



REVIEW

Non-invasive diagnosis of liver fibrosis: A review of current imaging modalities

Ling Wu, Yue Shen, Feng Li*



Department of Gastroenterology and Hepatology, Zhongshan Hospital, Fudan University, Shanghai, PR China

Received 20 June 2019; accepted 20 November 2019

KEYWORDS

Hepatic fibrosis;
Imageology diagnosis;
Noninvasive

Abstract Hundreds of millions of patients are suffering from cirrhosis and other chronic liver diseases worldwide, and this public health problem continues to grow. It has been proven that liver fibrosis is reversible after the elimination of the etiology, especially in the early stage. Thus, early diagnosis of liver fibrosis is of vital importance for clinical treatment. Liver biopsy remains the gold standard for both diagnosis and staging of fibrosis, but is suboptimal, due in large parts to its invasive nature and sundry associated complications. To overcome this, a number of non-invasive diagnosis based on serum biomarkers or imaging modalities have been developed. While diagnosis based on serum biomarkers is cheaper and more acceptable to patients, almost none developed to date are liver-specific, and may engender a false positive error. The imaging modalities have evolved rapidly and are taking on more and more important roles in the diagnosis of liver fibrosis.

© 2020 Elsevier España, S.L.U. All rights reserved.

PALABRAS CLAVE

Fibrosis hepática;
Técnicas de diagnóstico por imagen;
No invasivo

Diagnóstico no invasivo de la fibrosis hepática: una revisión de las técnicas de diagnóstico por imagen actuales

Resumen Cientos de millones de pacientes sufren cirrosis y otras enfermedades hepáticas crónicas en todo el mundo, y este problema de salud pública no cesa de crecer. Se ha demostrado que la fibrosis hepática es reversible tras la eliminación de su etiología, especialmente en una fase temprana. De este modo, el diagnóstico precoz de la fibrosis hepática resulta de crucial importancia para el tratamiento clínico. La biopsia de hígado sigue siendo el método de referencia tanto para el diagnóstico como para la estadificación de la fibrosis, pero se trata

Abbreviations: ADC, apparent diffusion coefficient; ARFI, acoustic radiation force impulse; CT, computed tomography; CTP, CT perfusion; DWI, diffusion-weighted imaging; ECM, extracellular matrix; MMPs, matrix metalloproteinases; MRE, magnetic resonance elastography; MRI, magnetic resonance imaging; NAFLD, non-alcoholic fatty liver disease; TE, transient elastography; US, ultrasound; 2D-SWE, two-dimensional shear wave elastography.

* Corresponding author.

E-mail address: 13661654285@163.com (F. Li).

de un enfoque mejorable, debido en gran medida a su naturaleza invasiva y a las diversas complicaciones asociadas. Para superar estas limitaciones se han desarrollado diversas técnicas diagnósticas no invasivas basadas en biomarcadores séricos o técnicas de diagnóstico por imagen. A pesar de que el diagnóstico basado en biomarcadores séricos es menos costoso y resulta más aceptable para los pacientes, hasta la fecha prácticamente no se ha desarrollado ningún método que sea específico para el hígado, y esto puede dar lugar a falsos positivos. Las técnicas de diagnóstico por imagen han evolucionado rápidamente y están adoptando un papel cada vez más importante en el diagnóstico de la fibrosis hepática.

© 2020 Elsevier España, S.L.U. Todos los derechos reservados.

Introduction

Liver fibrosis comes into being as a reversible result of a sustained or recurrent wound healing response to hepatic injury created by viral, toxic and/or metabolic insult, and represents an imbalance between the synthesis and degeneration of extracellular matrix (ECM).¹ Accompanied by the distortion of hepatic structure and function, cirrhosis is the result of the progression of liver fibrosis. Regardless of the etiology, activated hepatic stellate cells secrete ECM molecules and release matrix metalloproteinases (MMPs) and tissue inhibitors of MMPs in the space of Disse,² which can result in excessive matrix deposition and hepatic fibrogenesis.

