



## REVIEW ARTICLE

## Is there a halo-enzymopathy in Parkinson's disease?☆



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## PALABRAS CLAVE

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Oxiácido

**Abstract** Laboratory studies identified changes in the metabolism of halogens in the serum and cerebrospinal fluid (CSF) of patients with Parkinson's disease, which indicates the presence of "accelerated self-halogenation" of CSF and/or an increase in haloperoxidases, specifically serum thyroperoxidase and CSF lactoperoxidase. Furthermore, an excess of some halogenated derivatives, such as advanced oxygenation protein products (AOPP), has been detected in the CSF and serum. "Accelerated self-halogenation" and increased levels of haloperoxidases and AOPP proteins indicate that halogenative stress is present in Parkinson's disease. In addition, 3-iodo-L-tyrosine, a halogenated derivative, shows "parkinsonian" toxicity in experimental models, since it has been observed to induce  $\alpha$ -synuclein aggregation and damage to dopaminergic neurons in the mouse brain and intestine. The hypothesis is that patients with Parkinson's disease display halogenative stress related to a haloenzymatic alteration of the synthesis or degradation of oxyacid of halogens and their halogenated derivatives. This halogenative stress would be related to nervous system damage.

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## ¿Presenta la enfermedad de Parkinson una haloenzimopatía?

**Resumen** Los estudios en el laboratorio han permitido identificar cambios del metabolismo de halógenos en suero y líquido cefalorraquídeo (LCR) de pacientes con enfermedad de Parkinson, que indican la presencia de «autohalogenación acelerada» del LCR de los pacientes o aumento de haloperoxidases, en concreto, tiroperoxidasa en sangre y lactoperoxidasa en LCR. Además, se ha detectado un exceso en suero y LCR de algunos derivados halogenados, como proteínas con halogenación avanzada tipo *advanced oxidation protein products* (AOPP). Estos hechos,

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«autohalogenación acelerada» e incremento de haloperoxidases y proteínas AOPP, indican la presencia de estrés halogenativo en la enfermedad de Parkinson. Además, un derivado halogenado, la 3-yodo-L-tirosina, muestra toxicidad parkinsoniana en modelos experimentales, pues se ha observado que induce agregados de  $\alpha$ -sinucleína y daño de las neuronas de dopamina en cerebro e intestino en ratones. La hipótesis que se maneja es que en la enfermedad de Parkinson existe un exceso halogenativo, relacionado con una alteración haloenzimática de síntesis o degradación de oxiácidos de halógenos y sus derivados halogenados. Este estrés halogenativo se relacionaría con el daño del sistema nervioso.

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## Oxidative stress and Parkinson's disease

Oxidative stress is defined as an imbalance between the production of reactive oxygen species and the antioxidant system, and is currently thought to be an important pathogenic mechanism in Parkinson's disease (PD).<sup>1,2</sup> The different types of oxidative stress<sup>3</sup> include peroxidative stress, oxidative stress due to reactive nitrogen species (e.g., nitric oxide [ $\bullet\text{NO}$ ]), and stress due to halogen species (e.g., hypochlorous acid [HOCl]).

Peroxidative stress is caused by excessive superoxide anion ( $\bullet\text{O}_2^-$ ) or hydrogen peroxide ( $\text{H}_2\text{O}_2$ ), and can be detected in the brain tissue, blood, and cerebrospinal fluid (CSF) of patients with PD. The substantia nigra is known to undergo intense peroxidative stress, showing a considerable increase in levels of such oxidative markers as peroxidised lipids,<sup>4</sup> 8-hydroxyguanosine (a marker of oxidative stress to DNA),<sup>5</sup> carbonylated proteins,<sup>6</sup> and advanced lipoxidation end products.<sup>7</sup> CSF studies show significantly reduced activity of numerous antioxidant enzymes related to peroxidation in patients with PD.<sup>8</sup>

