



ORIGINAL ARTICLE

Human recombinant erythropoietin reduces sensorimotor dysfunction and cognitive impairment in rat models of chronic kidney disease[☆]



E.E. Reza-Zaldívar^a, S. Sandoval-Avila^a, Y.K. Gutiérrez-Mercado^a,
E. Vázquez-Méndez^a, A.A. Canales-Aguirre^{a,b}, H. Esquivel-Solís^{a,b},
U. Gómez-Pinedo^c, A.L. Márquez-Aguirre^{a,b,*}

^a Biotecnología Médica y Farmacéutica, Centro de Investigación y Asistencia en Tecnología y Diseño del Estado de Jalisco, Guadalajara, Jalisco, Mexico

^b Unidad de Evaluación Preclínica, Centro de Investigación y Asistencia en Tecnología y Diseño del Estado de Jalisco, Guadalajara, Jalisco, Mexico

^c Departamento de Neurología, Laboratorio de Neurociencias, IDISSC, Hospital Clínico San Carlos, Universidad Complutense, Madrid, Spain

Received 7 July 2017; accepted 18 July 2017

Available online 9 April 2019

KEYWORDS

Chronic kidney disease;
Sensorimotor function;
Cognitive impairment;
Erythropoietin

Abstract

Introduction: Chronic kidney disease (CKD) can cause anaemia and neurological disorders. Recombinant human erythropoietin (rHuEPO) is used to manage anaemia in CKD. However, there is little evidence on the effects of rHuEPO on behaviour and cognitive function in CKD. This study aimed to evaluate the impact of rHuEPO in sensorimotor and cognitive functions in a CKD model.

Methods: Male Wistar rats were randomly assigned to 4 groups: control and CKD, with and without rHuEPO treatment (1050 IU per kg body weight, once weekly for 4 weeks). The Morris water maze, open field, and adhesive removal tests were performed simultaneously to kidney damage induction and treatment. Markers of anaemia and renal function were measured at the end of the study.

Results: Treatment with rHuEPO reduced kidney damage and corrected anaemia in rats with CKD. We observed reduced sensorimotor dysfunction in animals with CKD and treated with rHuEPO. These rats also completed the water maze test in a shorter time than the control groups.

[☆] Please cite this article as: Reza-Zaldívar EE, Sandoval-Avila S, Gutiérrez-Mercado YK, Vázquez-Méndez E, Canales-Aguirre AA, Esquivel-Solís H, et al. La eritropoyetina humana recombinante reduce la disfunción sensoriomotora y el deterioro cognitivo en ratas con enfermedad renal crónica. Neurología. 2020;35:147–154.

* Corresponding author.

E-mail address: amarquez@ciatej.mx (A.L. Márquez-Aguirre).

PALABRAS CLAVE

Enfermedad renal crónica;
Función sensoriomotora;
Deterioro cognitivo;
Eritropoyetina

Conclusions: rHuEPO reduces kidney damage, corrects anaemia, and reduces sensorimotor and cognitive dysfunction in animals with CKD.

© 2017 Sociedad Española de Neurología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

La eritropoyetina humana recombinante reduce la disfunción sensoriomotora y el deterioro cognitivo en ratas con enfermedad renal crónica

Resumen

Introducción: La enfermedad renal crónica (ERC) puede provocar anemia e inducir afectaciones neurológicas. La eritropoyetina humana recombinante (rHuEPO) se utiliza en el tratamiento de la anemia en la ERC. Sin embargo, existe poca evidencia de los efectos de la rHuEPO sobre la conducta y las funciones cognitivas en la ERC. El objetivo de este estudio fue evaluar el efecto del tratamiento con rHuEPO sobre las funciones sensoriomotoras y cognitivas en un modelo de ERC.

Métodos: Ratas macho de la cepa Wistar fueron asignadas a 4 grupos: control y ERC, con y sin tratamiento con rHuEPO (1.050 UI/kg de peso, una vez por semana durante 4 semanas). Las pruebas conductuales de laberinto acuático de Morris, campo abierto y cinta adhesiva se realizaron de manera simultánea a la inducción del daño renal y el tratamiento. Mientras que la determinación de marcadores de función renal y anemia se realizaron al término del estudio.

Resultados: El tratamiento con rHuEPO redujo el daño en el riñón y corrigió la anemia en las ratas con ERC. En las pruebas conductuales, el tratamiento con rHuEPO redujo la disfunción sensoriomotora observada en los animales con ERC. Por otra parte, en los animales con ERC y tratamiento con rHuEPO resolvieron el laberinto en menor tiempo en comparación a los grupos control.

