

Effects of the administration of a catalase inhibitor into the fourth cerebral ventricle on cardiovascular responses in spontaneously hypertensive rats exposed to sidestream cigarette smoke

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OBJECTIVE: Previous studies have demonstrated a relationship between brain oxidative stress and cardiovascular regulation. We evaluated the effects of central catalase inhibition on cardiovascular responses in spontaneously hypertensive rats exposed to sidestream cigarette smoke.

METHODS: Male Wistar Kyoto (WKY) rats and spontaneously hypertensive rats (SH) (16 weeks old) were implanted with a stainless steel guide cannula leading into the fourth cerebral ventricle (4th V). The femoral artery and vein were cannulated for arterial pressure and heart rate measurement and drug infusion, respectively. The rats were exposed to sidestream cigarette smoke for 180 minutes/day, 5 days/week for 3 weeks (CO: 100-300 ppm). The baroreflex was tested using a pressor dose of phenylephrine (8 μ g/kg, bolus) and a depressor dose of sodium nitroprusside (50 μ g/kg, bolus). Cardiovascular responses were evaluated before and 5, 15, 30 and 60 minutes after injection of a catalase inhibitor (3-amino-1,2,4-triazole, 0.001 g/100 μ L) into the 4th V.

RESULTS: Vehicle administration into the 4th V did not affect the cardiovascular response, whereas administration of the central catalase inhibitor increased the basal HR and attenuated the bradycardic peak (p<0.05) to a greater extent in WKY rats exposed to sidestream cigarette smoke than in WKY rats exposed to fresh air. However, in spontaneously hypertensive rats, the effect of the catalase inhibitor treatment was stronger in the fresh air condition (p<0.05).

CONCLUSION: Administration of a catalase inhibitor into the 4th V combined with exposure to sidestream cigarette smoke has a stronger effect in WKY rats than in SH rats.

KEYWORDS: Oxidative Stress; Catalase; Medulla Oblongata; Tobacco; Air Pollutants.

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■ INTRODUCTION

The effects of cigarette smoke on the cardiovascular system underlie the adverse effects of smoking on cardiovascular (1,2) and brain health, (3-5) in addition to detrimental effects in different systems (6-8). Cigarette

No potential conflict of interest was reported. **DOI:** 10.6061/clinics/2013(06)21 smoke is classified into 2 categories: the mainstream smoke usually inhaled by active smokers and the sidestream smoke emitted from a cigarette and inhaled by so-called "passive smokers." Sidestream cigarette smoke (SSCS) is known to contain greater amounts of various oxidants and other harmful compounds than mainstream smoke (9). Thus, passive smokers are exposed to nearly the same chemicals in cigarette smoke as active smokers, and passive smoking has been found to increase the risk of cardiac or other related diseases in nonsmokers (10).

Increased production of reactive oxygen species (ROS) by cigarette smoke occurs as a direct effect of the radicals present in smoke (11-13). For instance, it was previously shown that cigarette exposure over 24 consecutive days

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increased mRNA levels of catalase in the heart by two fold relative to only 1 day of exposure (14). ROS, such as superoxide anions (O_2) and hydrogen peroxide (H_2O_2) , were once thought only to be harmful byproducts of oxidative metabolism but are now recognized as critical second messengers in a wide range of cellular processes (15). ROS are produced by the incomplete reduction of oxygen to O_2^- , which is spontaneously or enzymatically dismutated to H_2O_2 by superoxide dismutase (SOD). H_2O_2 is transformed to H_2O and O_2 by the activity of catalase (16). Previous investigations have associated brain ROS with increased sympathetic activity (17,18), and systemic ROS have also been associated with an impaired baroreflex (19). Drugs injected into the fourth cerebral ventricle (4th V) may easily reach structures surrounding the ventricular system such as the area postrema (20). Previous studies have indicated that ROS in the 4^{th} V influence cardiovascular responses (21). Moreover, administration of a catalase inhibitor into the 4th V has also been demonstrated to influence cardiovascular responses in normotensive rats (22,23). Luchese et al. (24) indicated that acute cigarette smoke exposure increases oxidative stress in the brain by increasing the activity of reactive species and reducing the activity of superoxide dismutase and catalase. Bartoli et al. (25) suggested that increased baroreceptor reflex sensitivity may compensate for particle-induced alterations in blood pressure in dogs. In addition, our group previously demonstrated that SSCS affects the cardiovascular responses induced by central catalase inhibition in normotensive rats (26). However, to the best of our knowledge, no previous study has evaluated the effects of SSCS and central catalase inhibition in spontaneously hypertensive (SH) rats. Thus, to study the detailed mechanism of catalase inhibition, we investigated the effects of administration of a catalase inhibitor into the 4th V on cardiovascular responses in SH rats exposed to SSCS.

METHODS

Animals

We used male Wistar Kyoto (WKY) rats and SH rats (16 weeks old) that were kept in the Animal Care Unit of our university. The rats were housed individually in plastic cages under standard laboratory conditions. The animals were divided into 4 groups: WKY rats exposed to fresh air (N = 7), WKY rats exposed to SSCS (N = 7), SH rats exposed to fresh air (N = 7) and SH rats exposed to SSCS (N = 7). The rats were kept under a 12-h light/dark cycle (lights on at 07:00 h) and had free access to food and water. The institution's Animal Ethics Committee authorized the housing conditions and experimental procedures (number 0255/10). Efforts were made to minimize the number of animals used.