Excessive  $\bullet\text{NO}$  activity induces 2 types of stress: nitrative stress and S-nitrosylation. In nitration, protein oxidation occurs in tyrosine residues, whereas S-nitrosylation involves cysteine residues. In patients with PD, nitrative stress affects proteins that are highly relevant in the disease, such as manganese superoxide dismutase, tyrosine hydroxylase (TH), and  $\alpha$ -synuclein ( $\alpha\text{SYN}$ ). Nitrated manganese superoxide dismutase is detected in Lewy bodies and in the CSF of these patients.<sup>9</sup> Nitrated superoxide dismutase presents loss of function and causes mitochondrial vacuolation and lipid peroxidation in mice; these phenomena are observed in PD.<sup>10</sup> TH is selectively nitrated in PD, inhibiting its enzymatic function; this may play a pathogenic role.<sup>11,12</sup> The protein 3-nitrotyrosine  $\alpha\text{SYN}$  (3NT-SYN) is a component of Lewy bodies, and represents an anomalous form of  $\alpha\text{SYN}$  that is also involved in PD pathogenesis.<sup>13,14</sup> Recent studies analysing the serum of patients with PD detected nitrative stress, indicated by significant increases in 3-nitrotyrosine and 3NT- $\alpha\text{SYN}$  proteins.<sup>15</sup> Finally, in S-nitrosylation, nitric oxide binds to cysteine thiols, giving rise to nitrosothiol derivatives. This modifies the functionality of numerous proteins involved in PD, such as protein disulfide-isomerase and parkin.<sup>16,17</sup> When

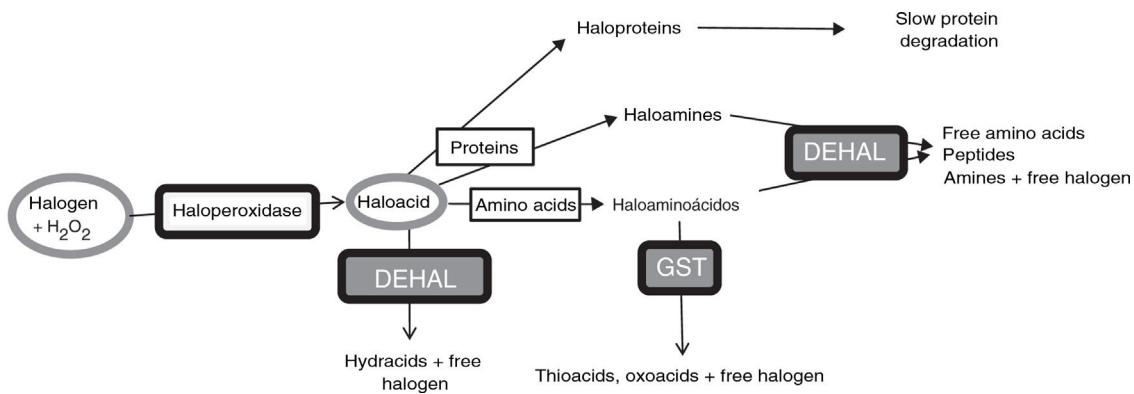
they are nitrosylated, both proteins lose their antioxidant and ubiquitination effects, facilitating protein aggregation and deposition.<sup>18,19</sup>

## Halogenative stress and Parkinson's disease

Halogenative stress is caused by an excess of reactive halogen species, mainly such halogen oxoacids as hypochlorous acid and hypoiodous acid. Haloperoxidases are enzymes that catalyse the conversion of  $\text{H}_2\text{O}_2$  into oxoacids through the incorporation of halogens (Fig. 1).<sup>20–23</sup> Halogen species are increasingly important in understanding the aetiology of PD and other neurodegenerative diseases.<sup>1,3,24–26</sup> Important examples of haloperoxidases are those expressed in white blood cells and microglia, such as myeloperoxidase and eosinophil peroxidase, and in cells presenting iodine/bromine uptake (e.g., cells from the thyroid, salivary glands, or breast), which include thyroid peroxidase (TPO), salivary peroxidase, and lactoperoxidase (LPO). Oxoacids, in turn, halogenate proteins and amino acids, increasing the levels of these derivatives when there is an excess of oxoacids or haloperoxidase activity.

