTLs determined are Lith1 (chromosome 2), Lith2 (chromosome 19), Lith3 (chromosome 17), Lith4 (chromosome X) and Lith5 (chromosome 5; Muc3).²¹ The most important Lith genes associated with GSD are Lith-1 and Lith-2. These inbred mice present alterations in some of the cholesterol regulatory genes mentioned above as SR-BI, ACAT2, HMG-CoAR, CYP7A1(1) and Hepatic Lipase.¹⁰⁷

Conclusions

Genetic factors identified in animal models suggest plays an important role accounting 25% in the development of GSD.⁴ This review summarizes the most important genes implicated in such pathology. We conclude that it is necessary to study the polymorphic genes (for example: 3HMGCoA, CYP7A1, CETP, Apo A1/B/E, ABCG5/ 8) due to variability and higher prevalence present in each population. However, the presence of environmental factors (diet, physical activity and emotions) of each culture could determine gene expression and exerts an independent effect in the development of GSD. Therefore, integral studies about the interaction between specific genetic and environmental factors in each population will be a very important approach to develop new strategies for prevention, diagnosis and management of patients with GSD based on personalized medicine (genomic medicine).

References

- Acalovschi M. Cholesterol gallstones: from epidemiology to prevention. *Postgrad Med J* 2001; 77: 221-9.
- Everheart JE, Khare M, Hill M, Maurer KR. Prevalence and ethnic differences in gallbladder disease in the United States. *Gastroenterology* 1999; 117: 632-9.
- Mendez-Sanchez N, Jessurun J, Ponciano-Rodriguez G, Alonso-de-Ruiz P, Uribe M, Hernandez-Avila M. Prevalence of gallstone disease in Mexico. A necropsy study. *Dig Dis Sci* 1993; 38: 680-3.
- Puppala S, Dodd GD, Fowler S, Arya R, Schneider J, Farook VS, Granato R, et al. A genomewide search finds major susceptibility loci for gallbladder disease on chromosome 1 in Mexican Americans. *Am J Hum Genet* 2006; 78: 377-9.
- Katsika D, Grjibovski A, Einarsson C, Lammert F, Lichtenstein P, Marschall HU. Genetic and environmental influences on symptomatic gallstone disease: a Swedish study of 43,141 twin pairs. *Hepatology* 2005; 41: 1138-43.
- Johnson C D. ABC of the upper gastrointestinal tract. Upper abdominal pain: Gall bladder. *BMJ* 2001; 7: 1170-3.
- Portincasa P, Moschetta A, Berardino M, Di-Ciaula A, Vacca M, Baldassarre G, Pietrapertosa A, et al. Impaired gallbladder motility and delayed orocecal transit contribute to pigment gallstone and biliary sludge formation in beta-thalassemia major adults. *World J Gastroenterol* 2004; 10: 2383-90.
- Méndez-Sánchez N, Chavez-Tapia NC, Motola-Kuba D, Sanchez-Lara K, Ponciano-Rodríguez G, Baptista H, Ramos MH, et al. Metabolic syndrome as a risk factor for gallstone disease. *World J Gastroenterol* 2005; 11: 1653-7.
- Leitzmann MF, Rimm EB, Willett WC, Spiegelman D, Grodstein F, Stampfer MJ, Colditz GA, et al. Recreational physical activity and the risk of cholecystectomy in women. *N Engl J Med* 1999; 341: 777-84.
- Curiel-López F, González M, Vázquez M, Román S, Panduro A. Prevalence of insulin resistance syndrome in Mexican population with gallstone disease. *Diabetes & Vascular Research* 2005; 2: 167.
- Xiao ZL, Chen Q, Amaral J, Biancani P, Jensen RT, Behar J. CCK receptor dysfunction in muscle membranes from human gallbladders with cholesterol stones. *Am J Physiol* 1999; 276: G1401-7.
