

## BASIC RESEARCH

# Sildenafil preserves diastolic relaxation after reduction by L-NAME and increases phosphodiesterase-5 in the intercalated discs of cardiac myocytes and arterioles

Silvia Elaine Ferreira-Melo,<sup>1</sup> Caroline Demacq,<sup>1</sup> Silvia Lacchini,<sup>II</sup> José Eduardo Krieger,<sup>III</sup> Maria Cláudia Irigoyen,<sup>III</sup> Heitor Moreno<sup>1</sup>

<sup>1</sup>Department of Internal Medicine, Faculty of Medical Sciences, Unicamp, Campinas, São Paulo, Brazil. <sup>II</sup>Department of Anatomy, Institute of Biology Sciences (ICB), USP, São Paulo, Brazil. <sup>III</sup>Heart Institute (InCor), Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil.

**OBJECTIVES:** We investigated the influence of sildenafil on cardiac contractility and diastolic relaxation and examined the distribution of phosphodiesterase-5 in the hearts of hypertensive rats that were treated with by NG-nitro-L-arginine methyl ester (L-NAME).

**METHODS:** Male Wistar rats were treated with L-NAME and/or sildenafil for eight weeks. The Langendorff method was used to examine the effects of sildenafil on cardiac contractility and diastolic relaxation. The presence and location of phosphodiesterase-5 and phosphodiesterase-3 were assessed by immunohistochemistry, and cGMP plasma levels were measured by ELISA.

**RESULTS:** In isolated hearts, sildenafil prevented the reduction of diastolic relaxation (dp/dt) that was induced by L-NAME. In addition, phosphodiesterase-5 immunoreactivity was localized in the intercalated discs between the myocardial cells. The staining intensity was reduced by L-NAME, and sildenafil treatment abolished this reduction. Consistent with these results, the plasma levels of cGMP were decreased in the L-NAME-treated rats but not in rats that were treated with L-NAME + sildenafil.

**CONCLUSION:** The sildenafil-induced attenuation of the deleterious hemodynamic and cardiac morphological effects of L-NAME in cardiac myocytes is mediated (at least in part) by the inhibition of phosphodiesterase-5.

**KEYWORDS:** Hypertension; Phosphodiesterase-5 Inhibitor; Immunohistochemistry.

Ferreira-Melo SE, Demacq C, Lacchini S, Krieger JE, Irigoyen MC, Moreno H. Sildenafil preserves diastolic relaxation after reduction by L-NAME and increases phosphodiesterase-5 in the intercalated discs of cardiac myocytes and arterioles. *Clinics*. 2011;66(7):1253-1258.

Received for publication on November 19, 2011; First review completed on December 28, 2011; Accepted for publication on April 4, 2011

E-mail: hmoreno@uol.com.br

Tel.: 55 19 3521-9538

## INTRODUCTION

The inhibition of phosphodiesterase type 5 (PDE5) by selective inhibitors such as sildenafil enhances intracellular levels of cGMP, which can be beneficial in restoring physiological function in situations in which nitric oxide (NO) formation is reduced. Although the cyclic GMP-selective PDE5 has been thought to play a minor role in cardiac myocytes, recent studies using selective inhibitors have suggested that PDE5 can modulate chronic cardiac stress responses.<sup>1-3</sup> In addition, recent studies have demonstrated PDE5 expression and activity in cardiac myocytes and its targeted inhibition by sildenafil; moreover, a role for

this PDE in cardiomyocyte hypertrophy modulation has been reported.<sup>4-5</sup>

The chronic inhibition of NO biosynthesis by the oral administration of the nonselective NO synthase (NOS) inhibitor NG-nitro-L-arginine methyl ester (L-NAME) is a well-established hypertension model<sup>6-8</sup> that is associated with reduced cardiac output, cardiac hypertrophy, extensive areas of fibrosis and myocardial necrosis, changes in myocardial contractility, and cardiomyocyte and vascular smooth muscle remodeling.<sup>8-13</sup> In a previous study, we demonstrated that sildenafil confers cardiovascular protection by inhibiting PDE5, thereby increasing the bioavailability of cGMP.<sup>14</sup>

Recently, PDE5 distribution in the heart was reported to be compartmentalized in the Z bands of myocardial tissue<sup>15-16</sup>, and sildenafil has been reported to affect cardiac performance and vascular function in L-NAME-treated rats.<sup>14</sup> These latter findings seem to be related to sildenafil's vasodilatory effects that both reduce afterload and improve

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted noncommercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

cardiac output. However, it is unknown whether sildenafil affects cardiac contractility and diastolic relaxation. Therefore, the aim of this study was to examine the effect of sildenafil on cardiac contractility and diastolic relaxation in isolated hearts and to confirm the association between the cardiovascular effects of sildenafil and the presence and distribution of PDE5 in the hearts of hypertensive L-NAME-treated rats.