Halogenated amino acids derived from tyrosine, such as chlorotyrosines and iodoxyrosines, are of great interest in the study of PD, as they present dopaminergic neurotoxicity and inhibit tyrosine hydroxylase.<sup>1,25,27–29</sup> Other chloroamino acids, such as chlorocysteine and chlorolysine, also damage dopaminergic neurons by altering membrane proteins.<sup>30</sup> Key members of the family of halogenated proteins are hypochlorite-modified proteins, haloamines, and advanced oxidation protein products (AOPP).<sup>31</sup> Hypochlorite-modified proteins present intense chlorination in cysteine residues. Haloamines are halogenated oligopeptides. AOPPs are anomalous proteins presenting strong halogenation, and particularly chlorination, of tyrosine or lysine residues.

A study conducted at our laboratory detected increased serum TPO concentration in approximately 35% of patients with PD<sup>15</sup> and increased CSF LPO concentration in approximately 43% (unpublished data). These patients often present elevated AOPP levels in serum and CSF.<sup>15</sup> All these findings indicate that patients with PD present halogenative stress. Degradation of halogen oxoacids and their amine derivatives



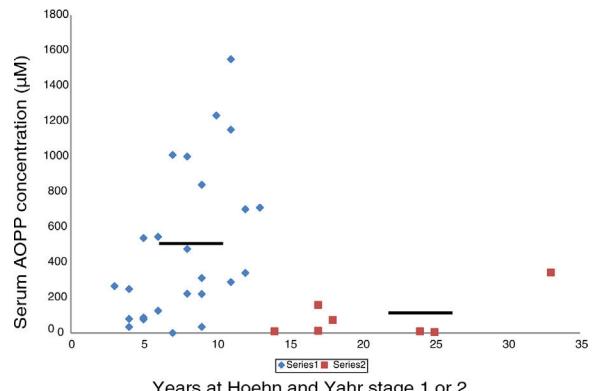
**Figure 1** Metabolic pathways associated with halogenative stress and the proteins and amino acids involved. Haloperoxidases produce oxoacids from halogens and hydrogen peroxide. Oxoacids are degraded to hydracids by the action of dehalogenases, or may halogenate proteins and amino acids. Proteins are converted into haloproteins or haloamines, and amino acids are converted into haloamino acids. Haloproteins degrade slowly. Haloamines and haloamino acids are degraded by dehalogenases, generating amino acids, peptides, and amines; and by glutathione-S-transferase, generating thioacids and oxoacids. Therefore, excessive haloperoxidase activity or a defect of enzymatic degradation may lead to halogenative stress, with elevated levels of oxoacids and halogenated derivatives. DEHAL: dehalogenases; GST: glutathione-S-transferase; H<sub>2</sub>O<sub>2</sub>: hydrogen peroxide.

(haloamino acids and haloamines) is mainly catalysed by dehalogenases (DEHAL), although glutathione-S-transferase is also involved. Iodotyrosine dehalogenase 1 (DEHAL 1) is an important member of this group of enzymes. This protein presents 2 main isoforms (DEHAL 1 and DEHAL 1B); these oxidoreductase enzymes are encoded by the *IYD* gene in humans.<sup>32</sup> Halogen protein derivatives, which usually appear in the context of prolonged situations of halogenative stress (e.g., chronic inflammation or metabolic disorders), degrade slowly and remain present in bodily fluids.<sup>31</sup> Fig. 1 shows the metabolic and halogenative stress pathways described above.

### Anomalous halogenative activity in Parkinson's disease

Laboratory studies have identified halogenation changes in the serum and CSF of patients with PD. First, elevated AOPP levels were detected in the serum. Serum AOPP levels also correlate with the length of time that the disease remains at Hoehn and Yahr stage 2 without passing to more advanced stages (Fig. 2). Patients at stage 2 with over 13 years' disease progression (i.e., those with relatively good motor status) present significantly lower serum AOPP levels. These patients may have more effective antihalogenative mechanisms or less severe halogenative stress. No relevant changes were detected in serum or CSF levels of myeloperoxidase, the haloperoxidase with greatest involvement in AOPP production.<sup>25</sup>

Furthermore, AOPPs were observed in the CSF of patients with PD, whereas they were undetectable in controls. AOPPs were detected in the CSF of approximately 53% of patients, with a mean (standard error of the mean [SEM]) AOPP concentration of 11.4 (2) µM. Hoehn and Yahr stage and



