

Exercise training prevents skeletal muscle damage in an experimental sepsis model

Carla Werlang Coelho,^{I,II,III} Paulo R. Jannig,^{IV} Arlete B. de Souza,^V Hercilio Fronza, Jr.,^V Glauco A Westphal,^{II} Fabricia Petronilho,^{VI} Larissa Constantino,^{VI} Felipe Dal-Pizzol,^{VI} Gabriela K. Ferreira,^{VI} Emilio E. Streck,^{VI} Eliezer Silva^{I,VII}

^IFaculdade de Medicina da Universidade de São Paulo, Anesthesiology Strict Sensus Post Graduation Program of the Medical School, São Paulo/SP, Brazil.

^{II}Universidade da Região de Joinville (UNIVILLE), Physical Education and Medicine Department, Joinville/SC, Brazil. ^{III}Universidade do Estado de Santa Catarina (UDESC-Joinville), Chemistry Department, Joinville/SC, Brazil. ^{IV}Faculdade de Medicina da Universidade de São Paulo, Experimental Pathophysiology Department, São Paulo/SP, Brazil. ^VCentro de Diagnóstico Anatomo Patológicos (CEDAP), Pathology Department, Joinville/SC, Brazil.

^{VI}Universidade do Extremo Sul Catarinense (UNESC), Experimental Pathophysiology Laboratory, Criciúma/SC, Brazil. ^{VII}Hospital Israelita Albert Einstein, Intensive Care Unit, São Paulo/SP, Brazil.

OBJECTIVE: Oxidative stress plays an important role in skeletal muscle damage in sepsis. Aerobic exercise can decrease oxidative stress and enhance antioxidant defenses. Therefore, it was hypothesized that aerobic exercise training before a sepsis stimulus could attenuate skeletal muscle damage by modulating oxidative stress. Thus, the aim of this study was to evaluate the effects of aerobic physical preconditioning on the different mechanisms that are involved in sepsis-induced myopathy.

METHODS: Male Wistar rats were randomly assigned to either the untrained or trained group. The exercise training protocol consisted of an eight-week treadmill program. After the training protocol, the animals from both groups were randomly assigned to either a sham group or a cecal ligation and perforation surgery group. Thus, the groups were as follows: sham, cecal ligation and perforation, sham trained, and cecal ligation and perforation trained. Five days after surgery, the animals were euthanized and their soleus and plantaris muscles were harvested. Fiber cross-sectional area, creatine kinase, thiobarbituric acid reactive species, carbonyl, catalase and superoxide dismutase activities were measured.

RESULTS: The fiber cross-sectional area was smaller, and the creatine kinase, thiobarbituric acid reactive species and carbonyl levels were higher in both muscles in the cecal ligation and perforation group than in the sham and cecal ligation and perforation trained groups. The muscle superoxide dismutase activity was higher in the cecal ligation and perforation trained group than in the sham and cecal ligation and perforation groups. The muscle catalase activity was lower in the cecal ligation and perforation group than in the sham group.

CONCLUSION: In summary, aerobic physical preconditioning prevents atrophy, lipid peroxidation and protein oxidation and improves superoxide dismutase activity in the skeletal muscles of septic rats.

KEYWORDS: CLP; Skeletal Muscle; Myopathy; Oxidative Stress; Antioxidant Enzymes; Aerobic Exercise.

Coelho CW, Jannig PR, Souza AB, Fronza Jr H, Westphal GA, Petronilho F, et al. Exercise training prevents skeletal muscle damage in an experimental sepsis model. *Clinics*. 2013;68(1):107-114.

Received for publication on August 1, 2012; First review completed on August 24, 2012; Accepted for publication on September 22, 2012

E-mail: silva.eliezer@einstein.br

Tel.: 55 11 2151-1521

INTRODUCTION

Severe sepsis has been associated with an uncontrolled systemic inflammatory response to infection that is associated with developing organ dysfunction (1). Among these

dysfunctions, severe myopathy plays an important role in prolonged intensive care unit stays and late sequelae (2-6).

The main signs of sepsis-induced myopathy are generalized weakness, fatigue, atrophy (7), and a delay in weaning from mechanical ventilation (8). These symptoms are associated with a pronounced catabolic response in skeletal muscle that results from proteolytic stimulation and inhibited protein synthesis (9,10) (particularly in the myofibrillar proteins) and can negatively impact patient recovery (11).

Oxidative stress plays an important role in skeletal muscle dysfunction in different clinical settings, including sepsis (12,13). Reactive oxygen species (ROS) production is exacerbated in sepsis disease mainly because of mitochondrial

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

No potential conflict of interest was reported.

DOI: 10.6061/clinics/2013(01)OA17



dysfunction (13), which in turn promotes an imbalance in the redox status because the antioxidant defenses do not increase in the same proportion. This altered redox balance causes cell damage, particularly in proteins, lipids and DNA (14).

Because aerobic exercise training induces higher oxygen consumption in parallel with increased ROS production (15), an antioxidant system is launched during such training to maintain an adequate redox balance (16). In addition, exercise training improves mitochondrial function and even increases the number of muscle mitochondria (17,18). The net effects of training are improved efficiency of the antioxidant defenses and oxygen consumption, which could be useful for scavenging ROS in sepsis (16). Therefore, aerobic exercise training enhances the antioxidant defenses in various tissues, particularly in the skeletal and cardiac muscles (19).

As an ROS scavenger strategy, the impact of exercise training on sepsis-induced muscle damage has never been investigated (to the best of our knowledge). Therefore, it was hypothesized that aerobic exercise training before the presentation of a sepsis stimulus could attenuate skeletal muscle damage by modulating oxidative stress. Thus, the aim of this study was to evaluate the effects of aerobic physical preconditioning on the different mechanisms that are involved in sepsis-induced myopathy.

■ MATERIALS AND METHODS

Animals

Adult male Wistar rats (70 days old) were obtained from the UNIVALI (Universidade do Vale de Itajai, Itajai, Brazil) breeding colony. The rats were maintained in a 12/12-hr light-dark cycle in a temperature-controlled (22°C) environment with free access to standard laboratory chow (protein 20 kcal%, carbohydrates 70 kcal%, and lipids 10 kcal%, Nuvital Nutrientes, Curitiba, Paraná, Brazil) and tap water. Initially, the animals were randomly assigned to the untrained and trained groups. After completing the eight-week aerobic exercise training protocol, animals in the trained group were randomly assigned to either the sham or CLP (cecal ligation and perforation) surgery group. The untrained group was subjected to the same surgical procedures. At this phase, there were four groups: 1) sham trained (ShamT), 2) CLP trained (CLPT), 3) sham and 4) CLP. All of the rats were euthanized five days post-surgery. This study was conducted in accordance with the ethical principles in animal research adopted by the Brazilian College of Animal Experimentation (www.cobea.org.br) and was approved by the UNIVILLE (Universidade da Região de Joinville, Joinville, Brazil) Ethics Committee (Protocol No. 008/08 – COEA).