According to the latest Global Burden of Disease Study, the global incidence of cirrhosis and other chronic liver diseases in 2017 was 5,154,900.³ Moreover, from 2007 to 2017, the years lived with disability (or 'YLDs') and all-age deaths from cirrhosis have increased by 34.8% and 15.0%,^{3,4} respectively. Hence, the clinical burden of cirrhosis is substantial. It has been validated that liver fibrosis can be reversed after proper treatment of the underlying etiologies, especially at the early stage of fibrosis.⁵ Therefore, early diagnosis and staging of liver fibrosis will benefit the treatment of patients, serving as a determinant in the prognosis of chronic liver disease, and acquiring the dynamic changes of liver fibrosis in a timely manner will be a boost for the clinical treatment approach.

Imaging modalities

The medical imaging is a non-invasive tool with robust diagnostic function. Basic imaging methods, including computed tomography (CT), ultrasound (US) and magnetic resonance imaging (MRI), can provide dependable information about decompensated cirrhosis, while the performance for diagnosing early fibrosis is not so good.⁶ Over the last few decades, quite a number of more advanced imaging modalities have been developed, enabling a prompt evaluation of liver fibrosis and cirrhosis. And, combining different imaging modalities or combining them with serum biomarkers can be a realistic substitution for liver biopsy. Advantages and flaws of different imaging modalities are summarized briefly in [Table 1](#).

CT

Traditionally, CT has often been used in the diagnosis of advanced liver fibrosis and cirrhosis, especially for assessment of liver cirrhosis-associated complications, such as portal hypertension. Based on simplified indices for liver remodeling and attenuation, quantitative CT scores have been shown to have a good performance in predicting significant liver fibrosis, with high areas (0.96–0.97) under the ROC curves.⁷ However, considering the reliance on radiation and inherent lower accuracy compared to other diagnostic methods such as fibroScan and MRE, CT remains an inferior choice for assessing early stages of liver fibrosis. Countless efforts have been taken to improve its accuracy and diagnostic value, for example: employing iodine density measurement in 8-cm detector dual-energy CT, clinicians could assess liver parenchyma hemodynamic changes and evaluate the severity through quantitative indices which correlate positively with Child-Paugh Score.⁸

Contrast-enhanced CT (CECT)

Many researchers have found that CECT may be a pretty good choice in the evaluation of liver fibrosis. Choi et al. developed a deep learning system (based upon a data set including portal venous phase CT images from 7461 patients with pathologically-confirmed liver fibrosis) for staging liver fibrosis with CECT images of the liver. With a high accuracy as it diagnosed significant or more severe fibrosis patients, it did not assess moderate fibrosis.⁹ While, from the study on the rat model through micro-CT, the performance of CECT in assessing early and intermediate fibrosis is satisfying with strong correlations to both the Ishak fibrosis score ($R^2=0.751$, $P<0.01$) and the fibrotic area ($R^2=0.801$, $P<0.01$).¹⁰ Additionally, monitoring pathological angiogenesis and microvasculature alterations could be realized by contrast-enhanced micro-CT.¹¹ Since a sharp increase in sinusoid angiogenesis has been observed during early-stage fibrosis and the vascular reconstruction would happen due to the portal hypertension,¹² CECT might be of potential use in assessing liver fibrosis and the complications.

Table 1 Summary characteristics of imaging modalities.

Imaging modalities	Advantages	Flaws
<i>CT</i>		
Contrast-enhanced CT ¹⁰	Enormous potential in the early diagnosis of liver fibrosis, monitoring pathological angiogenesis.	Lower repeatability compared to US, radiation
CTP ¹³	Short scanning time, high recognition rate, high accuracy in advanced fibrosis and cirrhosis.	
<i>US</i>		
TE ^{22,24}	User-friendly, time-saving, widely-used, low cost, evaluating steatosis by CAP.	Expensive equipment requirements, influenced by many factors such as obesity and food intake.
ARFI ¹⁸	User-friendly, more reliable for severe liver fibrosis or cirrhosis than TE	Expensive equipment requirements, influenced by obesity, sex and age.
CEUS ^{31,34}	Safe for patients with renal failure, real-time evaluation, high temporal and in-plane spatial resolution	false positive HCC diagnosis, microbubble disruption
<i>MRI</i>		
MRE ⁴⁰	High applicability and repeatability, evaluates the whole liver, more accurate than US for significant fibrosis.	Expensive, time-consuming, specialized knowledge requirement, patients with metal implants and psychological illness are precluded.
DWI ^{43,44}	Good performance in patients with sclerosing cholangitis	Influenced by common biologic factors such as inflammation, time-consuming controversial.
T1 ρ mapping ⁴⁹	Resistant to the interference of fatty liver	Novel, lack of robust clinical data.