**Figure 2** Relationship between serum levels of advanced oxidation protein products and duration of Parkinson's disease. All patients ( $n=34$ ) were classed as stage 1 or 2 on the Hoehn and Yahr scale. Patients with disease duration longer than 13 years present low serum levels of advanced oxidation protein products (AOPP) (rhombi: duration < 13 years; mean [standard error of the mean] AOPP, 487 [60] µM; squares: duration > 13 years; mean AOPP, 87 [9] µM;  $P<.01$  [ $t$  test]). Continuous lines show the mean value for each patient group. The Hoehn and Yahr scale is commonly used to characterise symptom progression in Parkinson's disease, with scores ranging from 1 (unilateral involvement only) to 5 (confinement to bed or wheelchair unless assisted). Source: modified from García-Moreno et al.<sup>25</sup>

progression time were not correlated with this marker; however, CSF positivity for AOPPs clearly indicates halogenative stress in the central nervous system.

As discussed above, excess haloperoxidase levels may induce halogenative stress, hence the selection of these enzymes as the subject of analysis. We initially studied levels of TPO, which is usually detectable in the blood and

**Table 1** Levels of thyroperoxidase, lactoperoxidase, and myeloperoxidase in the serum and CSF of patients with Parkinson's disease and healthy controls.

	Patients	Controls
<i>TPO, pg/mL</i>		
Serum	1736 (425)*	364 (212)
CSF	Ud	Ud
<i>LPO, ng/mL</i>		
Serum	765.8 (67)	788.2 (65)
CSF	13.2 (1.4)*	8.5 (0.8)
<i>MPO, pg/mL</i>		
Serum	39 638 (6950)	29 202 (9845)
CSF	110 (28)	91 (11)

Data are expressed as mean (standard error of the mean). CSF: cerebrospinal fluid; LPO: lactoperoxidase; MPO: myeloperoxidase; TPO: thyroperoxidase; Ud: undetectable. Source: modified from Fernández et al.<sup>15</sup> and García-Moreno et al.<sup>25</sup>

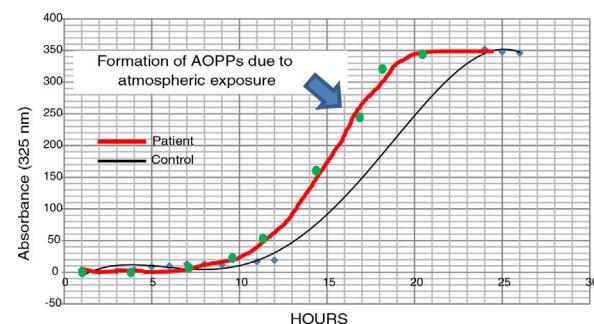
\* P < .05 with respect to controls (t test).

other fluids, as well as in the thyroid gland. Our findings indicate that the mean (SEM) serum TPO concentration is higher in patients with PD than in controls (1736 [425] pg/mL vs 364 [212] pg/mL; P < .05), which was explained by elevated serum TPO levels in approximately 35% of the patients studied (the cut-off point established was 1000 pg/mL, which was not reached in controls). We subsequently analysed levels of LPO, a haloperoxidase that is usually detected in blood, breast milk, and brain tissue. CSF LPO concentration was elevated in approximately 43% of patients studied (13.2 [1.4] ng/mL in patients and 8.5 [0.8] ng/mL in controls; P < .05; unpublished data). Finally, we analysed serum and CSF levels of myeloperoxidase; no significant differences were observed. **Table 1** shows levels of these haloperoxidases in the serum and CSF.

The possible presence of halogenative stress in the CSF of patients with PD was studied using spectrophotometry curves for spontaneous halogenation due to atmospheric exposure. Samples were exposed to the atmosphere and the presence of AOPPs was measured at 325 nm.<sup>31,33</sup> As shown in **Fig. 3**, halogenation of CSF is more rapid in samples from patients than in samples from controls, with the curve displaced to the left. This indicates an "accelerated self-halogenation" effect in the CSF of patients with PD.<sup>26</sup> These findings were observed in approximately 50% of the patients analysed. Levels of AOPPs and LPO were not correlated with accelerated halogenation in the CSF of patients with PD.

