

## REVIEW ARTICLE

## Are conjugated linolenic acid isomers an alternative to conjugated linoleic acid isomers in obesity prevention?☆

Jonatan Miranda<sup>a,b,\*</sup>, Noemi Arias<sup>a,b</sup>, Alfredo Fernández-Quintela<sup>a,b</sup>,  
María del Puy Portillo<sup>a,b</sup>

<sup>a</sup> Grupo Nutrición y Obesidad, Departamento de Farmacia y Ciencias de los Alimentos, Facultad de Farmacia, Universidad del País Vasco, Vitoria, Spain

<sup>b</sup> CIBERobn, Fisiopatología de la Obesidad y Nutrición, Instituto de Salud Carlos III, Madrid, Spain

Received 21 December 2012; accepted 14 April 2013

Available online 14 April 2014

### KEYWORDS

Conjugated linolenic acid;  
Obesity;  
Conjugated linoleic acid;  
Functional ingredients

### PALABRAS CLAVE

Ácido linolénico conjugado;  
Obesidad;  
Ácido linoleico conjugado;  
Ingredientes funcionales

**Abstract** Despite its benefits, conjugated linoleic acid (CLA) may cause side effects after long-term administration. Because of this and the controversial efficacy of CLA in humans, alternative biomolecules that may be used as functional ingredients have been studied in recent years. Thus, conjugated linolenic acid (CLNA) has been reported to be a potential anti-obesity molecule which may have additional positive effects related to obesity.

According to the results reported in obesity, CLNA needs to be given at higher doses than CLA to be effective. However, because of the few studies conducted so far, it is still difficult to reach clear conclusions about the potential use of these CLNAs in obesity and its related changes (insulin resistance, dyslipidemia, or inflammation).

© 2012 SEEN. Published by Elsevier España, S.L. All rights reserved.

### ¿Son los isómeros del ácido linolénico conjugado una alternativa a isómeros del ácido linoleico conjugado en la prevención de la obesidad?

**Resumen** Pese a sus efectos beneficiosos, se desconoce si el ácido linoleico conjugado (conjugated linoleic acid, CLA) podría producir efectos adversos al ser administrado de forma crónica. Considerando este hecho y dada la controvertida eficacia del CLA en humanos, en los últimos años el ácido linolénico conjugado (CLNA, *conjugated linolenic acid*) se ha descrito como alternativa al CLA, con un potencial funcional para la prevención de la obesidad, además de tener otros efectos positivos relacionados con la misma.

A la vista de los resultados descritos, en lo que respecta a la obesidad, no parece que el CLNA sea una molécula más prometedora que el CLA, dado que el efecto generalmente tiene lugar

☆ Please cite this article as: Miranda J, Arias N, Fernández-Quintela A, del Puy Portillo M. ¿Son los isómeros del ácido linolénico conjugado una alternativa a isómeros del ácido linoleico conjugado en la prevención de la obesidad?. Endocrinol Nutr. 2014;61:209–219.

\* Corresponding author.

E-mail address: [jonatan.miranda@ehu.es](mailto:jonatan.miranda@ehu.es) (J. Miranda).

a dosis más elevadas que las dosis efectivas de CLA. No obstante, dado el escaso número de estudios realizados hasta la fecha, todavía resulta difícil llegar a conclusiones claras acerca del potencial uso de estos CLNA en obesidad y alteraciones relacionadas con ella (resistencia a la insulina, dislipidemias o inflamación).

© 2012 SEEN. Publicado por Elsevier España, S.L. Todos los derechos reservados.

## Introduction

The general term conjugated fatty acids refers to positional and geometric isomers of fatty acids with double conjugated bonds. Conjugated linoleic acid isomers, grouped under the acronym CLA (conjugated linoleic acid), are the most widely studied to date. Many physiological benefits have been attributed to this group of fatty acids (anticancer and antiatherogenic effects, immune function improvement, decreased fat accumulation, decreased inflammation, and increased muscle mass). These effects have been associated with the double conjugated bond and are commonly due to the individual actions of its two most abundant isomers, *cis-9,trans-11*, and *trans-10,cis-12*.

It should be noted that, despite its beneficial effects, CLA may have adverse effects. Although few studies have analyzed this potential toxicity, several authors have reported in experimental animals effects such as insulin resistance, increased C-reactive protein levels, or hepatic steatosis.<sup>1,2</sup> However, the European Food Safety Authority (EFSA) demonstrated in 2010 that, in humans with normal weight or overweight with no history of diabetes, the administration of an equimolecular mixture of the *cis-9,trans-11* and *trans-10, -12* isomers of CLA at a dose of 3.5 g for six months had no adverse effects on insulin sensitivity levels and blood glucose control or liver function.<sup>3</sup> Further studies are, however, needed to ascertain its long term effects, as well as its safety in patients with type 2 diabetes. In fact, as concluded by the EFSA expert panel, in these specific patients, the equimolecular mixture of CLA isomers appeared to negatively affect dynamic (ISI, OGIS) and static (HOMA-IR) markers of insulin sensitivity and increased some subclinical inflammation markers (15-keto-dihydroprostaglandin F2 and C-reactive protein).

Multiple studies have shown that CLA, and more specifically the *cis-9,trans-11* and *trans-10,cis-12* isomers, have benefits in different animal species (mice, rats, hamsters, pigs).<sup>4,5</sup> By contrast, the effectiveness of CLA in humans is more controversial (Table 1).<sup>6-15</sup> As an example, it may be noted that while several authors found effects such as decreased fat accumulation, others did not.<sup>6-9</sup> Moreover, when this effect was reported, it was less prominent than in rodents.

Because of the controversial efficacy of CLA in humans and the lack of reliable information on its effects after long-term administration (longer than six months), it appears appropriate to search for alternative biomolecules that may be used as functional ingredients for the prevention of obesity. One such biomolecule, conjugated linolenic acid (CLNA), has been reported to be a molecule able to decrease body fat, as well as having other positive effects on health.

Actually, CLNA is a collective term for a group of positional and geometric isomers of linolenic acid (C18:3) in which at least two double bonds are conjugated (contiguous), rather than separated by methylene groups, as occurs in linolenic acid (Fig. 1).<sup>16</sup> These isomers only differ from CLA isomers in that they have a third double bond.