SSCS exposure

The rats were placed in a transparent chamber with a volume of approximately $95\times80\times65$ cm³, with 4 rats per chamber. The rats were maintained at 23 ± 1 °C and 50-60% relative humidity. The carbon monoxide (CO) concentration of the smoke in the chamber was maintained between 100-300 ppm. Cigarettes were placed inside the chamber in a small box, which prevented the rats from touching the cigarettes. SSCS was produced by burning the cigarettes inside the chamber without filtering. When the CO

concentration reached 100 ppm, the 180-minute interval began. The cigarettes were replaced by new cigarettes to maintain a CO concentration between 100-300 ppm. The rats were exposed to SSCS for 180 minutes on 5 days per week, and the total duration of the experiment was 3 weeks. All of the exposures per performed in the morning, between 8 a.m. and 12 p.m. The cigarettes used were of a commercial brand with the following composition: 1.1 mg of nicotine, 14 mg of tar and 15 mg of carbon monoxide. The control animals were maintained in the same manner and same conditions as the SSCS group but exposed to fresh air (27-29).

Surgical preparation

Five days before the experiment (one day after the last SSCS exposure), the rats were anesthetized with ketamine (50 mg/kg i.p.) and xylazine (50 mg/kg i.m.). After anesthesia was applied to the scalp using 2% lidocaine, the skull was exposed, and stainless steel guide cannulas (26 G) were implanted into the 4th V 1 mm above the site of injection using a stereotaxic apparatus (Stoelting, USA). The stereotactic coordinates for implantation of the cannula into the 4th V were as follows: AP = -13 mm from the bregma, L = 0 mm from the medial suture and V = -6 mm from the skull. The cannulas were fixed to the skull using dental cement and 1 metal screw (30).

One day before the experiment, the rats were anesthetized with ketamine (50 mg/kg i.p.) and xylazine (50 mg/kg i.m.), and a catheter was inserted into the abdominal aorta through the femoral artery for monitoring of blood pressure and heart rate. The catheters consisted of 4-cm segments of PE-10 polyethylene tubing (Clay Adams, USA) that were heat-bound to a 13-cm segment of PE-50 polyethylene tubing. The catheters were tunneled under the skin and exteriorized at the animal's dorsum (31,32). After each surgery, the animals received a single dose of an antibiotic (ampicillin, 100 mg/kg) and a single dose of the analgesic ketorolac (0.6 mg/kg).

Recording of arterial pressure and heart rate

After surgery, the animals were kept in the individual cages used in their transport to the experiment room. The animals were allowed 60 minutes to adapt to the conditions of the experimental room, such as sound and illumination, before the recording of the blood pressure and heart rate was initiated. The experiment room was acoustically isolated and had a constant background noise produced by an air exhauster. At least another 30-minute period was allowed before the experiments were initiated. The pulsatile arterial pressure of the freely moving animals was recorded using an HP-7754A preamplifier at a sampling frequency of 1,000 Hz (Hewlett Packard, USA) and an acquisition board (MP100A, Biopac Systems Inc., USA) connected to a computer. The mean arterial pressure (MAP) and heart rate (HR) were derived from the pulsatile arterial pressure recordings and processed on-line (33).

Baroreflex evaluation

The baroreflex was activated by intravenous phenylephrine (PHE, 8 μ g/kg, bolus) or sodium nitroprusside (SNP, 50 μ g/kg, bolus). The baroreflex gain was calculated as the derivation of HR in the function of the MAP variation (Δ HR/ Δ MAP, maximum changes in MAP and HR). The sympathetic baroreflex gain (SBG) was defined as the Δ HR/ Δ MAP ratio in response to i.v. SNP, and parasympathetic



Table 1 - Baseline mean arterial pressure (MAP) and heart rate (HR), bradycardic and tachycardic peak and sympathetic (SBG) and parasympathetic baroreflex gain (PBG) in Wistar Kyoto rats exposed to fresh air and treated with ATZ administered into the 4th V. N=7. Mean \pm SEM. *p<0.05 for comparison with 0'.

Variable	0′	5′	15′	30′	60 ′
MAP (mmHg)	112 ± 13	116 ± 15	124 ± 16	127 ± 14	113±14
HR (bpm)	315 ± 29	308 ± 24	318±22	370±22*	323 ± 25
Bradycardic Peak (bpm)	230 ± 28	260 ± 20	263 ± 29	318±22*	252 ± 21
Tachycardic Peak (bpm)	417 ± 25	419±23	454 ± 28	469 ± 31	425 ± 36
HR range (bpm)	187 ± 19	166 ± 11	191 ± 16	151 ± 18	173 ± 17
PBG (bpm x mmHg ⁻¹)	-1.73±0.24	-1.54 ± 0.03	-1.56 ± 0.14	-1.9±0.6	-1.35 ± 0.04
SBG (bpm x mmHg ⁻¹)	-2.25 ± 0.23	-2.24 ± 0.43	$\textbf{-1.93} \pm \textbf{0.16}$	-2.64 ± 0.45	$\textbf{-2.5} \pm \textbf{0.26}$

baroreflex gain (PBG) was defined as the Δ HR/ Δ MAP ratio in response to i.v. PHE. We also analyzed the bradycardic and tachycardic peak and HR range (i.e., the difference between the bradycardic and tachycardic peaks) (34).