### "Parkinsonian" neurotoxicity of 3-iodo-L-tyrosine

The fact that elevated serum TPO and AOPP levels, excessive halogenation, and elevated CSF LPO and AOPP levels have been detected in some patients with PD may suggest increased presence in the blood and CSF of halogenated amine products derived from halogen oxoacids. As pre-



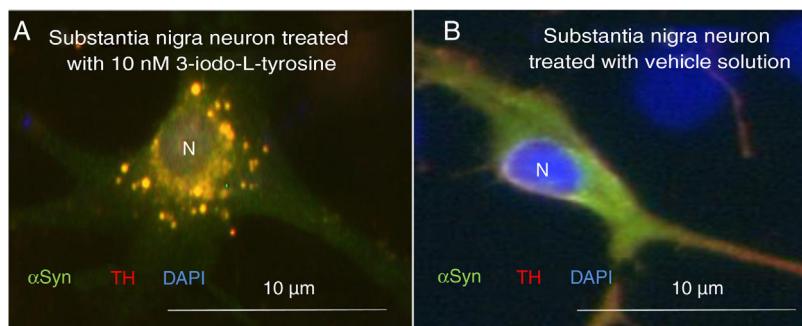
**Figure 3** Spectrophotometry curves for spontaneous halogenation of cerebrospinal fluid due to atmospheric exposure for 27 hours; results from 2 individuals with similar baseline levels and formation of advanced oxidation protein products. The bold line represents data from a patient with Parkinson's disease, and the fine line shows data from a healthy control. Data from the patient is displaced to the left, indicating accelerated self-halogenation.

viously discussed, halogenated amino acids derived from tyrosine, such as chlorotyrosines and iodoxytyrosines, are of great interest in the study of PD, as they present toxicity to dopaminergic neurons and inhibit TH.<sup>25,27,28</sup> At our laboratory, we have studied the possible parkinsonian action of 3-iodo-L-tyrosine in cellular and animal models. Our findings suggest that this molecule induces parkinsonian effects in cellular and animal models, with the appearance of  $\alpha$ SYN inclusions and the death of TH-positive neurons. The fact that 3-iodo-L-tyrosine seems to promote  $\alpha$ SYN aggregation highlights its potential parkinsonian effects, as this mechanism is considered a crucial factor in PD pathogenesis.<sup>34,35</sup> These results were published in a recent study.<sup>29</sup>

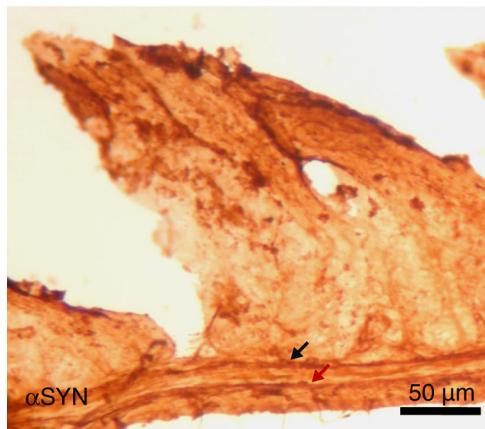
Exposure to 3-iodo-L-tyrosine induced the formation of intraneuronal aggregates expressing  $\alpha$ SYN and TH in dopaminergic neurons from the mouse substantia nigra (**Fig. 4**).

Unilateral injection of 3-iodo-L-tyrosine into the striatum in mice induced damage to the nigro-striatal pathway. The density of TH in the striatum decreased by approximately 30% after the injections, and the number of TH-positive neurons decreased by approximately 35%. The animals also displayed "parkinsonian" behavioural alterations, such as induced spinning behaviour and akinesia/b Bradykinesia.