## METHODS

The animal experiments described here were approved by the Institutional Committee for Ethics in Animal Experimentation (CEE/IB/Unicamp) and were performed in accordance with the *National Institutes of Health Guide for the Care and Use of Laboratory Animals* and under the ethical guidelines that have been established by the Brazilian College for Animal Experimentation (Cobea).

### Experimental design

Male Wistar rats (specific-pathogen free) weighing  $180 \pm 20$  g were obtained from the Central Animal House Services (Cemib, a facility that is engaged in the production and reproduction of laboratory animals and is affiliated with the International Council for Laboratory Animal Science - ICLAS) at Unicamp and were divided into the following groups, each containing 10-15 rats: control (water alone); L-NAME (20 mg/rat/day);<sup>17</sup> sildenafil (45 mg/kg/day);<sup>18</sup> and L-NAME + sildenafil (20 mg/rat/day and 45 mg/kg/day, respectively). Each group was treated for eight weeks. Before starting the treatment, the mean volume of liquid that was ingested by the five rats in each cage was determined by measuring the volume of water that the rats drank and dividing this volume by five. This calculated volume was then used to determine the amount of each drug to directly dilute in the drinking water to deliver the desired dose per rat or kg of body weight per day. The L-NAME and sildenafil citrate were dissolved in the drinking water at concentrations of 1.1 mM and 1 mM to provide daily intakes of approximately 74  $\mu\text{mol/rat/day}$  and 67  $\mu\text{mol/rat/day}$ , respectively. In the L-NAME + sildenafil group, the two drugs were diluted in the same drinking bottle. The average daily intake of both water and food did not differ significantly between the L-NAME-treated and untreated rats. The control rats received the same volume of tap water alone. In addition, we monitored the water consumption by the animals in each group daily to verify that the correct dose was administered.

### Non-Invasive (tail-cuff) blood pressure and body weight measurements

The systolic arterial blood pressure (SBP) of each rat was measured twice a week for eight weeks using the tail-cuff method (Codal system), and the mean of these two measurements was considered as the value for that week. The rats were also weighed twice a week after obtaining the blood pressure measurements, and the mean weekly weight gain was calculated as described for the blood pressure.

### Isolated heart preparation (Langendorff)

The method for isolating the beating heart was originally described by Langendorff.<sup>19</sup> At the end of study, the rats were anesthetized with sodium pentobarbital (50 mg/kg, i.p.). After opening the chest, the heart was isolated and

perfused in a Langendorff apparatus (Isolated Heart Perfusion Apparatus, Harvard Apparatus, Hollister, MA, USA.) under a constant pressure of 70 mmHg. A collapsed latex balloon was placed in the left ventricular cavity via an incision in the left atrium, and the initial intraballoon pressure was adjusted to 4-6 mmHg. The left ventricular pressure was monitored via a pressure transducer (YS100, Transonic Systems Inc., NY, USA). Both pressure parameters (the left ventricular development pressure, or LVDP,  $dP/dt+$ ,  $dP/dt-$ , the maximum rate of rise or fall in LVDP) and heart rate (HR) were continually recorded; the signals were acquired, amplified, and analyzed using an analog-to-digital interface (Dataq Instruments, Akron, OH, USA). The hearts were perfused with a Krebs-Henseleit solution.

### PDE immunohistochemistry

PDE5 and PDE3 were measured by immunohistochemistry performed in slices of the left ventricle or vessels that were stained with IgG anti-PDE5 or anti-PDE3 antibodies (Zymed, Laboratories, South San Francisco, CA). In brief, the slices were deparaffinized with Citrosolv (Fisher Scientific, Fair Lawn, NJ). Before tissue rehydration, the endogenous peroxidase activity was blocked with hydrogen peroxide and methanol (1:9) for 20 minutes. Following rehydration, the samples were rinsed with phosphate-buffered saline (PBS). Fetal bovine serum (10% FBS in PBS) was used to block the nonspecific sites for 60 minutes at room temperature, followed by incubation in 2.5% fat-free dry milk (Molico, Nestlé, Brazil) for 30 minutes. The primary antibody—either rabbit anti-PDE5 or goat anti-PDE3—was diluted to 1:250 in 2% BSA in PBS and applied to the sections for 16-18 hours at 4°C. Subsequently, the samples were washed and incubated with the biotinylated secondary antibody (Zymed Laboratories, South San Francisco, CA) for 60 minutes at room temperature, followed by incubation with streptavidin-peroxidase complex (1:1000) for 60 minutes at room temperature. Finally, a chromogen solution comprised of 3,3'-diaminobenzidine (DAB) (6 mg), hydrogen peroxide (150  $\mu\text{l}$ ) and PBS (10 ml) was applied for two minutes in the dark at room temperature.