- Ahlberg J, Angelin B, Einarsson K. Hepatic 3-hydroxy-3metylglutaryl coenzyme A reductase activity and biliary lipid composition in man: relation to cholesterol gallstone disease and effects of cholic acid and chenodeoxycholic acid treatment. *J Lipid Res* 1981; 22: 410-22.
- Beckingham IJ. ABC of diseases of liver, pancreas, and biliary system: Gallstone disease. *BMJ* 2001; 322: 91-4.
- Zanlugo S, Rogotti A, Nervi F. Hepatic cholesterol transport from plasma in to bile: implications for gallstone disease. *Curr Opin Lipidol* 2004; 15: 279-86.
- Curiel-López F, Ruíz B, Román S, Panduro A. Predisposición Genética de la Litiasis Biliar. *Investigación en Salud* 2005; 7: 79-84.
- Maurer KJ, Rogers AB, Ge Z, Wiese AJ, Carey MC, Fox JG. *Helicobacter pylori* and cholesterol gallstone formation in C57L/J mice: a prospective study. *Am J Physiol Gastrointest Liver Physiol* 2006; 290: G175-G182.
- Suggi S, Lin S, Ohgami N, Chang CCY, Chang TY. Roles of endogenously synthesized sterols in the endocytic pathway. *J Biol Chem* 2006: in press May 31: 1-30.

- Buhman KK, Accad M, Novak S, Choi RS, Wong JS, Hamilton RL, Turley S, et al. Resistance to diet-induced hypercholesterolemia and gallstone formation in ACAT2-deficient mice. *Nat Med* 2000; 6: 1341-7.
- Chang TY, Chang CC, Lin S, Yu C, Li BL, Miyazaki A. Roles of acyl-coenzyme A: cholesterol acyltransferase-1 and -2. *Curr Opin Lipidol* 2001; 12: 289-96.
- Nervi F, Bronfman M, Allalon W, Depiereux E, del Pozo E. Regulation of biliary cholesterol secretion in the rat: role of hepatic cholesterol esterification. *J Clin Invest* 1984; 74: 2226-37.
- Smith JL, Hardie IR, Pillay SP, de Jersey J. Hepatic acyl-coenzyme A: cholesterol acyltransferase activity is decreased in patients with cholesterol gallstones. *J Lipid Res* 1990; 31: 1993-2000.
- 22. Stromsten A, von Bahr S, Bringman S, Saeki M, Sahlin S, Bjorkhem I, Einarsson C. Studies on the mechanism of accumulation of cholesterol in the gallbladder mucosa. Evidence that sterol 27-hydroxylase is not a pathogenetic factor. *J Hepatol* 2004; 40: 8-13.
- Yokoyama C, Wang X, Briggs MR, Admon A, Wu J, Hua X, Goldstein JL, et al. SREBP-1, a basic-helix-loop-helix-leucine zipper protein that controls transcription of the low density lipoprotein receptor gene. *Cell* 1993; 75: 187-97.
- 24. Rigotti A, Marzolo MP, Nervi F. Lipid transport from the hepatocyte into the bile. *Curr Top Membr* 1994; 40: 579–615.
- Oude Elferink RP, Groen AK. Mechanisms of biliary lipid secretion and their role in lipid homeostasis. *Semin Liver Dis* 2000; 20: 293-305.
- Kosters A, Jirsa M, Groen AK. Genetic background of cholesterol gallstone disease. *Biochim Biophys Acta* 2003; 1637: 1-19.
- Wang X, Sato R, Brown MS, Hua X, Goldstein JL. SREBP-1, a membrane-bound transcription factor released by sterol-regulated proteolysis. *Cell* 1994; 77: 53-62.
- DeBose-Boyd RA, Brown MS, Li WP, Nohturfft A, Goldstein JL, Espenshade PJ. Transport-dependent proteolysis of SREBP: relocation of Site-1 protease from Golgi to ER obviates the need for SREBP transport to Golgi. *Cell* 1999; 99: 703-12.
- Goodwin B, Jones SA, Price RR, Watson MA, McKee DD, Moore LB, Galardi C, et al. A regulatory cascade of the nuclear receptor FXR, SHP-1 and LRH-1 represses bile acid biosynthesis. *Mol Cell* 2000; 6: 517-26.