The presence of three double bonds in the linolenic acid molecule leads to multiple positional and geometric isomers. It should be noted that studies by Bassaganya-Riera et al. suggest that the presence of double conjugated bonds in fatty acids increases their biological activity and enables them to act as agonists of nuclear receptors.<sup>17</sup> More specifically, in vitro studies have suggested that certain plant-derived CLNA isomers (such as punicic acid or  $\alpha$ -eleostearic acid) may act as natural agonists of peroxisome proliferator-activated receptors (PPAR).<sup>18,19</sup>

While CLA naturally occurs in food from ruminants (mainly cattle, sheep, and goats), and in milk and dairy products in very small amounts which account for up to 0.65% of all fatty acids,<sup>20</sup> CLNA isomers occur in greater amounts in plant-derived food and products. As documented by Takagi and Itabashi (1981),<sup>21</sup> Tung (*Vernicia fordii*) and bitter melon (*Momordica charantia*) seed oils contain 67.7% and 56.2% of  $\alpha$ -eleostearic acid (*cis-9,trans-11,trans-13* CLNA) respectively, while those extracted from pomegranate (*Punica granatum*), catalpa (*Catalpa ovata*), and pot marigold (*Calendula officinalis*) contain 83% of punicic acid (*cis-9,trans-11,cis-13* CLNA), 42.3% of catalpic acid (*trans-9,trans-11,cis-13* CLNA), and 62.2% of calendic acid (*trans-8,trans-10,cis-12* CLNA) respectively (Table 2).

## Effects of conjugated linolenic acid isomers on obesity

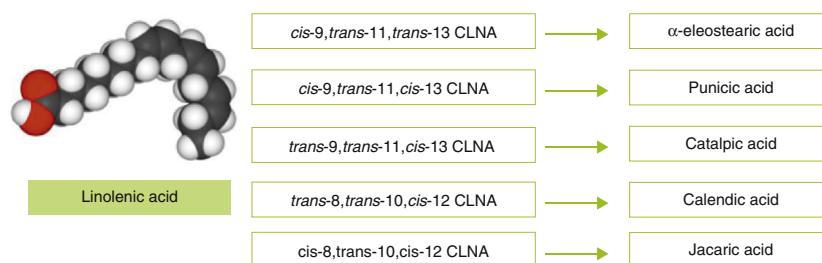
Since, as previously noted, the efficacy of conjugated linoleic acid as an anti-obesity molecule in human beings continues to be controversial, some scientific studies have started to focus on the search for CLNA isomers which can prevent this condition (Table 3).

In vitro studies conducted on 3T3-L1 adipocytes and HepG2 cells have demonstrated the activating effect of punicic acid and  $\alpha$ -eleostearic acid on nuclear receptors PPAR $\alpha$  and PPAR $\gamma$ .<sup>17-19</sup> Research conducted by our group found similar effects with a mixture of CLNA isomers *cis-9,trans-11,cis-15*, and *cis-9,trans-13,cis-15* in HEK293 cells. These isomers were able to activate the PPAR response element in cells overexpressing the PPAR $\alpha$  protein.<sup>22</sup> These results suggest that CLNA isomers may increase glucose tolerance and fatty acid oxidation, and decrease inflammation related to obesity.

**Table 1** Effects of CLA in humans.

Isomer	Treatment time/dose	Effect	Author	Study model
<i>Effects on fat reduction in obese humans</i>				
<i>cis-9,trans-11-CLA</i>	12 weeks; 3 g CLA/d	Significant body weight increase	Risérus et al. (2004) <sup>6</sup>	25M
Mixture of CLA isomers (Clarinol®)	6 months; 3.4 g CLA/d	Fat mass decrease by 1% at 3 months and 3.4% at 6 months	Gaullier et al. (2007) <sup>7</sup>	93F/93M
Mixture of CLA isomers	16 weeks; 6.4 g CLA/d	BMI and fat mass reduction	Norris et al. (2009) <sup>8</sup>	35F
CLA-enriched meat and dairy products from grazing livestock	56 days; 1.17 g CLA/d	No effect on body weight and fat and muscle mass	Brown et al. (2011) <sup>9</sup>	18F
<i>Effects on lipid levels</i>				
Mixture of CLA isomers	8 weeks; 0.7 g CLA/d: weeks 1–4 and 1.4 g CLA/d: weeks 5–8	Significant decrease in HDL cholesterol	Mougios et al. (2001) <sup>10</sup>	13M/9F
Mixture of CLA isomers or 80:20 <i>cis-9,trans-11</i> and <i>trans-10,cis-12-CLA</i>	8 weeks; 3 g 50:50 CLA/d or 3 g 80:20 CLA/d	TG decrease with the 50:50 mixture and VLDL decrease with the 80:20 mixture	Noone et al. (2002) <sup>11</sup>	18M/33F
CLA-enriched meat and dairy products from grazing livestock	56 days; 1.17 g CLA/d	No changes in TC, TG, HDL, LDL, VLDL, or IDL	Brown et al. (2011) <sup>9</sup>	18F
<i>Effects on insulin resistance and diabetes</i>				
Mixture of CLA isomers or <i>trans-10,cis-12-CLA</i> <i>cis-9,trans-12-CLA</i>	12 weeks; 3.4 g CLA/d or <i>t-10,c12-CLA</i> /d	<i>t10,c12-CLA</i> : increased insulin resistance and blood glucose	Risérus et al. (2002) <sup>12</sup>	57M
	12 weeks; 3 g CLA/d	<i>c9,t11-CLA</i> : increased insulin resistance in obese males	Risérus et al. (2004) <sup>6</sup>	25M
Mixture of CLA isomers	8 weeks; 4 g CLA/d	Insulin resistance improvement through decreased fasting insulin release	Eyjolfson et al. (2004) <sup>13</sup>	4M/12F
<i>Effects on inflammation</i>				
<i>cis-9,trans-11-CLA</i> or <i>trans-10,cis-12-CLA</i>	13 weeks; 3 g <i>c9,t11-CLA</i> or <i>t10,c12-CLA</i> /d	No changes in CRP, IL-6, IL-8, and TNF $\alpha$	Ramakers et al. (2005) <sup>14</sup>	38M/38F
Mixture of CLA isomers	12 weeks; 3 g CLA/d	Decreases in pro-inflammatory cytokines, TNF $\alpha$ , and IL-1 $\beta$ . Increase in anti-inflammatory cytokine: IL-10	Song et al. (2005) <sup>15</sup>	8M/20F

TC: total cholesterol; H: males; IL: interleukin; BMI: body mass index; F: females; mixture of CLA isomers; 50:50 *cis-9, trans-11-*, and *trans-10, cis-12-CLA*; TG: triglycerides, TNF: tumor necrosis factor.



**Figure 1** Chemical structure of linolenic acid and several of its most widely studied conjugated isomers. CLNA, conjugated linolenic acid.

Lai et al.<sup>23</sup> recently showed a decreased expression of genes controlling the differentiation process, such as PPAR $\gamma$  and C/EBP $\beta$ , and fatty acid synthase, an enzyme that increases triglyceride accumulation in 3T3-L1 preadipocytes treated with puniceic acid. The abovementioned study of our research group allowed us to propose another mechanism of action for CLNA isomers. Specifically, different doses (10–100  $\mu$ M) of the CLNA isomers *cis*-9,*trans*-11,*cis*-15 and *cis*-9,*trans*-13,*cis*-15 have been shown to decrease triglyceride contents in 3T3-L1 adipocytes, increasing the gene expression of key enzymes in lipolysis.<sup>22</sup>

In a study conducted to compare the effect of CLA and CLNA in rats, Koba et al.<sup>24</sup> noted that the addition of a mixture of unidentified CLNA isomers at a 1% concentration in diet induced a reduction in adipose tissue weight even greater than that caused by CLA. Similarly, CLNA intake had a much greater effect than CLA on lipid metabolism in the liver. Specifically, CLNA increased to a greater extent the oxidation of both mitochondrial and peroxisomal fatty acids.