The samples were coded and then assessed by two independent blinded observers using an optical microscope (Q500YW, LEICA, UK) equipped with a 40x objective and coupled to an image analyzer (Quantimet Q500YW, LEICA, UK).

### cGMP concentrations

The plasma cGMP concentrations were measured by enzyme-linked immunosorbent assay (ELISA) using a commercial kit (Cayman Chemical Co., Ann Arbor, MI). The plasma samples were initially precipitated with trichloroacetic acid, extracted with water-saturated ethyl ether, evaporated to dryness, and then reconstituted in an assay buffer. The standards and samples were acetylated to allow the detection of nucleotides in the picomolar range.<sup>20</sup>

### Statistical analysis

The results are expressed as the mean  $\pm$  SEM. An analysis of variance (ANOVA) for repeated measurements model was used to assess the differences in body weight and tail-cuff pressure. A two-way ANOVA was used to compare the heart weight, left ventricular weight, left ventricular weight index, mean arterial pressure, cardiac output, and total peripheral vascular resistance. When the

ANOVA results were deemed significant, the Bonferroni test was used to examine the differences among the groups. In all cases, a two-sided *p*-value of <0.05 was considered to be significant.

**RESULTS**

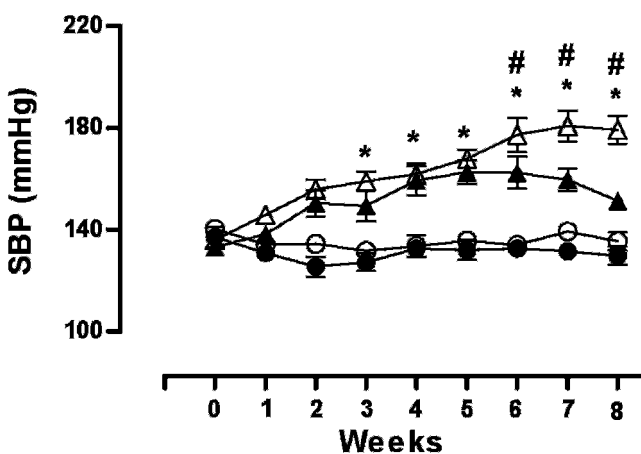
**Systolic blood pressure**

The systolic blood pressure (SBP) increased to a similar extent in the L-NAME and L-NAME + sildenafil groups until the sixth week; in addition, the SBP was higher in both groups than in the control and sildenafil groups from two weeks onwards. After six weeks, the rats in the L-NAME group had higher blood pressures than did the other three groups (*p*<0.05), while blood pressure in the L-NAME + sildenafil group essentially returned to normal (Figure 1).

**Isolated heart (Langendorff)**

The systolic contractility of the isolated hearts was measured by the first temporal derivative of the LVDP positive development (dP/dt+, in mmHg/s), and the isovolumetric relaxation was measured by the first temporal derivative of the LVDP negative pressure (dP/dt-, in mmHg/s). We analyzed all the hearts using the same diastolic pressure range (4–6 mmHg).

A significant decrease in dP/dt+ was found in the L-NAME group (4028±77 mmHg/s) relative to the control and sildenafil groups (4514±102 and 4466±116 mmHg/s, respectively; *p*<0.05). Sildenafil prevented this dP/dt+ reduction in the L-NAME-treated rats (4165±82 mmHg/s; *p*<0.05). The L-NAME group showed relatively decreased dP/dt- (3090±95 mmHg/s) when compared to the control and sildenafil groups (3930±96 and 4079±113 mmHg/s, respectively; *p*<0.05); however, sildenafil prevented this impaired cardiac relaxation (the dP/dt- in the L-NAME + sildenafil group was 3768±121 mmHg/s; *p*<0.01) (Table 1).



**Figure 1** - The changes in systolic blood pressure (SBP, in mmHg) during the eight weeks of the study. Control (squares, n = 15); L-NAME (diamonds, n = 15); sildenafil (circles, n = 15) and L-NAME + sildenafil (triangles, n = 15). The data points and error bars represent the mean ± SEM. \**p*<0.05 for L-NAME and L-NAME + sildenafil groups vs. control group. #*p*<0.05 for L-NAME group vs. L-NAME + sildenafil group.

**Table 1** - Isolated heart (Langendorff technique). Development of left ventricular systolic (LVSP, mmHg) and diastolic (LVDP, mmHg) pressures, first temporal derivative of LVDP positive development (dP/dt+, mmHg/s), first temporal derivative of the LVDP negative pressure (dP/dt-, mmHg/s) and heart rate (HR, bpm).