- Lu TT, Makishima M, Repa JJ, Schoonjans K, Kerr TA, Awerx J, Mangelsdorf DJ, et al. Molecular basis for feedback regulation of bile acid synthesis by nuclear receptors. *Mol Cell* 2000; 6: 507-15.
- Brendel C, Schoonjans K, Botrugno OA, Treuter E, Auwerx J. The small heterodimer partner interacts with the liver X receptor alpha and represses its transcriptional activity. *Mol Endocrinol* 2002; 16: 2065-76.
- 32. Watanabe M, Houten SM, Wang L, Moschetta A, Mangelsdorf DJ, Heyman RA, Moore DD, et al. Bile acids lower triglyceride levels via a pathway involving FXR, SHP, and SREBP-1c. *J Clin Invest* 2004; 113: 1408-18.
- Chiang JY. Regulation of bile acid synthesis: pathways, nuclear receptors, and mechanisms. J Hepatol 2004; 40: 539-51.
- Lambert G, Amar MJ, Guo G, Brewer HB Jr, Gonzalez FJ, Sinal CJ. The farnesoid X-receptor is an essential regulator of cholesterol homeostasis. *J Biol Chem* 2003; 278: 2563-70.
- 35. Guo GL, Lambert G, Negishi M, Ward JM, Brewer HB Jr, Kliewer SA, Gonzalez JC, et al. Complementary roles of farnesoid X receptor, pregnane X receptor, and constitutive androstane receptor in protection against bile acid toxicity. *J Biol Chem* 2003; 278: 45062-71.
- 36. Kast HR, Nguyen CM, Sinal CJ, Jones SA, Laffitte BA, Reue K, Gonzalez FJ, et al. Farnesoid X-activated receptor induces apolipoprotein C-II transcription: a molecular mechanism linking plasma triglyceride levels to bile acids. *Mol Endocrinol* 2001; 15: 1720-8.
- Claudel T, Sturm E, Duez H, Torra IP, Sirvent A, Kosykh V, Fruchart JC, et al. Bile acid-activated nuclear receptor FXR suppresses apolipoprotein A-I transcription via a negative FXR response element. J Clin Invest 2002; 109: 961-71.
- Shoonjans K, Dubuquoy L, Mebis J, Fayard E, Wendling O, Haby C, Geboes K, et al. Liver receptor homolog 1 contributes to intestinal tumor formation through effects on cell cycle and inflammation. *Proc Nat Acad Sci* 2005; 102: 2058-62.

- Pan GD, Wu H, Liu JW, Cheng NS, Xiong XZ, Li SF, Zhang GF, et al. Effect of peroxisome proliferator-activated receptor-gamma ligand on inflammation of human gallbladder epithelial cells. *World J Gastroenterol* 2005; 11: 6061-5.
- Bertolotti M, Gabbi C, Anzivino C, Mitro N, Godio C, De Fabiani E, Crestani M, et al. Decreased hepatic expression of PPAR-gamma coactivator-1 in cholesterol cholelithiasis. *Eur J Clin Invest* 2006; 36: 170-5.
- 41. De Vree JML, Jacquemin E, Sturm E, Cresteil D, Bosma PJ, AtenJ, Deleuze JF, et al. Mutations in the MDR3 gene cause progressive familial intrahepatic cholestasis. *Proc Nat Acad Sci* 1998; 95: 282-7.
- 42. Deleuze JF, Jacquemin E, Dubuisson C, Cresteil D, Dumont M, Erlinger S, Bernard O, et al. Defect of multidrug-resistance 3 gene expression in a subtype of progressive familial intrahepatic cholestasis. *Hepatology* 1996; 23: 904-8.
- Rosmorduc O, Hermelin B, Poupon R. MDR3 gene defect in adults with symptomatic intrahepatic and gallbladder cholesterol cholelithiasis. *Gastroenterology* 2001; 120: 1459-67.