The same research group subsequently conducted additional studies with CLNA, but using a specific isomer, puniceic acid (*cis*-9,*trans*-11,*cis*-13 CLNA).<sup>25</sup> A study conducted on ICR CD-1 mice for four weeks found that, in order to see a reducing effect on body fat, this fatty acid should be included at a 5% concentration in diet. This concentration is higher than the CLA dose usually causing significant body fat reductions (0.5–1%). Puniceic acid was also seen to decrease

leptin production and to increase the activity of the enzyme carnitine palmitoyltransferase (CPT-I).<sup>25</sup>

Arao et al.<sup>26</sup> conducted an additional two-week study on Otsuka Long Evans Tokushima Fatty (OLETF) rats with a 5% supplementation in the diet, not with puniceic acid, but with a pomegranate seed oil rich in this fatty acid. With this dose, a significant reduction was seen in omental adipose tissue; by contrast, a 2% supplement was not sufficient to induce changes in this tissue. Yamasaki et al.<sup>27</sup> also found no effects in adipose tissue from C57BL/6N mice fed diets supplemented with pomegranate oil providing 0.12% and 1.2% of puniceic acid for three weeks. Our research group found similar results in rats fed on a high-fat diet supplemented with 0.5% pomegranate seed oil for six weeks (data submitted for publication).

With this lower dose range, McFarlin et al.<sup>28</sup> also found no changes in fat mass, but did find weight gains in CD-1 mice fed with a fat-rich diet containing 61.79 mg of pomegranate seed oil for 14 weeks. By contrast, another study of similar duration (12 weeks) conducted in mice which were fed a diet supplemented with 1% of pomegranate seed oil found both fat mass and body weight reductions.<sup>29</sup> Improved insulin sensitivity was also documented.

Although puniceic acid is one of the most widely studied isomers, there are other isomers which also have this effect. Thus, Hontecillas et al.<sup>17</sup> noted that the addition to the diet of 1% catalpic acid, *trans*-9,*trans*-11,*cis*-13 CLNA, decreased

**Table 2** Contents of conjugated linoleic acid and conjugated linolenic acid isomers in several seed oils.

Fatty acid	<i>V. fordii</i>	<i>Prunus</i> sp.	<i>M. charantia</i>	<i>T. anguina</i>	<i>P. granatum</i>	<i>C. ovata</i>	<i>C. officinalis</i>
18:2	7.07	38.01	8.60	17.22	4.23	39.95	27.92
18:3							
<i>cis</i> -9, <i>trans</i> -11, <i>cis</i> -13; puniceic acid	1.33	–	0.50	48.48	82.99	Traces	–
<i>cis</i> -9, <i>trans</i> -11, <i>trans</i> -13; $\alpha$ -eleostearic acid	67.69	10.63	56.24	3.43	3.16	–	–
<i>trans</i> -8, <i>trans</i> -10, <i>cis</i> -12; calendic acid	–	–	–	–	–	–	62.17
<i>trans</i> -9, <i>trans</i> -11, <i>cis</i> -13; catalpic acid	0.17	–	–	0.41	0.20	42.25	–
<i>trans</i> -9, <i>trans</i> -11, <i>trans</i> -13	11.27	tr	0.32	–	–	0.55	–
<i>trans</i> -8, <i>trans</i> -10, <i>cis</i> -12; calendic acid	–	–	–	–	–	–	0.24

Values are percentages. Modified from Takagi and Itabashi.<sup>21</sup>

**Table 3** Summary of the positive effects of CLNA isomers on fat reduction.

Isomer	Effect	Mechanism	Author	Study model
1% CLNA	Decreased weight of adipose tissue. Increased lipid metabolism in liver	↓ PPAR $\gamma$ expression ↑ Mitochondria and peroxisome oxidation	Koba et al. (2002)	Sprague-Dawley rats
5% punicic acid	Decreased perirenal and epididymal fat	↑ CPT $\text{I}$ activity	Koba et al. (2007)	ICR CD-1 mice
5% pomegranate seed (punicic acid)	Decreased omental adipose tissue	↓ $\Delta 9$ desaturase activity	Arao et al. (2004)	OLEFT rats
1% pomegranate seed (punicic acid)	Decreased weight of adipose tissue	Not reported	Vroegrijk et al. (2011)	C57Bl/J6 mice
1% catalpic acid	Decreased abdominal fat	↑ PPAR $\alpha$ activation	Hontecillas et al. (2008)	C57BL6N mice
Calendic acid	Decreased weight of adipose tissue	Not reported	Chardigny et al. (2003)	ICR mice
$\alpha$ -Eleostearic		↑ PPAR $\alpha$ activation ↑ ACO activity	Chuang et al. (2006)	H4IIEC3 and CHOK1 cells
10–100 $\mu\text{M}$ mixture of CLNA isomers <i>cis-9,trans-11,cis-15</i> , and <i>cis-9,trans-13,cis-15</i>	Decreased triglyceride content	↑ HSL and ATGL gene expression	Miranda et al. (2011)	3T3-L1 cells
10 and 50 $\mu\text{g}/\text{mL}$ pomegranate seed (punicic acid)	Decreased adipogenesis and preadipocyte differentiation	↓ PPAR $\gamma$ and C/EBP $\beta$ ↓ FAS	Lai et al. (2012)	3T3-L1 cells

ACO, acyl-CoA oxidase; ATGL, triglycerides lipase; C/EBP, CCAAT/enhancer binding protein beta; CPT $\text{I}$ , carnitine palmitoyltransferase- $\text{I}$ ; FAS, fatty acid synthase; HSL, hormone-sensitive lipase; PPAR, peroxisome proliferator-activated receptor.

abdominal fat accumulation in C57BL/6N mice after 78 days of treatment.

Chardigny et al.<sup>30</sup> also found that the CLNA isomer *trans*-8,*trans*-10,*cis*-12 (calendic acid) achieved a greater body fat decrease in male mice when compared to animals fed on a control diet. It should be noted, however, that CLNA isomer *trans*-8,*trans*-10,*cis*-12 caused a lower body fat reduction than isomer *trans*-10,*cis*-12 of CLA.

Jacaric acid deserves special mention. Like calendic acid, this fatty acid maintains the *trans*-10,*cis*-12 as well as the carboxyl group in the first carbon. It has been suggested that this structure is indispensable for the inhibition of lipoprotein lipase (LPL), which is one of the most important mechanism involved in the body fat reduction induced by CLA.<sup>31</sup> Despite this, jacaric acid has no reducing effect on body fat.<sup>32</sup> Because of this lack of effect, added to the fact that this fatty acid alters insulin function, jararic acid cannot be proposed as an anti-obesity molecule.