Graded treadmill exercise test

Before the first exercise test, the rats were conditioned to exercise on a treadmill over a week-long period (10 min of exercise per session). During the graded treadmill exercise test, the rats were placed on a treadmill and allowed to acclimatize for at least 15 min. The exercise intensity was then increased by 3 m/min (6-33 m/min) every three minutes at a 0% grade until exhaustion (the point of maximum running speed). The graded treadmill exercise test was performed prior to the exercise training and then during the fourth and eighth weeks of exercise training. Exercise capacity was estimated by the total distance run and correlated with the skeletal muscle capacity, which is a

method used to detect exercise intolerance. Thus, exercise capacity was evaluated using a graded treadmill exercise protocol, as previously described (20).

Aerobic exercise training protocol

The aerobic exercise training protocol consisted of an eight-week program of running on a motorized treadmill (KT-4000 model INBRAMED, RS, Brazil), five days a week, for 60 minutes per sessions at 60% of the maximum running speed that was obtained in the graded treadmill test, which corresponded to the maximal lactate steady-state workload (MLSSw), as described elsewhere (20). All of the untrained rats were exposed to treadmill exercises (5 minutes) three times per week to become accustomed to the exercise protocol and handling.

Cecal ligation and perforation surgery

The animals in both the untrained and trained groups were subjected to the surgical procedure 72 hours after the last treadmill exercise test, as previously described (21,22). For the CLP surgery, the rats were anesthetized with ketamine (80 mg/kg), which was administered intraperitoneally. Under aseptic conditions, a 3-cm midline laparotomy was performed to expose the cecum with the adjoining intestine. The cecum was tightly ligated with a 3.0-silk suture at its base (below the ileocecal valve) and was perforated once with a 14-gauge needle. The cecum was then gently squeezed to extrude a small amount of feces from the perforation site before being returned to the peritoneal cavity. The laparotomy was then closed with 3.0 silk sutures. All of the animals received isotonic saline solution (30 ml/kg subcutaneously) immediately after the CLP surgery, as well as antibiotics (ceftriaxone 30 mg/kg and clindamycin 25 mg/kg) starting 6 hr after the CLP surgery and again every 6 hr up to 24 hr after the CLP surgery. Afterward, the animals were returned to their cages with free access to food and water. In the sham group, the rats were submitted to all of the surgical procedures and received isotonic saline solution immediately after the surgery. They also received the same antibiotics as the CLP groups, but their ceca were not ligated or perforated.

To minimize the possibility that the animals did not truly develop sepsis, the CLP procedure was always performed by the same investigator. In addition, the animals were observed after the CLP surgeries for signs of infection, such as piloerection, lethargy, tachypnea, and weight loss (21,22). The animals were weighed daily for five days after the surgery, and their soleus and plantaris muscles were harvested at the time of euthanization for later analysis. The rate of sepsis survival in this model was approximately 40%.

Fiber CSA

The fiber cross-sectional area (CSA) in the soleus and plantaris muscles was used as an indicator of muscle atrophy. Liquid nitrogen-frozen muscles were vertically mounted and serially sectioned. The muscle sections were then incubated in alkali (mATPase, pH 10.3) or acid (mATPase, pH 4.6), as described by Brooke & Kaiser (23), to assess myofibrillar ATPase activity. The myosin ATPase reaction was used to identify the muscle fiber type (Figure 1). Fiber CSA was evaluated in five areas at 200× magnification and further analyzed using a digitizing unit connected to a computer (Image-Pro Plus; Media

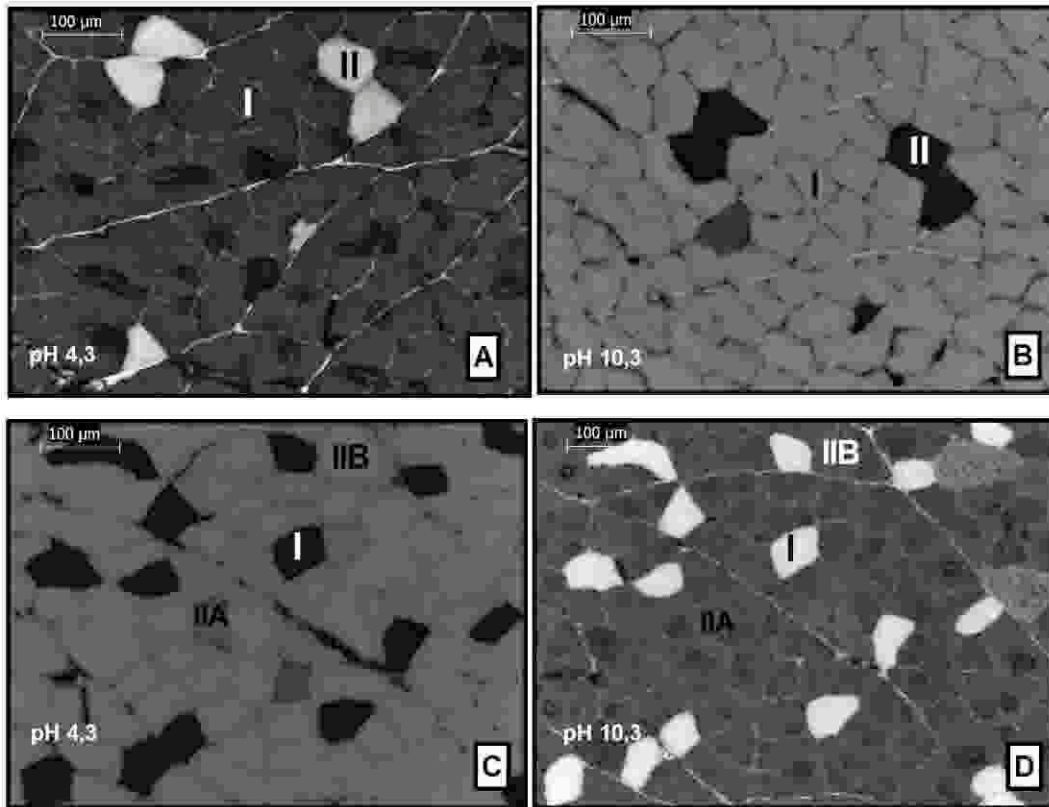


Figure 1 - Skeletal muscle histochemical staining for myosin ATPase. Fiber type I appears dark and type II is beige or white in the soleus (A) and plantaris (C) muscles at pH 4.6. The opposite is shown at pH 10.3 in the soleus (B) and plantaris (D) muscles.

