

REVIEW ARTICLE

Current concepts on the pathophysiology of idiopathic chronic adult hydrocephalus: Are we facing another neurodegenerative disease?☆



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Diffusion tractography

Abstract

Introduction: Since its description five decades ago, the pathophysiology of idiopathic chronic adult hydrocephalus (iCAH) has been traditionally related to the effect that ventricular dilatation exerts on the structures surrounding the ventricular system. However, altered cerebral blood flow, especially a reduction in the CSF turnover rate, are starting to be considered the main pathophysiological elements of this disease.

Development: Compression of the pyramidal tract, the frontostriatal and frontoreticular circuits, and the paraventricular fibres of the superior longitudinal fasciculus have all been reported in iCAH. At the level of the corpus callosum, gliosis replaces a number of commissural tracts. Cerebral blood flow is also altered, showing a periventricular watershed region limited by the subependymal arteries and the perforating branches of the major arteries of the anterior cerebral circulation. The CSF turnover rate is decreased by 75%, leading to the reduced clearance of neurotoxins and the interruption of neuroendocrine and paracrine signalling in the CSF.

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PALABRAS CLAVE

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del adulto idiopática;
Recambio licuoral;
Tractografía por
tensor de difusión

Conclusions: iCAH presents as a complex nosological entity, in which the effects of subcortical microangiopathy and reduced CSF turnover play a key role. According to its pathophysiology, it is simpler to think of iCAH more as a neurodegenerative disease, such as Alzheimer disease or Binswanger disease than as the classical concept of hydrocephalus.

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Actualización en la fisiopatología de la hidrocefalia crónica del adulto idiopática: ¿nos enfrentamos a otra enfermedad neurodegenerativa?

Resumen

Introducción: Desde la descripción hace 5 décadas de la hidrocefalia crónica del adulto idiopática (HCAi), su fisiopatología ha sido considerada básicamente relacionada con el efecto que la dilatación ventricular ejerce sobre las estructuras adyacentes al sistema ventricular. Sin embargo, las alteraciones en el flujo sanguíneo cerebral (FSC) y, sobre todo, la reducción en el recambio licuoral parecen emerger como componentes fisiopatológicos principales de esta enfermedad.

Desarrollo: En la HCAi se observa una compresión del tracto piramidal, de los circuitos cortico-subcorticales fronto-estriatales y fronto-reticulares, y de las fibras profundas del fascículo longitudinal superior. En el cuerpo calloso se objetiva un descenso en el número de fibras comisurales, que son reemplazadas por gliosis. El FSC se encuentra alterado, con un patrón de última pradera en la región subcortical adyacente a los ventrículos, correspondiente a la intersección entre las arterias subependimarias y las arterias perforantes dependientes de los grandes troncos arteriales de la circulación anterior. El recambio diario del LCR se ve disminuido en un 75%, lo que conlleva una reducción del aclaramiento de neurotóxicos y la interrupción de las señalizaciones neuroendocrinas y paracrinas que ocurren a través del LCR.

Conclusiones: La HCAi emerge como una entidad nosológica compleja, en la que los efectos de la microangiopatía subcortical y la disminución del recambio de LCR desempeñan un papel fundamental. Esta base fisiopatológica aleja la HCAi del concepto clásico de hidrocefalia y la acerca al perfil de otras enfermedades neurodegenerativas, como la enfermedad de Alzheimer o la enfermedad de Binswanger.

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Introduction

Idiopathic normal-pressure hydrocephalus (iNPH) is a nosological entity characterised by the clinical triad of gait disturbance, cognitive impairment, and urinary incontinence, with neuroimaging findings of ventricular dilatation (Fig. 1) and in the absence of any other cause that may explain clinical findings.

Although this clinical syndrome had previously been described in the literature^{1–5} (particular emphasis should be placed on the descriptions made by French neurologist Etienne Mouline⁶ in 1819 and German pathologist Friedrich Dörner⁷ in 1826), it was the late Colombian neurosurgeon Salomón Hakim Dow who provided a systematic description of the clinical and radiological features of iNPH in his doctoral thesis, written over 50 years ago.⁸ Hakim, together with 2 renowned neurologists from the Massachusetts General Hospital, Raymond D. Adams and Charles M. Fisher, disclosed his findings in 2 articles, which were published simultaneously in the *New England Journal of Medicine*⁹ and the *Journal of Neurological Sciences*.¹⁰

The classic triad of symptoms has traditionally been thought to be caused by the effect of ventricular dilatation on periventricular nerves^{11–17} and vessels.^{18–25} However, recent studies also suggest an inability of the CSF to remove waste products from the extracellular fluid as a causal factor for iNPH.^{26–29}

We provide updated information on the pathophysiology of the disease, placing special emphasis on decreased CSF turnover, a novel factor which may have an impact on long-term prognosis. These findings challenge the classic concept of hydrocephalus, suggesting that iNPH is a neurodegenerative disease.

Development**Compression of periventricular subcortical fibres**

Compression of the frontal projections descending close to the frontal horns of the lateral ventricles alters the