## Other benefits of conjugated linolenic acid isomers related to obesity

### Insulin resistance

Obesity is a chronic disorder associated with various conditions, most of them chronic, which may increase mortality as compared to the non-obese population and cause a significant decrease in patient quality of life. Epidemiological studies have shown a clear relationship between obesity and insulin resistance.<sup>33</sup> This metabolic change is defined as a decreased ability of insulin to exert its biological actions in target tissues such as skeletal muscle, liver, or adipose tissue. It has been found that increased free fatty acid levels and decreased adiponectin levels (insulin-sensitizing hormone) contribute to this resistance state.

As previously noted, CLNA isomers, in addition to reducing body fat, may also have other benefits for health. In this regard, it may be noted that one of the most significant effects of bitter melon is its hypoglycemic potential, shown in normal<sup>34</sup> and diabetic<sup>35,36</sup> rats and in patients with type 2 diabetes.<sup>24</sup> Although its mechanism of action has not been fully elucidated yet, it appears that it inhibits intestinal glucose absorption, promotes insulin utilization in the liver, and even increases the number of beta cells in the pancreas. Similarly,  $\alpha$ -eleostearic acid has recently been identified in bitter melon as a PPAR $\alpha$  activator and as potentially responsible for the hypoglycemic effects.<sup>18</sup> It is important to note that studies have related PPAR $\alpha$  agonists to the protection of pancreatic beta cells against the hyperactivation of insulin production derived from a diet rich in saturated fatty acids,<sup>37</sup> thus preventing type 2 diabetes mellitus as a result of hyperinsulinemia. Finally, it should be noted that, as stated above, Vroegrijk et al.<sup>29</sup> found improved insulin sensitivity in mice fed on a diet supplemented with 1% pomegranate seed oil for 12 weeks.

Similarly, catalpic acid<sup>17</sup> also appears to improve serum fasting glucose and insulin levels, and to increase the ability of mice to normalize plasma glucose levels after a glucose tolerance test. The effects of this CLNA isomer on glucose are believed to be mediated by a PPAR $\alpha$ -dependent mechanism. In fact, catalpic acid increases the expression

of this transcription factor and of the genes controlled by it.<sup>17,18</sup>

The pro-inflammatory cytokine TNF- $\alpha$  is one of the main factors responsible for insulin resistance induced by inflammation. Specifically, TNF- $\alpha$  inhibits the insulin signal by the phosphorylation of serine at the insulin receptor, thus inhibiting the ability of the receptor to bind to insulin receptor substrate 1 (IRS1). According to the results reported by the Bassaganya Riera group, the effectiveness of puniceic acid in the treatment of insulin resistance associated with obesity is due to its activating PPAR $\gamma$  and therefore decreasing TNF- $\alpha$  expression.<sup>19</sup>

Various research studies have shown that body fat reduction induced by CLA may be associated with a dramatic decrease in the serum levels of both leptin and adiponectin in mice<sup>38</sup> (adipokines related to blood glucose control). Studies conducted with puniceic acid appear to suggest that it does not modify the serum levels of these adipokines nor change plasma insulin or glucose levels in mice,<sup>25</sup> which could mean that this molecule is safer than CLA. However, it should not be forgotten that the different CLNA isomers may be metabolized to CLA.<sup>39,40</sup>

### Inflammation

In recent years, obese patients have been reported as having a low-grade chronic inflammatory state. This condition appears to be the consequence of an increased adipose tissue mass that leads to the increased production of pro-inflammatory mediators which are jointly stimulated by signals of exogenous and/or endogenous origin.<sup>41</sup> Adipose tissue contains fibroblasts, preadipocytes, adipocytes, and macrophages. The latter contribute significantly to the systemic inflammatory process by the production of pro-inflammatory mediators such as inflammatory cytokines (TNF- $\alpha$ , IL-6, IL-1 $\beta$ , IL-8, IL-18).<sup>42</sup>

It has been noted that excess calorie intake, some infections, and oxidative stress may cause the increased secretion of these adipokines leading to chronic inflammation in white adipose tissue which promotes the activation and infiltration of mature macrophages. The fact that these macrophages, as well as adipocytes and other cell types, start to secrete cytokines and chemokines makes possible the maintenance of an endless loop of macrophage recruitment and the production of inflammatory mediators, which initially leads to local primary inflammation in adipose tissue and may trigger the low-grade systemic inflammation seen in obesity.<sup>43</sup>

Conjugated fatty acids have been directly related to anti-inflammatory properties. While these properties were mostly attributed to CLA isomers in the past, according to recent research there are CLNA isomers which have the same effect. As for CLAs (in which discrepancies now exist), three mechanisms have been proposed as accounting for the effect of CLNA on inflammatory response:

- (a) Decreased genesis of inducible eicosanoids involved in inflammatory response, including prostaglandins and leukotrienes.

Prostaglandins are a group of oxygenated fatty acids occurring in most mammalian tissues positively correlated to inflammation.<sup>44</sup> As documented by Nugteren

and Christ-Hazelhof in 1987,<sup>45</sup> various CLNA isomers (jacaric acid, calendic acid, punicic acid, catalpic acid, and eleostearic acid) have anti-inflammatory activity mediated by inhibition of the activity of cyclooxygenases, enzymes that catalyze prostaglandin synthesis. Some of these data were confirmed by a subsequent study which concluded that pomegranate extract rich in punicic acid markedly inhibits both lipogenase and cyclooxygenase activity.<sup>46</sup>

(b) Interaction with PPARs $\gamma$ .

PPAR $\gamma$  is expressed in adipose tissue and macrophages, and plays a significant role in the regulation of inflammation.<sup>47</sup> In fact, PPAR has been found to be a negative regulator of macrophage activation and to inhibit the expression of genes related to immune response.<sup>48</sup> PPAR $\gamma$  activation similarly inhibits the production of pro-inflammatory cytokines such as TNF- $\alpha$  and IL-6.<sup>49</sup> The activating and agonist effect of punicic acid on PPAR $\gamma$  receptors has been demonstrated in this review. Punicic acid, as shown by a recent report, may therefore be effective in reducing the chronic inflammation underlying obesity.<sup>19</sup>

(c) Inactivation of the transduction signal of NF- $\kappa$ B (nuclear factor kappa light chain enhancer of activated  $\beta$  cells).

NF- $\kappa$ B is a nuclear transcription factor. Its activation upon cell exposure to external stimuli, such as oxidative stress or ultraviolet radiation, induces the expression of cell genes associated with inflammation,<sup>50</sup> including different cytokines (TNF- $\alpha$ , IL-1, IL-6) and chemokines (IL-8 and macrophage inflammatory protein).