Cybernetics, Silver Spring, MD). All of the analyses were conducted by a single observer (CWC) who was blinded to the rat group assignments.

CK activity

The creatine kinase (CK) activity was measured in the tissue homogenate of the soleus and plantaris muscles. The reaction mixture for the creatine kinase assay contained 100 mM Tris-HCl buffer (pH 7.5), 30 mM phosphocreatine, 20 mM glucose, 12 mM magnesium acetate, 10 mM diadenosine pentaphosphate, 15 mM sodium azide, 20 mM n-acetylcysteine, 2 mM ADP, 5 mM AMP, 2 mM NADP, 3500 U/L hexokinase, 2000 U/L glucose-6-phosphate dehydrogenase and approximately 1.5 mg of protein in a final volume of 1200 μ L. The creatine kinase activity was calculated based on reduced nicotinamide adenine dinucleotide (NADH) formation, which was monitored with a spectrophotometer at a wavelength of 340 nm at 37°C (24).

TBARS

Lipid peroxidation in the soleus and plantaris muscles was measured by the formation of thiobarbituric acid reactive species (TBARS) during an acid-heating reaction, as previously described (25). Briefly, the samples (200 μ L) were mixed with trichloroacetic acid (10%) (400 μ L) and centrifuged for 10 minutes (4000 \times g), and the supernatant was mixed with an equal volume of 0.67% thiobarbituric acid (TBA). The mixture was then heated in a boiling water bath for 15 minutes. The formation of TBARS was determined by measuring the absorbance at 535 nm.

Protein carbonyls

The oxidative damage to the protein in the soleus and plantaris muscles was measured by determining levels of the carbonyl groups based on the reaction of the groups with dinitrophenylhydrazine (DNPH) (26). The proteins were precipitated by adding 20% trichloroacetic acid and reacted with DNPH. The samples were then re-dissolved in 6 M guanidine hydrochloride, and the carbonyl contents were determined by measuring the absorbance at 370 nm.

CAT and SOD activity

To determine catalase (CAT) activity, the soleus and plantaris muscles were sonicated in a 50-mM phosphate buffer, and the resulting suspension was centrifuged at 3000 \times g for 10 minutes. The supernatant was used for the enzymatic assay. CAT activity was measured by the rate of decrease in hydrogen peroxide absorbency at 240 nm (27). Superoxide dismutase (SOD) activity was assayed by measuring the inhibition of adrenaline auto-oxidation, as previously described (28).

Statistical analysis

The data are presented as the means and standard errors of the means (mean \pm SEM). A two-way ANOVA with Fisher's post hoc test (Statistica software, StatSoft, Tulsa, OK) was used to compare the effects of training and surgery in all of the analyses except for the analyses of body weight and running distance, for which a repeated-measures two-way ANOVA with Fisher's post hoc test and Student's *t*-test, respectively, were used. Statistical significance was set at $p < 0.05$.



RESULTS

In the trained group, there was a significant increase in the running distance (375.3 ± 37.9 m) compared with the untrained group (175.3 ± 13.3 m) ($p < 0.05$).

The pre-training weights of the trained (333.9 ± 5.8 g) and untrained (332.8 ± 5.8 g) groups were similar. After eight weeks of aerobic exercise training, during the pre-surgery time interval, the trained groups (ShamT and CLPT) exhibited an 18.7-g increase in body weight compared with the untrained groups (Sham and CLP), but this increase was not statistically significant ($p = 0.11$). Seventy-two hours after surgery, the CLP group exhibited a significant decrease in body weight compared with their weight during the pre-surgery period, and the CLPT group exhibited a similar decrease after 96 hours (Figure 2).

The CSA of muscle fibers was smaller in the CLP group than in the other groups. Myosin ATPase staining of the skeletal muscle sections showed that in the soleus, only the type I fibers from the CLP group were smaller than those from the other groups ($p < 0.05$). In the plantaris muscle, only the type II fibers were smaller in the CLP group than in the other groups ($p < 0.05$) (Figure 3).

The soleus muscle CK activity was higher in the CLP group than in the sham and CLPT groups. However, the plantaris CK activity was higher in the CLP group than in the ShamT and CLPT groups (Figure 4).

In the soleus and plantaris muscles, TBARS formation was evaluated as an index of lipid peroxidation and carbonyl group levels were evaluated as an index of protein oxidation. As shown in Figures 5-A and 5-C, the TBARS levels in the soleus and plantaris muscles of the rats in the CLP group were significantly increased compared with those of the sham and CLPT groups ($p < 0.05$). Figures 5-B and 5-D demonstrate a greater increase in the carbonyl

levels of both muscle types in the CLP group compared with the levels in the sham and CLPT groups ($p < 0.05$).

Regulating antioxidant enzymes is important for maintaining the balance between ROS generation and consumption; therefore, the SOD and CAT activity levels were evaluated in the soleus and plantaris muscles. As shown in Figures 6A and 6C, the CLP group exhibited no difference in SOD activity compared with the sham group for both muscle types, whereas the CLPT group exhibited a significant increase in SOD activity in both muscle types compared with the sham and CLP groups ($p < 0.05$). However, there was a significant decrease in CAT activity in the CLP group for both muscle types compared with the sham group ($p < 0.05$). In contrast, the CAT activity in the soleus muscles of the CLPT group was similar to that of the sham group, although it differed from that of the CLP group in the plantaris muscle (Figures 6B and 6D).

DISCUSSION

The present study contributes valuable information regarding the effects of aerobic physical preconditioning on different mechanisms that are involved in sepsis-induced skeletal myopathy, including preventing atrophy, lowering lipid peroxidation and protein oxidation, and improving antioxidant defenses.

Several molecular mechanisms of inflammation and cellular damage are involved in the pathogenesis of sepsis, including the excessive generation of ROS (29). These key mediators of cellular injury greatly contribute to the development of sepsis-induced multiple organ dysfunction (29-31). The proinflammatory properties of ROS include endothelial cell damage, chemotactic factor formation, lipid peroxidation and oxidation, neutrophil recruitment, DNA damage, the release of interleukin (IL) and tumor necrosis