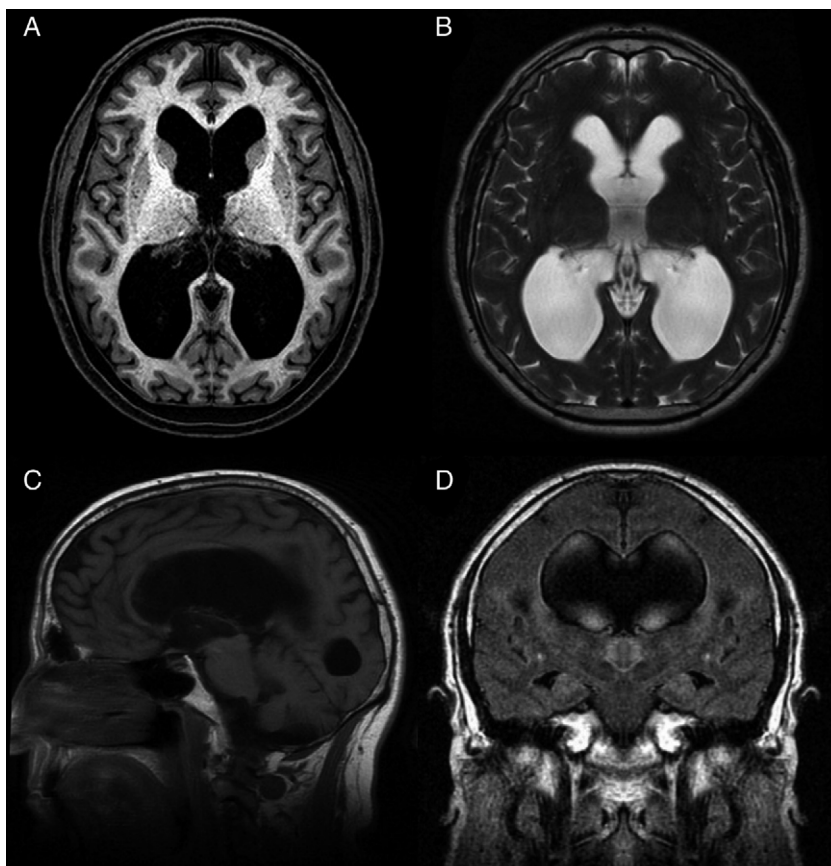


Figure 1 Preoperative MR image of a patient with idiopathic normal-pressure hydrocephalus who responded well to ventriculo-peritoneal shunting. (A) Axial T1-weighted sequence. Moderate dilatation of the lateral ventricles and the third ventricle. (B) Axial T2-weighted sequence. CSF flow artefacts in the third ventricle; absence of hyperintensity at the periventricular and subcortical levels. (C) Sagittal T1-weighted sequence. Descent of the third ventricle floor, rounding of the third ventricle, and decreased mamillopontine distance. The image shows no obstruction in the aqueduct of Sylvius that may explain ventricular dilatation. (D) Coronal FLAIR sequence. Typical pattern of effacement of convexity sulci, especially in the midline. The CSF flow signal void on the T2-weighted sequence extends towards both Monro foramina, reaching the lower and middle portions of the ventricular cavities. Absence of periventricular and subcortical hyperintensities.

function of the projections. These alterations may be completely reversible when dysfunction is caused by slowing or interruption of axonal transport at that level, or permanent in the case of demyelination or loss of axonal integrity.^{16,17} Different MRI techniques have increased our knowledge of the fascicles affected in patients with iNPH, including the cortical and subcortical projections of these fascicles, and help determine whether lesions are reversible. Diffusion-weighted imaging and diffusion tensor imaging (DTI) are the forms of MRI most frequently used to assess these patients. The former evaluates the presence of free interstitial water by calculating apparent diffusion coefficient (ADC) values (these values are higher in the extracellular oedema). However, axon regeneration by gliosis, which invariably occurs in the chronic phase of axonal damage, may cause an even greater increase in ADC values. DTI, including fibre tractography, is much more sensitive than ADCs for evaluating the integrity, density, and potential displacement of nerve fascicles, as it detects anisotropic changes in water molecules due to the unidirectional propagation of action potentials. Mean diffusivity (MD) may be analogous to ADC values,

whereas fractional anisotropy (FA) decreases significantly when axonal disruption occurs (even in the areas where ADC maps show no significant increase of ADC coefficients or where these are normal due to the artefact generated by residual gliosis), and increases with axon density, for example as a consequence of axon compaction due to the effect of a force perpendicular to the axons' trajectory. Consequently, an increase in ADC or MD values in the presence of normal or increased FA values suggests interstitial oedema, whereas a decrease in FA values is suggestive of axonal damage regardless of MD or ADC values.³⁰

The most important studies conducted to date using these techniques are consistent in that increased ADC and/or MD values are observed both in the corpus callosum and in the internal capsule. FA, however, increases in the internal capsule, especially in its anterior limb, and decreases in the corpus callosum, especially at the level of the genu.^{11–14,31} Other findings reported in the literature include increases in FA in the caudate nucleus,^{11,31} increases in MD with no FA changes in the white matter associated with the precentral cortex,¹⁵ and increased

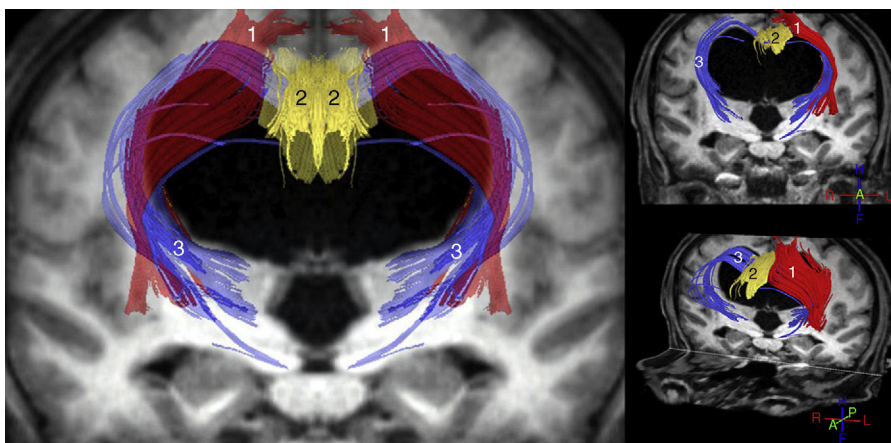


Figure 2 Tractography reconstruction of DTI sequences of the patient shown in the previous figure. The image shows the distortion generated by ventricular dilatation along the pyramidal tract (1) and in the corticostriatal and corticoreticular connections (3). The corticostriatal and corticoreticular tracts are considerably thinner than normal; thinning is also observed in the cingulate fasciculus (2) and, to a lesser extent, in the pyramidal tract (1).

axonal density in the corticospinal tract at the paraventricular level, as a consequence of compaction in the areas adjacent to the ventricle.³² In the light of these findings, we may conclude that iNPH involves compression of the pyramidal tract and the frontostriatal and frontoreticular cortico-subcortico-cortical circuits (Fig. 2), in addition to dysfunction of the deep fibres of the superior longitudinal fascicle. In the corpus callosum, decreased FA and increased ADC and/or MD values are suggestive of a reduced number of commissural fibres, which would have been replaced by gliosis.

Reduced cerebral blood flow

Multiple studies have shown reduced cerebral blood flow (CBF) in patients with iNPH, although global involvement is rather discreet, with CBF decreases ranging from 20% to 30%.^{18–25,33} A recent study conducted at Osaka University found patients with confirmed iNPH and those with ventricular dilatation displaying the radiological signs but no symptoms of iNPH to have lower CBF than do healthy controls.²⁵ The literature reports conflicting results regarding the severity of clinical symptoms and its association with CBF; most studies do not show a direct relationship,^{18,22,25} whereas some do suggest a correlation between clinical severity and a progressive decrease in CBF.^{19,21,34}

The mechanisms underlying CBF alterations are yet to be understood. If decreases in CBF were due only to the distortion caused by ventricular dilatation and the forces exerted on cerebral microcirculation, we may expect to observe a negative correlation between decreases in CBF and the distance to the ventricular system, with greater CBF decreases in the most internal and inferior portion of the centrum semiovale, in the thalamus, and in the head and tail of the caudate nucleus. Some recent studies have overcome the limitations of poor spatial resolution of traditional techniques for measuring CBF by co-registering $H_2^{15}O$ -PET images onto MRI images, demonstrating a CBF gradient from the periventricular region to the cortex; the relationship