The finding that pomegranate extract is able to decrease and inhibit the activation of NF- $\kappa$ B, and to inhibit the phosphorylation of cytokines related to inflammation and

mitogen-activated protein kinases (MAPK) is, therefore, of great interest.<sup>51,52</sup> The fact that patients with periodontitis (inflammation in the mouth) respond to treatment with pomegranate extract with a reduction in cytokines with a marked inflammatory nature (IL-1 $\beta$  and IL-6) should also not be overlooked.<sup>53</sup> The molecule in pomegranate extract responsible for the inflammatory effect has not yet been identified, but it should be borne in mind that punicic acid is one of the most important functional molecules contained in this extract. In fact, as confirmed by Hontecillas et al.,<sup>19</sup> punicic acid decreases the inflammation related to obesity by activating PPAR $\gamma$  and therefore inhibiting TNF- $\alpha$  expression and NF- $\kappa$ B activity. Since PPAR agonists have been shown to decrease the expression of pro-inflammatory cytokines by antagonizing NF- $\kappa$ B activity, it is natural to think that other CLNA isomers which are agonists of these PPAR receptors, such as  $\alpha$ -eleostearic acid, may be effective in inhibiting this nuclear transcription factor and, thus, in reducing the inflammation associated with obesity.<sup>48,54,55</sup>

A clear example of this is found in the experiments conducted by Saha et al.<sup>56,57</sup> in diabetic rats (streptozotocin-induced diabetes) treated with 0.5%  $\alpha$ -eleostearic acid and 0.5% punicic acid. In these rats, the increased expression of inflammatory cytokines such as tumor necrosis factor (TNF- $\alpha$ ) and interleukin 6 (IL-6) in blood, and the hepatic expression of transcription factor NF- $\kappa$ B after treatment with streptozotocin, due to increased inflammation, was restored by the administration of CLNA isomers.

## Lipidemia

In obese people, increased body fat, especially visceral fat, may lead to hyperglycemia and insulin resistance, but may also affect lipid metabolism. Thus, the metabolic changes

**Table 4** Summary of positive effects of CLNA isomers on diabetes and insulin resistance.

Isomer	Effect	Mechanism	Author	Study model
Bitter gourd ( $\alpha$ -eleostearic acid and catalpic acid)	Improved fasting serum glucose and insulin levels	Activation of PPAR $\alpha$ expression	Welihinda et al. (1986)	Rats
Bitter gourd ( $\alpha$ -eleostearic acid and catalpic acid)	Decreased serum glucose levels	Increased glucose utilization in liver	Sarkar et al. (1996)	Diabetic rats
Bitter gourd ( $\alpha$ -eleostearic acid and catalpic acid)	Improved insulin resistance. Decreased serum insulin and leptin levels	Not reported	Chen et al. (2003)	Rats
Bitter gourd ( $\alpha$ -eleostearic acid and catalpic acid)	Improved glucose tolerance. Hypoglycemic effect	Not reported	Leatherdale et al. (1981)	Diabetic rats
Catalpic acid	Improved fasting serum glucose and insulin levels	PPAR $\alpha$ activation	Hontecillas et al. (2008)	Obese mice
Bitter gourd ( $\alpha$ -eleostearic acid and catalpic acid)	Improved glucose and lipid homeostasis	PPAR $\alpha$ activation	Chuang et al. (2006)	H4IIEC3 cells
Pomegranate seed (punicic acid)	Improved peripheral insulin sensitivity	Not reported	Vroegrijk et al., 2011	C57Bl/J6 mice
Punicic acid	Improved glucose tolerance, with diabetes improvement	PPAR $\alpha$ and PPAR $\gamma$ activation	Hontecillas et al. (2009)	3T3-L1 cells and obese mice

PPAR, peroxisome proliferator-activated receptor.

usually associated with central obesity may include massive access to the liver of free fatty acids, the stimulation of hepatic synthesis of triglycerides, and the secretion of very low density lipoproteins (VLDL) as well as the decreased clearance of triglyceride-rich lipoproteins, the presence of small, dense low density lipoproteins (LDL), and decreased levels of high density lipoproteins (HDL). This decrease in HDLs, together with the increase in LDLs, which are able to enter the arterial wall, where they are oxidized, creates the adequate metabolic conditions for the development of the atherogenic process.<sup>58</sup>

In relation to this subject, research conducted in recent years suggests that some CLNA isomers could be effective in lipid profile control. Thus, the *cis-9,trans-11,cis-13* isomer of CLNA appears to reduce the secretion of apoB100 by human HepG2 liver cells, which could be due to decreased triglycerides in this type of cell.<sup>59</sup> Studies such as the one reported by Koba et al.<sup>25</sup> suggest that this decrease is due to a marked increase in the  $\beta$ -oxidation of fatty acids. This

is not an unexpected effect, considering the potential activating effect shown by CLNA isomers at PPAR $\alpha$ , the main regulator of  $\beta$ -oxidation in peroxisomes and mitochondria.<sup>18</sup> Thus, a reduction in apoB100 production could be related to in vivo VLDL reduction.

However, it appears that CLNA does not only affect apoB100 secretion, but also plays a protective role against the peroxidation of plasma lipids. Plasma lipids, LDL, and erythrocyte membrane lipids may experience peroxidation, which may in turn lead to the development of atherosclerosis and diabetic vascular complications.<sup>60</sup> According to a study conducted by Dhar et al.,<sup>61</sup> in which lipid peroxidation and the antioxidant effect of two concentrations (0.1% and 0.05%) of  $\alpha$ -eleostearic acid in diabetic and non-diabetic subjects were assessed, this substance was shown to be effective for reducing oxidation at both doses, the 0.1% concentration being the more effective.

In addition, in the study conducted by Arao et al.,<sup>59</sup> the *cis-9,trans-11,cis-13* isomer of CLNA was associated with