Finally, the effect of repeated intraperitoneal injection of 3-iodo-L-tyrosine on the jejunal wall was studied in mice. In humans,  $\alpha$ SYN aggregates and degradation of TH-positive neurons are also detected in other peripheral locations of the nervous system, such as in the enteric nervous system, from the oesophagus to the rectum.<sup>34,36,37</sup> As is shown in **Fig. 5**, repeated injection of 10  $\mu$ M of 3-iodo-L-tyrosine induces  $\alpha$ SYN aggregation in the Auerbach plexus and the Meissner plexus, with thickening of nerve fibres; this change was not observed in control subjects injected with vehicle solution. Degeneration of TH-positive neurons and their fibres is also observed. These results are consistent with those reported for experimental models of PD, such as MPTP.



**Figure 4** Immunocytochemistry images showing the distribution of tyrosine hydroxylase (TH) and  $\alpha$ -synuclein ( $\alpha$ SYN) in a culture of dopaminergic neurons from the substantia nigra, treated with 10  $\mu$ M of 3-iodo-L-tyrosine (A) or vehicle solution (B). (A) The cells present numerous round inclusions expressing both  $\alpha$ SYN and TH, hence the light colouration. Numerous aggregates are observed surrounding the nucleus. (B) A neuron treated with vehicle solution, displaying diffuse light signal mainly in the soma, with darker signal for TH in the neurites. In other words, the expression of both proteins is diffuse in the soma, with TH expression being more intense in the neurites. The sample does not present inclusions like those shown in panel A. The nucleus was stained with DAPI. N: nucleus. Source: taken from Fernández-Espejo.<sup>26</sup> Copyright © 2018, *Fisiología*, journal of the Spanish Society of Physiological Sciences.



**Figure 5** Image of the jejunal wall after immunohistochemistry staining for  $\alpha$ -synuclein ( $\alpha$ SYN); sample taken from a mouse treated with 4 weekly intraperitoneal injections of 10  $\mu$ M 3-iodo-L-tyrosine. The Meissner plexus and Auerbach plexus (arrows) are positive for  $\alpha$ SYN, and present thickening, with aggregates of the protein. These findings were not observed in control mice, in which these plexi were thin and expressed little  $\alpha$ SYN.

MPTP is known to cause a loss of 40%–80% of TH-positive neurons in the intestines of mice, as well as the appearance of  $\alpha$ SYN aggregates in intramural plexi in the enteric nervous system.<sup>38–40</sup>

## Conclusions

Laboratory studies have identified changes in the metabolism of halogens in the serum and CSF of patients with PD, suggesting a process of “accelerated self-halogenation” in the CSF and increased levels of haloperoxidase enzymes, which synthesise halogen

oxoacids, and specifically TPO in the serum and LPO in the CSF. Serum analysis has also shown elevated levels of some molecular derivatives of excessive halogenation, including AOPPs. The accelerated self-halogenation process observed and the increased levels of haloperoxidases and AOPPs indicate that halogenative stress is present in PD. Furthermore, 3-iodo-L-tyrosine, a halogen derivative, has shown parkinsonian toxicity in experimental models, inducing  $\alpha$ SYN aggregation and damage to dopaminergic neurons in the mouse brain and intestine.

Based on this evidence, we believe that excess halogenation occurs in PD due to alterations to enzymes responsible for synthesising or degrading halogen oxoacids and their halogenated derivatives. In other words, one of the processes involved in PD pathogenesis may be a halo-enzymopathy that would result in halogenative stress-induced damage to the nervous system.

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## Conflicts of interest

The author has no conflicts of interest to declare.