Conclusiones: El tratamiento con rHuEPO reduce el daño en el riñón, corrige la anemia y reduce la disfunción sensoriomotora y cognitiva en los animales con ERC.

© 2017 Sociedad Española de Neurología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

Cognitive impairment is a frequent complication of chronic kidney disease (CKD), causing such alterations as attention deficits and difficulties with verbal communication, learning, and memory.¹ According to epidemiological data, over 60% of patients with CKD are highly likely to develop cerebrovascular diseases, which promote cognitive impairment and dementia.^{2–4}

The main pathogenic factors involved in neurological complications of CKD are high concentrations of circulating uraemic toxins,⁵ proinflammatory cytokines, and reactive oxygen species,² which cause alterations in serotonergic and cholinergic neurotransmission, and endothelial inflammation and dysfunction.⁶ These factors can affect all levels of the nervous system, promoting such neurological alterations as uraemic encephalopathy, cognitive problems, and such neuromuscular disorders as mononeuropathy, polyneuropathy, and myopathy.⁷

At advanced stages, patients with kidney disease may present severe anaemia, known as renal anaemia, caused by a deficiency in renal erythropoietin production.^{8–10} Numerous studies have reported that anaemia negatively affects cognitive function^{11,12} due to insufficient cerebral oxygen supply caused by a decrease in the oxygen

transport capacity.¹³ Chronic hypoxia causes toxicity secondary to oxidative stress; combined with the accumulation of uraemic toxins,^{7,14} this accelerates neuronal degeneration.^{12,15} In patients with CKD, anaemia is treated with erythropoiesis-stimulating agents, such as recombinant human erythropoietin (rHuEPO).¹⁰

While erythropoietin plays an essential role in erythropoiesis, it has been reported to have other functions, such as anti-inflammatory¹⁶ and anti-apoptotic activities,¹⁷ stimulating neurogenesis and neuronal differentiation in early developmental stages,¹⁸ and tissue protection in the kidneys¹⁹ and brain.^{20,21}

Both in vitro^{17,22} and in vivo models have demonstrated the neuroprotective effects of this growth factor; in animal models, rHuEPO has promoted angiogenesis, neurogenesis, and the migration of neural precursor cells to the lesion site, and inhibited apoptosis by suppressing glutamate secretion and modulating intracellular calcium.^{23–28} Strikingly, clinical trials have reported that administration of rHuEPO reduces cognitive impairment, with a positive impact on memory and learning problems.^{29–31}

In addition to learning and memory impairment, motor problems are reported in some animal models of CKD.^{32,33} However, there is little evidence on the neuroprotective effects of erythropoietin in CKD. This study aims to

evaluate the effect of rHuEPO on sensorimotor activity, memory, and learning when the agent is administered simultaneously with the induction of kidney damage in a model of CKD.

Methods

Subjects

We used 24 male Wistar rats (Envigo RMS) weighing 250 to 300 g at the beginning of the study. Rats were housed in translucent polycarbonate cages in a vivarium with a temperature of $25 \pm 2^\circ\text{C}$ and a 12:12 hour light-dark cycle; rats had free access to food and water. Rats were handled according to the international ethical standards established in the *Guide for care and use of laboratory animals* and official Mexican guidelines established in regulation NOM-062-ZOO-1999. The institutional ethical approval code for the study is CICUAL-2016-019.

Experimental groups and treatments

The animals were randomly assigned to the following study groups: (1) control + sham ($n=5$); (2) control + rHuEPO ($n=5$); (3) CKD + sham ($n=7$); and (4) CKD + rHuEPO ($n=7$). CKD was induced with adenine, administered orogastrically at 100 mg/kg/day for 28 days.^{34,35} Groups 1 and 3 received a sham treatment (saline solution) and groups 2 and 4 were treated with rHuEPO administered at 1050 IU/kg via dorsal subcutaneous injection once weekly for 4 weeks. All behavioural tests were performed simultaneously with CKD induction and treatment.

Sample collection and determination of biochemical and haematological parameters

At the end of the treatment and behavioural testing period, animals were placed for 24 hours in metabolic cages for urine collection. They were subsequently euthanised with an overdose of sodium pentobarbital (40–60 mg/kg, intraperitoneal administration), and we collected total blood with ethylenediaminetetraacetic acid and serum. Finally, kidneys were dissected, measured, and weighed. Creatinine and urea levels were determined by dry chemistry using a Vitros 250 system (Ortho Clinical Diagnostics); haemoglobin, haematocrit, erythrocyte, and reticulocyte levels were measured with a KX-21N haematology analyser (Sysmex Corporation).