**Table 5** Summary of positive effects of CLNA isomers on lipidemia and inflammation underlying obesity.

Isomer	Effect	Mechanism	Author	Study model
Punicic acid	↓ Plasma triglycerides. Improved saturated-monounsaturated fatty acid ratio	↓ apoB100 secretion. Inhibition of stearoyl CoA desaturase	Arao et al. (2004)	HepG2 cells
Mixture of CLNA isomers	↓ Plasma cholesterol	Inhibition of hydroxymethylglutaryl coenzyme A reductase	Saha et al. (2007)	Charles Foster rats
Pomegranate seeds	↓ Plasma triglycerides ↓ Triglycerides/HDL cholesterol and total cholesterol/HDL cholesterol	Not reported	Mirmiran et al. (2010)	Patients with dyslipidemia
Bitter gourd ( $\alpha$ -eleostearic acid)	Protective against peroxidation of LDL and erythrocyte membrane lipids	Not reported	Dhar et al. (2006)	Red blood cells
Jacaric acid, calendric acid, punicic acid, catalpic acid, and eleostearic acid	Anti-inflammatory activity	Inhibition of cyclooxygenase activity (inhibition of prostaglandin synthesis)	Nugteren and Christ-Hazelhof (1987)	Microsomes from sheep vesicular glands
Pomegranate extract (punicic acid)	Anti-inflammatory activity	Inhibition of lipogenase and cyclooxygenase activity	Schubert et al. (1999)	Sheep
Pomegranate extract (punicic acid)	Decreased chronic inflammation	PPAR $\gamma$ receptor activator and agonist (inhibition of TNF- $\alpha$ and IL-6 production and NF-activity $\kappa\beta$ )	Hontecillas et al. (2009)	3T3-L1 cells
Pomegranate extract (punicic acid)	Decreased inflammation associated with obesity	Inhibition of NF- $\kappa\beta$ and MAPK activation	Afaq et al. (2005)	Normal human epidermal keratinocytes
Pomegranate extract	Anti-inflammatory activity	Inhibition of NF- $\kappa\beta$ and MAPK activation (inhibits IL-1 production)	Ahmed et al. (2005)	Human chondrocytes from cartilage with osteoarthritis
$\alpha$ -Eleostearic and punicic acids	Anti-inflammatory activity	Inhibition of NF- $\kappa\beta$ expression. Decreased serum TNF- $\alpha$ and IL-6	Saha et al. (2012)	Diabetic rats

IL, interleukin; MAPK, mitogen-activated protein kinases; NF- $\kappa\beta$ , nuclear factor kappa light chain enhancer of activated  $\beta$  cells; PPAR, peroxisome proliferator-activated receptor; TNF, tumor necrosis factor.



an improved ratio between saturated and monounsaturated fatty acids, a significant marker not only of cardiovascular markers, but also of metabolic syndrome. The authors of this research postulated that, as occurred with the *trans*-10,*cis*-12 isomer of CLA, this improvement was the result of the inhibition of stearoyl CoA desaturase by the *cis*-9,*trans*-11,*cis*-13 isomer of CLNA.

In the Saha and Ghosh study,<sup>56</sup> plasma cholesterol levels and cholesterol content in tissues returned to normal in rats with induced hypercholesterolemia on feeding a diet enriched with 0.5% and 1% concentrations of mixed CLNA isomers. This effect was due to decreased cholesterol synthesis in the liver, associated with the inhibition of hydroxymethylglutaryl coenzyme A reductase, the rate-limiting enzyme in cholesterol synthesis.

In the Mirmiran et al. study,<sup>62</sup> although patients with dyslipidemia treated for four weeks with 400mg of pomegranate seed oil experienced no changes in total and LDL cholesterol levels in plasma, reductions did occur in triglyceride levels and triglyceride/HDL cholesterol and total cholesterol/LDL cholesterol ratios, with a resultant decrease in cardiovascular risk.

It should, however, be noted that studies have been conducted in rats and hamsters where the CLNA isomers tested had no effect on serum lipid profile<sup>63</sup> or even worsened it, increasing total cholesterol levels, the LDL/HDL ratio, and triglycerides.<sup>64</sup> These discrepancies may have been due to the use of different experimental models and different doses of CLNA isomers.

## Conclusions

Research conducted in recent years has shown that conjugated linolenic acid isomers are potential functional substances for the prevention of obesity and its associated conditions. The results achieved in the in vitro studies conducted show that punicic acid and  $\alpha$ -eleostearic acid act as natural agonists of PPAR $\alpha$ , which suggests that they may be effective in body fat reduction, in addition to having other positive effects related to obesity. These encouraging in vitro results are supported by the in vivo study conducted by Koba et al., where a mixture of unidentified CLNA isomers was added to the diet at a 1% concentration. It should be noted, however, that this mixture contained CLA isomers, which may have masked the actual effect of CLNA isomers on body fat reduction.

Subsequent studies with isolated CLNA isomers have reported that the fat-reducing effect usually occurs at doses higher than the effective CLA doses (0.5–1%). Therefore, CLNA does not appear to be a more promising molecule than CLA with regard to obesity. However, because of the multiple isomers encompassed by the term conjugated linolenic acid, additional experimental studies are needed to definitively rule out CLNA isomers as potential functional elements in the prevention of obesity through body fat reduction.

Obesity is a chronic disorder associated with various conditions, most of them chronic, which may increase mortality as compared to the non-obese population and may cause a significant decrease in patient quality of life. As regards the use of these conjugated fatty acids for the prevention of underlying changes in obesity, it should be borne in mind that only a few studies have been conducted to date

(Tables 4 and 5). It is, therefore, difficult to draw clear-cut conclusions concerning the potential use of CLNA in insulin resistance, dyslipidemia, or inflammation.

## Conflicts of interest

The authors state that they have no conflicts of interest.

## References

1. Kelley DS, Erickson KL. Modulation of body composition and immune cell functions by conjugated linoleic acid in humans and animal models: Benefits vs. risks. *Lipids*. 2003;38:377–86.
2. Risérus U, Basu S, Jovinge S, Fredrikson G, Arnlöv J, Vessby B. Supplementation with conjugated linoleic acid causes isomer-dependent oxidative stress and elevated C-reactive protein: a potential link to fatty acid-induced insulin resistance. *Circulation*. 2002;106:1925–9.
3. EFSA Panel on Dietetic Products NaAN. Scientific opinion on the safety of 'conjugated linoleic acid (CLA)-rich oil' (Clarinol®) as a novel food ingredient. *EFSA J*. 2010;8:1–41.
4. Pariza M, Park Y, Cook M. The biologically active isomers of conjugated linoleic acid. *Prog Lipid Res*. 2001;40:283–98.
5. Evans M, Brown J, McIntosh M. Isomer-specific effects of conjugated linoleic acid (CLA) on adiposity and lipid metabolism. *J Nutr Biochem*. 2002;13:508.
6. Risérus U, Vessby B, Arnlöv J, Basu S. Effects of *cis*-9,*trans*-11 conjugated linoleic acid supplementation on insulin sensitivity, lipid peroxidation, and proinflammatory markers in obese men. *Am J Clin Nutr*. 2004;80:279–83.
7. Gaullier JM, Halse J, Høivik HO, Høye K, Syvertsen C, Nurminiemi M, et al. Six months supplementation with conjugated linoleic acid induces regional-specific fat mass decreases in overweight and obese. *Br J Nutr*. 2007;97:550–60.
8. Norris LE, Collene AL, Asp ML, Hsu JC, Liu LF, Richardson JR, et al. Comparison of dietary conjugated linoleic acid with safflower oil on body composition in obese postmenopausal women with type 2 diabetes mellitus. *Am J Clin Nutr*. 2009;90:468–76.
9. Brown AW, Trenkle AH, Beitz DC. Diets high in conjugated linoleic acid from pasture-fed cattle did not alter markers of health in young women. *Nutr Res*. 2011;31:33–41.
10. Mougios V, Matsakas A, Petridou A, Ring S, Sagredos A, Melissopoulou A, et al. Effect of supplementation with conjugated linoleic acid on human serum lipids and body fat. *J Nutr Biochem*. 2001;12:585–94.
11. Noone EJ, Roche HM, Nugent AP, Gibney MJ. The effect of dietary supplementation using isomeric blends of conjugated linoleic acid on lipid metabolism in healthy human subjects. *Br J Nutr*. 2002;88:243–51.
12. Risérus U, Arner P, Brismar K, Vessby B. Treatment with dietary *trans*10*cis*12 conjugated linoleic acid causes isomer-specific insulin resistance in obese men with the metabolic syndrome. *Diabetes Care*. 2002;25:1516–21.
13. Eyjolfson V, Spriet LL, Dyck DJ. Conjugated linoleic acid improves insulin sensitivity in young, sedentary humans. *Med Sci Sports Exerc*. 2004;36:814–20.
14. Ramakers JD, Plat J, Sébédio JL, Mensink RP. Effects of the individual isomers *cis*-9,*trans*-11 vs. *trans*-10,*cis*-12 of conjugated linoleic acid (CLA) on inflammation parameters in moderately overweight subjects with LDL-phenotype B. *Lipids*. 2005;40:909–18.
15. Song HJ, Grant I, Rotondo D, Mohede I, Sattar N, Heys SD, et al. Effect of CLA supplementation on immune function in young healthy volunteers. *Eur J Clin Nutr*. 2005;59:508–17.