- 44. Young SG, Fielding CJ. The ABCs of cholesterol efflux. *Nature Genet* 1999; 22: 316-8.
- 45. Vaisman BL, Lambert G, Amar M, Joyce C, Ito T, Shamburek RD, Cain WJ, et al. ABCA1 overexpression leads to hyperalphalipoproteinemia and increased biliary cholesterol excretion in transgenic mice. J Clin Invest 2001; 108: 303-9.
- 46. Lawn RM, Wade DP, Garvin MR, Wang X, Schwartz K, Porter JG, Seilhamer JJ, et al. The Tangier disease gene product ABC1 controls the cellular apolipoprotein-mediated lipid removal pathway. *J Clin Invest* 1999; 104: R25-31.
- Lee J, Tauscher A, Seo DW, Oram JF, Kuver R. Cultured gallbladder epithelial cells synthesize apolipoproteins A-I and E. Am J Physiol Gastrointest Liver Physiol 2003; 285: G630-41.
- Repa JJ, Turley SD, Lobaccaro JMA, Medina J, Li L, Lustig K, Shan B, Heyman RA, et al. Regulation of absorption and ABC1-mediated efflux of cholesterol by RXR heterodimers. *Science* 2000; 289: 1524-29.
- Goldiner I, van der Velde AE, Vandenberghe KE, van Wijland MA, Halpern Z, Gilat T, Konikoff FM, et al. ABCA1-dependent but apoA-I-independent cholesterol efflux mediated by fatty acid-bile acid conjugates (FABACs). *Biochem J* 2006; 396: 529-36.
- Yu L, Hammer RE, Li-Hawkins J, von Bergmann K, Lutjohann D, Cohen JC, Hobbs HH, et al. Disruption of Abcg5/Abcg8 in mice reveals their crucial role in biliary cholesterol secretion. *Proc Nat Acad Sci* 2002; 99: 16237-42.
- Hazard SE, Patel SB. Sterolins ABCG5 and ABCG8: regulators of whole body dietary sterols. *Pflugers Arch* 2006; 27: 1-8.
- Berge KE, Tian H, Graf GA, Yu L, Grishin NV, Schultz J, Kwiterovich P, et al. Accumulation of dietary cholesterol in sitosterolemia caused by mutations in adjacent ABC transporters. *Science* 2000; 290: 1771-5.
- 53. Kosters A, Kunne C, Looije N, Patel SB, Oude Elferink RP, Groen AK. The mechanism of Abcg5/Abcg8 in biliary cholesterol secretion in mice. *J Lipid Res* 2006: In press June 12.
- Plosch T, Kosters A, Groen AK, Kuipers F. The ABC of hepatic and intestinal cholesterol transport. *Handb Exp Pharmacol* 2005; 170: 465-82.
- Small DM. Role of ABC transporters in secretion of cholesterol from liver into bile. (Commentary) Proc Nat Acad Sci 2003; 100: 4-6.
- Miettinen TA, Klett EL, Gylling H, Isoniemi H, Patel SB. Liver transplantation in a patient with sitosterolemia and cirrhosis. *Gastroenterology* 2006; 130: 542-7.
- vanBerge-Henegouwen GP, Venneman NG, Portincasa P, Kosters A, van Erpecum KJ, Groen AK. Relevance of hereditary defects in lipid transport proteins for the pathogenesis of cholesterol gallstone disease. *Scand J Gastroenterol* Suppl 2004; 241: 60-9.
- Acton S, Osgood D, Donoghue M, Corrella D, Pocovi M, Cenarro A, Mozas P, et al. Association of Polymorphisms at the SR-BI Gene Locus with Plasma Lipid Levels and Body mass Index in a White Population. *Artherioscler Thromb Vasc Biol* 1999; 19: 1734-43.
- Johnson MS, Svensson PA, Boren J, Billig H, Carlsson LM, Carlsson B. Expression of scavenger receptor class B type I in gallbladder columnar epithelium. J Gastroenterol Hepatol 2002; 17: 713-20.