Open field test

The open field test enables general assessment of animals' locomotor activity and anxiety.³⁶ The test was performed in a white open-field arena measuring 50 cm \times 50 cm \times 50 cm. Rats were placed in the centre of the arena and allowed to explore freely for 3 minutes. Video footage was recorded for subsequent analysis. The entire arena was cleaned with 70% ethanol after each rat completed the test. The behavioural parameters evaluated were: (1) exploration

time, (2) latency time, (3) number of escape attempts, and (4) instances of grooming.

Adhesive removal test

This test assesses somatosensory and motor function. Adhesive tape strips are placed on the rat's fore or hind paws and the animal's performance is assessed by measuring the time taken for it to feel and remove the tape.^{37,38} The test was performed 3 times per day for 3 days. Two adhesive tape strips of equal size were placed on the hind paws. Tactile response was determined by measuring the time between initial contact with the tape and removal of the tape. Rats that did not complete the task within 60 seconds were considered unable to remove the tape.

Morris water maze test

The water maze test was performed with a circular pool (110 cm diameter by 60 cm depth) filled with water (mean temperature [SD], 23°C [2°C]), as specified by Tóthová et al.,³⁹ with the modifications described below. The maze was divided virtually into 4 quadrants and a geometric shape was placed on the wall in each quadrant to act as an external clue for orientation. A 10 cm \times 10 cm polycarbonate platform was placed in the centre of one quadrant. In an initial training phase (4 sessions daily for 3 days), rats were allowed to swim freely for one minute (beginning in a different quadrant each time), enabling them to see the platform, 1 cm above the surface of the water; in the final test, the platform was submerged 1 cm below the water level. Animals that were unable to reach the platform within 60 seconds were guided and placed on top of the platform for 20 seconds to enable spatial orientation. Video footage was recorded for subsequent analysis; the parameters evaluated were time taken to reach the platform and distance swum.

Statistical analysis

Data on kidney function and anaemia were analysed using one-way ANOVA; data from the open-field, adhesive removal, and water maze tests were analysed using two-way repeated measures ANOVA with treatment and training days as factors. Statistical significance was set at $P < .05$, and the Tukey post hoc test was applied. Results are expressed as mean (standard deviation).

Results

Treatment with recombinant human erythropoietin reduces kidney damage and corrects renal anaemia in rats with chronic kidney disease

Statistical analysis found that animals with CKD had significantly lower body weight (data not shown), reduced urine creatinine levels ($P < .05$), and increased serum creatinine and urea levels ($P < .05$). These rats also had increased urine volume ($P < .05$) and increased kidney size

Table 1 Kidney function markers.

Parameter	Group			
	Control		CKD	
	Sham	rHuEPO	Sham	rHuEPO
Mean kidney weight (g)	1.14 (0.05)	1.09 (0.05)	2.8 (0.2) ^a	1.8 (0.11) ^b
Mean kidney size (cm)	1.7 (0.09)	1.6 (0.02)	2.3 (0.08) ^a	1.9 (0.04) ^b
Urine volume (mL)	16.7 (1.9)	13.4 (1.7)	66 (7.0) ^a	59.8 (2.8) ^a
Urea (mg/dL)	43.5 (1.5)	38.2 (2.5)	196.6 (33.4) ^a	78.2 (5.3) ^b
Serum creatinine (mg/dL)	0.62 (0.04)	0.63 (0.04)	2.19 (0.42) ^a	0.93 (0.09) ^b
Urine creatinine (mg/dL)	46.75 (8.4)	48.9 (3.7)	19 (1.4) ^a	19.3 (2.2) ^a
GFR (mL/min/g)	0.73 (0.05)	0.67 (0.04)	0.24 (0.08) ^a	0.47 (0.08)

Data are expressed as mean (standard deviation).

Statistical significance was set at $P < .05$ (one-way ANOVA).

^a CKD vs. control.

^b CKD + rHuEPO vs. CKD.

ANOVA: analysis of variance; CKD: chronic kidney disease; GFR: glomerular filtration rate; rHuEPO: recombinant human erythropoietin.

Table 2 Haematological data.

Parameter	Group			
	Control		CKD	
	Sham	rHuEPO	Sham	rHuEPO
Haemoglobin (g/dL)	16.3 (0.14)	17.5 (0.37)	13.3 (0.42) ^a	17.8 (0.89) ^b
Reticulocytes (%)	3.2 (0.4)	3.6 (0.5)	1.2 (0.2) ^a	4 (0.3) ^b
Erythrocytes ($\times 10^6 \mu\text{L}^{-1}$)	8.5 (0.4)	8.4 (0.2)	6.7 (0.1) ^a	8.8 (0.5) ^b
Haematocrit (%)	52.8 (0.3)	57.3 (1.6)	41.3 (1.3) ^a	58.2 (3.4) ^b

Data are expressed as mean (standard deviation).