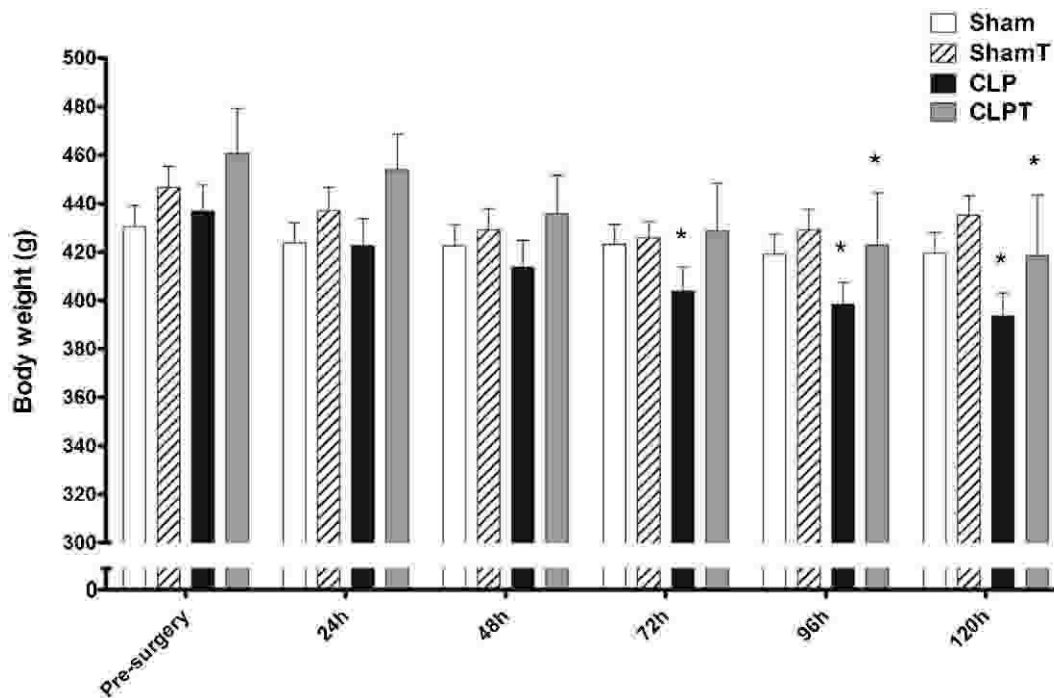


Figure 2 - Time course of the body weight changes from pre-surgery until 5 days after surgery in the sham (n=8), ShamT (n=8), CLP (n=6) and CLPT (n=6) groups. The data are presented as the means \pm SEM. * $p < 0.05$ vs. the same group pre-surgery.

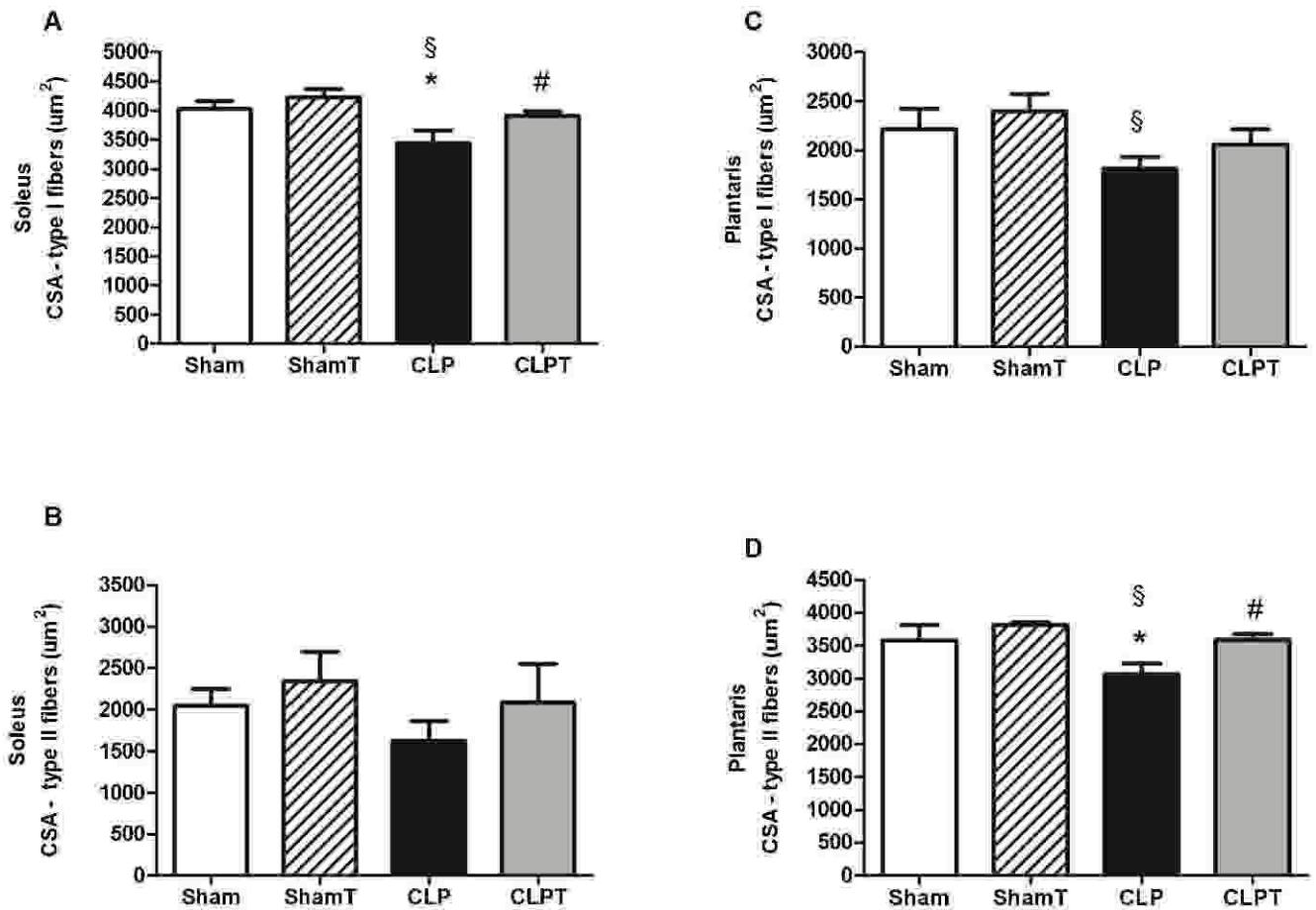


Figure 3 - Cross-sectional area (CSA) of type I and II fibers in the soleus (A and B) and plantaris (C and D) muscles at five days after surgery. The animals were divided into four groups: sham (n=5), ShamT (n=5), CLP (n=5) and CLPT (n=5). The data are presented as the means \pm SEM. *, $p < 0.05$ vs. sham; #, $p < 0.05$ vs. CLP; and §, $p < 0.05$ vs. ShamT.

factor (TNF α), and the formation of peroxynitrite (31). The results of the present study indicated that lipid peroxidation and protein oxidation increased in the oxidative and glycolytic skeletal muscles of rats in the untrained group (CLP) and that tissue damage could be attenuated through exercise training, as observed in the trained group (CLPT) (Figure 5).

