

## CLINICAL SCIENCE

# The role of oxidative stress and antioxidants in the pathogenesis of age-related macular degeneration

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**OBJECTIVE:** To investigate the role of oxidant/antioxidant status and protein oxidation in the development of age-related macular degeneration.

**METHOD:** The activities of serum superoxide dismutase and glutathione peroxidase and the levels of serum malondialdehyde, advanced oxidation protein products, glutathione and vitamin C were measured in 25 patients with age-related macular degeneration and 25 control subjects without age-related macular degeneration.

**RESULT:** The malondialdehyde and advanced oxidation protein product levels in the serum were significantly higher in the age-related macular degeneration patient group than in the control group ( $p < 0.05$ ). The superoxide dismutase activity in the serum was significantly lower in the age-related macular degeneration patient group than in the control group ( $p < 0.05$ ). The levels of vitamin C and glutathione and the activity of glutathione peroxidase in the serum were unchanged between groups ( $p > 0.05$ ).

**CONCLUSION:** The results of the present study suggest that decreased effectiveness of the antioxidant defense system and increased oxidative stress may play a role in the pathogenesis of age-related macular degeneration.

**KEYWORDS:** Oxidative stress; Antioxidants; Protein oxidation; AMD.

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## INTRODUCTION

Age-related macular degeneration (AMD) is the leading cause of irreversible visual impairment and blindness among people aged 60 years and older.<sup>1</sup> Although the disease presents a serious social and economic problem, its pathogenesis and etiology are still unclear.<sup>2,3</sup> Based on the disease's clinical and pathological features, there are two subtypes of late AMD: atrophic (dry) and neovascular (wet), both of which can lead to significant visual loss.<sup>4</sup>

Over the past decade, the body of literature regarding the modifiable factors associated with AMD has grown considerably and includes cigarette smoking, age,<sup>5</sup> nutritional factors,<sup>6-8</sup> obesity<sup>9</sup> and insufficient antioxidants in the diet.<sup>10</sup>

The eye is an exceptional organ because of its continuous exposure to environmental chemicals, radiation, and atmospheric oxygen.<sup>11</sup> These oxidative stresses have been implicated in the possible pathophysiology of various ocular diseases, such as AMD, cataracts, glaucoma, uveitis, and pseudoexfoliation syndrome. Reactive oxygen species

(ROS) are involved in this process. Several ocular degenerative disorders have been studied, and the presence of oxidative stress has been demonstrated through markers of lipid peroxidation, the activity of antioxidant enzymes, and the levels of low-molecular-weight antioxidants.<sup>12</sup>

Non-enzymatic lipid peroxidation is an example of a free radical-associated process through which oxidative stress promotes cellular damage. Serum malondialdehyde (MDA) is the end product of the primary reactions that lead to the significant oxidation of such polyunsaturated fatty acids in cellular membranes and, thus, serves as a reliable marker of oxidative stress.<sup>13</sup> Protein oxidation is currently considered to be an important factor in a variety of diseases, such as Alzheimer's and Parkinson's diseases, cancer, hypertension, cardiovascular disease, diabetes, ischemia-reperfusion injury and aging.<sup>14,15</sup>

Advanced oxidation protein products (AOPP) are described as dityrosines that contain cross-linked protein products. Importantly, this definition excludes protein aggregates that form as a result of disulfide links following low-level oxidative stress. Therefore, the presence of AOPP may be a better and more accurate marker of oxidative stress than lipid peroxidation products.<sup>16</sup>

Endogenous antioxidants, including such non-enzymatic scavengers as glutathione (GSH) and such antioxidant enzymes as superoxide dismutase (SOD), glutathione peroxidase (GPx), and catalase (CAT), are the first lines of

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defense against oxidative stress and act by scavenging potentially damaging free radical moieties.<sup>17</sup>