Injections into the 4th V

Injections into the 4th V were performed using 10-µl Hamilton syringes connected by polyethylene tubing (PE-10) to an injector needle. The injector, when completely inserted, protruded 2 mm beyond the tip of the guide cannula. The injections into the 4th V consisted of a volume of 1.0 µl for approximately 5-10 s (35).

Protocol

Baroreflex and cardiovascular responses were evaluated before (control) and 5, 15, 30 and 60 minutes after injection of the catalase inhibitor (3-amino-1,2,4-triazole, ATZ, 0.001 g/100 μ L) or vehicle (0.9% NaCl) into the 4th V of conscious rats (23).

Histology

At the end of the experiments, the animals were anesthetized using urethane (1.25 g/kg, i.p.), and 200 ml of 1% Evan's blue dye was injected in the 4th V as a marker of the injection site. The chest was surgically opened, the descending aorta occluded, the right atrium severed and the brain perfused with 10% formalin through the left ventricle. The brains were post-fixed for 24 h at 4°C, and 40-µm sections were cut in a cryostat (model CM 1900, Leica, Germany). The brain sections were stained with 1% neutral red. The actual placement of the injection needle was verified using serial sections (36).

Statistical analysis

The results were reported as the mean \pm standard error of the mean (SEM). Analyses of variance (ANOVA) for repeated measures followed by the Tukey post-test were

applied to compare all variables (basal MAP and HR, bradycardic and tachycardic peak, HR range, SBG, PBG, PHE-induced increase and SNP-induced decrease in MAP and bradycardic and tachycardic reflex). We compared the variables at baseline (control) with the 5-, 15-, 30- and 60-minute time points after injection of ATZ into the 4th V in the same rat. We applied Student's t-test to compare the cardiovascular responses between WKY groups and SH groups exposed to SSCS. Differences were considered significant when the probability of a Type I error was less than 5% (p<0.05).

RESULTS

Effect of vehicle injection into the 4th V

Injection of vehicle (0.9% NaCl) into the 4th V did not affect the basal MAP and HR, tachycardic and bradycardic peak, HR range, SBG and PBG in WKY or SH rats exposed to fresh air.

Effect of ATZ injection into the 4th V

Injection of ATZ into the 4th V did not affect the basal MAP; however, the basal HR was increased at 30 minutes (p<0.05) after ATZ administration in WKY rats exposed to fresh air. Furthermore, the bradycardic peak was also attenuated at 30 minutes. In contrast, we did not observe significant changes in the bradycardic and tachycardic peak, HR range, PBG and SBG after central catalase inhibition in WKY rats exposed to fresh air (Table 1).

As shown in Table 4, injection of ATZ into the 4th V produced a strong response in SH rats exposed to fresh air. The basal HR increased was at 60 minutes and the bradycardic peak was attenuated at 30 minutes after ATZ administration. PBG was increased at 60 minutes after ATZ treatment, whereas SBG was decreased at 30 and 60 minutes (Table 2).

Among the groups exposed to SSCS, we observed stronger effects for catalase inhibition in WKY rats. Central ATZ

Table 2 - Baseline mean arterial pressure (MAP) and heart rate (HR), bradycardic and tachycardic peak and sympathetic (SBG) and parasympathetic baroreflex gain (PBG) in SH rats exposed to fresh air and treated with ATZ administered into the 4th V. N=7. Mean \pm SEM. *p<0.05 for comparison with 0'.

Variable	0′	5′	15′	30′	60 ′
MAP (mmHg)	176 ± 16	192 ± 14	180 ± 11	175 ± 15	175 ± 16
HR (bpm)	322 ± 22	407±22*	340±21*	343±27*	341±29*
Bradycardic Peak (bpm)	273 <u>+</u> 19	$352\pm19*$	305±21*	308±28*	309 ± 28
Tachycardic Peak (bpm)	442 ± 37	463 ± 31	445 ± 31	455 ± 34	445 ± 32
HR range (bpm)	154 ± 17	111 ± 12	142 ± 11	147 ± 11	148 ± 12
PBG (bpm x mmHg⁻¹)	-0.35 ± 0.03	-1.2±0.14*	-0.8±0.05*	-0.67±0.13*	-0.7±0.12*
SBG (bpm x mmHg ⁻¹)	-2±0.18	-1.96 ± 0.01	-2.16 ± 0.08	-1.39±0.13*	-1.36±0.1*



Table 3 - Baseline mean arterial pressure (MAP) and heart rate (HR), bradycardic and tachycardic peak and sympathetic (SBG) and parasympathetic baroreflex gain (PBG) in Wistar Kyoto rats exposed to SSCS and treated with ATZ administered into the 4th V. N=7. Mean \pm SEM. *p<0.05 for comparison with 0'.

