## P-14 OPTIMIZATION OF MOLECULAR METHODS FOR SARS-CoV-2 QUALITATIVE DETECTION AND GENOTYPING IN RESPIRATORY SPECIMENS FROM PATIENTS WITH LIVER DISEASE

Vanessa Duarte da Costa<sup>1</sup>, Alanna Calheiros Santos<sup>1</sup>, Lucas Limas da Silva<sup>1</sup>, Wilian Jean Wiggers<sup>2</sup>, Claudia Alexandra Pontes Ivantes<sup>2</sup>, Danielle Malta Lima<sup>3</sup>, Jeová Keny Baima Colares<sup>3</sup>, Deusilene Souza Vieira Dallacqua<sup>4</sup>, Ana Rita Coimbra Motta-Castro<sup>5</sup>, Vanessa Salete de Paula<sup>6</sup>, Alberto Martín Rivera Dávila<sup>7</sup>, Priscilla Pollo-Flores<sup>8</sup>, Lia Laura Lewis-Ximenez<sup>1</sup>, Livia Melo Villar<sup>1</sup>

<sup>1</sup> Brazilian Reference Laboratory of Viral Hepatitis, Oswaldo Cruz Institute, FIOCRUZ, Rio de Janeiro, Brazil <sup>2</sup> Service of Gastroenterology, Hepatology and Liver Transplantation, Hospital Nossa Senhora das Graças, Curitiba, Paraná, Brazil <sup>3</sup> Postgraduate Program in Pathology, Federal University of Ceará, Fortaleza, Ceará, Brazil <sup>4</sup> Molecular Virology Laboratory, Oswaldo Cruz Foundation, FIOCRUZ, Porto Velho, Rondônia, Brazil <sup>5</sup> Federal University of Mato Grosso do Sul, Campo Grande, Mato Grosso do Sul, Brazil; Oswaldo Cruz Foundation, Campo Grande, Mato Grosso do Sul, Brazil <sup>6</sup> Molecular Virology Laboratory, Oswaldo Cruz Institute, FIOCRUZ, Rio de Janeiro, Brazil <sup>7</sup> Computational and Systems Biology Laboratory, Graduate Program in Biodiversity and Health, Oswaldo Cruz Institute, Fiocruz, Rio de Janeiro, Brazil <sup>8</sup> Internal Medicine Department, Fluminense Federal University, Niterói, Rio de Janeiro, Brazil

**Introduction and Objectives:** SARS-CoV-2 active infection diagnosis is currently performed through RT-qPCR. Despite the fact that PCR-based assays can provide results relatively fast, these techniques require capable professionals, specific equipment and adequate infrastructure. In order to facilitate COVID-19 diagnosis in remote areas, an alternative to RT-qPCR would be loop-mediated isothermal (RT-LAMP) amplification. SARS-CoV-2 variant genotyping through high-throughput sequencing (HTS) allows SARS-CoV-2 genomic surveillance, especially for patients with a higher vulnerability. This study aimed to optimize RT-LAMP and HTS methods for SARS-CoV-2 RNA detection and genotyping, respectively, in respiratory samples from patients with liver disease.

**Materials and Methods:** A total of 142 respiratory secretions were obtained from individuals with SARS-CoV-2 RNA detectable by RT-qPCR (N1 Ct  $\leq$  30), divided into groups with (n=18) or without (n=124) liver disease. The study also enrolled 55 individuals who had SARS-CoV-2 RNA undetectable at RT-qPCR. For RT-LAMP methodology, primers were used for ORF1 gene amplification. As for HTS genotyping, the steps of cDNA synthesis, complete SARS-CoV-2 genome PCR amplification, preparation of genomic libraries and sequencing in MinION device were performed for 26 swab samples.

**Results:** Samples with viral RNA detectable by RT-qPCR had a mean Ct value of  $24.3\pm3.75$ . Referring to RT-LAMP, it was observed a sensitivity of 71.1% (101/142). When considering RT-qPCR mean Ct value, RT-LAMP sensitivity was 88.9% (16/18), associated with a mean Ct of  $23.3\pm3.5$  for patients with COVID and hepatitis. A specificity of 100% (55/55) was observed since all negative swabs tested by RT-qPCR were negative at RT-LAMP. Through sequencing by MinION, SARS-CoV-2 lineages gamma (7/26; 27%), zeta (1/26; 3.9%), delta (6/26; 23%) and omicron (12/26; 46.1%) were genotyped and detected by RT-LAMP.