In agreement with the results of the present study, some authors have demonstrated that septic patients produce a large amount of ROS without any corresponding antioxidant defenses, which is accompanied by lipid peroxidation (32). Note that one of the mechanisms causing this oxidative stress is the reduction of altered tetravalent oxygen as a consequence of endotoxic, hypoxic, and acidic conditions, which occur in sepsis (14). Thus, in skeletal muscle tissue, acidosis increases the quantity of free iron that is released from myoglobin and hemoglobin (33), thereby activating leukocytes and resulting in superoxide generation (34). Thus, the outcome is a vicious cycle that increases cellular dysfunction, promotes muscle wasting, and is associated with a more serious prognosis (11).

Septic patients frequently develop severe myopathy (2-7) that results in significant weight loss. In the present study, as expected, the CLP group exhibited significant decreases in body weight after 72 hours of sepsis, while the CLPT group exhibited this significant decrease only after 96 hours

(Figure 2). Importantly, 120 hours after sepsis induction, the skeletal muscle fiber CSA revealed atrophic conditions only in the soleus and plantaris muscles from the untrained CLP group (Figure 3); in addition, there was significant skeletal muscle injury, as observed by the increase in the TBARS and carbonyl activity (Figure 5). Additionally, there was a significant increase in skeletal muscle anaerobic metabolism in the soleus muscle and a trend toward this increase in the plantaris muscles from rats in the CLP group, as demonstrated by the increase in CK activity (Figure 4). However, in the rats submitted to the aerobic exercise protocol before sepsis induction (CLPT group), this early weight loss was prevented, and skeletal muscle fiber CSA and regular levels of CK, lipid peroxidation and protein oxidation were maintained. These results suggest that the training effects, such as maintaining muscle mass and improving the enzymatic antioxidant system, could be preserved even after 120 hours of sepsis.

It has been shown that exercise training, particularly aerobic exercise, protects skeletal muscles against a variety of stressors, including oxidative stress (15,35). In this context, the elevation of several cytoprotective proteins occurs as a consequence of exercise. For instance, the antioxidant enzymes SOD, CAT and glutathione peroxidase (GPX) in skeletal muscles were shown to be up-regulated (19,36,37), and these enzymes were subsequently able to

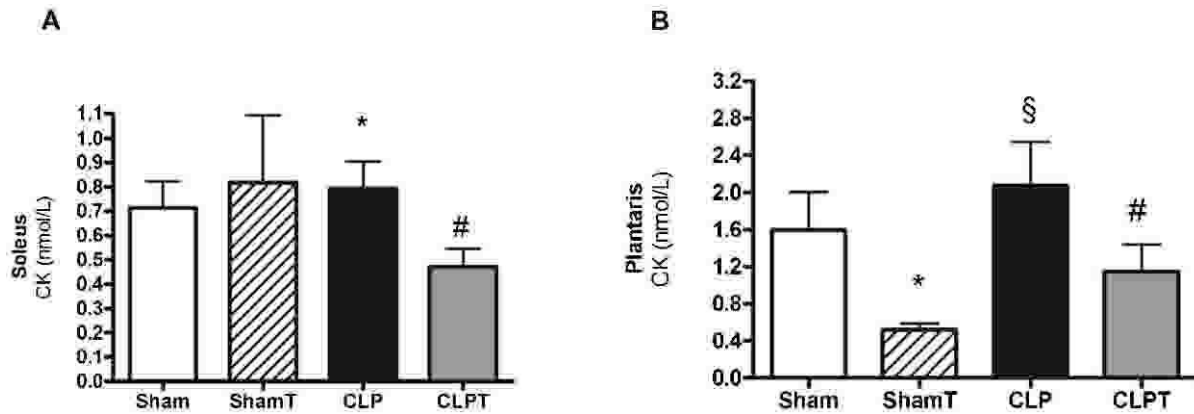


Figure 4 - The creatine kinase (CK) activity. **(A)** The soleus and **(B)** plantaris muscles five days after surgery. The animals were divided into four groups: sham (n=8), ShamT (n=8), CLP (n=6) and CLPT (n=6). The data are presented as the means \pm SEM. *, $p < 0.05$ vs. sham; #, $p < 0.05$ vs. CLP; and §, $p < 0.05$ vs. ShamT.

protect skeletal muscle against ROS-mediated damage (15,35).

The impact of aerobic physical preconditioning on sepsis has not been fully evaluated. Some authors have suggested that 12-wk treadmill training maintains the integrity of the beta-adrenergic receptor adenylate cyclase system, which can be depressed by *in vivo* endotoxin administration (38). Moreover, Bagby et al. demonstrated that exercise to near exhaustion before intravenous LPS challenge in rats markedly suppressed the systemic TNF response that is normally observed in response to LPS challenge (39). Some studies have shown that appropriate exercise in humans

increases anti-inflammatory cytokine IL-6 plasma levels and suppresses the endotoxemia-induced elevation in TNF α (40). For instance, Chen et al. reported that four weeks of exercise training before a sepsis challenge attenuated sepsis-induced systemic hypotension and tachycardia, decreased the number of blood cells, increased the levels of proinflammatory cytokines (TNF α and IL-1 β); and, also protected organs from sepsis damage (41). However, there are conflicting studies regarding the effects of transcutaneous electrical muscle stimulation (TEMS) on the quadriceps muscle in septic patients. Gerovasili et al. demonstrated that TEMS can preserve muscle mass; however, Poulsen et al.