	CONTROL (n = 12)	SILDENAFIL (n = 10)	L-NAME (n = 10)	L-NAME+ SILDENAFIL (n = 12)
LVSP (mmHg)	138 ± 16	132 ± 17	127 ± 12	129 ± 18
LVDP (mmHg)	4.5 ± 0.9	5.3 ± 0.5	4.6 ± 0.9	4.6 ± 0.8
dP/dt+ (mmHg/s)	4514 ± 102	4466 ± 116	4028 ± 77*	4165 ± 82**
dP/dt- (mmHg/s)	3930 ± 96	4079 ± 113	3090 ± 95*	3768 ± 121**
HR (bpm)	275 ± 25	291 ± 19	270 ± 28	297 ± 18

\**p*<0.05 vs. CONTROL and sildenafil.  
\*\**p*<0.01 vs. L-NAME.

**PDE immunohistochemistry**

**PDE5.** The PDE5 immunohistochemistry revealed positive staining in the tunicae media and intima but not in the adventitia of both arteries and veins. The L-NAME treatment markedly reduced the media staining in the 10-50-µm diameter arterioles when compared to the control group. However, this reduction was abolished in the L-NAME + sildenafil group. Conversely, the rats that were treated only with sildenafil showed an increase in staining when compared to the other groups (Figure 2A). Larger arteries (larger than 100 µm in diameter) did not show any relevant alterations in PDE5 staining and localization in the four groups.

Staining was also observed in the intercalated discs of the myocardial cells, and this was reduced by the L-NAME treatment. Sildenafil administered concomitantly with the L-NAME reversed the L-NAME effect to the control levels. In addition, sildenafil alone increased PDE5 intensity when compared to the control group (Figure 2B).

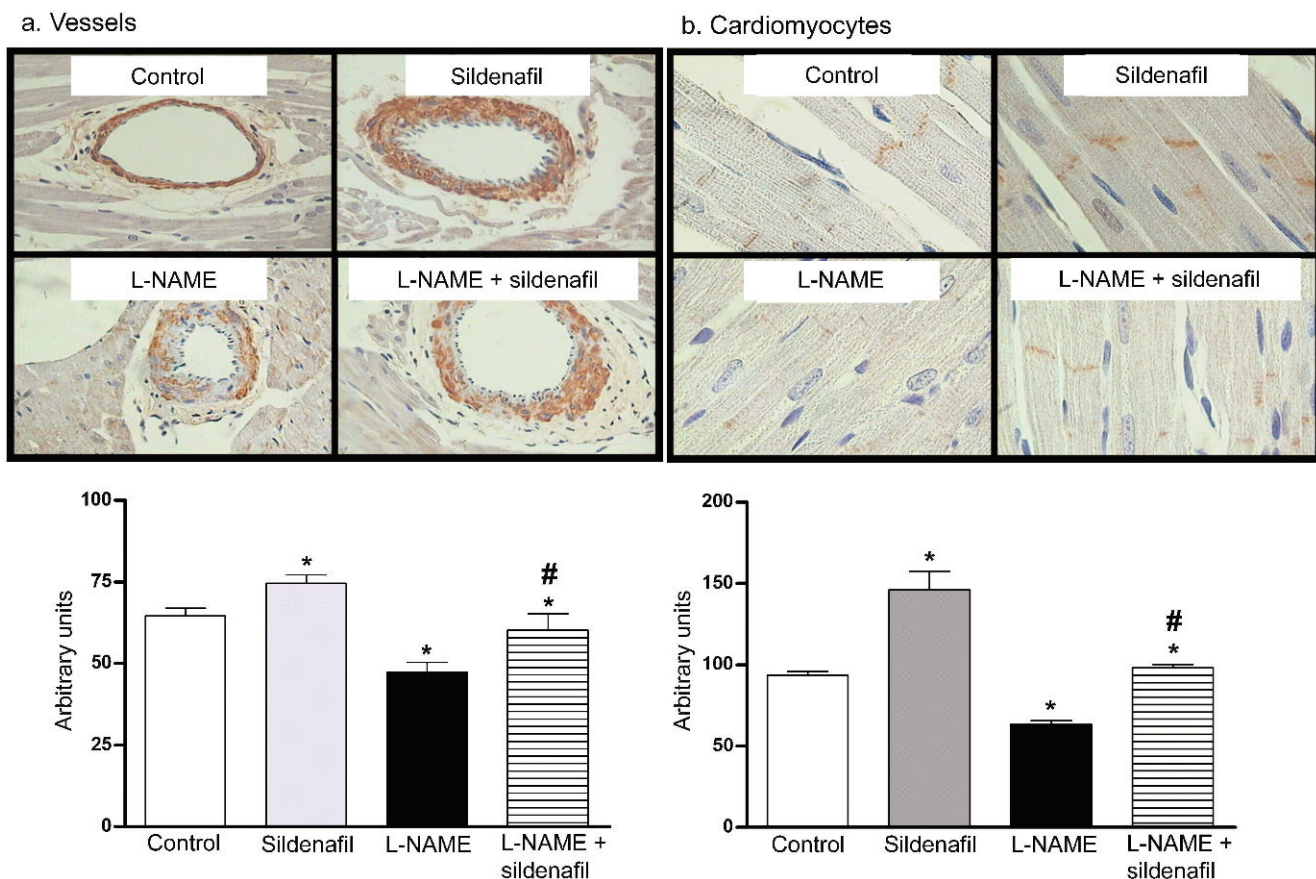
**PDE3.** The PDE3 immunohistochemistry revealed positive staining in the adventitia of both arteries and veins that was intensified in the group that received only sildenafil when compared to the control, L-NAME, and L-NAME + sildenafil groups. The endothelium also showed positive (albeit less intense) staining for PDE3. The cardiac myocytes had light positive staining for PDE3 in all four experimental groups (data not shown). Interestingly, areas of lesions showed an intense staining that was not affected by the sildenafil treatment (data not shown).