16. Plourde M, Destaillets F, Chouinard PY, Angers P. Conjugated alpha-linolenic acid isomers in bovine milk and muscle. *J Dairy Sci.* 2007;90:5269–75.
17. Hontecillas R, Diguardo M, Duran E, Orpi M, Bassaganya-Riera J. Catalpic acid decreases abdominal fat deposition, improves glucose homeostasis and upregulates PPAR alpha expression in adipose tissue. *Clin Nutr.* 2008;27:764–72.
18. Chuang C, Hsu C, Chao C, Wein Y, Kuo Y, Huang C. Fractionation and identification of 9c, 11t, 13t-conjugated linolenic acid as an activator of PPARalpha in bitter melon (*Momordica charantia* L.). *J Biomed Sci.* 2006;13:763–72.
19. Hontecillas R, O'Shea M, Einerhand A, Diguardo M, Bassaganya-Riera J. Activation of PPAR gamma and alpha by punical acid ameliorates glucose tolerance and suppresses obesity-related inflammation. *J Am Coll Nutr.* 2009;28:184–95.
20. Stanton C, Lawless F, Kjellmer G, Harrington R, Devery R, Connolly JF, et al. Dietary influences on bovine milk cis-9,trans-11-conjugated linoleic acid content. *J Food Sci.* 1997;62:1083–6.
21. Takagi T, Itabashi Y. Occurrence of mixtures of geometrical isomers of conjugated octadecatrienoic acids in some seed oils: analysis by open tubular gas liquid chromatography and high performance liquid chromatography. *Lipids.* 1981;16:546–51.
22. Miranda J, Lasa A, Fernández-Quintela A, García-Marzo C, Ayo J, Dentin R, et al. cis-9,trans-11,cis-15 and cis-9,trans-13,cis-15 CLNA mixture activates PPAR $\alpha$  in HEK293 and reduces triacylglycerols in 3T3-L1 cells. *Lipids.* 2011;46:1005–12.
23. Lai CS, Tsai ML, Badmaev V, Jimenez M, Ho CT, Pan MH. Xanthigen suppresses preadipocyte differentiation and adipogenesis through down-regulation of PPAR $\gamma$  and C/EBPs and modulation of SIRT-1, AMPK, and FoxO pathways. *J Agric Food Chem.* 2012;60:1094–101.
24. Koba K, Akahoshi A, Yamasaki M, Tanaka K, Yamada K, Iwata T, et al. Dietary conjugated linolenic acid in relation to CLA differently modifies body fat mass and serum and liver lipid levels in rats. *Lipids.* 2002;37:343–50.
25. Koba K, Imamura J, Akashoshi A, Kohno-Murase J, Nishizono S, Iwabuchi M, et al. Genetically modified rapeseed oil containing cis-9,trans-11,cis-13-octadecatrienoic acid affects body fat mass and lipid metabolism in mice. *J Agric Food Chem.* 2007;55:3741–8.
26. Arao K, Wang Y, Inoue N, Hirata J, Cha JY, Nagao K, et al. Dietary effect of pomegranate seed oil rich in 9cis, 11trans, 13cis conjugated linolenic acid on lipid metabolism in obese, hyperlipidemic OLETF rats. *Lipids Health Dis.* 2004;3:24.
27. Yamasaki M, Kitagawa T, Koyanagi N, Chujo H, Maeda H, Kohno-Murase J, et al. Dietary effect of pomegranate seed oil on immune function and lipid metabolism in mice. *Nutrition.* 2006;22:54–9.
28. McFarlin BK, Strohacker KA, Kueht ML. Pomegranate seed oil consumption during a period of high-fat feeding reduces weight gain and reduces type 2 diabetes risk in CD-1 mice. *Br J Nutr.* 2009;102:54–9.
29. Vroegrijk IO, van Diepen JA, van den Berg S, Westbroek I, Keizer H, Gambelli L, et al. Pomegranate seed oil, a rich source of punical acid, prevents diet-induced obesity and insulin resistance in mice. *Food Chem Toxicol.* 2011;49:1426–30.
30. Chardigny J, Hasselwander O, Genty M, Kraemer K, Ptock A, Sébédio J. Effect of conjugated FA on feed intake, body composition, and liver FA in mice. *Lipids.* 2003;38:895–902.
31. Park Y, Storkson J, Liu W, Albright K, Cook M, Pariza M. Structure–activity relationship of conjugated linoleic acid and its cognates in inhibiting heparin-releasable lipoprotein lipase and glycerol release from fully differentiated 3T3-L1 adipocytes. *J Nutr Biochem.* 2004;15:561–8.
32. Miranda J, Fernández-Quintela A, Macarulla M, Churruga I, García C, Rodríguez VM, et al. A comparison between CLNA and CLA effects on body fat, serum parameters and liver composition. *J Physiol Biochem.* 2009;65:25–32.
33. Qatanani M, Lazar M. Mechanisms of obesity-associated insulin resistance: many choices on the menu. *Genes Dev.* 2007;21:1443–55.
34. Welihinda J, Karunanayake E. Extra-pancreatic effects of *Momordica charantia* in rats. *J Ethnopharmacol.* 1986;17:247–55.
35. Sarkar S, Pranava M, Marita R. Demonstration of the hypoglycemic action of *Momordica charantia* in a validated animal model of diabetes. *Pharmacol Res.* 1996;33:1–4.
36. Chen Q, Chan L, Li E. Bitter melon (*Momordica charantia*) reduces adiposity, lowers serum insulin and normalizes glucose tolerance in rats fed a high fat diet. *J Nutr.* 2003;133:1088–93.
37. Hellemans K, Kerckhofs K, Hannaert J, Martens G, van Veldhoven P, Pipeleers D. Peroxisome proliferator-activated receptor alpha-retinoid X receptor agonists induce beta-cell protection against palmitate toxicity. *FEBS J.* 2007;274:6094–105.
38. Tsuboyama-Kasaoka N, Takahashi M, Tanemura K, Kim HJ, Tange T, Okuyama H, et al. Conjugated linoleic acid supplementation reduces adipose tissue by apoptosis and develops lipodystrophy in mice. *Diabetes.* 2000;49:1534–42.
39. Tsuzuki T, Tokuyama Y, Igarashi M, Nakagawa K, Ohsaki Y, Komai M, et al. Alpha-eleostearic acid (9Z11E13E-18:3) is quickly converted to conjugated linoleic acid (9Z11E-18:2) in rats. *J Nutr.* 2004;134:2634–9.
40. Tsuzuki T, Igarashi M, Komai M, Miyazawa T. The metabolic conversion of 9,11,13-eleostearic acid (18:3) to 9,11-conjugated linoleic acid (18:2) in the rat. *J Nutr Sci Vitaminol (Tokyo).* 2003;49:195–200.
41. Berg A, Scherer P. Adipose tissue, inflammation, and cardiovascular disease. *Circ Res.* 2005;96:939–49.
42. Weisberg S, McCann D, Desai M, Rosenbaum M, Leibel R, Ferrante AJ. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest.* 2003;112:1796–808.
43. Moreno-Aliaga M, Campión J, Milagro F, Berjón A, Martínez J. Adiposity and proinflammatory state: the chicken or the egg. *Adipocytes.* 2005;1:1–13.
44. Kuehl FJ, Egan R. Prostaglandins, arachidonic acid, and inflammation. *Science.* 1980;210:978–84.
45. Nugteren D, Christ-Hazelhof E. Naturally occurring conjugated octadecatrienoic acids are strong inhibitors of prostaglandin biosynthesis. *Prostaglandins.* 1987;33:403–17.
46. Schubert S, Lansky E, Neeman I. Antioxidant and eicosanoid enzyme inhibition properties of pomegranate seed oil and fermented juice flavonoids. *J Ethnopharmacol.* 1999;66:11–7.
47. Cuzzocrea S, Pisano B, Dugo L, Ianaro A, Maffia P, Patel NS, et al. Rosiglitazone, a ligand of the peroxisome proliferator-activated receptor-gamma, reduces acute inflammation. *Eur J Pharmacol.* 2004;483:79–93.
48. Ricote M, Li A, Willson T, Kelly C, Glass C. The peroxisome proliferator-activated receptor-gamma is a negative regulator of macrophage activation. *Nature.* 1998;391:79–82.
49. Jiang C, Ting A, Seed B. PPAR-gamma agonists inhibit production of monocyte inflammatory cytokines. *Nature.* 1998;391:82–6.
50. Lamas O, Moreno-Aliaga M, Martínez J, Martí A. NF-kappa B-binding activity in an animal diet-induced overweightness model and the impact of subsequent energy restriction. *Biochem Biophys Res Commun.* 2003;311:533–9.
51. Afaq F, Malik A, Syed D, Maes D, Matsui M, Mukhtar H. Pomegranate fruit extract modulates UV-B-mediated phosphorylation of mitogen-activated protein kinases and activation of nuclear factor kappa B in normal human epidermal keratinocytes paragraph sign. *Photochem Photobiol.* 2005;81:38–45.
52. Ahmed S, Wang N, Hafeez B, Cheruvu V, Haqqi T. *Punica granatum* L. extract inhibits IL-1beta-induced expression of