Variable	0′	5′	15′	30′	60′
MAP (mmHg)	106 ± 13	118 ± 14	111 ± 15	107 ± 12	107±13
HR (bpm)	344 ± 16	412 \pm 19*	440 ± 13 *	$429\pm17*$	397 ± 13
Bradycardic Peak (bpm)	279 ± 14	323 ± 19	$364 \pm 26^*$	$\textbf{380} \pm \textbf{18}^{\star}$	318 ± 10
Tachycardic Peak (bpm)	489 ± 11	506 ± 18	525 ± 19	527 ± 14	502 ± 12
HR range (bpm)	203 ± 12	183 ± 13	161±22	169 ± 25	183 ± 11
PBG (bpm x mmHg ⁻¹)	-1.2 ± 0.34	-1.19 ± 0.11	-1.08 ± 0.22	-0.85±0.13	-1.11 ± 0.19
SBG (bpm x mmHg ⁻¹)	-1.48 ± 0.2	-1.26 ± 0.14	-1.39 ± 0.26	-1.13 ± 0.1	-1.84 ± 0.21

administration increased the basal HR at 5, 15 and 30 minutes and reduced the bradycardic peak at 15 and 30 minutes (Table 3).

In the SH rats exposed to SSCS, we observed that ATZ treatment increased the basal HR during the first 30 minutes, reduced the bradycardic peak at 5 and 15 minutes and reduced the tachycardic peak at 60 minutes (Table 4).

Comparison of cardiovascular responses between WKY rats and SH rats exposed to SSCS and treated with ATZ

In Table 5, we present a comparison of the cardiovascular responses induced by central ATZ administration between the WKY and SH rats exposed to SSCS at each time point. It should be noted that almost all variables were different between the groups before ATZ injection. However, the differences were not significant after the injection of ATZ.

DISCUSSION

This study was undertaken to evaluate the effects of central catalase inhibitor administration on cardiovascular responses in WKY and SH rats exposed to SSCS. We observed that administration of a catalase inhibitor into the 4th V produced a strong effect on cardiovascular responses in WKY rats exposed to SSCS but not in SH rats exposed to SSCS. The lack of any change in the vehicle-treated group is consistent with these findings.

Based on our data, injection of ATZ into the 4th V did not affect the sympathetic and parasympathetic baroreflex gain in WKY rats. A previous study published by our group (22) reported that central ATZ administration into the 4th V did not affect the same components of the baroreflex in Wistar rats. We also reported that central catalase inhibition increased the parasympathetic baroreflex gain and reduced the sympathetic baroreflex gain in SH rats. The present study provides additional information because a different response was observed in the SH group. We therefore consider that central catalase inhibition affects the parasympathetic and sympathetic baroreflex gain to a greater extent in SH rats than in WKY rats. This hypothesis is supported by our previous study that investigated the cardiopulmonary reflex responses to catalase inhibitor administration into the 4th V in SH and WKY rats (37).

In the present study, administration of a catalase inhibitor into the 4th V strongly attenuated the bradycardic peak in WKY rats exposed to SSCS, whereas this response was not enhanced in SH rats exposed to SSCS. The parasympathetic activity of the baroreflex response is represented by the bradycardic peak, whereas the sympathetic activity during the baroreflex is represented by the tachycardic peak, and the difference between both peaks corresponds to the HR range (38). Our findings indicate that SSCS exposure has a stronger effect on parasympathetic activity in WKY rats than in SH rats. A recent study demonstrated that vagal modulation of the heart is blunted in heavy smokers, particularly during parasympathetic modulation (39). Considering that SH rats present increased levels of ROS in the brain relative to normotensive rats (40,41), we hypothesize that 3 weeks of exposure to SSCS was not sufficient to increase ROS production in the 4th V and affect cardiovascular responses.

We observed that acute administration of the catalase inhibitor into the 4th V had a strong effect on the parasympathetic regulation of the cardiovascular system in WKY rats exposed to SSCS because it strongly increased the basal HR. Conversely, in SH rats, the central catalase inhibitor treatment strongly affected cardiovascular responses in the group exposed to fresh air. In particular, administration of the catalase inhibitor into the 4th V increased the parasympathetic baroreflex gain and reduced the sympathetic baroreflex gain in the group exposed to fresh air, whereas it did not affect these components in SH rats exposed to SSCS. As mentioned above, we believe that 3 weeks of exposure to SSCS is insufficient to induce changes in ROS in the 4th V of the SH rats, possibly because the SH

Table 4 - Baseline mean arterial pressure (MAP) and heart rate (HR), bradycardic and tachycardic peak and sympathetic (SBG) and parasympathetic baroreflex gain (PBG) in SH rats exposed to SSCS and treated with ATZ administration into the 4th V. N=7. Mean \pm SEM. *p<0.05 for comparison with 0'.

Variable	0′	5′	15′	30′	60′
MAP (mmHg)	168 ± 13	175 ± 15	176 ± 13	173 ± 12	170 ± 11
HR (bpm)	317 ± 21	429 ± 22 *	398±24*	380±24*	344±28
Bradycardic Peak (bpm)	213 ± 17	$307\pm32^{\star}$	$\textbf{298} \pm \textbf{14}^{\textbf{*}}$	270 ± 17	26 ± 18
Tachycardic Peak (bpm)	504 ± 28	522 ± 28	519±27	519 ± 29	475±25*
HR range (bpm)	272 ± 11	216 ± 29	222 ± 11	249 ± 15	213 ± 11
PBG (bpm x mmHg ⁻¹)	-1.9±0.23	-2.87 ± 0.5	-2.77 ± 0.9	-2.4±0.52	-1.65±0.19
SBG (bpm x mmHg ⁻¹)	-3.16 ± 0.3	-2.46 ± 0.45	-3.03 ± 0.59	-2.73 ± 0.31	-3.31 ± 0.43

Table 5 - p-value for the intergroup comparison between the WKY and SH groups exposed to SSCS at each time point.