**Plasma cGMP concentrations**

After eight weeks, the plasma cGMP levels (in pmol/ml) were decreased in the L-NAME group and increased in the sildenafil group (*p*<0.05 vs. the control group). The cGMP levels were restored to the control values in the L-NAME + sildenafil group (Figure 3).

**DISCUSSION**

Using the isolated heart preparation, the decrease in dP/dt- that was observed in the L-NAME group was prevented in the L-NAME + sildenafil group. This result may be interpreted as an effect of sildenafil on the L-NAME-induced diastolic relaxation because the cardiac

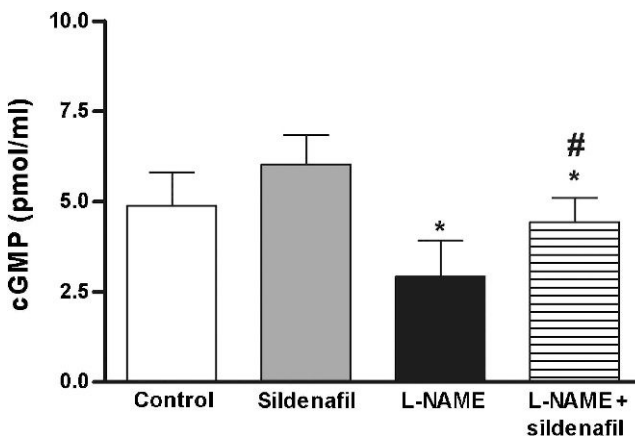


**Figure 2 - A)** The PDE5 immunohistochemistry in the cardiac arteries after 8 weeks of treatment. Representative photomicrographs and a summary graph of the quantitative analysis of the staining intensity in the vascular smooth muscle cells of small arteries are shown. **B)** The PDE5 immunohistochemistry in the cardiac tissue after 8 weeks of treatment. Representative photomicrographs and a summary graph of the quantitative analysis of staining intensity in the intercalary discs between myocytes. The data are expressed in arbitrary units; n = 5/group; \*p < 0.05 vs. control; #p < 0.05 vs. L-NAME.

load was pre-controlled and there were no significant differences in heart rates among the various groups. In cardiac myocytes, cAMP mediates catecholamine signaling, while cGMP mediates the effects of nitric oxide and atrial natriuretic peptide. cGMP activates PKG, which is then capable of countering the cAMP-PKA contractile stimulation.<sup>21</sup> Moreover, the duration and magnitude of these actions are determined by cGMP generation and by its hydrolysis, which is catalyzed by PDE5 that is compartmentalized within the cell, thereby facilitating the regulation of specific targeted proteins.<sup>22</sup>

While sildenafil had no effect on blood pressure in the normotensive (control) group, the compound produced a small but significant attenuation of the increase in blood pressure that was seen in the rats that were treated with L-NAME alone. Moreover, we have previously showed that enalapril and amlodipine decrease blood pressure in L-NAME-treated rats but do not prevent cardiac lesions.<sup>17,36</sup> Therefore, we suggest that this hemodynamic effect may have been at least partially mediated by an inhibition of PDE5 in arterial resistance vessels<sup>23-24</sup> and that it was associated with a slight reduction in the total peripheral vascular resistance and with enhanced cardiac output.<sup>14</sup> An alternative explanation could be sildenafil's effect of preventing the impaired diastolic relaxation (dP/dt-) that was diminished by L-NAME.