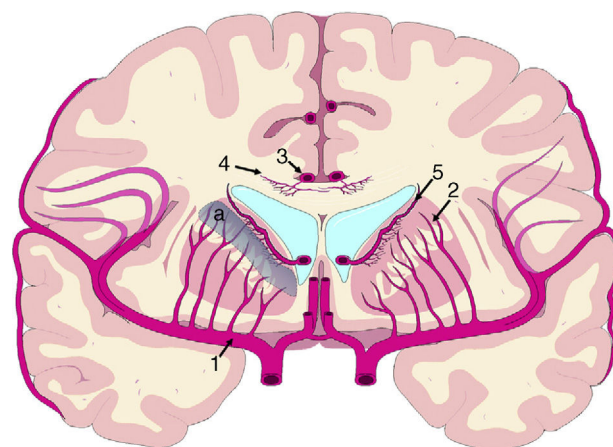


Figure 3 Vascularisation of periventricular structures. The basal ganglia and internal capsule are located in a last meadow area (a) between the territory of the perforating branches (2) of the middle cerebral artery (1) and the branches of the subependymal arteries (5). The corpus callosum is irrigated mainly by the short callosal arteries (4) from the pericallosal artery (3), which belong to the terminal circulation.

between changes in CBF and distance is not proportional, however.^{18,19} The study by Momjian et al.¹⁸ is particularly interesting: the researchers located the area of maximal CBF decrease in the subcortical white matter, 1 cm from the ventricular wall, observing a 50% decrease in the cerebrovascular reserve (CVR) in that location (Fig. 3). These findings are compatible with the concept of last meadow in the subcortical region adjacent to the ventricles. This phenomenon has a coherent microanatomical basis, since the tissue closest to the ependyma is irrigated by subependymal arteries and the tissue farthest from the ependyma by perforating branches of the major arteries of the anterior circulation (Fig. 4).^{35,36} The area of greatest sensitivity to ischaemia is the area where both vascular territories intersect and where CBF and CVR alterations are most marked.

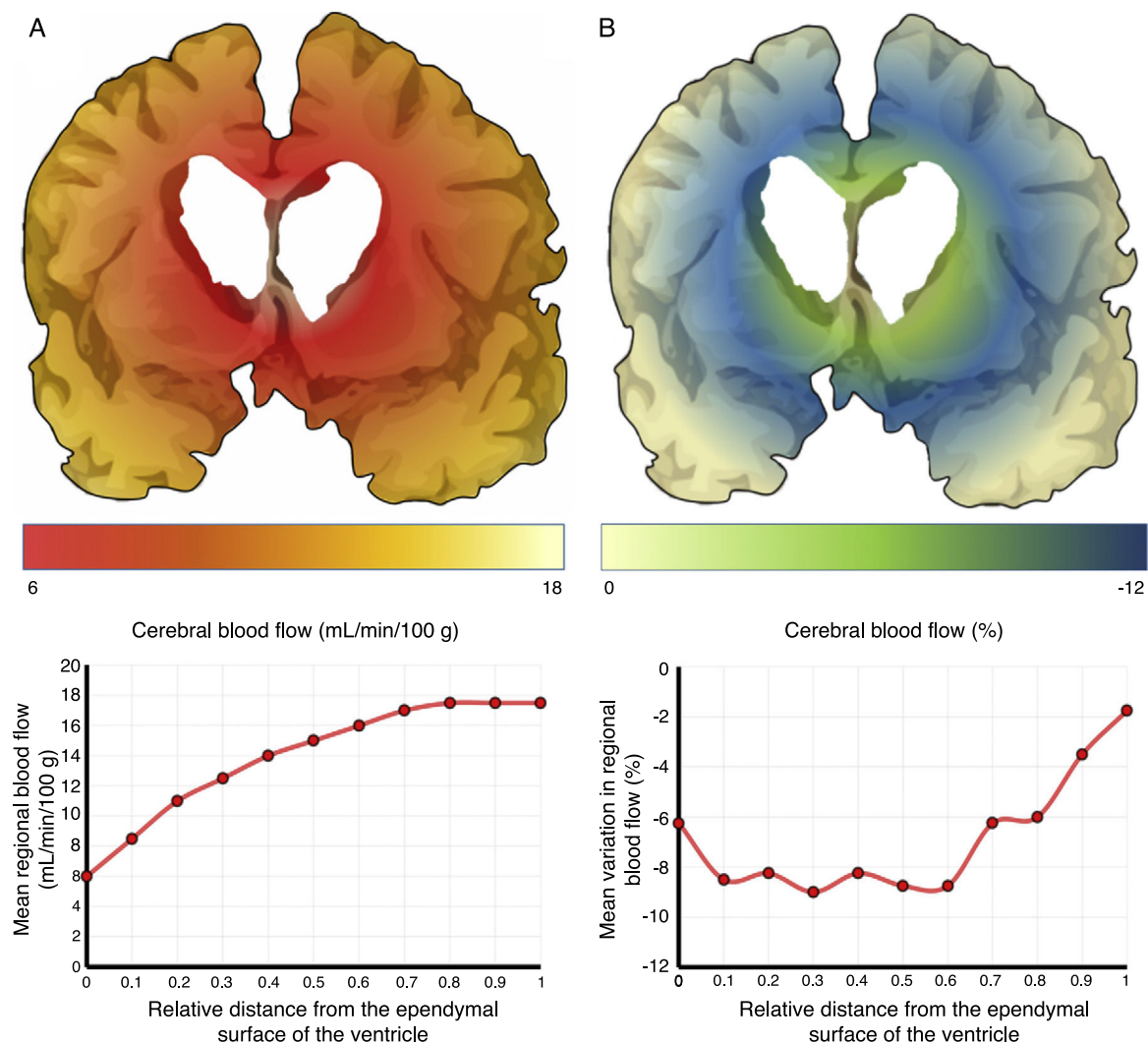


Figure 4 Cerebral blood flow (CBF) in the periventricular region in patients with idiopathic normal-pressure hydrocephalus. (A) The relationship between CBF and distance to the ependymal surface of the ventricle follows a logarithmic curve.¹⁸ (B) When patients experience fluid overload, CBF drops in the subependymal last meadow area, located 5-15 mm from the ventricular surface.

In any case, basal ganglia alterations seem to be a constant in most clinical studies of iNPH.^{19,34,37}

Other studies have also detected cortical alterations which may not be explained by a merely mechanical phenomenon, given the relative distance from the ventricular system. Hypoperfusion is more marked in the anterior and inferior mesial regions of the frontal lobe than in other structures.^{22,34} Several studies have also described alterations in such areas as the left anterior temporal cortex,²² the hippocampus and parahippocampus,³⁷ the frontal lobe white matter corresponding to the superior longitudinal fasciculus,³⁴ and parietal association areas.³⁴

Cerebrovascular involvement associated with iNPH is considerable, also affecting cerebral vasoreactivity, which is reduced as a result of exhaustion of the CVR. To evaluate the CVR, Chang et al.²¹ studied CBF in 167 patients diagnosed with iNPH using ^{99m}Tc-HMPAO SPECT before and after administering 1 g acetazolamide. The researchers found that the CBF increased by 50%-80% less in patients with iNPH than in healthy individuals after the infusion of acetazolamide;

this may reflect decreased vasoreactivity in response to increased partial pressure of CO₂ in arterial blood or a significant decrease in the CVR. Fortunately, these changes are not accompanied by alterations in the metabolic coupling of the areas involved, since the cerebral metabolic rate decreases in line with decreases in CBF, and the oxygen extraction fraction remains within normal limits.^{38,39}