CT perfusion (CTP)

Based on the changes of substantial microcirculation occurring in liver fibrosis: an increase in the arterial perfusion and a drop in portal and total liver perfusion, CTP allows for a quantitative assessment of the hepatic perfusion.¹³ And according to the transient time (a parameter of CTP) increased significantly between minimal fibrosis and intermediate fibrosis ($P=0.025$), discriminating mild fibrosis from intermediate fibrosis could be realized by CTP.¹³ Combining the measurements of liver and spleen might improve the accuracy of the assessment for liver fibrosis, and the splenic mean transient time (a parameter of splenic perfusion) also changes significantly between different fibrosis stages revealed under the Kruskal–Wallis test ($P<0.001$).¹⁴

US

On the basis of gray-scale findings such as surface nodularity, altered parenchymal echogenicity and heterogeneous echotexture, which reflect the presence of regenerative nodules and fibrous septa, US could help diagnosis liver cirrhosis.¹⁵ And combining the spleen longitudinal diameter, doctors could evaluate portal hypertension in patients with viral hepatitis.¹⁶ It is noteworthy that splenomegaly does not equal portal hypertension, other mechanisms such as reduced lysosomal lipase in patients with non-alcoholic fatty liver disease (NAFLD) could result in splenomegaly.¹⁷ So in case of the splenomegaly detected by US, combining other

examinations is needed for the right estimation of portal hypertension, especially in patients with NAFLD. The advantages of low cost, non-invasiveness, reproducibility, simple application, and non-reliance on ionizing radiation make US an attractive alternative to liver biopsy.¹⁸ However, like traditional CT, its accuracy is not reliable for the diagnosis of early fibrosis, which continues to limit the utility of US.

Transient elastography (TE)

Based on the rationale that the collagen deposits and imparts parenchymal rigidity in livers during fibrogenesis, TE converts this rigidity into a stiffness value to evaluate the fibrosis degree,¹⁹ and it has to be noted that different from other imaging methods, TE presents us with the stiffness measurement of the liver tissue instead of an intuitive picture of the liver, therefore, it is not per se an imaging method.

Numerous researches have validated its accuracy in diagnosing and staging liver fibrosis under a recognized pre-determined set of cut-off values,¹⁹ as well as in identifying the improvement of liver fibrosis under effective treatment of different etiologies.²⁰ Furthermore, the value of TE is a good reference for identifying patients at the risk of possible negative outcomes of liver cirrhosis such as liver decompensation, liver failure, hepatocellular carcinoma,²¹ and at worst, death. Moreover, no more than 5 min is needed to perform TE at patient bedside or in an outpatient clinic, with results obtained immediately,²² doctors often choose TE to

monitor patients longitudinally for evaluating therapeutic effectiveness and modulating treatment plan promptly.²³ Meanwhile, it is important to note that a non-cirrhotic liver may also have a high TE value. Non-cirrhotic portal hypertension conditions such as portal vein thrombosis and nodular regenerative hyperplasia might result in an elevated TE value.²⁴ Recent food intake, abdominal fat and elevated liver enzymes have all been reported as factors with potential for impairing the accuracy of TE.²⁵

Acoustic radiation force impulse (ARFI)