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## References

- Jenner P. Oxidative stress in Parkinson's disease. *Ann Neurol.* 2003;53 Suppl. 3:S26–36.
- Schapira AH, Olanow CW, Greenamyre JT, Bezard E. Slowing of neurodegeneration in Parkinson's disease and Huntington's disease: future therapeutic perspectives. *Lancet.* 2014;384:545–55.
- Navarro-Yepes J, Burns M, Anandhan A, Khalimonchuk O, del Razo LM, Quintanilla-Vega B, et al. Oxidative stress, redox signaling, and autophagy: cell death versus survival. *Antioxid Redox Signal.* 2014;21:66–85.
- Dexter D, Carter C, Agid F, Agid Y, Lees AJ, Jenner P, et al. Lipid peroxidation as cause of nigral cell death in Parkinson's disease. *Lancet.* 1986;2:639–40.
- Alam ZI, Jenner A, Daniel SE, Lees AJ, Cairns N, Marsden CD, et al. Oxidative DNA damage in the parkinsonian brain: an apparent selective increase in 8-hydroxyguanine levels in substantia nigra. *J Neurochem.* 1997;69:1196–203.
- Floor E, Wetzel MG. Increased protein oxidation in human substantia nigra pars compacta in comparison with basal ganglia and prefrontal cortex measured with an improved dinitrophenylhydrazine assay. *J Neurochem.* 1998;70:2675–82.
- Castellani R, Smith MA, Richey PL, Perry Q. Glycoxidation and oxidative stress in Parkinson disease and diffuse Lewy body disease. *Brain Res.* 1996;737:195–200.
- Martín de Pablos A, García-Moreno JM, Fernández E. Does the cerebrospinal fluid reflect altered redox state but not neurotrophic support loss in Parkinson's disease? *Antioxid Redox Signal.* 2015;23:893–8.
- Aoyama K, Matsubara K, Fujikawa Y, Nagahiro Y, Shimizu K, Umegae N, et al. Nitration of manganese superoxide dismutase in cerebrospinal fluids is a marker for peroxynitrite-mediated oxidative stress in neurodegenerative diseases. *Ann Neurol.* 2000;47:524–7.
- Lebovitz RM, Zhang H, Vogel H, Cartwright J Jr, Dionne L, Lu N, et al. Neurodegeneration myocardial injury, and perinatal death in mitochondrial superoxide dismutase-deficient mice. *Proc Natl Acad Sci USA.* 1996;93:9782–7.
- Blanchard-Fillion B, Souza JM, Friel T, Jiang GC, Vrana K, Sharov V, et al. Nitration and inactivation of tyrosine hydroxylase by peroxynitrite. *J Biol Chem.* 2001;276:46017–23.
- Scholz J, Toska K, Luborzewski A, Maass A, Schünemann V, Haavik J, et al. Endogenous tetrahydroisoquinolines associated with Parkinson's disease mimic the feedback inhibition of tyrosine hydroxylase by catecholamines. *FEBS J.* 2008;275:2109–21.
- Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, et al. Mutation in the  $\alpha$ -synuclein gene identified in families with Parkinson's disease. *Science.* 1997;276:2045–7.
- Papadimitriou A, Veletza V, Hadjigeorgiou GM, Patrikiou A, Hirano M, Anastasopoulos I. Mutated  $\alpha$ -synuclein gene in two Greek kindreds with familial PD: incomplete penetrance? *Neurology.* 1999;52:651–4.
- Fernández E, García-Moreno JM, Martín de Pablos A, Chacón J. May the thyroid gland and thyroperoxidase participate in nitrosylation of serum proteins and sporadic Parkinson's disease? *Antioxid Redox Signal.* 2014;21:2143–8.
- Lipton SA, Choi YB, Pan ZH, Lei SZ, Chen HS, Sucher NJ, et al. A redox-based mechanism for the neuroprotective and neurodestructive effects of nitric oxide and related nitroso-compounds. *Nature.* 1993;364:626–32.
- Stamler JS. Redox signaling: nitrosylation and related target interactions of nitric oxide. *Cell.* 1994;78:931–6.
- Chung KK, Thomas B, Li X, Pletnikova O, Troncoso JC, Marsh L, et al. S-nitrosylation of parkin regulates ubiquitination and compromises parkin's protective function. *Science.* 2004;304:1328–31.
- Uehara T, Nakamura T, Yao D, Shi ZQ, Gu Z, Ma Y, et al. S-nitrosylated protein-disulphide isomerase links protein misfolding to neurodegeneration. *Nature.* 2006;441:513–7.