Decreased CSF turnover

As occurs with the lymph in the rest of the body, CSF excretes the macromolecules present in the interstitial fluid that cannot be reabsorbed by venous capillaries. Under normal circumstances, CSF contains over 2000 different proteins, which amount to less than 4% of its weight.⁴⁰ Reabsorption of CSF water may be unrelated to macromolecule excretion: the former takes place in the venous ends of the capillaries, whereas the latter occurs in arachnoid granulations or in extracranial lymphatic vessels. Alterations in

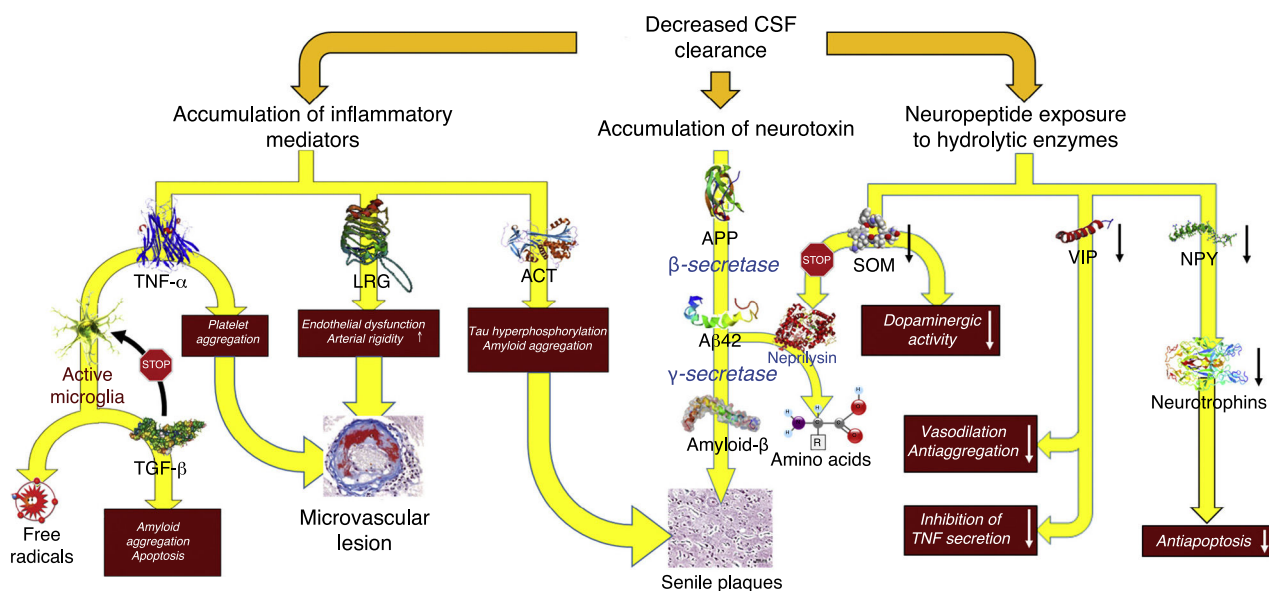


Figure 5 Pathophysiological consequences of decreased CSF clearance: accumulation of neurotoxins and inflammatory mediators, and exposure of the neuropeptides responsible for neuroendocrine signalling to the activity of hydrolytic enzymes.

A β -42: 42-residue fragment of amyloid- β peptide; ACT: antichymotrypsin; APP: amyloid precursor protein; LRG: leucine-rich glycoprotein; NPY: neuropeptide Y; TGF: transforming growth factor; TNF: tumour necrosis factor; VIP: vasoactive intestinal peptide.

macromolecule excretion may persist even in the case of volumetric balance in the production and absorption of CSF water (Fig. 5).

In patients with iNPH, the CSF production rate may drop to 0.25 ± 0.08 mL/min, which represents a decrease of over 30%.⁴¹ Furthermore, increased ventricle size results in a 30% increase in distribution volume (an increase of over 200 mL in most patients). This, combined with impaired CSF reabsorption, results in a 75% decrease in the CSF turnover rate. Adaptive changes in the choroid plexus and the vascular feet of the astrocytes prevent CSF production-absorption imbalances. Aquaporin-1 expression decreases in the choroid plexus^{42–44} as a result of the activation of the natriuretic peptide system of the circumventricular organs and hypothalamic nuclei^{45,46}; this results in decreased water transport in the apical membrane of the choroid epithelium and, consequently, reduced CSF production. On the other hand, aquaporin-4 expression increases in white matter astrocytes^{42,43,47–49} probably in order to increase CSF water reabsorption into the venous capillaries.^{42,50} However, increased absorption into venous capillaries may reduce periventricular interstitial fluid pressure, promoting the appearance of a pressure gradient between the interstitium and the ventricle and maintaining ventricular dilatation.^{51,52} There is insufficient evidence for conclusions to be drawn on how this situation affects protein clearance in patients with iNPH, although it may be hypothesised that CSF turnover involves at least 2 processes: the reduction of neurotoxin clearance and the interruption of neuroendocrine and paracrine signalling in the CSF.⁵³

Although there is no direct evidence of decreased neurotoxin clearance in patients with iNPH, this may be observed indirectly in the results from biomarker studies and CSF proteomic analysis. Most studies report decreased concentration of the majority of metabolites of the amyloid

proteolytic processing pathway, including amyloid precursor protein (APP) and amyloid- β -42 peptide (A β -42).^{54–58} Total and phosphorylated tau protein levels remain within normal ranges or are lower than normal.^{54,57,59} Decreased CSF turnover results in deficient APP clearance from the interstitial space: APP would therefore be processed by β -secretase and then by γ -secretase, resulting in A β -42 aggregation into amyloid plaques, and decreasing the concentration of these 2 components in the CSF.^{26–29} A study by Fagan et al.,⁶⁰ including patients without dementia, provides evidence in support of this hypothesis. The researchers found that patients with cerebral amyloid deposition in ¹¹C-PiB PET images had lower CSF A β -42 levels. Pyyk \ddot{o} et al.⁶¹ confirmed these results in patients with iNPH, observing a linear, inversely proportional relationship between APP levels in brain biopsy samples and A β -42 concentrations in both ventricular and lumbar intrathecal CSF. Moriya et al.⁶² and Jeppsson et al.⁵⁸ report increased A β -42 levels in the CSF of patients with iNPH after shunt implantation; this increase was correlated with the patients' neurological improvements. These findings suggest that normalisation of CSF flow dynamics after shunting promotes a shift from oligomeric to monomeric A β as a result of increased concentration of A β -38, an isomer with low aggregability.⁶²