Akin to TE, ARFI is shear wave-based technique, and it is based on an acoustic internal push, known as liver shear wave velocity, that assesses liver stiffness using focused US beams. Converted by Young's modulus: $3\rho v^2$ (ρ and v represent the tissue density and the speed of shear wave respectively), the measured shear wave speed could be represented by the value in kilopascals (kPa) that positively correlated with tissue stiffness.¹⁸ Several researches have thoroughly compared the diagnostic performance of ARFI and TE for the evaluation of liver fibrosis with the conclusion that ARFI is more reliable than TE, especially in patients with ascites,²⁶ the AUROC values and sensitivity of ARFI in diagnosing significant fibrosis could reach over 85%.²⁷ Overall, there are 2 types of ARFI: point shear wave elastography (pSWE) and two-dimensional shear wave elastography (2D-SWE),¹⁸ sampling area and focused energy are the main differences, and the 2D-SWE is the latest elastography used in clinic.

The 2D-SWE creates a real-time, 2D quantitative map of liver stiffness superimposed on a B-mode image, characterized by a higher frame rate to record the shear wave than the conventional US device.²⁸ It integrates the information from the anatomy and stiffness: the shear wave velocities distribution reflects liver elastic properties, and the B-mode imaging represents the liver morphology, presenting us the anatomy specific elastograms of the liver.²⁹ The reliability of it to assess the liver fibrosis is now being researched widely. It has a better performance than TE, probably due to the larger area of interest in 2D-SWE, which is more reflective of the whole picture of the liver; moreover, it could localize to the area of interest exactly. Because the liver stiffness could be influenced by the inflammation due to different etiologies,³⁰ the threshold values that defines should be defined according to etiologies, and different cut-off values according to the etiologies are shown in Table 2. However, the area of interest which could be representative of the whole liver is depended on the operator, and it might cause deviations. It is noteworthy that the fibrosis is not reflected by the stiffness entirely, as the confounding factors such as the inflammation and fluctuations of liver enzymes might influence the stiffness value,³⁰ so it is necessary to combine other examinations such as medical history, clinical symptoms, serum biomarkers and so on.

Contrast-enhanced ultrasound (CEUS)

Akin to CTP, it evaluates liver fibrosis and cirrhosis is based on the liver hemodynamics alteration. Using microbubble as contrast agents, CEUS has intrinsic advantages: On

the one hand, it is safety for patients with renal failure for whom conventional CT or MRI contrast agents are contraindicated³¹; on the other hand, free of interstitial or equilibrium phase, it has a better temporal and in-plane spatial resolution than contrast-enhanced CT and MRI. On the basis of its wash-in and washout patterns, focal liver lesions such as hepatocellular carcinoma can be detected – hypervascularity in the arterial phase and washout in the portal venous or delayed phase.³² For cirrhotic patients that are at a high risk for hepatocellular carcinoma, CEUS could be used during the follow up for the detection of new lesions. Based on portal vein maximum signal intensity, CEUS could diagnosis both and early fibrosis with high specificities in rabbits.³³ CEUS also has limitations: based on US, the value of it depends on skills of operators, breath-hold of the patients and so on. Moreover, factors such as continuous imaging and inappropriate frame rate could result in excessive disruption of microbubble agent.³⁴

MRI

Like US, MRI is another radiation-free imaging modality, and its specificity, sensitivity and accuracy are similar to CT in the diagnosis of liver cirrhosis. In the experimental fibrosis animal model (tetrachloride-induced), T1 and T2 mapping in MRI were found to associated with the severity of liver fibrosis.³⁵ Moreover, a prospective study showed that, by calculating the hepatocyte fraction, the AUROC values of T1 mapping for diagnosis of any ($\geq F1$), significant ($\geq F2$), advanced ($\geq F3$) and cirrhosis ($\geq F4$) were 0.837, 0.890, 0.957, 0.957, respectively.³⁶