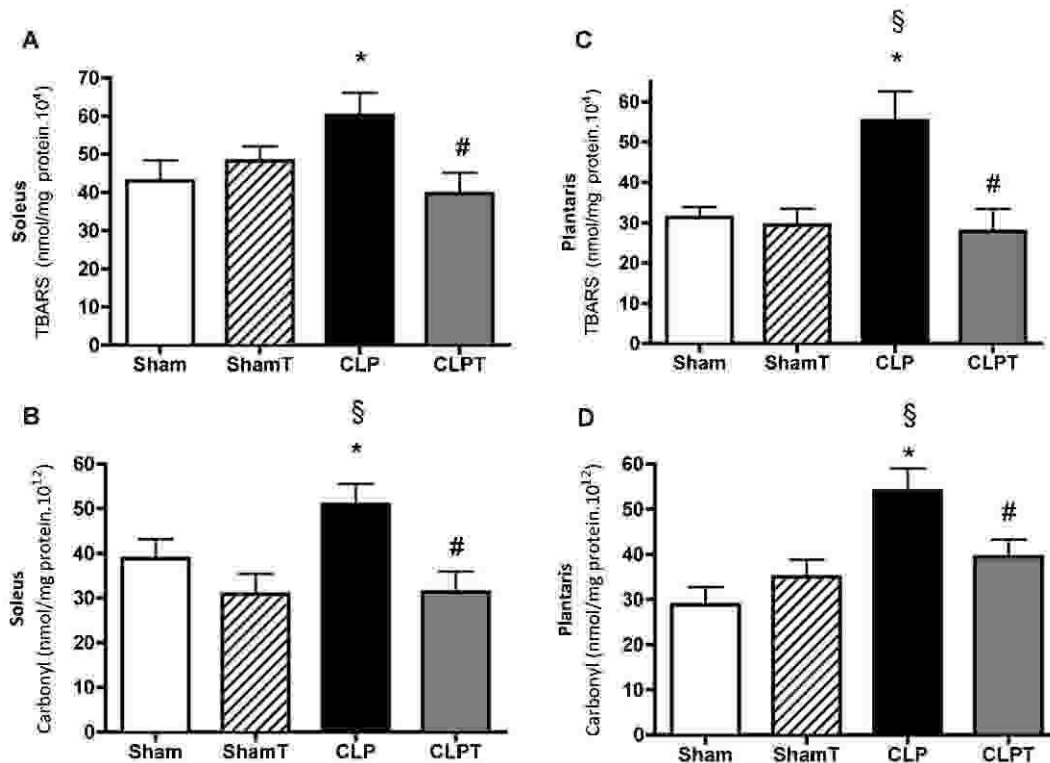


Figure 5 - Oxidative stress parameters. **(A, C)** Thiobarbituric acid reactive substances (TBARS) and **(B, D)** carbonyl in the soleus and plantaris muscles at five days after surgery. The animals were divided into four groups: sham (n=8), ShamT (n=8), CLP (n=6) and CLPT (n=6). The data are presented as the means \pm SEM. *, $p < 0.05$ vs. sham; #, $p < 0.05$ vs. CLP; and §, $p < 0.05$ vs. ShamT.

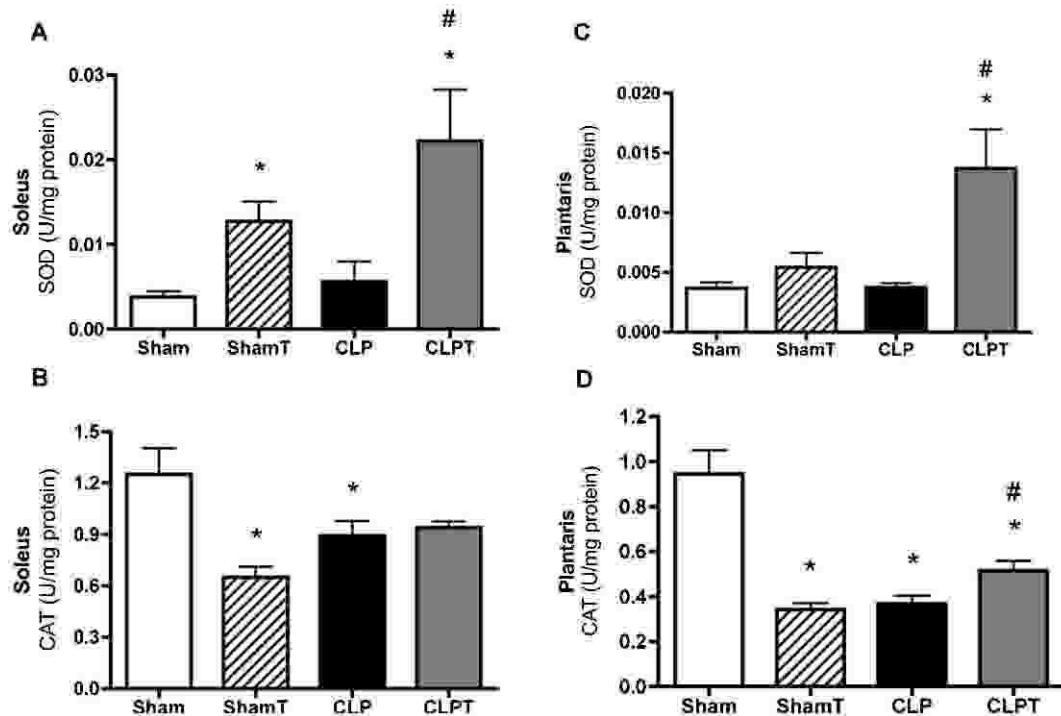


Figure 6 - Antioxidant enzymes. (A, C) Superoxide dismutase (SOD) and (B, D) catalase (CAT) in the soleus and plantaris muscles at five days after surgery. The animals were divided into four groups: sham (n=8), ShamT (n=8), CLP (n=6) and CLPT (n=6). The data are presented as the means \pm SEM. *, $p < 0.05$ vs. sham; #, $p < 0.05$ vs. CLP; and § $p < 0.05$ vs. ShamT.

concluded that the loss of muscle mass was unaffected by TEMS (42,43).

Remarkably, the present study demonstrated that aerobic exercise training was able to prevent atrophy, oxidative stress and muscle damage in skeletal muscle in a rat sepsis model (Figures 3 and 5). The exercise-induced increase in the antioxidant defense system of muscles is most likely the underlying mechanism responsible for protecting muscle cells against oxidative damage. Regarding the removal of ROS from muscle fibers, SOD activity is the first-line of defense against superoxide radicals (15). Subsequently, the present findings show that the ShamT and CLPT groups exhibited significant increases in SOD activity compared with the sham and CLP groups (Figure 6). Other researchers have shown that SOD expression can be induced in skeletal muscle by exercise, which can provide cellular protection against ROS (19,44). For instance, Pinho et al. investigated the effect of 12 weeks of treadmill exercise on the skeletal muscle of rats, and they found that aerobic exercise could increase the SOD/CAT ratio (45).

The results of the present study suggest that exercise increased SOD activity and could be a contributory factor in preventing sepsis-related myocyte injury. CAT, another cellular antioxidant enzyme, is responsible for the catalytic reduction of hydrogen peroxide to water. In the present study, CAT activity levels did not decrease in the CLPT group, whereas a significant decrease in these levels was found in the CLP group compared with the sham group (Figure 6). The trained groups did not exhibit an increase in CAT activity levels, which could be associated with reduced hydrogen peroxide levels due to increased GPX enzyme activity. Moreover, in this study, the CAT activity data from the trained group was in accordance with the results of the

study by Powers et al., who showed that there was no change in oxidative and glycolytic muscle CAT activity in response to chronic exercise training (19). Thus, one limitation of the present study is that the exact mechanism that is involved in preventing lipid peroxidation and protein oxidation could not be clarified.

In conclusion, aerobic physical preconditioning prevents atrophy, lipid peroxidation and protein oxidation and improves SOD activity in the skeletal muscle of septic rats. Furthermore, this study suggested that regular exercise may minimize the muscle damage that results from serious bacterial infections.