Ascorbate (vitamin C) is the most effective aqueous-phase antioxidant found in human blood. Increasing evidence suggests that, within the aqueous phase, vitamin C plays a vital role in the antioxidant defense mechanism of the eye, protecting ocular tissues against photooxidative damage by acting as a free radical scavenger.<sup>18</sup>

Determining the level of oxidative stress encountered by the human eye has not yet been attempted in clinical diagnoses and/or treatment.<sup>19</sup> The aim of the present study was to investigate the role of antioxidants and protein and lipid peroxidation in the development of AMD. Therefore, we measured the activities of the SOD and GPx enzymes and the serum levels of MDA, AOPP, GSH and vitamin C in patients with AMD and in control subjects not exhibiting AMD.

## MATERIALS AND METHODS

This study included 15 men and 10 women (mean age  $\pm$  SD;  $65.16 \pm 13.96$  years) with AMD and choroidal neovascular (CNV) membrane, which is secondary to AMD. The control group included 15 men and 10 women (mean age  $\pm$  SD;  $65.72 \pm 12.62$  years). No statistically significant differences between the groups were observed in terms of age and sex. Patients with other ophthalmic conditions (e.g., glaucoma, uveitis, pseudoexfoliation syndrome, other progressive retinal diseases) and systemic diseases (e.g., diabetes, arthritis, coronary arterial disease, peripheral vascular disease) were excluded.

All subjects, both in the control group and the patient group, completed a questionnaire confirming the following information: age, gender, non-smoker status, non-consumption of supplements, such as vitamins and/or antioxidants. All patients with AMD were first tested to ensure that they did not ingest antioxidant vitamins and minerals. After obtaining blood samples from the patients, we initiated anti-vascular endothelial growth factor therapy.

All patients underwent a comprehensive ophthalmic examination. CNV was diagnosed by slit-lamp biomicroscopy of the fundi, color fundus photographs, fundus fluorescein angiographies, and optical coherence tomographies. The control subjects exhibited entirely normal ophthalmic conditions. For this study, the rules of the Human Ethics Committee were followed. Blood samples, which were drawn from patients and control subjects after an overnight fast and before the operation, were centrifuged at  $2,000 \times g$  for 10 min at  $4^\circ\text{C}$ . Serum samples were stored at  $-70^\circ\text{C}$  for further analysis.

AOPP levels were measured by a spectrophotometric method (Shimadzu UV 1601 spectrophotometer, Shimadzu, Tokyo, Japan) in the presence of potassium iodide at  $340 \text{ nm}$ <sup>16</sup> and using chloramine-T solutions for calibration. AOPP levels were expressed in micromoles of chloramine-T equivalents per liter.

Lipid peroxidation was estimated using the thiobarbituric acid-reactive substances (TBARS) test, as described previously.<sup>20</sup> Briefly, the formation of TBARS was quantitated using 1,1,3,3-tetraethoxypropane as a standard, and the absorbances of the TBARS were read at  $532 \text{ nm}$  using a Shimadzu UV 1601 spectrophotometer (Tokyo, Japan).

The GSH levels were determined as total sulfhydryl group (R-SH).<sup>21</sup> A 0.5 ml aliquot of each sample was mixed with 1 ml of a solution containing 100 mM Tris-HCl pH 8.2, 1% sodium dodecyl sulfate (SDS) and 2 mM EDTA. The

mixture was incubated for 5 min at  $25^\circ\text{C}$  and centrifuged to remove any precipitate. Next, 5,5-dithiobis (2-nitrobenzoic acid)/DTNB 0.3 mM was added to each reaction volume, and the reaction was incubated for 15 min at  $37^\circ\text{C}$ . The absorbance of each sample was determined at  $412 \text{ nm}$ . The R-SH levels were calculated assuming a molar extinction coefficient of  $13,000 \text{ mol}^{-1} \text{ cm}^{-1}$  at  $412 \text{ nm}$ .