- Bernlohr DA, Simpson MA, Hertzel AV, Banaszak LJ. Intracellular lipid-binding proteins and their genes. *Annu Rev Nutr* 1997; 17: 277-303.
- Sweetser DA, Birkenmeier EH, Klisak IJ, Zollman S, Sparkes RS, Mohandas T, Lusis AJ, et al. The human and rodent intestinal fatty acid binding protein genes. A comparative analysis of their structure, expression, and linkage relationships. *J Biol Chem* 1987; 262: 16060-71.
- 62. Pratley RE, Baier L, Pan DA, Salbe AD, Storlien L, Ravussin E, Bogardus C. Effects of an Ala54Thr polymorphism in the intestinal fatty acid-binding protein on responses to dietary fat in humans. J Lipid Res 2000; 41: 2002-8.
- Levy E, Menard D, Delvin E, Stan S, Mitchell G, Lambert M, Ziv E, et al. The polymorphism at codon 54 of the FABP2 gene increases fat absorption in human intestinal explants. *J Biol Chem* 2001; 276: 39679-84.
- Randle PJ, Garland PB, Hales CN, Newsholme EA. The glucose fattyacid cycle. Its role in insulin sensitivity and the metabolic disturbances of diabetes mellitus. *Lancet* 1963; 13: 785-9.
- 65. Dworatzek PDN, Hegele RA, Wolever TMS. Postprandial lipemia in subjects with the threonine 54 variant of the fatty acid– binding protein 2 gene is dependent on the type of fat ingested. *Am J Clin Nutr* 2004; 79: 1110-7.
- Bernlohr DA, Coe NR, Simpson MA, Hertzel AV. Regulation of gene expression in adipose cells by polyunsaturated fatty acids. *Adv Exp Med Biol* 1997; 422: 145-56.
- 67. Davis HR Jr, Zhu LJ, Hoos LM, Tetzloff G, Maguire M, Liu J, Yao X, et al. Niemann-Pick C1 Like 1 (NPC1L1) is the intestinal phytosterol and cholesterol transporter and a key modulator of whole-body cholesterol homeostasis. *J Biol Chem* 2004; 279: 33586-92
- Anwer MS. Cellular regulation of hepatic bile acid transport in health and cholestasis. *Hepatology* 2004; 39: 581-90.
- Maxfield FR, Wüstner D. Intracellular cholesterol transport. J Clin Invest 2002; 110: 891-98.
- Liscum L, Munn NJ. Intracellular cholesterol transport. *Biochim Biophys Acta* 1999; 1438: 19-37.
- Amigo L, Mendoza H, Castro J, Quinones V, Miquel JF, Zanlungo S. Relevance of Niemann-Pick type C1 protein expression in controlling plasma cholesterol and biliary lipid secretion in mice. *Hepatology* 2002; 36: 819-28.
- Murphy EJ. Sterol carrier protein-2: not just for cholesterol any more. Mol Cell Biochem 2002; 239: 87-93.
- Huang H, Gallegos AM, Zhou M, Ball JM, Schroeder F. Role of the sterol carrier protein-2 N-terminal membrane binding domain in sterol transfer. *Biochemistry* 2002; 41: 12149-62.
- 74. Kriska T, Levchenko VV, Korytowski W, Atshaves BP, Schroeder F, Girotti AW. Intracellular dissemination of peroxidative stress: Internalization, transport and lethal targeting of a cholesterol hyderoperoxide species by SCP-2-overexpressing hepatoma cells. J Biol Chem 2006; in press June 12.
- Vila A, Levchenko VV, Korytowski W, Girotti AW. Sterol carrier protein-2-facilitated intermembrane transfer of cholesterol- and phospholipid-derived hydroperoxides. *Biochemistry* 2004; 43: 12592-605.
- Gallegos AM, Atshaves BP, Storey SM, Starodub O, Petrescu AD, Huang H, McIntosh AL, et al. Gene structure, intracellular localization, and functional roles of sterol carrier protein-2. *Prog Lipid Res* 2001; 40: 498-563.