Statistical significance was set at $P < .05$ (one-way ANOVA).

^a CKD vs. control.

^b CKD + rHuEPO vs. CKD.

ANOVA: analysis of variance; CKD: chronic kidney disease; rHuEPO: recombinant human erythropoietin.

and weight (39% and 152% greater, respectively) compared to control rats. This represents a 66% reduction in glomerular filtration rate (GFR), demonstrating a loss of kidney function in these animals (Table 1). Haematological findings indicated significant reductions in haemoglobin concentration (21%), haematocrit (25%), erythrocytes (21%), and reticulocytes (65%) in CKD rats vs. controls ($P < .05$) (Table 2).

Treatment with rHuEPO significantly reduced the increase in serum creatinine and urea levels in CKD rats ($P < .05$). The treatment also prevented kidney hypertrophy: kidney weight and size were 1.5 times and 1.2 times smaller than in CKD rats not receiving rHuEPO (Table 1). In addition, animals receiving rHuEPO presented decreased urine volume and GFR (9.4% and 33% less, respectively), although these differences were not statistically significant (Table 1). Treatment with rHuEPO in rats with CKD also corrected renal anaemia, with haemoglobin levels, reticulocyte and erythrocyte counts, and haematocrit returning to the levels observed in the control group (Table 2).

Treatment with recombinant human erythropoietin reduces sensorimotor dysfunction in rats with chronic kidney disease

Animals with CKD showed increased latency times in the open field and adhesive removal tests, with increased periods of inactivity compared to control animals ($P < .05$) (Table 3). They also showed significant reductions in exploration time, escape attempts, and instances of grooming in the open-field test ($P < .05$); similarly, they took longer to remove adhesive strips, demonstrating reduced tactile response compared to control rats ($P < .05$) (Table 3).

Treatment with rHuEPO was associated with significant increases in the number of escape attempts and instances of grooming in the open field test ($P < .05$) and reduced time taken to remove adhesive tape strips, compared to animals receiving the sham treatment (Table 3). Interestingly, rHuEPO treatment was associated with decreased latency times in both tests and increased exploration time in CKD rats ($P < .05$) (Fig. 1A).

Table 3 Tests of sensorimotor function.

Test	Parameter	Group			
		Control		CKD	
		Sham	rHuEPO	Sham	rHuEPO
Open-field test	Latency time (s)	47.0 (3.0)	89.8 (7.4)	129.8 (6.0) ^a	97.4 (3.2) ^b
	Exploration time (s)	133.0 (3.0)	90.2 (7.4)	44.2 (1.4) ^a	82.6 (3.2) ^b
	Escape attempts	14.7 (0.8)	10.5 (1.7)	5.8 (0.9) ^a	10.8 (0.7) ^b
	Grooming (instances)	2.3 (0.2)	2.0 (0.3)	0.75 (0.2) ^a	2.3 (0.1) ^b
Adhesive removal test	Latency time (s)	0.75 (0.4)	0	1.9 (0.6) ^a	0
	Removal time (s)	9.0 (0.8)	9.0 (1.1)	22.2 (2.2) ^a	14.1 (1.6) ^b

Data are expressed as mean (standard deviation) for the final test session. Statistical significance was set at $P < .05$ (two-way ANOVA).

^a CKD vs. control.

^b CKD + rHuEPO vs. CKD.

ANOVA: analysis of variance; CKD: chronic kidney disease; rHuEPO: recombinant human erythropoietin.