The SOD activity was measured by inhibiting this activity through nitroblue tetrazolium (NBT) reduction. Xanthine-xanthine oxidase was used as a superoxide generator, and one IU was defined as the quantity of SOD required to produce a 50% inhibition.<sup>22</sup>

The level of GPx activity was determined spectrophotometrically as described in the literature.<sup>23</sup> The reaction mixture, containing a 50 mM phosphate buffer pH 7.4, 7.7 units of GSH reductase, 5 mM GSH, and crude extract, was preincubated for 5 min at  $37^\circ\text{C}$ . Next, 20  $\mu\text{l}$  of a nicotinamide adenine dinucleotide phosphate solution in the reduced form (NADPH) (0.3 mM) was added, and the hydroperoxide-independent consumption of NADPH was monitored for approximately 5 min. The overall reaction was started by adding 20  $\mu\text{l}$  of prewarmed hydroperoxide solution (0.025 mM), and the reduction in absorption at  $340 \text{ nm}$  was monitored.

The level of total ascorbate was determined by the modified Roe and Kuether method.<sup>24</sup> Serum samples were added to a trichloroacetic acid solution and centrifuged at  $3,000 \times g$  for 10 min. Next, a 2,4-dinitrophenylhydrazine (DNPH)-thiourea-copper sulfate reagent was added to the serum sample tubes. The contents of each tube were mixed, capped with parafilm, and placed in a water bath at  $37^\circ\text{C}$  for 4 h, after which the tubes were removed and cooled in ice water. An ice-cold 65% sulfuric acid solution was added to each tube, and the sample was mixed thoroughly. The mixture was allowed to stand at room temperature for 30 min, and the absorbance of each tube was read in a Shimadzu UV 1601 model spectrophotometer at 515 and 520 nm. The lower limit of detection for vitamin C was  $0.05 \mu\text{mol/L}$ .

**Statistical Analysis.** Data are presented as the mean  $\pm$  SD. Statistical analyses were conducted by Kruskal Wallis test and Mann-Whitney *U*-test (SPSS for Windows 11.5; SPSS, Chicago, IL, USA). A value of  $p < 0.05$  was defined as significant.

## RESULTS

This study included a total of 50 subjects who were equally divided among two groups, an AMD group and a control group. The mean age was  $65.16 \pm 13.96$  years (40-89 years) (for 15 men and 10 women) in the AMD group and  $65.72 \pm 12.62$  years (36-89 years) (for 15 men and 10 women) in the control group. All of the groups were matched for age and gender, and no statistically significant differences were observed. The levels of MDA and AOPP in the serum were significantly higher in the AMD patient group than in the control group ( $p < 0.05$ ) (Table 1). Furthermore, the level of SOD activity in the serum was significantly lower in the AMD patient group than in the control group ( $p < 0.05$ ) (Table 1). In addition, the levels of vitamin C and GSH and the activities of GPx in the serum were unchanged between the two groups ( $p > 0.05$ ) (Table 1).

## DISCUSSION

AMD is a complex, multifactorial disease of aging for which several theories of pathogenesis have been proposed,

**Table 1 - Oxidation markers are down-regulated in AMD patients and Control Subjects (mean  $\pm$  SD).**

	AMD group (n=25)	Control group (n=25)
SOD (U/mL)	4.99 $\pm$ 3.09*	9.91 $\pm$ 4.42
GPx (U/mL)	12.97 $\pm$ 11.23	15.79 $\pm$ 11.20
MDA (nmole/L)	6.39 $\pm$ 1.80*	4.90 $\pm$ 1.97
AOPP ( $\mu$ mole/L)	251.89 $\pm$ 69.58*	181.43 $\pm$ 43.11
GSH (nmole/mL)	310.05 $\pm$ 102.51	331.03 $\pm$ 129.45
Vitamin C ( $\mu$ mole/L)	2.54 $\pm$ 1.50	3.14 $\pm$ 1.54

\* $p < 0.05$ , as compared with the control group.

including oxidative damage<sup>25</sup> and ocular perfusion abnormalities.<sup>26</sup>

Oxidative stress may cause injury to the retinal pigment epithelium (RPE), the Bruch's membrane, and the choroid, which are layers in the eye involved in the pathophysiology of AMD.<sup>27-31</sup>

Liang et al.<sup>32</sup> demonstrated that human RPE cells exposed to oxidative stress or rod outer segments exhibited damage primarily to mitochondrial (mt) DNA and that damaged mtDNA was not efficiently repaired.