■ ACKNOWLEDGMENTS

We thank Hospital Israelita Albert Einstein for covering the publication fees.

■ AUTHOR CONTRIBUTIONS

Coelho CW, Westphal GA, Dal-Pizzol F, Streck EE, and Silva E conceived and designed the study, conducted the experiment, performed the data analysis, reviewed the drafts of the manuscript, and approved the final version of the manuscript. Coelho CW, Jannig PR, Souza AB, and Fronza Jr H conducted the experiment and muscle morphological analysis, reviewed the drafts of the manuscript, and approved the final version of the manuscript. Petronilho F, Constantino L, and Ferreira GK conducted the biochemical analyses, reviewed the drafts of the manuscript, and approved the final version of the manuscript.

■ REFERENCES

1. Bone RC, Balk RA, Cerra FB, Dellinger RP, Fein AM, Knaus WA, et al. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. The ACCP/SCCM Consensus Conference Committee. American College of Chest Physicians/Society of Critical Care Medicine. 1992. *Chest*. 2009;136(5 Suppl):e28.
2. Garnacho-Montero J, Madrazo-Osuna J, Garcia-Garmendia JL, Ortiz-Leyba C, Jimenez-Jimenez FJ, Barrero-Almodovar A, et al. Critical illness



polyneuropathy: risk factors and clinical consequences. A cohort study in septic patients. *Intensive Care Med.* 2001;27(8):1288-96.