As a part of the body unit, liver with pathological changes could impact other visceral organs such as spleen, kidney and cardiac. Measuring the volume, blood flow, perfusion of critical organs could help clinicians evaluate liver fibrosis and cirrhosis in a holistic fashion.³⁷ The hepatic venous pressure gradient (HVPG) is an important index for evaluating the patients with fibrosis or cirrhosis, while invasiveness, high cost and poor reproducibility limit its implement. However, from the research by Bradley et al., apart from the structural changes reflected by prolonged T1 values and hemodynamic changes within the liver reflected by increased total hepatic blood flow and decreased liver perfusion, the HVPG could be estimated by the combination of T1 relaxation time and splenic artery velocity.³⁸ Moreover, renal cortex T1 significantly reduced with disease severity ($P < 0.001$).³⁹ Therefore, MRI could help clinicians make an overall evaluation of cirrhotic patients. However, the accuracy of MRI for diagnosing liver fibrosis at early stages is still not satisfying. Thanks to great efforts of countless researchers, advanced imaging modalities based on MRI with improved performance in the evaluation of liver fibrosis are now being proposed and even developed, overcoming the limitations of diagnosis on the basis of morphological features alone.

Magnetic resonance elastography (MRE)

MRE is a novel non-invasive tool for mapping of tissue elasticity based upon phase contrast, something like the palpation of the liver which can assess its tissue stiffness physically.

Table 2 Optimal cut-off values of TE, 2D-SWE and MRE in classifying liver fibrosis according to different etiologies.

Diagnostic modalities	US														
	TE ^{19,55-58}						2D-SWE ²⁸				MRE ^{43,59,60}				
	All	CHB	CHC	NAFLD	ALD	AIH ¹⁹	All	CHB	CHC	NAFLD	All	CHB	CHC	NAFLD	PSC
Moderate fibrosis (\geq F1)	8	N/A	N/A	7.0	N/A	N/A	N/A	N/A	N/A	N/A	2.46	2.48	2.47	3.45	2.41
Significant fibrosis (\geq F2)	8.5	8.85	7.1	11.0	N/A	5.8	8.25	6.95	7.095	7.15	2.80	2.73	2.73	3.66	3.26
Advanced fibrosis (\geq F3)	8.5	10.80	9	11.4	12	10.4	9.15	8.15	9.15	9.15	3.77	3.76	3.71	4.11	N/A
Cirrhosis (F4)	14.6	17.05	12.2	14.0	15	16.0	9.89	10.90	13.3	11.0	4.09	4.16	3.83	4.71	4.93

US: ultrasonography; TE: transient elastography; 2D-SWE: two-dimensional shear wave elastography; MRE: magnetic resonance elastography; CHB: chronic hepatitis B; CHC: chronic hepatitis C; NAFLD: non-alcoholic fatty liver disease; AIH: auto-immune hepatitis; PSC: primary sclerosing cholangitis.

It can directly visualize and quantitatively measure propagating acoustic strain waves.³⁶ The diagnostic performance of MRE remains promising in clinical practice, not only for diagnosing liver fibrosis, but also for evaluating the patients' survival by clinical end-points such as hepatocellular carcinoma and hepatic decompensation.⁴⁰ Considering the less robust features of traditional MRI for detecting early fibrosis and the better performance of MRE in detecting morphological features in cirrhosis,³¹ MRE is superior for diagnosing patients with suspected liver fibrosis or for evaluating the effectiveness of a treatment. Furthermore, recent studies have reported that MRE is superior to ARFI and TE,⁴⁰ having a higher AUROC than either for identifying liver fibrosis ($P < 0.01$), especially in patients with non-alcoholic fatty liver disease.

Preceding studies and analyses have indicated satisfactory sensitivity, specificity, and accuracy rates of MRE in diagnosing and staging of liver fibrosis; moreover, just like TE, the high rates of repeatability and painlessness⁴¹ of MRE make it useful for longitudinal follow-up of patients with chronic liver disease. However, as every coin has two sides, MRE has some limitations too. Firstly, compared to other imaging modalities, such as TE, it is more expensive, though still cheaper than liver biopsy.⁴¹ Secondly, to obtain an accurate diagnosis through MRE, the clinical practitioner should possess specialized knowledge about MRE, and appropriate judgments should be made under different circumstances, such as severe obesity, massive ascites, liver iron deposition and the use of 3.0-Tesla (T), which can lead to the failure of MRE ($P < 0.004$).⁴² Additionally, metal implants or psychological illness, such as claustrophobia, preclude a patient's ability to be examined by MRE. Because the value from the elastography could be influenced by different etiologies, according to the highest Youden's index (sensitivity + specificity - 1), the optimal cut-off values for different fibrosis stages in different etiologies are shown in Table 2.