The inflammatory profile provides additional data: an excess of acute-phase reactants in the CSF (in the absence of abnormal cellularity according to biochemical analysis or of microglial activation in the periventricular region, which may point to an inflammatory process as the cause of these findings) suggests that decreased CSF clearance is responsible for the accumulation of astrocytic proinflammatory mediators. Several studies have reported increased levels of leucine-rich α -2-glycoprotein,^{63,64} α -1-antichymotrypsin,^{63,65} haptoglobin,⁶³ transferrin,⁶⁶ α -1- β glycoprotein,⁶⁵ and tumour necrosis factor α .⁶⁷ Other

researchers have also observed an increase in free-radical peroxidation products, which may also be associated with decreased CSF turnover.⁶⁸

Li et al.⁶⁹ described an increase in TGF- β -dependent signalling. Although excessive TGF- β may be partially due to deficient TGF- β clearance, this mechanism does not explain TGF- β type II receptor upregulation, another finding reported by these researchers. These findings may be explained by the presence of an adaptive mechanism in response to the increasing levels of inflammatory mediators and acute-phase reactants, since this cytokine has a protective effect, blocking inflammatory response in glial and endothelial cells.^{70–72}

The potential impact of abnormal accumulation of these proteins is evident:

- Reduced APP clearance promotes amyloid deposition in blood vessels and tissues, promoting the development of intercurrent neurodegenerative processes or accelerating their progression (e.g. Alzheimer disease).⁵³
- The diffusion of proinflammatory proteins, especially TNF- α (which promotes aggregation and cell adhesion in capillaries), in the periventricular region may alter microvascular dynamics, compounding the mechanical effect of ventricular dilatation in the periventricular region.¹⁸
- Free radicals damage neurons, glial cells, and endothelial cells, and their effects at the molecular level have been associated with the pathophysiology of a wide range of neurological diseases; it is therefore very likely that they play a role in tissue damage in iNPH.⁶⁸
- TGF- β induces neuronal and oligodendrocytic apoptosis.^{73,74} Delayed programmed cell death may explain the progressive clinical deterioration frequently seen in these patients despite initially successful shunting.

Alterations in neuroendocrine signalling in the CSF constitute another possible mechanism, despite the lack of objective evidence of the existence of this neuroendocrine signalling pathway. Over 100 neuropeptides are described in the literature as potentially using CSF circulation to reach distant regions of the CNS; these neuropeptides are similar in terms of their synthesis, release, and regulation, and their functions are radically different from those of conventional neurotransmitters.⁷⁵

Patients with iNPH have been found to have low CSF levels of somatostatin (SOM),^{76–80} vasoactive intestinal peptide (VIP),^{59,79,81} neuropeptide Y (NPY),^{59,79,80,82} cholecystinin,⁸³ δ sleep-inducing peptide,^{79,84} and corticotropin-releasing hormone.⁸⁰ The interpretation of these findings is not straightforward: they may result from a nearly global dysfunction of peptidergic neurotransmission secondary to iNPH, or may reflect the action of an active mechanism by which deficient CSF turnover would result in the exposure of peptides to the action of neuropeptidases, decreasing CSF peptide levels and interrupting neurotransmission in the CSF.^{85,86}

SOM is probably the peptide that has been studied most extensively in this context. This peptide is diffusely located throughout the brain, although it is mainly produced in the preoptic, paraventricular, arcuate, and

ventromedial nuclei of the hypothalamus. In addition to its involvement in regulating the endocrine system, acting as a growth hormone-releasing hormone antagonist, SOM promotes dopaminergic transmission in the striatum and contributes to normal neuronal function in ageing.⁸⁷ The latter function is linked to the proteolytic activity of neprilysin; this protease is involved in A β -42 catabolism and its activity induces SOM expression.⁸⁸ Reduced SOM expression secondary to impaired CSF turnover may promote A β -42 accumulation, worsening clinical symptoms. NPY is also diffusely distributed throughout the brain, including in such areas as the amygdala, hippocampus, basal ganglia, and of course the hypothalamus, where NPY colocalises with SOM-producing neurons in the paraventricular and arcuate nuclei.⁸⁷ A recent article shows that NPY balances the toxic effects of A β , promoting neurotrophin synthesis in cells.⁸⁹ Decreased NPY levels in patients with iNPH may therefore promote neuronal loss, especially due to the synergistic effect of SOM. VIP has a similar effect: in addition to its function in vasodilation and the synthesis of neurotrophin-3 and activity-dependent neurotrophic factor in glial cells, the peptide inhibits the inflammatory response in the neuroglia, mainly by blocking the production of TNF- α and free radicals in the microglia.⁹⁰

Conclusions

Compression of the periventricular subcortical fibres is not the only pathophysiological mechanism in iNPH. The characteristic, long-term clinical progression of this type of hydrocephalus may also be explained by CBF alterations in the last meadow areas between the perforating branches of the major arteries of the anterior circulation and the subependymal arteries, and by reduced CSF turnover, which leads to reduced neurotoxin clearance and altered neuroendocrine signalling in the CSF.

Conflicts of interest

The authors have no conflicts of interest to declare.

References

1. Morgagni GB. *De sedibus et causis morborum per anatomen indagatis libri quinque. Dissectiones, et animadversiones, nunc primum editas, complectuntur propemodum innumeras, medicis, chirurgis, anatomicis profuturas. Multiplex pr'fixus est index rerum, et nominum accuratissimus. Venetiis: Typographia Remondiniana; 1761.*
2. Gölis LA. *Kranfengeschichte vom Wasserschlage und von der hizigen Gehirnhöhlen wassersucht.* In: Gerold C, editor. *Praktische Abhandlungen über die vorzüglicheren Krankheiten des kindlichen Alters.* Wien; 1815. p. 268–70.
3. Riddoch G. *Progressive dementia without headaches or changes in the optic disks due to tumors of the third ventricle.* *Brain.* 1936;59:225–33.
4. Roger H, Paillas J, Roger J, Tamalet J. *Grande hydrocéphalie latente du vieillard chez une démente artérioscléreuse.* *Rev Neurol (Paris).* 1950;82:437–8.