- Stolowich NJ, Petrescu AD, Huang H, Martin GG, Scott AI, Schroeder F. Sterol carrier protein-2: structure reveals function. *Cell Mol Life Sci* 2002; 59: 193-212.
- Puglielli L, Rigotti A, Amigo L, Nunez L, Greco AV, Santos MJ, Nervi F. Modulation of intrahepatic cholesterol trafficking: evidence by in vivo antisense treatment for the involvement of sterol carrier protein-2 in newly synthesized cholesterol transport into rat bile. *Biochem J* 1996; 317: 681-7.
- Amigo L, Zanlungo S, Miquel JF, Glick JM, Hyogo H, Cohen DE, Rigotti A, et al. Hepatic overexpression of sterol carrier protein-2 inhibits VLDL production and reciprocally enhances biliary lipid secretion. *J Lipid Res* 2003; 44: 399-407.
- Ito T, Kawata S, Imai Y, Kakimoto H, Trzaskos JM, Matsuzawa Y. Hepatic cholesterol metabolism in patients with cholesterol gallstones:

enhanced intracellular transport of cholesterol. *Gastroenterology* 1996; 110: 1619-27.

- Fuchs M, Lammert F, Wang DQ, Paigen B, Carey MC, Cohen DE. Sterol carrier protein 2 participates in hypersecretion of biliary cholesterol during gallstone formation in genetically gallstone-susceptible mice. *Biochem J* 1998; 336: 33-7.
- 82. Olkkonen VM, Johansson M, Suchanek M, Yan D, Hynynen R, Ehnholm C, Jauhiainen M, et al. The OSBP-related proteins (ORPs): global sterol sensors for co-ordination of cellular lipid metabolism, membrane trafficking and signalling processes? *Biochem Soc Trans* 2006; 34: 389-91.
- Olkkonen VM, Levine TP. Oxysterol binding proteins: in more than one place at one time? *Biochem Cell Biol* 2004; 82: 87-98.
- Wang P, Weng J, Anderson RGW. OSBP is a cholesterol-regulated scaffolding protein in control of ERK1/2 activation. *Science* 2005; 307: 1472-76.
- Pelkmans L, Zerial M. Kinase-regulated quantal assemblies and kissand-run recycling of caveolae. *Nature* 2005; 436: 128-33.
- Cheng ZJ, Deep Singh R, Marks DL, Pagano RE. Membrane microdomains, caveolae, and caveolar endocytosis of sphingolipids. *Mol Membr Biol* 2006; 23: 101-10.
- Frank PG, Galbiati F, Razani B, Volante D, Razani B, Cohen DE, Marcel YL, et al. Influence of caveolin-1 on cellular cholesterol efflux mediated by high density lipoproteins. *Am J Physiol Cell Physiol* 2001; 280: C1204-14.
- Le Lay S, Hajduch E, Lindsay MR, Le Liepvre X, Thiele C, Ferre P, Parton RG, et al. Cholesterol-induced caveolin targeting to lipid droplets in adipocytes: a role for caveolar endocytosis. *Traffic* 2006; 7: 549-61.
- Pallottini V, Martini C, Cavallini G, Donati A, Bergamini E, Notarnicola M, Caruso MG, et al. Modified HMG-CoA reductase and LDLr regulation is deeply involved in age-related hypercholesterolemia. J Cell Biochem 2006; in press Jun 1.
- Llaverias G, Vazquez-Carrera M, Sanchez RM, Noe V, Ciudad CJ, Laguna JC, Alegret M. Rosiglitazone upregulates caveolin-1 expression in THP-1 cells through a PPAR-dependent mechanism. *J Lipid Res* 2004; 45: 2015-24.
- Chen SH, Van Tuinen P, Ledbetter DH, Smith LC, Chan L. Human liver fatty acid binding protein gene is located on chromosome 2. Somat cell. *Molec Genet* 1996; 12: 303-6.
- Corisco B, Liou HL, Storch J. The alpha-helical domain of liver fatty acid binding protein is responsible for the diffusion-mediated transfer of fatty acids to phospholipid membranes. *Biochemistry* 2004; 43: 3600-7.