- Harrison JE, Schultz J. Studies on the chlorinating activity of myeloperoxidase. *J Biol Chem.* 1976;251:1371–4.
- Dunfor HB. Myeloperoxidase and eosinophil peroxidase: phagocytosis and microbial killing. In: Dunfor HB. Nueva York: Heme Peroxidases, Wiley; 1999. p. pp. 349–385.
- Yap YW, Whiteman M, Cheung NS. Chlorinative stress: an under-appreciated mediator of neurodegeneration? *Cell Signal.* 2007;19:219–28.
- Curtis MP, Hicks AJ, Neidigh JW. Kinetics of 3-chlorotyrosine formation and loss due to hypochlorous acid and chloramines. *J Chem Res Toxicol.* 2011;24:418–28.
- Hallwell B, Gutteridge JM. Free radicals in biology and medicine. Nueva York: Oxford University Press; 1999.
- García-Moreno JM, Martín de Pablos A, García-Sánchez MI, Méndez-Lucena C, Damas-Hermoso F, Rus M, et al. May serum levels of advanced oxidized protein products serve as a prognostic marker of disease duration in patients with idiopathic Parkinson's disease? *Antioxid Redox Signal.* 2013;18:1296–302.
- Fernández-Espejo E. Actividad halogenativa anómala en la enfermedad de Parkinson. *Fisiología.* 2018;21:5–8.
- Roskoski R Jr, Wilgus H, Vrana KE. Inactivation of tyrosine hydroxylase by pterin substrates following phosphorylation by cyclic AMP-dependent protein kinase. *Mol Pharmacol.* 1990;38:6–541.
- Prokai D, Nguyen T, Kamrowski K, Chandra A, Talamantes T, Baxter LR, et al. An exploratory evaluation of tyrosine hydroxylase inhibition in planaria as a model for Parkinsonism. *Int J Mol Sci.* 2013;14:23289–96.
- Fernández-Espejo E, Bis-Humbert C. Excess amounts of 3-iodo-l-tyrosine induce Parkinson-like features in experimental approaches of Parkinsonism. *Neurotoxicology.* 2018;67:178–89.
- Robaszkiewicz A, Bartosz G, Soszyński M. N-chloroamino acids cause oxidative protein modifications in the erythrocyte membrane. *Mech Ageing Dev.* 2008;129:572–9.
- Witko-Sarsat V, Nguyen-Khoa T, Jungers P, Drüeke TB, Descamps-Latscha B. Advanced oxidation protein products as a novel marker of oxidative stress in uremia. *Kidney Int.* 1996;49:1304–13.
- Moreno JC, Klootwijk W, van Toor H, Pinto G, D'Alessandro M, Lèger A, et al. Mutations in the iodotyrosine deiodinase gene and hypothyroidism. *N Engl J Med.* 2008;358:1811–8.
- Kontush A, Beisiegel U. Measurement of oxidizability of blood plasma. *Methods Enzymol.* 1999;299:35–49.
- Arizona Parkinson's Disease ConsortiumBeach TG. Multi-organ distribution of phosphorylated alpha-synuclein histopathology in subjects with Lewy body disorders. *Acta Neuropathol.* 2010;119:689–702.
- Lee SJ, Desplats P, Lee HJ, Spencer B, Masliah E. Cell-to-cell transmission of  $\alpha$ -synuclein aggregates. *Methods Mol Biol.* 2012;849:347–59.

36. Fahn S, Halliday GM. Lesions associated with the classic triad of Parkinsonian motor features. In: Halliday G, Barker RA, Rowe DB, editors. Non-dopamine lesions in Parkinson's disease. Oxford: Oxford University Press; 2011. p. 3–17.
37. Del Tredici K, Braak H. Lewy pathology and neurodegeneration in premotor Parkinson's disease. *Mov Disord*. 2012;27:597–607.
38. Tian YM, Chen X, Luo DZ, Zhang XH, Xue H, Zheng LF, et al. Alteration of dopaminergic markers in gastrointestinal tract of different rodent models of Parkinson's disease. *Neuroscience*. 2008;153:634–44.
39. Natale G, Kastsiuchenka O, Pasquali L, Ruggieri S, Paparelli A, Fornai F. MPTP-but not methamphetamine-induced parkinsonism extends to catecholamine neurons in the gut. *Ann N Y Acad Sci*. 2008;1139:345–9.
40. Côté M, Bourque M, Poirier AA, Aubé B, Morissette M, Di Paolo T, et al. GPER1-mediated immunomodulation and neuroprotection in the myenteric plexus of a mouse model of Parkinson's disease. *Neurobiol Dis*. 2015;82:99–113.