5. McHugh PR. Occult hydrocephalus. *Q J Med.* 1964;33:297–308.
6. Moulin E. Hydrocéphale passive ou chronique. In: Baillière JB, editor. *Décrite pour la première fois. Paris: Traité de l'apoplexie, ou hémorragie cérébrale; 1819.* p. 117–215.
7. Dörner F [thesis] *De hydrocephalo chronico senili.* University of Würzburg; 1826.
8. Hakim S [Tesis doctoral] *Algunas observaciones sobre la presión del LCR. Síndrome hidrocefálico del adulto con presión normal del LCR.* Bogotá: Pontificia Universidad Javeriana; 1964.
9. Adams RD, Fisher CM, Hakim S, Ojemann RG, Sweet WH. Symptomatic occult hydrocephalus with normal cerebrospinal-fluid pressure, a treatable syndrome. *N Engl J Med.* 1965;273:117–26.
10. Hakim S, Adams RD. The special clinical problem of symptomatic hydrocephalus with normal cerebrospinal fluid pressure. Observations on cerebrospinal fluid hydrodynamics. *J Neurol Sci.* 1965;2:307–27.
11. Osuka S, Matsushita A, Yamamoto T, Saotome K, Isobe T, Nagatomo Y, et al. Evaluation of ventriculomegaly using diffusion tensor imaging: correlations with chronic hydrocephalus and atrophy. *J Neurosurg.* 2010;112:832–9.
12. Assaf Y, Ben-Sira L, Constantini S, Chang LC, Beni-Adani L. Diffusion tensor imaging in hydrocephalus: initial experience. *AJNR Am J Neuroradiol.* 2006;27:1717–24.
13. Hattingen E, Jurcoane A, Melber J, Blasel S, Zanella FE, Neumann-Haefelin T, et al. Diffusion tensor imaging in patients with adult chronic idiopathic hydrocephalus. *Neurosurgery.* 2010;66:917–24.
14. Air EL, Yuan W, Holland SK, Jones BV, Bierbrauer K, Altaye M, et al. Longitudinal comparison of pre- and postoperative diffusion tensor imaging parameters in young children with hydrocephalus. *J Neurosurg Pediatr.* 2010;5:385–91.
15. Lenfeldt N, Larsson A, Nyberg L, Birgander R, Eklund A, Malm J. Diffusion tensor imaging reveals supplementary lesions to frontal white matter in idiopathic normal pressure hydrocephalus. *Neurosurgery.* 2011;68:1586–93.
16. Del Bigio MR. Neuropathology and structural changes in hydrocephalus. *Dev Disabil Res Rev.* 2010;16:16–22.
17. Del Bigio MR. Pathophysiologic consequences of hydrocephalus. *Neurosurg Clin N Am.* 2001;12:639–49, vii.
18. Momjian S, Owler BK, Czosnyka Z, Czosnyka M, Pena A, Pickard JD. Pattern of white matter regional cerebral blood flow and autoregulation in normal pressure hydrocephalus. *Brain.* 2004;127:965–72.
19. Owler BK, Momjian S, Czosnyka Z, Czosnyka M, Pena A, Harris NG, et al. Normal pressure hydrocephalus and cerebral blood flow: a PET study of baseline values. *J Cereb Blood Flow Metab.* 2004;24:17–23.
20. Owler BK, Pickard JD. Normal pressure hydrocephalus and cerebral blood flow: a review. *Acta Neurol Scand.* 2001;104:325–42.
21. Chang CC, Asada H, Mimura T, Suzuki S. A prospective study of cerebral blood flow and cerebrovascular reactivity to acetazolamide in 162 patients with idiopathic normal-pressure hydrocephalus. *J Neurosurg.* 2009;111:610–7.
22. Klinge PM, Brooks DJ, Samii A, Weckesser E, van den Hoff J, Fricke H, et al. Correlates of local cerebral blood flow (CBF) in normal pressure hydrocephalus patients before and after shunting – a retrospective analysis of [(15)O]H(2)O PET-CBF studies in 65 patients. *Clin Neurol Neurosurg.* 2008;110:369–75.
23. Mori K, Maeda M, Asegawa S, Iwata J. Quantitative local cerebral blood flow change after cerebrospinal fluid removal in patients with normal pressure hydrocephalus measured by a double injection method with N-isopropyl-p-[(123)I] iodoamphetamine. *Acta Neurochir (Wien).* 2002;144:255–62.
24. Hertel F, Walter C, Schmitt M, Morsdorf M, Jammers W, Busch HP, et al. Is a combination of Tc-SPECT or perfusion weighted magnetic resonance imaging with spinal tap test helpful in the diagnosis of normal pressure hydrocephalus? *J Neurol Neurosurg Psychiatry.* 2003;74:479–84.
25. Takaya M, Kazui H, Tokunaga H, Yoshida T, Kito Y, Wada T, et al. Global cerebral hypoperfusion in preclinical stage of idiopathic normal pressure hydrocephalus. *J Neurol Sci.* 2010;298:35–41.
26. Silverberg GD, Messier AA, Miller MC, Machan JT, Majmudar SS, Stopa EG, et al. Amyloid efflux transporter expression at the blood-brain barrier declines in normal aging. *J Neuropathol Exp Neurol.* 2010;69:1034–43.
27. Silverberg GD, Miller MC, Machan JT, Johanson CE, Caralopoulos IN, Pascale CL, et al. Amyloid and Tau accumulate in the brains of aged hydrocephalic rats. *Brain Res.* 2010;1317:286–96.
28. Silverberg GD, Miller MC, Messier AA, Majmudar S, Machan JT, Donahue JE, et al. Amyloid deposition and influx transporter expression at the blood–brain barrier increase in normal aging. *J Neuropathol Exp Neurol.* 2010;69:98–108.
29. Nonaka Y, Miyajima M, Ogino I, Nakajima M, Arai H. Analysis of neuronal cell death in the cerebral cortex of H-Tx rats with compensated hydrocephalus. *J Neurosurg Pediatr.* 2008;1:68–74.
30. Hagmann P, Jonasson L, Maeder P, Thiran JP, Wedeen VJ, Meuli R. Understanding diffusion MR imaging techniques: from scalar diffusion-weighted imaging to diffusion tensor imaging and beyond. *Radiographics.* 2006;26 Suppl 1:S205–23.
31. Osuka S, Matsumura A, Ishikawa E, Matsushita A. Diffusion tensor imaging in patients with adult chronic idiopathic hydrocephalus. *Neurosurgery.* 2010;67:E1474.
32. Kamiya K, Hori M, Miyajima M, Nakajima M, Suzuki Y, Kamagata K, et al. Axon diameter and intra-axonal volume fraction of the corticospinal tract in idiopathic normal pressure hydrocephalus measured by q-space imaging. *PLoS ONE.* 2014;9:e103842.
33. Mamo HL, Meric PC, Ponsin JC, Rey AC, Luft AG, Seylaz JA. Cerebral blood flow in normal pressure hydrocephalus. *Stroke.* 1987;18:1074–80.
34. Mataro M, Poca MA, Salgado-Pineda P, Castell-Conesa J, Sahuquillo J, Diez-Castro MJ, et al. Postsurgical cerebral perfusion changes in idiopathic normal pressure hydrocephalus: a statistical parametric mapping study of SPECT images. *J Nucl Med.* 2003;44:1884–9.
35. Marinkovic S, Gibo H, Filipovic B, Dulejic V, Piscevic I. Microanatomy of the subependymal arteries of the lateral ventricle. *Surg Neurol.* 2005;63:451–8.
36. Marinkovic S, Gibo H, Milisavljevic M, Djulejic V, Jovanovic VT. Microanatomy of the intrachoroidal vasculature of the lateral ventricle. *Neurosurgery.* 2005;57:22–36.
37. Küstermann E, Ebke M, Dolge K, Schwendemann G, Leibfritz D, Herrmann M. Neural correlates of movement disorders in normal pressure hydrocephalus. In: Herrmann M, Thiel CM, editors. *Topics in advance imaging.* Oldenburg: BIS-Verlag; 2007. p. 171–4.
38. Miyamoto J, Imahori Y, Mineura K. Cerebral oxygen metabolism in idiopathic-normal pressure hydrocephalus. *Neurol Res.* 2007;29:830–4.
39. Miyamoto J, Tatsuzawa K, Inoue Y, Imahori Y, Mineura K. Oxygen metabolism changes in patients with idiopathic normal pressure hydrocephalus before and after shunting operation. *Acta Neurol Scand.* 2007;116:137–43.
40. Xu J, Chen J, Peskind ER, Jin J, Eng J, Pan C, et al. Characterization of proteome of human cerebrospinal fluid. *Int Rev Neurobiol.* 2006;73:29–98.
41. Silverberg GD, Huhn S, Jaffe RA, Chang SD, Saul T, Heit G, et al. Downregulation of cerebrospinal fluid production in patients with chronic hydrocephalus. *J Neurosurg.* 2002;97:1271–5.
42. Filippidis AS, Kalani MY, ReKate HL. Hydrocephalus and aquaporins: lessons learned from the bench. *Childs Nerv Syst.* 2011;27:27–33.
43. Paul L, Madan M, Rammling M, Chigurupati S, Chan SL, Pattisapu JV. Expression of aquaporin 1 and 4 in a congenital hydrocephalus rat model. *Neurosurgery.* 2011;68:462–73.