**Conclusions:** RT-LAMP demonstrated high sensitivity for molecular detection of SARS-CoV-2 RNA for patients with high viral load.

Besides, RT-LAMP was capable of detecting all SARS-CoV-2 lineages genotyped by MinION in both groups.

https://doi.org/10.1016/j.aohep.2023.100918

## P-15 MMP-2 AND MMP-9 LEVELS IN ALCOHOLIC LIVER DISEASE, NON-ALCOHOLIC FATTY LIVER DISEASE AND CHRONIC HEPATITIS C

María Lemus-Peña<sup>1</sup>, Abigail Hernandez-Barragan<sup>1</sup>, Daniel Montes de Oca-Ángeles<sup>1</sup>, Marisela Hernandez-Santillan<sup>1</sup>, Daniel Santana-Vargas<sup>2</sup>, Moisés Martinez-Castillo<sup>1</sup>, Zaira Medina-Avila<sup>1</sup>, Aldo Torre-Delgadillo<sup>2</sup>, José Luis Pérez-Hernández<sup>2</sup>, Fátima Higuera-De la Tijera<sup>2</sup>, Paula Cordero-Pérez<sup>3</sup>, Linda Muñoz-Espinosa<sup>3</sup>, David Kershenobich<sup>4</sup>, Gabriela Gutiérrez-Reyes<sup>1</sup>

<sup>1</sup> Liver. Pancreas and Motility Laboratory, Unit of Research in Experimental Medicine, School of Medicine, Universidad Nacional Autónoma de México (UNAM), Mexico City, Mexico

<sup>2</sup> Department of Gastroenterology, Hospital General de México "Dr. Eduardo Liceaga", Mexico City, Mexico <sup>3</sup> Hospital Universitario "Dr. José Eluterio González", School of Medicine, Universidad Autónoma de Nuevo León (UANL), Nuevo León, Mexico <sup>4</sup> Instituto Nacional de Ciencias Médicas y Nutrición "Salvador Zubirán", Mexico City, Mexico

**Introduction and Objectives:** It has already been reported that elevated serum levels were present in Hepatitis C patients, although they were found inactive. The behavior of gelatinases MMP-2 and -9 is yet unknown in other liver diseases. This study aimed to evaluate serum concentration of MMP-2 and -9 in different etiologies of liver disease also according to fibrosis stages.

**Materials and Methods:** Cross-sectional multicentric study, including subjects with no alcoholic fatty liver disease (NAFLD), chronic Hepatitis C (CHC), alcohol cirrhosis (CiOH) and alcoholism (OH), groups with alcohol drinking habits were classified according to WHO criteria, with clinical and biochemical evidence of alcoholic liver disease (ALD). Transient elastography (Fibroscan) was performed in NAFLD and CHC, considering mild fibrosis (FL: F0, F1, F2) and severe fibrosis (FA: F3, F4). As controls, subjects without alcohol consumption (CT) were recruited. Multiplex®-MERCK© was used for MMP-2 and -9 quantification. Statistical analysis was performed by Mann Whitney-U test, p<0.05, with SPSS V.22.

**Results:** The groups included were: 27 NAFLD (mild fibrosis: F0, F1, F2), 36 NAFLD (severe fibrosis: F3, F4), 48 CHC (mild fibrosis: F0, F1, F2), 54 CHC (severe fibrosis: F3, F4), 45 (CiOH), 99 (OH), and 138 CT. Both gelatinases, MMP-2 y MMP-9, were found elevated in CHC (mild and severe fibrosis) vs. CT; and decreased in OH, CiOH, HGNA (mild and severe fibrosis) vs. CT, plus there is significant differences between all etiologies, p<0.001.

**Conclusions:** In patients with CHC, MMP-2 y -9 serum concentration increases, particularly in severe fibrosis stages, although it has no effect on ECM (extracellular matrix) degradation, as they are inactive. Nevertheless, there is a significant decrease in these gelatinases in ALD and NAFLD. MMP-2 y MMP-9 are modulated according to etiological agents, which can be useful for the differential diagnosis of liver diseases.

**Funding:** This work was partially financed by CONACyT SALUD-2016-272579 and PAPIIT- UNAM TA200515

https://doi.org/10.1016/j.aohep.2023.100919