- matrix metalloproteinases by inhibiting the activation of MAP kinases and NF-kappaB in human chondrocytes in vitro. *J Nutr.* 2005;135:2096–102.
53. Sastravaha G, Gassmann G, Sangtherapitikul P, Grimm W. Adjunctive periodontal treatment with *Centella asiatica* and *Punica granatum* extracts in supportive periodontal therapy. *J Int Acad Periodontol.* 2005;7:70–9.
  54. Pascual G, Fong A, Ogawa S, Gamliel A, Li AC, Perissi V, et al. A SUMOylation-dependent pathway mediates transrepression of inflammatory response genes by PPAR-gamma. *Nature.* 2005;437:759–63.
  55. Kelly D, Campbell J, King T, Grant G, Jansson EA, Coutts AG, et al. Commensal anaerobic gut bacteria attenuate inflammation by regulating nuclear-cytoplasmic shuttling of PPAR-gamma and RelA. *Nat Immunol.* 2004;5:104–12.
  56. Saha SS, Ghosh M. Antioxidant and anti-inflammatory effect of conjugated linolenic acid isomers against streptozotocin-induced diabetes. *Br J Nutr.* 2012;108:974–83.
  57. Saha SS, Dasgupta P, Sengupta Bandyopadhyay S, Ghosh M. Synergistic effect of conjugated linolenic acid isomers against induced oxidative stress, inflammation and erythrocyte membrane disintegrity in rat model. *Biochim Biophys Acta.* 2012;1820:1951–70.
  58. Tchernof A, Després JP. Pathophysiology of human visceral obesity: an update. *Physiol Rev.* 2013;93:359–404.
  59. Arao K, Yotsumoto H, Han S, Nagao K, Yanagita T. The 9cis, 11trans, 13cis isomer of conjugated linolenic acid reduces apolipoprotein B100 secretion and triacylglycerol synthesis in HepG2 cells. *Biosci Biotechnol Biochem.* 2004;68:2643–5.
  60. Giugliano D, Ceriello A, Paolisso G. Oxidative stress and diabetic vascular complications. *Diabetes Care.* 1996;19:257–67.
  61. Dhar P, Chattopadhyay K, Bhattacharyya D, Roychoudhury A, Biswas A, Ghosh S. Antioxidative effect of conjugated linolenic acid in diabetic and non-diabetic blood: an in vitro study. *J Oleo Sci.* 2006;56:19–24.
  62. Mirmiran P, Fazeli MR, Asghari G, Shafiee A, Azizi F. Effect of pomegranate seed oil on hyperlipidaemic subjects: a double-blind placebo-controlled clinical trial. *Br J Nutr.* 2010;104:402–6.
  63. Yang L, Leung K, Cao Y, Huang Y, Ratnayake W, Chen Z. Alpha-linolenic acid but not conjugated linolenic acid is hypcholesterolaemic in hamsters. *Br J Nutr.* 2005;93:433–8.
  64. Dhar P, Bhattacharyya D. Nutritional characteristics of oil containing conjugated octadecatrienoic fatty acid. *Ann Nutr Metab.* 1998;42:290–6.