The retina is particularly susceptible to oxidative stress due to its high concentration of oxygen, its high proportion of polyunsaturated fatty acids, and its exposure to visible light.<sup>33</sup> Prior reports have suggested that the retina is susceptible to lipid peroxidation<sup>34,35</sup> and that this susceptibility also increases with aging in the macular region.<sup>35</sup>

Previous studies demonstrated that the plasma MDA levels were higher in an AMD patient group than in a control group.<sup>36,37</sup> In the present study, we found that the serum MDA levels were significantly higher in the AMD patient group than in the control group, in agreement with a previous study.<sup>38</sup>

Protein oxidation is also a useful marker for the evaluation of oxidative stress in vivo. Many different types of protein oxidative modifications can be induced by free radicals. A prior study demonstrated that the levels of protein carbonyl groups in the serum were higher in the AMD patient group than in the control group.<sup>38</sup>

AOPP measurements reflect the generation of free radicals and the degree of protein oxidation.<sup>16,39</sup> In the present study, we found that the levels of AOPP in the serum were significantly higher in the AMD patient group than in the control group. Therefore, this study demonstrated that protein oxidation, a useful oxidative stress marker, is upregulated in AMD, suggesting increased oxidative stress.

Antioxidant enzymes are a primary defense system that protects biological macromolecules from oxidative damage. SOD, CAT and GPx are antioxidant enzymes that form part of the complex system that protects the retina from oxidative damage, and all three of these enzymes are found in the photoreceptors and the RPE.<sup>40</sup> SOD is a key antioxidant enzyme involved in the metabolism of oxygen free radicals.<sup>41</sup> A previous report suggested that the activities of SOD in plasma and erythrocytes were lower in the AMD patient group than in the control group.<sup>37</sup> In the present study, we found that the activities of SOD in the serum were significantly lower in the AMD patient group than in the control group.

GSH is a major non-enzymatic antioxidant that is effective in protecting cells against to reactive oxygen products and toxins. A high concentration of GSH is present in most

living cells, and GSH is involved in the responses to various stresses. The redox condition of cells is associated with their degree of protection by GSH in the reduced state.<sup>42</sup>

A prior study demonstrated that the plasma GSH levels were lower in an AMD patient group than in a control group.<sup>43</sup> In the present study, however, we determined that the serum GSH levels were unchanged between our groups.

GPx uses GSH as an electron donor to reduce organic hydroperoxides. GPx is found in the human retina and is dependent on selenium as a cofactor. Extracellular GPx is found in the retina, ciliary epithelium, and aqueous humor, and GPx may act as an extracellular antioxidant.<sup>33</sup>

A previous study demonstrated that the activities of GPx in the plasma and erythrocytes were lower in an AMD patient group than in a control group.<sup>37,43</sup> In the present study, the GPx activities in the serum were unchanged between groups.

Vitamin C is the second major water-soluble antioxidant. An important function of vitamin C is protecting the lens by means of direct absorption of ultraviolet radiation by aqueous vitamin C.<sup>44</sup>

A prior study demonstrated that the low levels of vitamin C in the plasma were associated with an increased risk of AMD but that high levels were not protective.<sup>44</sup> In the present study, the serum vitamin C levels were unchanged between groups.

Much of the research into the relationship between oxidative stress and AMD has focused on the antioxidant status of subjects with and without the disease, and the limitations of these studies are worth noting.

Deficiencies in antioxidant enzymes may be related to the development of disease. Because the retina suffers oxidative damage, antioxidant nutrients are thought to be protective of the retina through their function as antioxidants.