- 93. Skrtic S, Carlsson L, Ljugberg A, Linden D, Michalik L, Wahli W, Oscarsson J. Decreased expression of peroxisome proliferator-activated receptor alpha and liver fatty acid binding protein after partial hepatectomy of rats and mice. *Liver Int* 2005; 25: 33-40.
- 94. Robitaille J, Brouillette C, Lemieux S, Perusse L, Gaudet D, Vohl MC. Plasma concentrations of apolipoprotein B are modulated by a gene-diet interaction effect between the LFABP T94A polymorphism and dietary fat intake in French-Canadian men. *Mol Genet Metab* 2004; 82: 296-303.
- Ito J, Kheirollah A, Nagayasu Y, Lu R, Kato K, Yokoyama S. Apolipoprotein A-I increases association of cytosolic cholesterol and caveolin-1 with microtubule cytoskeletons in rat astrocytes. J Neurochem 2006; 97: 1034-43.
- van Erpecum KJ, van Berge-Henegouwen GP. Gallstones: an intestinal disease? Gut 1999; 44: 435-38.
- Srivastava RA, Srivastava N, Averna M. Dietary cholic acid lowers plasma levels of mouse and human apolipoprotein A-I primarily via a transcriptional mechanism. *Eur J Biochem* 2000; 267: 4272-80.
- Mittal B, Mittal RD. Genetics of Gallstone Disease. J Postgrad Med 2002; 48: 149-52.
- Akita H, Chiba H, Tsuchihashi K, Tsuji M, Kumagai M, Matsuno K, Kobayashi K. Cholesteryl ester transfer protein gene: two common mutations and their effect on plasma high-density lipoprotein cholesterol content. *J Clin Endocr Metab* 1994; 79: 1615-18.
- 100. Pulai JI, Neuman RJ, Groenewegen AW, Wu J, Schonfeld G. Genetic heterogeneity in familial hypobetalipoproteinemia: Linkage and nonlinkage to the apoB gene in caucasian families. *American Journal of Medical Genetics* 1998; 78: 79-86.

- 101.Han T, Jiang Z, Suo G, Zhang S. Apolipoprotein B-100 gene Xba I polymorphism and cholesterol gallstone disease. *Clin Genet* 2000; 57: 304-8.
- 102.Boerwinkle E. Utermann G. Simultaneous effects of the apolipoprotein E polymorphism on apolipoprotein E, apolipoprotein B, and cholesterol metabolism. Am J Hum Genet 1998; 42: 104-12.
- 103. Juvonen T, Savolainen MS, Kairaluoma MI, Lajunen LH, Humphries SE, Kesaniemi YA. Polymorphism at the apoB, apoA-1 and cholesteryl ester transfer protein gene loci in patients with gallbladder disease. J Lipid Res 1995; 36: 804-10.
- 104. Bertomeu A, Ros E, Zambon D, Vela M, Perez-Ayuso RM, Torgorena E, Tris M, et al. Apoliprotein E Polymorphism and Gallstones. *Gastroenterology* 1996; 111: 1603-10.
- 105. Couture P, Otvos JD, Cupples LA, Wilson PW, Schaefer EJ, Ordovas JM. Association of the A-204C polymorphism in the cholesterol 7-acylcholesterol levels in the Framingham Offspring Study. J Lipid Res 1999; 40: 1883-89.
- 106. Wittenburg H, Lammert F, Wang DQ, Churchill GA, Li R, Bouchard G, Carey MC, et al. Interacting QTLs for cholesterol gallstones and gallbladder mucin in AKR and SWR strains of mice. *Physiol Genomics* 2002; 8: 67-77.
- 107.Lammert F, Wang Q-H D, Paigen B, Carey C. Phenotypic characterization of Lith genes that determine susceptibility to colesterol cholelithiasis in inbred mice: integrated activities of hepatic lipid regulatory enzymes. J Lipid Res 1999; 40: 2080-90.