44. Oshio K, Watanabe H, Song Y, Verkman AS, Manley GT. Reduced cerebrospinal fluid production and intracranial pressure in mice lacking choroid plexus water channel Aquaporin-1. *FASEB J*. 2005;19:76–8.
45. Johanson CE, Donahue JE, Spangenberg A, Stopa EG, Duncan JA, Sharma HS. Atrial natriuretic peptide: its putative role in modulating the choroid plexus-CSF system for intracranial pressure regulation. *Acta Neurochir Suppl*. 2006;96:451–6.
46. Preston JE, McMillan PN, Stopa EG, Nashold JR, Duncan JA, Johanson CE. Atrial natriuretic peptide induction of dark epithelial cells in choroid plexus: consistency with the model of CSF downregulation in hydrocephalus. *Eur J Pediatr Surg*. 2003;13 Suppl 1:540–2.
47. Skjolding AD, Rowland IJ, Sogaard LV, Praetorius J, Penkowa M, Juhler M. Hydrocephalus induces dynamic spatiotemporal regulation of aquaporin-4 expression in the rat brain. *Cerebrospinal Fluid Res*. 2010;7:20.
48. Skjolding AD, Holst AV, Broholm H, Laursen H, Juhler M. Differences in distribution and regulation of astrocytic aquaporin-4 in human and rat hydrocephalic brain. *Neuropathol Appl Neurobiol*. 2013;39:179–91.
49. Owler BK, Pitham T, Wang D. Aquaporins: relevance to cerebrospinal fluid physiology and therapeutic potential in hydrocephalus. *Cerebrospinal Fluid Res*. 2010;7:15.
50. Tourdias T, Dragonu I, Fushimi Y, Deloire MS, Boiziau C, Brochet B, et al. Aquaporin 4 correlates with apparent diffusion coefficient and hydrocephalus severity in the rat brain: a combined MRI-histological study. *Neuroimage*. 2009;47:659–66.
51. Wilkie KP, Nagra G, Johnston M. A mathematical analysis of physiological and molecular mechanisms that modulate pressure gradients and facilitate ventricular expansion in hydrocephalus. *Int J Numer Anal Model B*. 2012;316:65–81.
52. Pena A, Harris NG, Bolton MD, Czosnyka M, Pickard JD. Communicating hydrocephalus: the biomechanics of progressive ventricular enlargement revisited. *Acta Neurochir Suppl*. 2002;81:59–63.
53. Silverberg GD, Mayo M, Saul T, Rubenstein E, McGuire D. Alzheimer's disease, normal-pressure hydrocephalus, and senescent changes in CSF circulatory physiology: a hypothesis. *Lancet Neurol*. 2003;2:506–11.
54. Agren-Wilsson A, Lekman A, Sjoberg W, Rosengren L, Blennow K, Bergenheim AT, et al. CSF biomarkers in the evaluation of idiopathic normal pressure hydrocephalus. *Acta Neurol Scand*. 2007;116:333–9.
55. Tarnaris A, Toma AK, Pullen E, Chapman MD, Petzold A, Cipolotti L, et al. Cognitive, biochemical, and imaging profile of patients suffering from idiopathic normal pressure hydrocephalus. *Alzheimers Dement*. 2011;7:501–8.
56. Ray B, Reyes PF, Lahiri DK. Biochemical studies in normal pressure hydrocephalus (NPH) patients: change in CSF levels of amyloid precursor protein (APP), amyloid-beta (Abeta) peptide and phospho-tau. *J Psychiatr Res*. 2011;45:539–47.
57. Lins H, Wichart I, Bancher C, Wallesch CW, Jellinger KA, Rosler N. Immunoreactivities of amyloid beta peptide ([1-42]) and total tau protein in lumbar cerebrospinal fluid of patients with normal pressure hydrocephalus. *J Neural Transm*. 2004;111:273–80.
58. Jeppsson A, Zetterberg H, Blennow K, Wikkelso C. Idiopathic normal-pressure hydrocephalus: Pathophysiology and diagnosis by CSF biomarkers. *Neurology*. 2013;80:1385–92.
59. Tullberg M, Blennow K, Mansson JE, Fredman P, Tisell M, Wikkelso C. Cerebrospinal fluid markers before and after shunting in patients with secondary and idiopathic normal pressure hydrocephalus. *Cerebrospinal Fluid Res*. 2008;5:9.
60. Fagan AM, Mintun MA, Shah AR, Aldea P, Roe CM, Mach RH, et al. Cerebrospinal fluid tau and ptau (181) increase with cortical amyloid deposition in cognitively normal individuals: implications for future clinical trials of Alzheimer's disease. *EMBO Mol Med*. 2009;1:371–80.
61. Pyykko OT, Lumela M, Rummukainen J, Nerg O, Seppala TT, Herukka SK, et al. Cerebrospinal fluid biomarker and brain biopsy findings in idiopathic normal pressure hydrocephalus. *PLoS ONE*. 2014;9:e91974.
62. Moriya M, Miyajima M, Nakajima M, Ogino I, Arai H. Impact of cerebrospinal fluid shunting for idiopathic normal pressure hydrocephalus on the amyloid cascade. *PLoS ONE*. 2015;10:e0119973.
63. Li X, Miyajima M, Mineki R, Taka H, Murayama K, Arai H. Analysis of potential diagnostic biomarkers in cerebrospinal fluid of idiopathic normal pressure hydrocephalus by proteomics. *Acta Neurochir (Wien)*. 2006;148:859–64.
64. Nakajima M, Miyajima M, Ogino I, Watanabe M, Miyata H, Karagiozov KL, et al. Leucine-rich alpha-2-glycoprotein is a marker for idiopathic normal pressure hydrocephalus. *Acta Neurochir (Wien)*. 2011;153:1339–46.
65. Scollato A, Terreni A, Caldini A, Salvadori B, Gallina P, Francese S, et al. CSF proteomic analysis in patients with normal pressure hydrocephalus selected for the shunt: CSF biomarkers of response to surgical treatment. *Neurol Sci*. 2010;31:283–91.
66. Futakawa S, Nara K, Miyajima M, Kuno A, Ito H, Kaji H, et al. A unique N-glycan on human transferrin in CSF: A possible biomarker for iNPH. *Neurobiol Aging*. 2012;33:1807–15.
67. Tarkowski E, Tullberg M, Fredman P, Wikkelso C. Normal pressure hydrocephalus triggers intrathecal production of TNF-alpha. *Neurobiol Aging*. 2003;24:707–14.
68. Fersten E, Gordon-Krajcer W, Glowacki M, Mroziak B, Jurkiewicz J, Czernicki Z. Cerebrospinal fluid free-radical peroxidation products and cognitive functioning patterns differentiate varieties of normal pressure hydrocephalus. *Folia Neuropathol*. 2004;42:133–40.
69. Li X, Miyajima M, Jiang C, Arai H. Expression of TGF-betas and TGF-beta type II receptor in cerebrospinal fluid of patients with idiopathic normal pressure hydrocephalus. *Neurosci Lett*. 2007;413:141–4.
70. Finch CE, Laping NJ, Morgan TE, Nichols NR, Pasinetti GM. TGF-beta 1 is an organizer of responses to neurodegeneration. *J Cell Biochem*. 1993;53:314–22.
71. Tesseur I, Wyss-Coray T. A role for TGF-beta signaling in neurodegeneration: Evidence from genetically engineered models. *Curr Alzheimer Res*. 2006;3:505–13.
72. Tesseur I, Zou K, Esposito L, Bard F, Berber E, Can JV, et al. Deficiency in neuronal TGF-beta signaling promotes neurodegeneration and Alzheimer's pathology. *J Clin Invest*. 2006;116:3060–9.
73. Salins P, He Y, Olson K, Glazner G, Kashour T, Amara F. TGF-beta1 is increased in a transgenic mouse model of familial Alzheimer's disease and causes neuronal apoptosis. *Neurosci Lett*. 2008;430:81–6.
74. Schuster N, Bender H, Rossler OG, Philippi A, Dunker N, Thiel G, et al. Transforming growth factor-beta and tumor necrosis factor-alpha cooperate to induce apoptosis in the oligodendroglial cell line OLI-neu. *J Neurosci Res*. 2003;73:324–33.
75. Irani DN. Properties and composition of normal cerebrospinal fluid. In: Irani DN, editor. *Cerebrospinal fluid in clinical practice*. Philadelphia: Saunders-Elsevier; 2009. p. 69–89.
76. Molins A, Catalan R, Sahuquillo J, Castellanos JM, Codina A, Galard R. Somatostatin cerebrospinal fluid levels in dementia. *J Neurol*. 1991;238:168–70.
77. Cramer H, Schaudt D, Rissler K, Strubel D, Warter JM, Kuntzmann F. Somatostatin-like immunoreactivity and substance-P-like immunoreactivity in the CSF of patients with senile dementia of Alzheimer type, multi-infarct syndrome and communicating hydrocephalus. *J Neurol*. 1985;232:346–51.
78. Rissler K, Cramer H, Schaudt D, Strubel D, Gattaz WF. Molecular size distribution of somatostatin-like immunoreactivity in the