3. Hund E. Neurological complications of sepsis: critical illness polyneuropathy and myopathy. *J Neurol.* 2001;248(11):929-34.
4. De Jonghe B, Sharshar T, Lefaucheur JP, Authier FJ, Durand-Zaleski I, Boussarsar M, et al. Paresis acquired in the intensive care unit: a prospective multicenter study. *JAMA.* 2002;288(22):2859-67, <http://dx.doi.org/10.1001/jama.288.22.2859>.
5. Fredriksson K, Hammarqvist F, Strigard K, Hultenby K, Ljungqvist O, Wernerer J, et al. Derangements in mitochondrial metabolism in intercostal and leg muscle of critically ill patients with sepsis-induced multiple organ failure. *Am J Physiol Endocrinol Metab.* 2006;291(5):E1044-50, <http://dx.doi.org/10.1152/ajpendo.00218.2006>.
6. Fredriksson K, Tjader I, Keller P, Petrovic N, Ahlman B, Scheele C, et al. Dysregulation of mitochondrial dynamics and the muscle transcriptome in ICU patients suffering from sepsis induced multiple organ failure. *PLoS one.* 2008;3(11):e3686, <http://dx.doi.org/10.1371/journal.pone.0003686>.
7. Deem S. Intensive-care-unit-acquired muscle weakness. *Respir Care.* 2006;51(9):1042-52; discussion 52-3.
8. Reid WD, MacGowan NA. Respiratory muscle injury in animal models and humans. *Mol Cell Biochem.* 1998;179(1-2):63-80, <http://dx.doi.org/10.1023/A:1006803703128>.
9. Kazi AA, Pruznak AM, Frost RA, Lang CH. Sepsis-induced alterations in protein-protein interactions within mTOR complex 1 and the modulating effect of leucine on muscle protein synthesis. *Shock.* 2011;35(2):117-25, <http://dx.doi.org/10.1097/SHK.0b013e3181ecb57c>.
10. Cooney RN, Kimball SR, Vary TC. Regulation of skeletal muscle protein turnover during sepsis: mechanisms and mediators. *Shock.* 1997;7(1):1-16, <http://dx.doi.org/10.1097/00024382-199701000-00001>.
11. Hasselgren PO, Menconi MJ, Fareed MU, Yang H, Wei W, Evenson A. Novel aspects on the regulation of muscle wasting in sepsis. *Int J Biochem Cell Biol.* 2005;37(10):2156-68, <http://dx.doi.org/10.1016/j.biocel.2005.01.017>.
12. Brealey D, Karyampudi S, Jacques TS, Novelli M, Stidwill R, Taylor V, et al. Mitochondrial dysfunction in a long-term rodent model of sepsis and organ failure. *Am J Physiol Regul Integr Comp Physiol.* 2004;286(3):R491-7, <http://dx.doi.org/10.1152/ajpregu.00432.2003>.
13. Crouser ED, Julian MW, Blaho DV, Pfeiffer DR. Endotoxin-induced mitochondrial damage correlates with impaired respiratory activity. *Crit Care Med.* 2002;30(2):276-84, <http://dx.doi.org/10.1097/00003246-200202000-00002>.
14. Dal-Pizzol F, Ritter C, Cassol-Jr OJ, Rezin GT, Petronilho F, Zugno AI, et al. Oxidative mechanisms of brain dysfunction during sepsis. *Neurochem Res.* 2010;35(1):1-12, <http://dx.doi.org/10.1007/s11064-009-0043-4>.
15. Powers SK, Jackson MJ. Exercise-induced oxidative stress: cellular mechanisms and impact on muscle force production. *Physiol Rev.* 2008;88(4):1243-76, <http://dx.doi.org/10.1152/physrev.00031.2007>.
16. Smuder AJ, Kavazis AN, Min K, Powers SK. Exercise protects against doxorubicin-induced oxidative stress and proteolysis in skeletal muscle. *J Appl Physiol.* 2011;110(4):935-42, <http://dx.doi.org/10.1152/jappphysiol.00677.2010>.
17. Boveris A, Navarro A. Systemic and mitochondrial adaptive responses to moderate exercise in rodents. *Free Radic Biol Med.* 2008;44(2):224-9, <http://dx.doi.org/10.1016/j.freeradbiomed.2007.08.015>.
18. Ljubicic V, Joseph AM, Saleem A, Uguccioni G, Collu-Marchese M, Lai RY, et al. Transcriptional and post-transcriptional regulation of mitochondrial biogenesis in skeletal muscle: effects of exercise and aging. *Biochim Biophys Acta.* 2010;1800(3):223-34, <http://dx.doi.org/10.1016/j.bbagen.2009.07.031>.
19. Powers SK, Criswell D, Lawler J, Ji LL, Martin D, Herb RA, et al. Influence of exercise and fiber type on antioxidant enzyme activity in rat skeletal muscle. *Am J Physiol.* 1994;266(2 Pt 2):R375-80.
20. Ferreira JC, Rolim NP, Bartholomeu JB, Gobatto CA, Kokubun E, Brum PC. Maximal lactate steady state in running mice: effect of exercise training. *Clin Exp Pharmacol Physiol.* 2007;34(8):760-5, <http://dx.doi.org/10.1111/j.1440-1681.2007.04635.x>.
21. Ritter C, Andrades M, Frota Junior ML, Bonatto F, Pinho RA, Polydoro M, et al. Oxidative parameters and mortality in sepsis induced by cecal ligation and perforation. *Intensive Care Med.* 2003;29(10):1782-9, <http://dx.doi.org/10.1007/s00134-003-1789-9>.
22. Ritter C, Andrades ME, Reinke A, Menna-Barreto S, Moreira JC, Dal-Pizzol F. Treatment with N-acetylcysteine plus deferoxamine protects rats against oxidative stress and improves survival in sepsis. *Crit Care Med.* 2004;32(2):342-9, <http://dx.doi.org/10.1097/01.CCM.0000109454.13145.CA>.
23. Brooke MH, Kaiser KK. Muscle fiber types: how many and what kind? *Archives of neurology.* 1970;23(4):369-79, <http://dx.doi.org/10.1001/archneur.1970.00480280083010>.
24. Oliver IT. A spectrophotometric method for the determination of creatine phosphokinase and myokinase. *The Biochemical journal.* 1955;61(1):116-22.
25. Draper HH, Hadley M. Malondialdehyde determination as index of lipid peroxidation. *Methods Enzymol.* 1990;186:421-31, [http://dx.doi.org/10.1016/0076-6879\(90\)86135-I](http://dx.doi.org/10.1016/0076-6879(90)86135-I).
26. Levine RL, Garland D, Oliver CN, Amici A, Climent I, Lenz AG, et al. Determination of carbonyl content in oxidatively modified proteins. *Methods Enzymol.* 1990;186:464-78, [http://dx.doi.org/10.1016/0076-6879\(90\)86141-H](http://dx.doi.org/10.1016/0076-6879(90)86141-H).
27. Aebi H. Catalase in vitro. *Methods Enzymol.* 1984;105:121-6, [http://dx.doi.org/10.1016/S0076-6879\(84\)05016-3](http://dx.doi.org/10.1016/S0076-6879(84)05016-3).
28. Bannister JV, Calabrese L. Assays for superoxide dismutase. *Methods Biochem Anal.* 1987;32:279-312, <http://dx.doi.org/10.1002/9780470110539.ch5>.
29. Salvemini D, Cuzzocrea S. Oxidative stress in septic shock and disseminated intravascular coagulation. *Free Radic Biol Med.* 2002;33(9):1173-85, [http://dx.doi.org/10.1016/S0891-5849\(02\)00961-9](http://dx.doi.org/10.1016/S0891-5849(02)00961-9).
30. Kozlov AV, Szalay L, Umar F, Fink B, Kropik K, Nohl H, et al. Epr analysis reveals three tissues responding to endotoxin by increased formation of reactive oxygen and nitrogen species. *Free Radic Biol Med.* 2003;34(12):1555-62, [http://dx.doi.org/10.1016/S0891-5849\(03\)00179-5](http://dx.doi.org/10.1016/S0891-5849(03)00179-5).
31. Zimmerman JJ. Defining the role of oxyradicals in the pathogenesis of sepsis. *Crit Care Med.* 1995;23(4):616-7, <http://dx.doi.org/10.1097/00003246-199504000-00003>.
32. Bayir H. Reactive oxygen species. *Crit Care Med.* 2005;33(12 Suppl):S498-501, <http://dx.doi.org/10.1097/01.CCM.0000186787.64500.12>.
33. Linares E, Nakao LS, Augusto O, Kadiiska MB. EPR studies of in vivo radical production by lipopolysaccharide: potential role of iron mobilized from iron-nitrosyl complexes. *Free Radic Biol Med.* 2003;34(6):766-73, [http://dx.doi.org/10.1016/S0891-5849\(02\)01424-7](http://dx.doi.org/10.1016/S0891-5849(02)01424-7).
34. Hotchkiss RS, Karl IE. The pathophysiology and treatment of sepsis. *N Engl J Med.* 2003;348(2):138-50.
35. Noble EG, Milne KJ, Melling CW. Heat shock proteins and exercise: a primer. *Appl Physiol Nutr Metab.* 2008;33(5):1050-65, <http://dx.doi.org/10.1139/H08-069>.
36. McArdle A, Pattwell D, Vasilaki A, Griffiths RD, Jackson MJ. Contractile activity-induced oxidative stress: cellular origin and adaptive responses. *Am J Physiol Cell Physiol.* 2001;280(3):C621-7.
37. Jackson MJ. Redox regulation of skeletal muscle. *IUBMB Life.* 2008;60(8):497-501, <http://dx.doi.org/10.1002/iub.72>.
38. DeBlieux PM, McDonough KH, Barbee RW, Shepherd RE. Exercise training attenuates the myocardial dysfunction induced by endotoxin. *J Appl Physiol.* 1989;66(6):2805-10.
39. Bagby GJ, Sawaya DE, Crouch LD, Shepherd RE. Prior exercise suppresses the plasma tumor necrosis factor response to bacterial lipopolysaccharide. *J Appl Physiol.* 1994;77(3):1542-7.
40. Starkie R, Ostrowski SR, Jauffred S, Febbraio M, Pedersen BK. Exercise and IL-6 infusion inhibit endotoxin-induced TNF-alpha production in humans. *Faseb J.* 2003;17(8):884-6.
41. Chen HI, Hsieh SY, Yang FL, Hsu YH, Lin CC. Exercise training attenuates septic responses in conscious rats. *Med Sci Sports Exerc.* 2007;39(3):435-42.
42. Gerovasilis V, Stefanidis K, Vitzilaios K, Karatzanos E, Politis P, Koroneos A, et al. Electrical muscle stimulation preserves the muscle mass of critically ill patients: a randomized study. *Critical Care.* 2009;13(5):R161, <http://dx.doi.org/10.1186/cc8123>.
43. Poulsen JB, Moller K, Jensen CV, Weisdorf S, Kehlet H, Perner A. Effect of transcutaneous electrical muscle stimulation on muscle volume in patients with septic shock. *Crit Care Med.* 2011;39(3):456-61, <http://dx.doi.org/10.1097/CCM.0b013e318205c7bc>.
44. Leeuwenburgh C, Hollander J, Leichtweis S, Griffiths M, Gore M, Ji LL. Adaptations of glutathione antioxidant system to endurance training are tissue and muscle fiber specific. *Am J Physiol.* 1997;272(1 Pt 2):R363-9.
45. Pinho RA, Andrades ME, Oliveira MR, Pirola AC, Zago MS, Silveira PC, et al. Imbalance in SOD/CAT activities in rat skeletal muscles submitted to treadmill training exercise. *Cell Biol Int.* 2006;30(10):848-53, <http://dx.doi.org/10.1016/j.cellbi.2006.03.011>.