Inhibitors of nitric oxide synthesis may cause an afterload increment in hypertensive animals that can subsequently induce cardiac hypertrophy, fibrosis and/or ischemia.<sup>25</sup>



**Figure 3 -** The plasma cGMP levels after eight weeks of treatment in the control (open column, n = 15), L-NAME (black column, n = 15), sildenafil (grey column, n = 15) and L-NAME + sildenafil (horizontally striped column, n = 15) groups. The bars represent the mean ± SEM. \*p < 0.05 vs. control group; #p < 0.05 vs. L-NAME group.

These three changes, either alone or in combination, predispose the heart to impaired left ventricular relaxation. In addition, decreased left ventricular diastolic distensibility may arise from a dysfunction of the dynamics of ventricular relaxation.<sup>26</sup> Thus, the impaired cardiac output could be due to these alterations in ventricular diastolic function through a reduction in the isovolumic relaxation time (IVRT).

In support of this idea, another important finding in the present study is that sildenafil prevented the L-NAME-induced reduction in PDE5 staining in the myocardium and in the arterioles, while it did not alter the PDE3 staining. Consistent with previous studies,<sup>15-16</sup> we found that PDE5 is localized at the intercalated discs. Importantly, low concentrations of cGMP in unstimulated hearts can augment contractile function, and this effect is likely mediated by cross-talk with cAMP-dependent signaling.<sup>27</sup> However, PDE5 inhibition targets cGMP-PKG activity in a region that is strategically linked to adrenergic regulation. In this region, it depresses myofilament calcium sensitivity by increasing troponin I phosphorylation, thereby accounting for the positive lusitropic<sup>28-29</sup> and negative inotropic effects.<sup>30</sup> Thus, our immunohistochemical findings are consistent with previous findings in isolated hearts in which sildenafil prevented the L-NAME-induced impairment in diastolic relaxation (dp/dt-).

In contrast to our findings, however, a previous study has reported a shift in the intracellular localization of PDE5 from its normal Z band localization to a more diffuse cytosolic distribution following a chronic NOS inhibition by L-NAME that eliminated sildenafil's effectiveness even when exogenous NO was provided.<sup>16</sup> In the present study, we found no shift in PDE5 localization. These contradictory results may be explained by differences in the experimental protocols; the aforementioned study administered L-NAME for one or two weeks following the sildenafil treatment, while we studied the effect of co-treatment with L-NAME and sildenafil for eight weeks and found that sildenafil prevented the L-NAME-induced effects.

Another finding from our study is that sildenafil increased circulating cGMP and abolished the decrease that was caused by L-NAME. Importantly, although cGMP is considered to reflect natriuretic peptides in patients with cardiac dysfunction and to be an indicator of NO synthase activity in healthy subjects,<sup>31</sup> Castro et al has reported that PDE-5 regulation appears to be compartmentalized in cardiac myocytes, where it interacts with NO but not with natriuretic peptide-stimulated cGMP.<sup>32</sup> In addition, chronic L-NAME-induced NOS inhibition decreases cGMP,<sup>8,33-35</sup> suggesting that the natriuretic peptide pathway in this model is unable to rescue the impaired cGMP formation caused by NOS inhibition. Therefore, the changes in the plasma cGMP levels that we found in the present study are likely the result of NO pathway modulation.

## CONCLUSIONS

In conclusion, our results suggest that the sildenafil-mediated attenuation of L-NAME-induced deleterious hemodynamic and morphological alterations is at least partially modulated by PDE5 inhibition in cardiac myocytes, as supported by our immunohistochemical and isolated heart findings. Finally, sildenafil's effects on cardiac relaxation should be studied further as a new therapeutic

approach for treating hypertensive patients with diastolic dysfunction.

## ACKNOWLEDGMENTS

Financial support was provided by the State of São Paulo Research Foundation (Fapesp), the Coordination for Higher Level Graduates Improvement (Capes), and the National Council for Scientific and Technological Development (CNPq), Brazil. The sildenafil citrate was provided by Pfizer, Inc.