The results of the present study suggest that reductions in the antioxidant defense system and increased oxidative stress may play a role in the pathogenesis of AMD.

In conclusion, increased oxidative stress, which causes oxidative damage to lipids and proteins and decreases antioxidant capacity, may lead to irreversible damage in the form of AMD. Further studies that analyze samples obtained both from the serum and the aqueous humor are required to confirm our findings.

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## REFERENCES

- Seddon JM, George S, Rosner B, Rifai N. Progression of age-related macular degeneration: prospective assessment of C-reactive protein, interleukin 6 and, and other cardiovascular biomarkers. *Arch Ophthalmol*. 2005;123:774-82, doi: 10.1001/archophth.123.6.774.
- Drobek-Slowik M, Karczewicz D, Safranow K. The potential role of oxidative stress in the pathogenesis of the age-related macular degeneration (AMD). *Postepy Hig Med Dosw*. 2007;61:28-37.
- Ding X, Patel M, Chan CC. Molecular pathology of age-related macular degeneration. *Prog Retin Eye Res*. 2009;28:1-18, doi: 10.1016/j.preteyeres.2008.10.001.
- Tsai YY, Lin JM, Wan L, Lin HJ, Tsai Y, Lee CC, et al. Interleukin gene polymorphisms in age-related macular degeneration. *Invest Ophthalmol Vis Sci*. 2008;49:693-8, doi: 10.1167/iovs.07-0125.
- Smith W, Assink J, Klein R, Mitchell P, Klacar CC, Klein BE, et al. Risk factors for age-related macular degeneration: pooled findings from three continents. *Ophthalmology*. 2001;108:697-704, doi: 10.1016/S0161-6420(00)00580-7.