Variable	0′	5′	15′	30′	60 ′
MAP (mmHg)	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001
HR (bpm)	0.049	0.29	0.067	0.02	0.07
Bradycardic Peak (bpm)	0.005	0.38	0.15	0.04	0.1
Tachycardic Peak (bpm)	0.16	0.46	0.44	0.43	0.34
HR range (bpm)	0.0005	0.27	0.09	0.049	0.18
PBG (bpm x mmHg⁻¹)	0.04	0.14	0.33	0.43	0.48
SBG (bpm x mmHg⁻¹)	0.0001	0.21	0.34	0.43	0.48

rats present higher basal levels of ROS in the brain (40,41). Thus, a stronger stimulus may be necessary to induce changes in ROS in the brain of SH rats.

A recent study published by our group demonstrated that central catalase inhibition increased the basal HR and attenuated the bradycardic peak in Wistar rats exposed to SSCS (26). In our current investigation, WKY rats exposed to SSCS presented similar responses, which thus supports the results of our previous study (26). In this case, we aimed to determine whether SH rats exposed to SSCS would present increased responses. However, SH rats exposed to fresh air presented greater responses than SH rats exposed to SSCS.

In this context, we also found in this study that before injection of the catalase inhibitor into the 4th V, a difference existed between the WKY and SH groups exposed to SSCS regarding basal MAP and HR, bradycardic peak, HR range and sympathetic and parasympathetic baroreflex gain. This finding is in agreement with our previous studies (34). However, after central administration of the catalase inhibitor, these variables were no longer significantly different between the groups. Therefore administration of the catalase inhibitor into the 4th V combined with exposure to SSCS likely had a greater acute effect in WKY rats than SH rats.

Based on our findings, we suggest that SSCS exposure affects the oxidative stress balance in the 4th V in WKY rats. The morphology of the nose can be affected by substances able to achieve passage into the brain, and some substances may enter the brain through cranial nerves (42). We believe that the olfactory vector hypothesis for neurological diseases may explain our data (43). Prior studies have indicated that substances injected into the 4th V preferably reach the parasympathetic system. Areas such as the dorsal motor nucleus of the vagus and nucleus ambiguus receive glutamatergic projections from the nucleus of the solitary tract (20).

We used SSCS exposure in this investigation because it has been suggested that SSCS is more toxic than mainstream cigarette smoke (44). Indeed, the Philip Morris Company secretly performed in vivo toxicological tests of the effects of sidestream smoke (44) and observed that condensates and sidestream particulate matter present more mutagenic effects than the mainstream particulate matter. Moreover, SSCS contains approximately 200-fold more ammonia than mainstream smoke. SSCS presents a higher density of toxic substances than mainstream smoke because of its lower temperature of combustion and absence of filtering (45). For example, SSCS has five-fold more acrolein than mainstream smoke (46). On inhalation, fresh sidestream smoke is two- to six-fold more toxic per gram than mainstream smoke, depending on the end point (47). Taken together, our results and the findings of these other studies are relevant for directing and implementing public health policies.

In our research, the baroreflex was investigated in conscious, unanesthetized rats because anesthesia affects cardiovascular responses (48). We believe that our investigation provides reliable information regarding the effects of the administration of the catalase inhibitor ATZ into the 4th V on cardiovascular responses in conscious rats. Our results are useful because recently, the cardiovascular reflex has been studied in different rat models and strains (49-51) with the aim of preventing the development of cardiovascular disorders in humans (52-54), as reduced cardiovascular reflex responses are indicative of cardiovascular disease (55-57). However, some points must be considered. We did not quantify concentrations of H₂O₂ or other ROS in the 4th V during the procedures. Such data would reinforce our results by demonstrating that the ROS levels are changed in the 4^{th} V. A previous study (58) reported a technique for continuous on-line measurement of cerebral H₂O₂.

In conclusion, our findings indicate that administration of a catalase inhibitor into the 4th V has strong effects on WKY rats but not SH rats exposed to SSCS.

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AUTHOR CONTRIBUTIONS

Valenti VE, Abreu LC, Fonseca FL, Adami F and Sato MA designed the study and performed the experiments. Vanderlei LC, Ferreira LL, Rodrigues LM and Ferreira C drafted the manuscript and performed the English grammar and spelling review. Valenti VE, Vanderlei LC, Ferreira LL, Rodrigues LM, Abreu LC, Fonseca FL and Ferreira C developed the experimental design, interpreted the text and drafted the manuscript. All authors read and approved the final version submitted for publication.

REFERENCES

- Kuo WW, Wu CH, Lee SD, Lin JA, Chu CY, Hwang JM. Second-hand smoke-induced cardiac fibrosis is related to the Fas death receptor apoptotic pathway without mitochondria-dependent pathway involvement in rats. Environ. Health Perspect. 2005;113(10):1349-53, http://dx. doi.org/10.1289/ehp.7479.
- Probst-Hensch NM, Imboden M, Dietrich D, Barthélemy JC, Ackermann-Liebrich U. Glutathione S-transferase polymorphisms, passive smoking, obesity, and heart rate variability in nonsmokers. Environ. Health Perspect. 2008;116(11):1494-9, http://dx.doi.org/10.1289/ehp.11402.
 Parain K, Hapdey C, Rousselet E, Marchand V, Dumery B, Hirsch EC.
- Parain K, Hapdey C, Rousselet E, Marchand V, Dumery B, Hirsch EC. Cigarette smoke and nicotine protect dopaminergic neurons against the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine Parkinsonian toxin. Brain Res. 2003;984(1-2):224-32, http://dx.doi.org/10.1016/S0006-8993(03) 03195-0.
- Karolewicz B, Klimek V, Zhu H, Szebeni K, Nail E, Stockmeier CA, et al. Effects of depression, cigarette smoking, and age on monoamine oxidase B in amygdaloid nuclei. Brain Res. 2005;1043(1-2):57-64, http://dx.doi. org/10.1016/j.brainres.2005.02.043.
- 5. Lavezzi AM, Ottaviani G, Mingrone R, Matturri L. Analysis of the human locus coeruleus in perinatal and infant sudden unexplained