## REFERENCES

- Nagayama T, Hsu S, Zhang M, Koitabashi N, Bedja D, Gabrielson KL, et al. Pressure-overload magnitude-dependence of the anti-hypertrophic efficacy of PDE5A inhibition. *J Mol Cell Cardiol.* 2009;46:560-7, doi: 10.1016/j.yjmcc.2008.12.008.
- Kass DA, Champion HC, Beavo JA. Phosphodiesterase type 5: expanding roles in cardiovascular regulation. *Circ Res.* 2007;101:1084-95, doi: 10.1161/CIRCRESAHA.107.162511.
- Takimoto E, Champion HC, Li M, Belardi D, Ren S, Rodriguez ER, et al. Chronic inhibition of cyclic GMP phosphodiesterase 5A prevents and reverses cardiac hypertrophy. *Nat Med.* 2005;11:214-22, doi: 10.1038/nm1175.
- Zhang M, Koitabashi N, Nagayama T, Rambaran R, Feng N, Takimoto E, et al. Expression, activity, and pro-hypertrophic effects of PDE5A in cardiac myocytes. *Cell Signal.* 2008;20:2231-6, doi: 10.1016/j.cellsig.2008.08.012.
- Rossoni G, Manfredi B, De Gennaro Colonna V, Berti M, Guazzi M, Berti F. Sildenafil reduces L-NAME-induced severe hypertension and worsening of myocardial ischaemia-reperfusion damage in the rat. *Br J Pharmacol.* 2007; 150:567-76, doi: 10.1038/sj.bjp.0707131.
- Arnal JF, el Amrani AI, Chatellier G, Menard J, Michel JB. Cardiac weight in hypertension induced by nitric oxide synthase blockade. *Hypertension.* 1993;22:380-7.
- Moreno Junior H, Metzke K, Zatz R, De Nucci G. Chronic nitric oxide blockade causes cardiac ischaemia but not cardiac hypertrophy: An experiment of four weeks in rats. *Verh Dtsch Ges Path.* 1994;78:459.
- Pechanova O, Bernatova I, Pelouch V, Babal P. L-NAME-induced protein remodeling and fibrosis in the rat heart. *Physiol Res.* 1999;48:353-62.
- Amrani M, O'Shea J, Allen NJ, Harding SE, Jayakumar J, Pepper JR, et al. Role of basal release of nitric oxide on coronary flow and mechanical performance of the isolated rat heart. *J Physiol.* 1992;456:681-7.
- Jover B, Herizi A, Ventre F, Dupont M, Mimran A. Sodium and angiotensin in hypertension induced by long-term nitric oxide blockade. *Hypertension.* 1993;21:944-8.
- Moreno H, Metzke K, Antunes E, Zatz R, De Nucci G. Chronic nitric oxide inhibition as a model of hypertensive heart muscle disease. *Bas Res Cardiol.* 1996;91:248-55, doi: 10.1007/BF00788911.
- Li JS, Schiffrin EL. Resistance artery structure and neuroeffector mechanisms in hypertension induced by inhibition of nitric oxide synthase. *Am J Hypertens.* 1994;7:996-1004.
- Huang M, Manning RD, Jr., LeBlanc MH, Hester RL. Overall hemodynamic studies after the chronic inhibition of endothelial-derived nitric oxide in rats. *Am J Hypertens.* 1995;8:358-64, doi: 10.1016/0895-7061(94)00203-N.
- Ferreira-Melo SE, Yugar-Toledo JC, Coelho OR, De Luca IM, Tanus-Santos JE, Hyslop S, et al. Sildenafil reduces cardiovascular remodeling associated with hypertensive cardiomyopathy in NOS inhibitor-treated rats. *Eur J Pharmacol.* 2006;542:141-7, doi: 10.1016/j.ejphar.2006.04.039.
- Senzaki H, Smith CJ, Juang GJ, Isoda T, Mayer SP, Ohler A, et al. Cardiac phosphodiesterase 5 (cGMP-specific) modulates beta-adrenergic signaling in vivo and is down-regulated in heart failure. *Faseb J.* 2001; 15:1718-26, doi: 10.1096/fj.00-0538com.
- Takimoto E, Champion HC, Belardi D, Moslehi J, Mongillo M, Mergia E, et al. cGMP catabolism by phosphodiesterase 5A regulates cardiac adrenergic stimulation by NOS3-dependent mechanism. *Circ Res.* 2005; 96:100-9, doi: 10.1161/01.RES.0000152262.22968.72.
- Moreno H, Jr., Piovesan Nathan L, Pereira Costa SK, Metzke K, Antunes E, Zatz R, et al. Enalapril does not prevent the myocardial ischemia caused by the chronic inhibition of nitric oxide synthesis. *Eur J Pharmacol.* 1995; 287:93-6, doi: 10.1016/0014-2999(95)00625-X.
- Walker DK, Ackland MJ, James GC, Muirhead GJ, Rance DJ, Wastall P, et al. Pharmacokinetics and metabolism of sildenafil in mouse, rat, rabbit, dog and man. *Xenobiotica.* 1999;29:297-310, doi: 10.1080/004982599238687.
- Langendorff O. Untersuchungen am überlebenden Säugetierherzen. *Pflügers Arch Ges Physiol.* 1895;61:291-332, doi: 10.1007/BF01812150.
- Branka JE, Vallette G, Jarry A, Laboisie CL. Stimulation of mucin exocytosis from human epithelial cells by nitric oxide: evidence for a cGMP-dependent and a cGMP-independent pathway. *Biochem J.* 1997;323:521-4.