6. Seddon JM, Cote J, Rosner B. Progression of age-related macular degeneration: association with dietary fat, transunsaturated fat, nuts, and fish intake. *Arch Ophthalmol.* 2003;121:1728-37, doi: 10.1001/archophth.121.12.1728.
7. Seddon JM, Rosner B, Sperduto RD, Yannuzzi L, Haller JA, Blair NP, et al. Dietary fat and risk for advanced age-related macular degeneration. *Arch Ophthalmol.* 2001;119:1191-9.
8. Cho E, Hung S, Willett W, Spiegelman D, Rimm EB, Seddon JM, et al. Prospective study of dietary fat and the risk of age-related macular degeneration. *Am J Clin Nutr.* 2001;73:209-8.
9. Seddon JM, Cote J, Davis N, Rosner B. Progression of age-related macular degeneration: association with body mass index, waist circumference, and waist-hip ratio. *Arch Ophthalmol.* 2003;21:785-92, doi: 10.1001/archophth.121.6.785.
10. Seddon JM, Ajani UA, Sperduto RD, Dietary carotenoids, vitamins A, C, and E, and advanced age-related macular degeneration. Eye disease Case-Control Study Group. *JAMA.* 1994;272:1413-20, doi: 10.1001/jama.272.18.1413.
11. Ohia SE, Opere CA, Leday AM. Pharmacological consequences of oxidative stress in ocular tissues. *Mutat Res.* 2005;579:22-36.
12. Halliwell B, Gutteridge J. Free radicals in biology and medicine. 3rd ed. New York:Oxford University Press Inc, 1989:1-35.
13. Irmak MK, Fadillioglu E, Sogut S, Erdogan H, Gulec M, Ozer M, et al. Effects of caffeic acid phenethyl ester and alpha-tocopherol on reperfusion injury in rat brain. *Cell Biochem Funct.* 2003;21:283-9, doi: 10.1002/cbf.1024.
14. Touyz RM. Reactive oxygen species and angiotensin II signaling in vascular cells-implications in cardiovascular disease. *Braz J Med Biol Res.* 2004;37:1263-73, doi: 10.1590/S0100-879X2004000800018.
15. Olivares-Corichi IM, Ceballos G, Ortega-Camarillo C, Guzman-Grenfell AM, Hicks JJ. Reactive oxygen species (ROS) induce chemical and structural changes on human insulin in vitro, including alterations in its immunoreactivity. *Front Biosci.* 2005;10:834-43.
16. Witko-Sarsat V, Friedlander M, Capeillere-Blandin C, Nguyen-Khoa T, Nguyen AT, Zingraff J, et al. Advanced oxidation protein products as a novel marker of oxidative stress in uremia. *Kidney Int.* 1996;49:1304-13, doi: 10.1038/ki.1996.186.
17. Harman D. Free radical theory of aging: An update: Increasing the functional life span. *Ann N Y Acad Sci.* 2006;1067:10-21, doi: 10.1196/annals.1354.003.
18. Fairfield KM, Fletcher RH. Vitamins for chronic disease prevention in adults: scientific review. *JAMA.* 2002;287:3116-26, doi: 10.1001/jama.287.23.3116.
19. Pryor WA, Godber SS. Noninvasive measures of oxidative stress status in humans. *Free Radic Biol Med.* 1991;10:177-84, doi: 10.1016/0891-5849(91)90073-C.
20. Yoshioka T, Kawada K, Shimada T, Mori M. Lipid peroxidation in maternal and cord blood and protective mechanism against activated-oxygen toxicity in the blood. *Am J Obstet Gynecol.* 1979;135:372-6.
21. Kurtel H, Granger DN, Tso P, Grisham MB. Vulnerability of intestinal interstitial fluid to oxidant stress. *Am J Physiol.* 1992;263:G573-78.
22. Sun Y, Oberley LW, Li Y. A simple method for clinical assay of superoxide dismutase. *Clin Chem.* 1988;34:497-500.
23. Paglia DE, Valentine WN. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. *J Lab Clin Med.* 1967;70:158-69.
24. Mapson LW. The estimation of dehydro-L-ascorbic acid when present in low concentration in tissues, by the Roe and Kuether procedure. *Ann N Y Acad Sci.* 1961; 92:284-5, doi: 10.1111/j.1749-6632.1961.tb46127.x.
25. Cai J, Nelson KC, Wu M, Sternberg P Jr, Jones DP. Oxidative damage and protection of the RPE. *Prog Retin Eye Res.* 2000;19:205-21, doi: 10.1016/S1350-9462(99)00009-9.
26. Harris A, Chung HS, Ciulla TA, Kagemann L. Progress in measurement of ocular blood flow and relevance to our understanding of glaucoma and age-related macular degeneration. *Prog Retin Eye Res.* 1999;18:669-87, doi: 10.1016/S1350-9462(98)00037-8.
27. Johnson LV, Ozaki S, Staples MK, Erickson PA, Anderson DH. A potential role for immune complex pathogenesis in drusen formation. *Exp Eye Res.* 2000;70:441-9, doi: 10.1006/exer.1999.0798.