nucleus. Brain Res Dev Brain Res. 2005;154(1):71-80, http://dx.doi.org/ 10.1016/j.devbrainres.2004.10.007. 6. Dogan ÓT, Elagoz S, Ozsahin SL, Epozturk K, Tuncer E, Akkurt I.

- Pulmonary toxicity of chronic exposure to tobacco and biomass smoke in rats. Clinics. 2011;66:1081-7, http://dx.doi.org/10.1590/S1807-59322011 000600027
- 7. Moreira TL, Gomes AR, Dresch TR, Silva SM, Valderramas S. Effects of inhaled cigarette smoke on the myo-articular system of female rats with collagen-induced arthritis. Clinics. 2011;66(6):915-7, http://dx.doi.org/ 10.1590/S1807-59322011000500033.
- Prado GF, Lombardi EM, Bussacos MA, Arrabal-Fernandes FL, Terra-Filho M, Santos Ude P. A real-life study of the effectiveness of different pharmacological approaches to the treatment of smoking cessation: rediscussing the predictors of success. Clinics. 2011;66(1):65-71, http://dx. doi.org/10.1590/S1807-59322011000100012.
- 9. Ling PM, Glantz SA. Tobacco industry consumer research on socially acceptable cigarettes. Tob Control. 2005;14(5):e3, http://dx.doi.org/10. 1136/tc.2005.011239.
- 10 Wang L, Pinkerton KE. Air pollutant effects on fetal and early postnatal development. Birth Defects Res C Embryo Today. 2006;81:144-54, http:// dx.doi.org/10.1002/bdrc.20097.
- Ayaori M, Hisada T, Suzukawa M, Yoshida H, Nishiwaki M, Ito T. 11. Plasma levels and redox status of ascorbic acid and levels of lipid peroxidation products in active and passive smokers. Environ. Health Perspect. 2000;108(2):105-8, http://dx.doi.org/10.1289/ehp.00108105.
- 12 Alberg A. The influence of cigarette smoking on circulating concentrations of antioxidant micronutrients. Toxicology. 2002;180(2):121-37, http://dx.doi.org/10.1016/S0300-483X(02)00386-4.
- 13. Gilmour MI, Jaakkola MS, London SJ, Nel AE, Rogers CA. How exposure to environmental tobacco smoke, outdoor air pollutants, and increased pollen burdens influences the incidence of asthma. Environ Health Perspect. 2006;114(4):627-633, http://dx.doi.org/10.1289/ehp.8380.
- 14. Al-Arifi MN, Maayah ZH, Alshamrani AA, Korashy HM. Impact of cigarette smoke exposure on the expression of cardiac hypertrophic genes, cytochrome P450 enzymes, and oxidative stress markers in rats. Toxicol Sci. 2012;37(5):1083-90, http://dx.doi.org/10.2131/jts.37.1083.
- Thannickal VJ, Fanburg BL. Reactive oxygen species in cell signaling. 15. Am J Physiol. 2000;279(6):L1005-28.
- 16. Halliwell B. Reactive oxygen species and the central nervous system. J. Neurochem. 1992;59(5):1609-23, http://dx.doi.org/10.1111/j.1471-4159. 1992.tb10990.x.
- Wang G, Anrather J, Huang J, Speth RC, Pickel VM, Iadecola C. NADPH 17. oxidase contributes to angiotensin II signaling in the nucleus tractus solitarius. J Neurosci. 2004;24(24):5516-24, http://dx.doi.org/10.1523/ JNEUROSCI.1176-04.2004.
- 18. Gao L, Wang W, Liu D, Zucker IH. Exercise training normalizes sympathetic outflow by central antioxidant mechanisms in rabbits with pacing-induced chronic heart failure. Circulation. 2007;115(24):3095-102, http://dx.doi.org/10.1161/CIRCULATIONAHA.106.677989.
- 19. Bertagnolli M, Campos C, Schenkel PC, de Oliveira VL, De Angelis K, Belló-Klein A. Baroreflex sensitivity improvement is associated with decreased oxidative stress in trained spontaneously hypertensive rat. Hypertens. 2006;24(12):2437-2443, http://dx.doi.org/10.1097/01.hjh. 0000251905.08547.17
- Colombari E, Sato MA, Cravo SL, Bergamaschi CT, Campos RR Jr, Lopes 20. OU. Role of the medulla oblongata in hypertension. Hypertension. 2001;38(3 Pt 2):549-54, http://dx.doi.org/10.1161/01.HYP.38.3.549.
- Giusti MF, Sato MA, Cardoso LM, Braga VA, Colombari E. Central antioxidant therapy inhibits parasympathetic baroreflex control in conscious rats. Neurosci Lett. 2011;489(2):115-8, http://dx.doi.org/10. 1016/j.neulet.2010.11.077.
- Valenti VE, Abreu LC, Sato MA, Ferreira C. ATZ (3-amino-1,2,4-triazole) 22. injected into the fourth cerebral ventricle influences the Bezold-Jarisch reflex in conscious rats. Clinics. 2010;65(12):1339-43, http://dx.doi.org/ 10.1590/S1807-59322010001200018.
- Valenti VE, De Abreu LC, Sato MA, Fonseca FL, Riera AR, Ferreira C. 23. Catalase inhibition into the fourth cerebral ventricle affects bradycardic parasympathetic response to increase in arterial pressure without changing the baroreflex. J Integr Neurosci. 2011;10(1):1-14, http://dx. doi.org/10.1142/S0219635211002580.
- 24. Luchese C, Pinton S, Nogueira CW. Brain and lungs of rats are differently affected by cigarette smoke exposure: antioxidant effect of an organoselenium compound. Pharmacol Res. 2009;59(3):194-201, http://dx.doi.org/10.1016/j.phrs.2008.11.006.
- 25. Bartoli CR, Wellenius GA, Diaz EA, Lawrence J, Coull BA, Akiyama I. Mechanisms of inhaled fine particulate air pollution-induced arterial blood pressure changes. Environ Health Perspect. 2009;117(3):361-6.
- Valenti VE, de Abreu LC, Sato MA, Ferreira C, Adami F, Fonseca FL, et al. 26. Sidestream cigarette smoke effects on cardiovascular responses in conscious rats: involvement of oxidative stress in the fourth cerebral ventricle. BMC Cardiovasc Disord. 2012;12:22, http://dx.doi.org/10. 1186/1471-2261-12-22.