21. Tsai EJ, Kass DA. Cyclic GMP signaling in cardiovascular pathophysiology and therapeutics. *Pharmacol Ther.* 2009;122:216-38, doi: 10.1016/j.pharmthera.2009.02.009.
22. Fischmeister R, Castro LR, Abi-Gerges A, Rochais F, Jurevicius J, Leroy J, et al. Compartmentation of cyclic nucleotide signaling in the heart: the role of cyclic nucleotide phosphodiesterases. *Circ Res.* 2006;99:816-28, doi: 10.1161/01.RES.0000246118.98832.04.
23. Cremers B, Scheler M, Maack C, Wendler O, Schafers HJ, Sudkamp M, et al. Effects of sildenafil (viagra) on human myocardial contractility, in vitro arrhythmias, and tension of internal mammary arteries and saphenous veins. *J Cardiovasc Pharmacol.* 2003;41:734-43, doi: 10.1097/00005344-200305000-00010.
24. Kloner RA. Novel phosphodiesterase type 5 inhibitors: assessing hemodynamic effects and safety parameters. *Clin Cardiol.* 2004;27:120-5, doi: 10.1002/clc.4960271306.
25. Moreno H, Jr., Metze K, Bento AC, Antunes E, Zatz R, de Nucci G. Chronic nitric oxide inhibition as a model of hypertensive heart muscle disease. *Basic Res Cardiol.* 1996;91:248-55, doi: 10.1007/BF00788911.
26. Bonow RO, Udelson JE. Left ventricular diastolic dysfunction as a cause of congestive heart failure. Mechanisms and management. *Ann Intern Med.* 1992; 117:502-10.
27. Vila-Petroff MG, Younes A, Egan J, Lakatta EG, Sollott SJ. Activation of distinct cAMP-dependent and cGMP-dependent pathways by nitric oxide in cardiac myocytes. *Circ Res.* 1999;84:1020-31.
28. Fentzke RC, Buck SH, Patel JR, Lin H, Wolska BM, Stojanovic MO, et al. Impaired cardiomyocyte relaxation and diastolic function in transgenic mice expressing slow skeletal troponin I in the heart. *J Physiol.* 1999;517:143-57, doi: 10.1111/j.1469-7793.1999.0143z.x.
29. Massion PB, Feron O, Dessy C, Balligand JL. Nitric oxide and cardiac function: ten years after, and continuing. *Circ Res.* 2003;93:388-98, doi: 10.1161/01.RES.0000088351.58510.21.
30. Wegener JW, Nawrath H, Wolfgruber W, Kuhbandner S, Werner C, Hofmann F, et al. cGMP-dependent protein kinase I mediates the negative inotropic effect of cGMP in the murine myocardium. *Circ Res.* 2002;90:18-20, doi: 10.1161/hh0102.103222.
31. Kielstein JT, Impraïm B, Simmel S, Bode-Boger SM, Tsikas D, Frolich JC, et al. Cardiovascular effects of systemic nitric oxide synthase inhibition with asymmetrical dimethylarginine in humans. *Circulation.* 2004;109:172-7, doi: 10.1161/01.CIR.000105764.22626.B1.
32. Castro LR, Verde I, Cooper DM, Fischmeister R. Cyclic guanosine monophosphate compartmentation in rat cardiac myocytes. *Circulation.* 2006;113:2221-8, doi: 10.1161/CIRCULATIONAHA.105.599241.
33. Arnal JF, Warin L, Michel JB. Determinants of aortic cyclic guanosine monophosphate in hypertension induced by chronic inhibition of nitric oxide synthase. *J Clin Invest.* 1992;90:647-52, doi: 10.1172/JCI115906.
34. Deng LY, Thibault G, Schiffrin EL. Effect of hypertension induced by nitric oxide synthase inhibition on structure and function of resistance arteries in the rat. *Clin Exp Hypertens.* 1993;15:527-37, doi: 10.3109/10641969309041627.
35. Pechanova O, Bernatova I. Effect of long-term NO synthase inhibition on cyclic nucleotide content in rat tissues. *Physiol Res.* 1996;45:305-9.
36. de Oliveira CF, Nathan LP, Metze K, Moreno Jr H, de Luca IM, Sucupira M. Effect of Ca<sup>2+</sup> channel blockers on arterial hypertension and heart ischaemic lesions induced by chronic blockade of nitric oxide in the rat. *Eur J Pharmacol.* 1999;373:195-200, doi: 10.1016/S0014-2999(99)00267-8.