28. Mullins RF, Russell SR, Anderson DH, Hageman GS. Drusen associated with aging and age-related macular degeneration contain proteins common to extracellular deposits associated with atherosclerosis, elastosis, amyloidosis, and dense deposit disease. *FASEB J.* 2000;14:835-46.
29. Johnson LV, Leitner WP, Staples MK, Anderson DH. Complement activation and inflammatory processes in drusen formation and age related macular degeneration. *Exp Eye Res.* 2001;73:887-96, doi: 10.1006/exer.2001.1094.
30. Hageman GS, Luthert PJ, Victor Chong NH, Johnson LV, Anderson DH, Mullins RF. An integrated hypothesis that considers drusen as biomarkers of immune-mediated processes at the RPE-Bruch's membrane interface in aging and age-related macular degeneration. *Prog Retin Eye Res.* 2001;20:705-32, doi: 10.1016/S1350-9462(01)00010-6.
31. Anderson DH, Mullins RF, Hageman GS, Johnson LV. A role of for local inflammation in the formation of drusen in the aging eye. *Am J Ophthalmol.* 2002;134:411-31, doi: 10.1016/S0002-9394(02)01624-0.
32. Liang FQ, Godley BF. Oxidative stress-induced mitochondrial DNA damage in human retinal pigment epithelial cells: a possible mechanism RPE aging and age-related macular degeneration. *Exp Eye Res.* 2003;76:397-403, doi: 10.1016/S0014-4835(03)00023-X.
33. Beatty S, Koh H, Phil M, Henson D, Boulton M. The role of oxidative stress in the pathogenesis of age-related macular degeneration. *Surv Ophthalmol.* 2000;45:115-34, doi: 10.1016/S0039-6257(00)00140-5.
34. Ito T, Nakano M, Yamamoto Y, Hiramitsu T, Mizuno Y. Hemoglobin-induced lipid peroxidation in the retina: a possible mechanism for macular degeneration. *Arch Biochem Biophys.* 1995;316:864-72, doi: 10.1006/abbi.1995.1116.
35. De La Paz MA, Anderson RE. Lipid peroxidation in rod outer segments. Role of hydroxyl radical and lipid hydroperoxides. *Invest Ophthalmol Vis Sci.* 1992;33:2091-6.
36. Totan Y, Çağcı O, Borazan M, Uz E, Söğüt S, Akyol O. Plasma malondialdehyde and nitric oxide levels in age related macular degeneration. *Br J Ophthalmol.* 2001;85:1426-8, doi: 10.1136/bjo.85.12.1426.
37. Evereklioglu C, Er H, Doganay S, Cekmen M, Turkoz Y, Otlu B, Ozerol E. Nitric oxide and lipid peroxidation are increased and associated with decreased antioxidant enzyme activities in patients with age-related macular degeneration. *Doc Ophthalmol.* 2003;106:129-36, doi: 10.1023/A:1022512402811.
38. Totan Y, Yağcı R, Bardak Y, Ozyurt H, Kendir F, Yılmaz G, Sahin S, Sahin Tiğ U. Oxidative macromolecular damage in age-related macular degeneration. *Curr Eye Res.* 2009;34:1089-93, doi: 10.3109/02713680903353772.
39. Alderman CJ, Shah S, Foreman JC, Chain BM, Katz DR. The role of advanced oxidation protein products in regulation of dendritic cell function. *Free Radic Bio Med.* 2002;32:377-85, doi: 10.1016/S0891-5849(01)00735-3.
40. Beatty S, Koh H, Phil M, Henson D, Boulton M. The role of oxidative stress in the pathogenesis of age-related macular degeneration. *Surv Ophthalmol.* 2000;45:115-34, doi: 10.1016/S0039-6257(00)00140-5.
41. Enghild JJ, Thøgersen IB, Oury TD, Valnickova Z, Hojrup P, Crapo JD. The heparin-binding domain of extracellular superoxide dismutase is proteolytically processed intracellularly during biosynthesis. *J Biol Chem.* 1999;274:14818-22, doi: 10.1074/jbc.274.21.14818.
42. Penninckx M. A short review on the role of glutathione in the response of yeasts to nutritional, environmental, and oxidative stresses. *Enzyme Microb Technol.* 2000;26:737-42, doi: 10.1016/S0141-0229(00)00165-4.
43. Samiec PS, Drews-Botsch C, Flagg EW, Kurtz JC, Sternberg P Jr, Reed RL, Johns DP. Glutathione in human plasma decline: in association with aging, age-related macular degeneration, and diabetes. *Free Radic Biol Med.* 1998;24:699-704, doi: 10.1016/S0891-5849(97)00286-4.
44. Antioxidant status and neovascular age-related macular degeneration. Eye Disease Case-Control Study Group. *Arch Ophthalmol.* 1993;111:104-9 [Erratum in: *Arch Ophthalmol* 1993 Sep;111(9):1228, 1993 Oct;111(10):1366. *Arch Ophthalmol* 1993 Nov;111(11):1499.