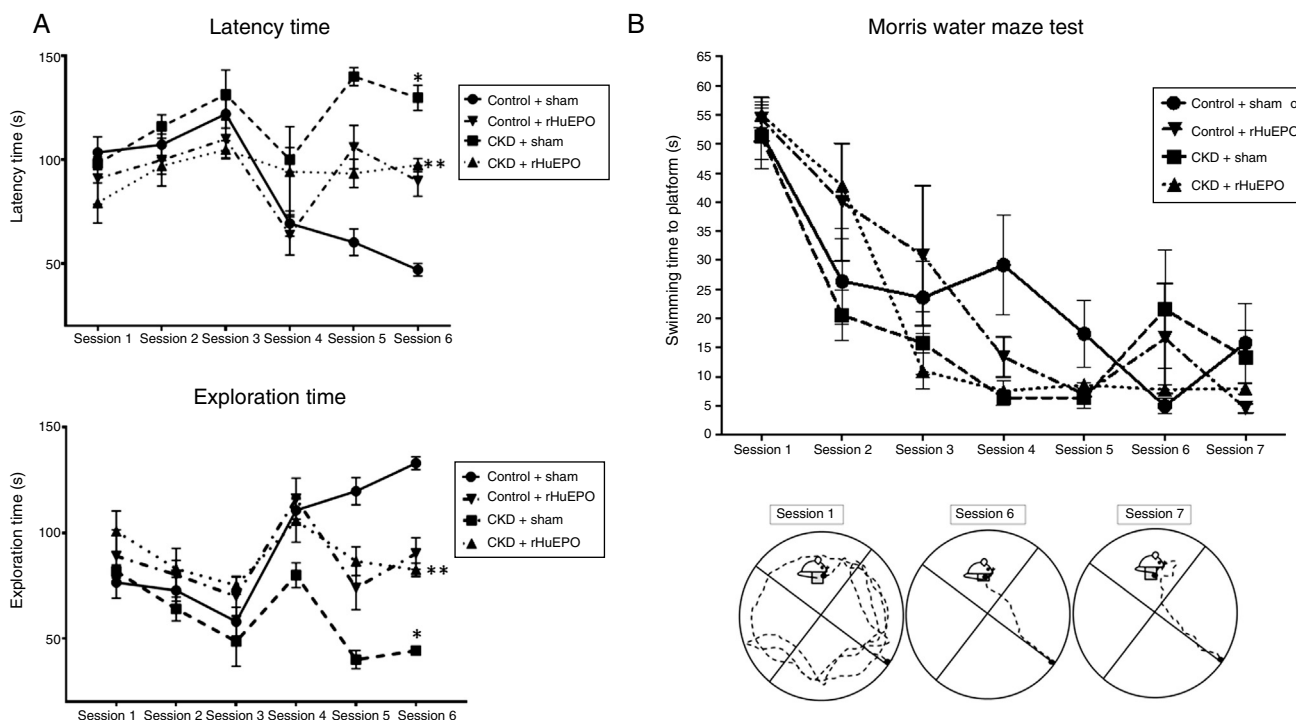


Figure 1 Behavioural tests. (A) Open field test. The graphs show latency and exploration times for each session. (B) Morris water maze test. The graph shows swimming time to platform for each session, and a representative illustration of paths taken to the platform.

Data are expressed as means (error bars denote standard deviation). Statistical significance was set at $P < .05$ (one-way ANOVA).

*Significant difference between CKD and control rats. **Significant difference between CKD + rHuEPO and CKD + sham rats.

ANOVA: analysis of variance; CKD: chronic kidney disease; rHuEPO: recombinant human erythropoietin.

Treatment with recombinant human erythropoietin reduces swimming time to platform in rats with chronic kidney disease

In the water maze test, all groups showed significant reductions in the time taken to swim to the platform between training sessions 1 and 4 ($P < .05$). While all animals showed a significant reduction in swimming time in tests 5 to 7, a consistent reduction was observed from session 3 in CKD rats treated with rHuEPO (Fig. 1B).

Discussion

This study aimed to evaluate the effects of treatment with rHuEPO on sensorimotor and cognitive function in a model of CKD. CKD is characterised by abnormalities in kidney structure and function.⁴⁰ Our findings show that administration of adenine (used to induce CKD) increased serum creatinine and urea concentrations, reduced GFR, and caused renal anaemia; these observations are consistent with those of previous studies of this model.^{32,41} In rats with CKD, rHuEPO

corrected anaemia and decreased kidney damage. Coldewey et al.⁴² report similar findings, with erythropoietin administration attenuating acute kidney dysfunction in a murine model of sepsis.

By performing behavioural tests in parallel with the induction of kidney damage, we were able to observe progressive sensorimotor dysfunction. Rats with CKD showed a gradual increase in latency times in these tests, as well as decreased ability to remove the adhesive tape and reductions in exploration times and the number of escape attempts and instances of grooming. Mazumder et al.⁴³ and Karthick et al.³³ make similar observations, reporting that inactivity times are 2-3 times greater in animals with kidney damage. Behavioural changes in rats with CKD may be attributed to increased plasma urea concentration and reduced GFR, which would cause neurotoxicity due to the accumulation of uraemic toxins, promoting neurodegeneration.^{2,44} They may also be related to anaemia, which contributes to reduced cerebral oxygen supply and consequently greater distribution of uraemic toxins, with an impact on brain metabolism.^{12,45}

While some studies do not report that treatment with erythropoietin has any effect on cognitive function,^{46,47} several studies do observe beneficial effects on motor function, learning, and memory.^{25,27,33,48}