- 27. Gairola CG, Drawdy ML, Block AE, Daugherty A. Sidestream cigarette smoke accelerates atherogenesis in apolipoprotein E-/-2001;156(1):49-55, http://dx.doi.org/10.1016/S0021-Atherosclerosis. 9150(00)00621-3.
- Valenti VE, Abreu LC, Saldiva PH, Carvalho TD, Ferreira C. Effects of 28. sidestream cigarette smoke exposure on baroreflex components in spontaneously hypertensive rats. Int J Environ. Health Res. 2010;20(6): 431-7.
- Valenti VE, Abreu LC, Ferreira C. Sidestream cigarette smoke exposure 29 effects on baroreflex in adult rats. Arq Bras Cardiol. 2010;96(2):148-53.
- 30 Paxinos G, Watson C. The rat brain in stereotaxic coordinates. Sydney: NSW: Academic Press; pp75; 1997.
- 31. Valenti VE, Ferreira C, Meneghini A, Ferreira M, Murad N, Ferreira Filho C. Evaluation of baroreflex function in Young spontaneous hypertensive rats. Arg Bras Cardiol. 2009;92:205-209, http://dx.doi.org/10.1590/ S0066-782X2009000300009.
- Vieira AA, Colombari E, De Luca Jr LA, Colombari DS, De Paula PM, 32. Menani JV. Importance of angiotensinergic mechanisms for the pressor response to l-glutamate into the rostral ventrolateral medulla. Brain Res. 2010;1322:72-80, http://dx.doi.org/10.1016/j.brainres.2010.01.066.
- Valenti VE, Imaizumi C, Abreu LC, Colombari E, Sato MA, Ferreira C. Intra-strain variations of baroreflex sensitivity in Young Wistar-Kyoto rats. Clin Invest Med. 2009;32(6):E251.
- 34. Cisternas JR, Valenti VE, Alves TB, Ferreira C, Petenusso M, Breda JR, et al. Cardiac baroreflex is already blunted in eight weeks old spontaneously hypertensive rats. Int Arch Med. 2010;3:2, http://dx. doi.org/10.1186/1755-7682-3-2.
- Cardoso LM, Colombari DSA, Menani JV, Alves Chianca Jr D, Colombari 35. E. Cardiovascular responses produced by central injection of hydrogen peroxide in conscious rats. Brain Res. Bull. 2006;71(1-3):37-44, http://dx. doi.org/10.1016/j.brainresbull.2006.07.013.
- Valenti VE, De Abreu LC, Sato MA, Saldiva PH, Fonseca FL, Giannocco 36. G, et al. Central N-acetylcysteine effects on baroreflex in juvenile spontaneously hypertensive rats. J Integr Neurosci. 2011;10(2):161-76, http://dx.doi.org/10.1142/S0219635211002671.
- 37. Cisternas JR, Valenti VE, Sato MA, Fonseca FL, Saldiva PH, De Mello Monteiro CB, et al. The effects of catalase inhibition into the fourth cerebral ventricle on the Bezold-Jarisch reflex in spontaneously hypertensive rats. J Integr Neurosci. 2011;10(4):475-87, http://dx.doi.org/10. 1142/S021963521100283X.
- Head GA, McCarty R. Vagal and sympathetic components of the heart 38. rate range and gain of the baroreceptor-heart rate reflex in conscious rats. Auton Nerv Syst. 1984;21(2-3):203-13.
- Barutcu I, Esen AM, Kaya D, Turkmen M, Karakaya O, Melek M. 39. Cigarette smoking and heart rate variability: dynamic influence of parasympathetic and sympathetic maneuvers. Ann Noninvasive Electrocardiol. 2005;10(3):324-9, http://dx.doi.org/10.1111/j.1542-474X. 2005.00636.x
- Adibhatla RM, Hatcher JF, Sailor K, Dempsey RJ. Polyamines and central 40. nervous system injury: spermine and spermidine decrease following transient focal cerébral ischemia in spontaneously hypertensive rats. Brain Res. 2002;938(1-2):81-86, http://dx.doi.org/10.1016/S0006-8993(02)02447-2.
- 41. Agarwal D, Welsch MA, Keller JN, Francis J. Chronic exercise modulates RAS components and improves balance between pro- and antiinflammatory cytokines in the brain of SHR. Basic Res Cardiol. 2011;106(6):1069-85, http://dx.doi.org/10.1007/s00395-011-0231-7.
- 42. Doty RL. Cranial nerve I: olfaction. In: Goltz CG, Pappert EJ, editors. Textbook of clinical neurology. Philadelphia: WB Saunders; p.90-101; 1998.
- Doty RL. The olfactory vector hypothesis of neurodegenerative disease: is it viable? Ann. Neurol. 2008;63(1):7-15, http://dx.doi.org/10.1002/ ana.21327.
- 44. Diethelm P, Rielle J, McKee M. The whole truth and nothing but the truth? The research that Philip Morris did not want you to see. Lancet. 2004:366(9479):86-92.
- 45. Rickert WS, Wright WG, Trivedi AH, Momin RA, Lauterbach JH. A comparative study of the mutagenicity of various types of tobacco products. Regul Toxicol Pharmacol. 2007;48(3):320-30, http://dx.doi. org/10.1016/j.yrtph.2007.05.003.
- Talbot P. In vitro assessment of reproductive toxicity of tobacco smoke 46 and its constituents. Birth Defects Res C Embryo Today. 2008;84(1):61-72, http://dx.doi.org/10.1002/bdrc.20120.
- 47. Schick S, Glantz S. Philip Morris toxicological experiments with fresh sidestream smoke: more toxic than mainstream smoke. Tob Control. 2005;14(6):396-404, http://dx.doi.org/10.1136/tc.2005.011288.
- Fluckiger JP, Sonnay M, Boillat N, Atkinson J. Attenuation of the 48. baroreceptor reflex by general anesthetic agents in the normotensive rat. Eur. J. Pharmacol. 1985;109(1):105-9, http://dx.doi.org/10.1016/0014-2999(85)90545-X
- Vanderlei LC, Pastre CM, Freitas Júnior IF, Godoy MF. Analysis of 49. cardiac autonomic modulation in obese and eutrophic children. Clinics. 2010;65(8):789-92, http://dx.doi.org/10.1590/S1807-59322010000800008.