- cerebrospinal fluid of patients with degenerative brain disease. *Neurosci Res.* 1986;3:213–25.
79. Wikkelso C, Ekman R, Westergren I, Johansson B. Neuropeptides in cerebrospinal fluid in normal-pressure hydrocephalus and dementia. *Eur Neurol.* 1991;31:88–93.
 80. Poca MA, Mataro M, Sahuquillo J, Catalan R, Ibanez J, Galard R. Shunt related changes in somatostatin, neuropeptide Y, and corticotropin releasing factor concentrations in patients with normal pressure hydrocephalus. *J Neurol Neurosurg Psychiatry.* 2001;70:298–304.
 81. Wikkelso C, Fahrenkrug J, Blomstrand C, Johansson BB. Dementia of different etiologies: vasoactive intestinal polypeptide in CSF. *Neurology.* 1985;35:592–5.
 82. Catalan R, Sahuquillo J, Poca MA, Molins A, Castellanos JM, Galard R. Neuropeptide Y cerebrospinal fluid levels in patients with normal pressure hydrocephalus syndrome. *Biol Psychiatry.* 1994;36:61–3.
 83. Galard R, Poca MA, Catalan R, Tintore M, Castellanos JM, Sahuquillo J. Decreased cholecystokinin levels in cerebrospinal fluid of patients with adult chronic hydrocephalus syndrome. *Biol Psychiatry.* 1997;41:804–9.
 84. Ernst A, Cramer H, Strubel D, Kuntzmann F, Schoenenberger GA. Comparison of DSIP- (delta sleep-inducing peptide) and P-DSIP-like (phosphorylated) immunoreactivity in cerebrospinal fluid of patients with senile dementia of Alzheimer type, multi-infarct syndrome, communicating hydrocephalus and Parkinson's disease. *J Neurol.* 1987;235:16–21.
 85. Waters SM, Davis TP. Alterations of peptide metabolism and neuropeptidase activity in senile dementia of the Alzheimer's type. *Ann N Y Acad Sci.* 1997;814:30–9.
 86. Saito T, Takaki Y, Iwata N, Trojanowski J, Saido TC. Alzheimer's disease, neuropeptides, neuropeptidase, and amyloid-beta peptide metabolism. *Sci Aging Knowledge Environ.* 2003;2003:PE1.
 87. Geraciotti TD, Strawn JR, Ekhaton NN, Wortman M, Kasckow J. Neuroregulatory peptides of central nervous system origin: from laboratory to clinic. In: Pfaff DW, Etgen AM, Fahrbach SE, Rubin RT, editors. *Hormones, brain and behavior.* Amsterdam: Academic press/Elsevier Inc.; 2009. p. 2541–96.
 88. Saito T, Iwata N, Tsubuki S, Takaki Y, Takano J, Huang SM, et al. Somatostatin regulates brain amyloid beta peptide Abeta42 through modulation of proteolytic degradation. *Nat Med.* 2005;11:434–9.
 89. Croce N, Dinallo V, Ricci V, Federici G, Caltagirone C, Bernardini S, et al. Neuroprotective effect of neuropeptide Y against beta-amyloid 25-35 toxicity in SH-SY5Y neuroblastoma cells is associated with increased neurotrophin production. *Neurodegener Dis.* 2011;8:300–9.
 90. Dedja ASP, Nowak JZ. Neuroprotective potential of three peptides: PACAP, VIP and PHI. *Pharmacol Rep.* 2005;57:307–20.