Our results demonstrate that rHuEPO treatment reduces latency time and tape removal time and increases exploration time; this reflects an improvement in the sensorimotor impairment observed in rats with CKD. Similarly, other studies report that preventive treatment with a single dose of erythropoietin improves animals' motor performance in models of traumatic brain injury^{28,46} and ischaemia.⁴⁹ One study of an adenine-induced CKD model found that simultaneous and subsequent treatment with erythropoietin improved the behavioural alterations.³³

Finally, it should be noted that slight alterations were observed in memory and learning during the induction of CKD. As explained by Tóthová et al.,³⁹ the duration of the study period is an important factor in observing cognitive alterations in rats with kidney damage. Other studies report increases in memory and learning alterations several weeks after kidney failure is established.^{50,51} This is explained by the association between cognitive impairment and CKD severity due to uraemia and anaemia, which lead to a state of chronic hypoxia of the brain.⁵²

In conclusion, despite the slight changes in cognitive function at 4 weeks, adenine-induced kidney damage was associated with progressive sensorimotor alterations. Simultaneous treatment with rHuEPO corrected anaemia and reduced uraemia and kidney damage, which may be associated with reductions in sensorimotor dysfunction in animals with CKD.

Funding

This project was supported by the Sectoral Fund for Research in Education (Ciencia Básica SEP-CONACyT; project code 243118); the technical supervisor is Dr Ana Laura Márquez.

Conflicts of interest

The authors have no conflicts of interest to declare.

References

1. Singh NP, Sahni V, Wadhwa A, Garg S, Bajaj SK, Kohli R, et al. Effect of improvement in anemia on electroneurophysiological markers (P300) of cognitive dysfunction in chronic kidney disease. *Hemodial Int.* 2006;10:267–73.
2. Arnold R, Issar T, Krishnan AV, Pussell BA. Neurological complications in chronic kidney disease. *JRSM Cardiovasc Dis.* 2016;5:1–13.
3. Krishnan AV, Kiernan MC. Neurological complications of chronic kidney disease. *Nat Rev Neurol.* 2009;5:542–51.
4. Miranda AS, Cordeiro TM, dos Santos Lacerda Soares TM, Ferreira RN, Simoes ESAC. Kidney-brain axis inflammatory cross-talk: from bench to bedside. *Clin Sci (Lond).* 2017;131:1093–105.
5. Arnold R, Kwai NC, Krishnan AV. Mechanisms of axonal dysfunction in diabetic and uraemic neuropathies. *Clin Neurophysiol.* 2013;124:2079–90.
6. Siassi F, Wang M, Kopple JD, Swendseid ME. Brain serotonin turnover in chronically uremic rats. *Am J Physiol.* 1977;232:E526–8.
7. Brouns R, de Deyn PP. Neurological complications in renal failure: a review. *Clin Neurol Neurosurg.* 2004;107:1–16.
8. Section 2: AKI definition. *Kidney Int Suppl.* 2012;2:19–36.
9. Grimm G, Stockenhuber F, Schneeweiss B, Madl C, Zeitlhofer J, Schneider B. Improvement of brain function in hemodialysis patients treated with erythropoietin. *Kidney Int.* 1990;38:480–6.
10. Bonomini M, del Vecchio L, Sirolli V, Locatelli F. New treatment approaches for the anemia of CKD. *Am J Kidney Dis.* 2016;67:133–42.
11. Mathew RJ, Rabin P, Stone WJ, Wilson WH. Regional cerebral blood flow in dialysis encephalopathy and primary degenerative dementia. *Kidney Int.* 1985;28:64–8.
12. Kurtz P, Schmidt JM, Claassen J, Carrera E, Fernandez L, Helbok R, et al. Anemia is associated with metabolic distress and brain tissue hypoxia after subarachnoid hemorrhage. *Neurocrit Care.* 2010;13:10–6.
13. Kuwabara Y, Sasaki M, Hirakata H, Koga H, Nakagawa M, Chen T, et al. Cerebral blood flow and vasodilatory capacity in anemia secondary to chronic renal failure. *Kidney Int.* 2002;61:564–9.
14. National Guideline C. KDIGO clinical practice guideline for anemia in chronic kidney disease. Rockville MD: Agency for Healthcare Research and Quality (AHRQ); 2012. Available