- Valenti VE, de Abreu LC, Imaizumi C, Petenusso M, Ferreira C. Strain differences in baroceptor reflex in adult Wistar Kyoto rats. Clinics. 2010;65(2):203-8, http://dx.doi.org/10.1590/S1807-59322010000200013.
- Farah VM, De Angelis K, Joaquim LF, Candido GO, Bernardes N, Fazan Jr R. Autonomic modulation of arterial pressure and heart rate variability in hypertensive diabetic rats. Clinics. 2007;62(4):477-82, http://dx.doi. org/10.1590/S1807-59322007000400015.
- Monahan KD, Eskurza I, Seals DR. Ascorbic acid increases cardiovagal baroreflex sensitivity in healthy older men. Am J Physiol Heart Circ Physiol. 2004;286(6):H2113-7, http://dx.doi.org/10.1152/ajpheart.01054. 2003.
- Wright CI, Ruediger H, Kroner CI, Janssen BJ, Draijer R. Acute autonomic effects of vitamins and fats in male smokers. Eur J Clin Nutr. 2009;63(2):246-52.
- 54. Mainardi L, Corino V, Belletti S, Terranova P, Lombardi F. Low frequency component in systolic arterial pressure variability in patients

with persistent atrial fibrillation. Auton Neurosci. 2009;151(2):147-53, http://dx.doi.org/10.1016/j.autneu.2009.06.008.

- Pinheiro CH, Medeiros RÁ, Pinheiro DG, Marinho MJ. Spontaneous respiratory modulation improves cardiovascular control in essential hypertension. Arq Bras Cardiol. 2007;88(6):651-9, http://dx.doi.org/10. 1590/S0066-782X2007000600005.
- Lopes HF, Consolim-Colombo FM, Hachul D, Carvalho ME, Pileggi F, Silva HB. Hormonal and cardiovascular reflex assessment in a female patient with pure autonomic failure. Arq Bras Cardiol. 2000;75(3):235-42.
- Rodrigues FL, de Oliveira M, Salgado HC, Fazan R Jr. Effect of baroreceptor denervation on the autonomic control of arterial pressure in conscious mice. Exp Physiol. 2011;96(9):853-62.
- in conscious mice. Exp Physiol. 2011;96(9):853-62.
 58. Mao L, Osborne PG, Yamamoto K, Kato T. Continuous on-line measurement of cerebral hydrogen peroxide using enzyme-modified ring-disk plastic carbon film electrode. Anal Chem. 2002;74(15):3684-9, http://dx.doi.org/10.1021/ac011261+.