- from: <https://www.guideline.gov/summaries/summary/38245> [accessed 17.05.17].
15. Hong CH, Falvey C, Harris TB, Simonsick EM, Satterfield S, Ferrucci L, et al. Anemia and risk of dementia in older adults: findings from the Health ABC study. *Neurology*. 2013;81:528–33.
 16. Nairz M, Schroll A, Moschen AR, Sonnweber T, Theurl M, Theurl I, et al. Erythropoietin contrastingly affects bacterial infection and experimental colitis by inhibiting nuclear factor-kappaB-inducible immune pathways. *Immunity*. 2011;34:61–74.
 17. Renzi MJ, Farrell FX, Bittner A, Galindo JE, Morton M, Trinh H, et al. Erythropoietin induces changes in gene expression in PC-12 cells. *Brain Res Mol Brain Res*. 2002;104:86–95.
 18. Arcasoy MO. Non-erythroid effects of erythropoietin. *Haematologica*. 2010;95:1803–5.
 19. Nangaku M. Tissue protection by erythropoietin: new findings in a moving field. *Kidney Int*. 2013;84:427–9.
 20. Arcasoy MO. The non-haematopoietic biological effects of erythropoietin. *Br J Haematol*. 2008;141:14–31.
 21. Lund A, Lundby C, Olsen NV. High-dose erythropoietin for tissue protection. *Eur J Clin Invest*. 2014;44:1230–8.
 22. Yoo SJ, Cho B, Moon C, Yu SW, Moon C. Neuroprotective effects of an erythropoietin-derived peptide in PC12 cells under oxidative stress. *CNS Neurol Disord Drug Targets*. 2016;15:927–34.
 23. Nguyen AQ, Cherry BH, Scott GF, Ryou M-G, Mallet RT. Erythropoietin: powerful protection of ischemic and post-ischemic brain. *Exp Biol Med*. 2014;239:1461–75.
 24. Wang R, Zhao H, Li J, Duan Y, Fan Z, Tao Z, et al. Erythropoietin attenuates axonal injury after middle cerebral artery occlusion in mice. *Neurol Res*. 2017;39:545–51.
 25. Fan X, Heijnen CJ, van der KM, Groenendaal F, van Bel F. Beneficial effect of erythropoietin on sensorimotor function and white matter after hypoxia-ischemia in neonatal mice. *Pediatr Res*. 2011;69:56–61.
 26. Zhu L, Huang L, Wen Q, Wang T, Qiao L, Jiang L. Recombinant human erythropoietin offers neuroprotection through inducing endogenous erythropoietin receptor and neuroglobin in a neonatal rat model of periventricular white matter damage. *Neurosci Lett*. 2017;650:12–7.
 27. Ning R, Xiong Y, Mahmood A, Zhang Y, Meng Y, Qu C, et al. Erythropoietin promotes neurovascular remodeling and long-term functional recovery in rats following traumatic brain injury. *Brain Res*. 2011;1384:140–50.
 28. Ponce LL, Navarro JC, Ahmed O, Robertson CS. Erythropoietin neuroprotection with traumatic brain injury. *Pathophysiology*. 2013;20:31–8.
 29. Van der Kooij MA, Groenendaal F, Kavelaars A, Heijnen CJ, van Bel F. Neuroprotective properties and mechanisms of erythropoietin in in vitro and in vivo experimental models for hypoxia/ischemia. *Brain Res Rev*. 2008;59:22–33.
 30. Kristensen PL, Pedersen-Bjergaard U, Kjaer TW, Olsen NV, Dela F, Holst JJ, et al. Influence of erythropoietin on cognitive performance during experimental hypoglycemia in patients with type 1 diabetes mellitus: a randomized cross-over trial. *PLOS ONE*. 2013;8:e59672.
 31. Kristensen PL, Hoi-Hansen T, Olsen NV, Pedersen-Bjergaard U, Thorsteinsson B. Erythropoietin during hypoglycaemia in type 1 diabetes: relation to basal renin-angiotensin system activity and cognitive function. *Diabetes Res Clin Pract*. 2009;85:75–84.
 32. Ali BH, Al Za'abi M, Ramkumar A, Yasin J, Nemmar A. Anemia in adenine-induced chronic renal failure and the influence of treatment with gum acacia thereon. *Physiol Res*. 2014;63:351–8.
 33. Karthick N, Alwin D, Poornima K, Chitra V, Saravanan A, Balakrishnan D, et al. Neurobehavioral alterations and brain creatine kinase system changes in chronic renal failure induced male wistar rats: impact of erythropoietin supplementation. *J Bioequiv Availab*. 2015;7:74.
 34. Philips FS, Bendich A, Thiersch JB. Adenine intoxication in relation to in vivo formation and deposition of 2,8-dioxyadenine in renal tubules. *J Pharmacol Exp Ther*. 1952;104:20–30.
 35. Rivera-Valdés JJ, García-Benavides L, Armendáriz-Borunda J, Sandoval-Rodríguez A. 311-Human adipose derived stem cells reduce fibrosis in an experimental model of chronic kidney damage. *Cytotherapy*. 2017;19:S226, 5, Suppl.
 36. Gould TD, Dao DT, Kovacsics CE. The open field test. In: Gould TD, editor. *Mood and anxiety related phenotypes in mice: characterization using behavioral tests*. Totowa, NJ: Humana Press; 2009. p. 1–20.
 37. Franco ECS, Cardoso MM, Gouvêia A, Pereira A, Gomes-Leal W. Modulation of microglial activation enhances neuroprotection and functional recovery derived from bone marrow mononuclear cell transplantation after cortical ischemia. *Neurosci Res*. 2012;73:122–32.
 38. Bouet V, Freret T, Toutain J, Divoux D, Boulouard M, Schumann-Bard P. Sensorimotor and cognitive deficits after transient middle cerebral artery occlusion in the mouse. *Exp Neurol*. 2007;203:555–67.
 39. Tothova L, Babickova J, Borbelyova V, Filova B, Sebekova K, Hodosy J. Chronic renal insufficiency does not induce behavioral and cognitive alteration in rats. *Physiol Behav*. 2015;138:133–40.
 40. Inker LA, Astor BC, Fox CH, Isakova T, Lash JP, Peralta CA, et al. KDOQI US commentary on the 2012 KDIGO clinical practice guideline for the evaluation and management of CKD. *Am J Kidney Dis*. 2014;63:713–35.
 41. Diwan V, Mistry A, Gobe G, Brown L. Adenine-induced chronic kidney and cardiovascular damage in rats. *J Pharmacol Toxicol Methods*. 2013;68:197–207.
 42. Coldewey SM, Khan AI, Kapoor A, Collino M, Rogazzo M, Brines M, et al. Erythropoietin attenuates acute kidney dysfunction in murine experimental sepsis by activation of the beta-common receptor. *Kidney Int*. 2013;84:482–90.
 43. Mazumder MK, Giri A, Kumar S, Borah A. A highly reproducible mice model of chronic kidney disease: evidences of behavioural abnormalities and blood-brain barrier disruption. *Life Sci*. 2016;161:27–36.
 44. Watanabe K, Watanabe T, Nakayama M. Cerebro-renal interactions: impact of uremic toxins on cognitive function. *Neurotoxicology*. 2014;44:184–93.
 45. Marsh JT, Brown WS, Wolcott D, Carr CR, Harper R, Schweitzer SV, et al. rHuEPO treatment improves brain and cognitive function of anemic dialysis patients. *Kidney Int*. 1991;39:155–63.
 46. Bramlett HM, Dietrich WD, Dixon CE, Shear DA, Schmid KE, Mondello S, et al. Erythropoietin treatment in traumatic brain injury: operation brain trauma therapy. *J Neurotrauma*. 2016;33:538–52.
 47. Robertson CS, Hannay HJ, Yamal JM, Gopinath S, Goodman JC, Tilley BC, et al. Effect of erythropoietin and transfusion threshold on neurological recovery after traumatic brain injury: a randomized clinical trial. *JAMA*. 2014;312:36–47.
 48. Lu D, Mahmood A, Qu C, Goussev A, Schallert T, Chopp M. Erythropoietin enhances neurogenesis and restores spatial

- memory in rats after traumatic brain injury. *J Neurotrauma*. 2005;22:1011–7.
49. Sadamoto Y, Igase K, Sakanaka M, Sato K, Otsuka H, Sakaki S, et al. Erythropoietin prevents place navigation disability and cortical infarction in rats with permanent occlusion of the middle cerebral artery. *Biochem Biophys Res Commun*. 1998;253:26–32.
 50. Ballesta JJ, del Pozo C, Castelló-Banyuls J, Faura CC. Selective down-regulation of $\alpha 4\beta 2$ neuronal nicotinic acetylcholine receptors in the brain of uremic rats with cognitive impairment. *Exp Neurol*. 2012;236:28–33.
 51. Fujisaki K, Tsuruya K, Yamato M, Toyonaga J, Noguchi H, Nakano T, et al. Cerebral oxidative stress induces spatial working memory dysfunction in uremic mice: neuroprotective effect of tempol. *Nephrol Dial Transplant*. 2014;29:529–38.
 52. Chandanathil MI, Upadhya S, Upadhya S, Bhat G. Psychomotor functions at various weeks of chronic renal failure in rats. *Cogn Neurodyn*. 2015;9:201–11.