Diffusion-weighted imaging (DWI)

DWI is a kind of imaging modality based on the assumption of the free (unrestricted) diffusion of water protons, which measures the apparent diffusion coefficient (ADC). Since excessive ECM deposition hinders the free movement of water molecules,⁴³ the ADC value is inversely

correlated with fibrosis stages as reflected by differing quantifiable measures of the random microscopic motion of water molecules in biological tissue. Conflicting results have been published regarding the accuracy of DWI. Furthermore, findings from the meta-analysis by Wang et al.⁴⁴ challenged the value of DWI, with its accuracy being inferior to MRE when a 1.5-T MRI scanner is used (z test, $P < 0.05$).

Use of a 3.0-T MRI scanner with consecutive b values of 0, 50, 100, 200, 400, 800 s/mm² has shown DWI to perform well in diagnosing and staging of liver fibrosis for primary sclerosing cholangitis cohorts,⁴⁵ to be superior to Gd-EOB-DTPA-enhanced MRI, and to be capable of sufficiently discriminating mild or no fibrosis from moderate fibrosis and cirrhosis ($P < 0.001$, sensitivity of 0.917 and 0.8, respectively). However, a contradictory conclusion was made in another study,⁴⁶ in which use of the same 3.0-T magnetic field at a b value of 500 ns/mm² showed the ADC to be only weakly correlated with fibrosis stages. It has also been reported that when a 3.0-T MRI scanner was used, fibrosis stages were poorly associated with ADC at a b value of 500 s/mm² ($P = 0.27$), while were significantly associated with ADC at a b value of 1000 s/mm² ($P = 0.01$).⁴⁷

Thus, the collective findings reported in the literature to date suggested that use of a 3.0-T MRI scanner with a higher b value may benefit the ability of DWI to assess liver fibrosis more reliably. Disappointingly, it can be influenced by several common biologic, pathologic and physiologic factors, such as liver inflammation, steatosis and perfusion effects, and it is not sensitive enough in differentiating mild fibrosis from moderate fibrosis.⁴⁶

T1 ρ mapping

T1 ρ is the spin-lattice relaxation time constant in the rotating frame in MRI, and is sufficiently sensitive for identifying the motion related to tissue macromolecular composition, such as interaction among water molecules.⁴⁸ In recently years, its application has been extended to detecting and staging liver fibrosis with 1.5-T or 3.0-T MRI scanner. Several preliminary studies have indicated the capabilities of T1 ρ mapping to diagnose and stage fibrosis⁴⁸: T1 ρ values in fibrotic livers were significantly higher compared to healthy livers ($P < 0.05$); at a threshold of 49.5 ms, the sensitivity and specificity of a 3T scanner in predicting liver fibrosis could reach to 77.8% and 100%, respectively, as well as

Table 3 Indexes of different techniques assess different fibrosis degrees.

(A) Diagnostic modalities														
Index	CT Contrast-enhanced CT ⁹													
	Sensitivity (%)				Specificity (%)				Accuracy (%)				AUROC	
Disease	Any CLD related liver fibrosis													
Moderate fibrosis (\geq F1)	N/A				N/A				N/A				N/A	
Significant fibrosis (\geq F2)	95.5				89.9				94.1				0.96	
Advanced fibrosis (\geq F3)	94.6				95.4				95				0.97	
Cirrhosis (F4)	84.6				96.6				92.1				0.95	
(B) Diagnostic modalities														
Index	US													
	TE ²⁴				pSWE ²⁷					2D-SWE ³⁹				
Disease	Cut-off (kPa)	Sensitivity (%)	Specificity (%)	AUROC	Cut-off (m/s)	Sensitivity (%)	Specificity (%)	Accuracy (%)	AUROC	Cut-off (kPa)	Sensitivity (%)	Specificity (%)	Accuracy (%)	AUROC
	NAFLD				CHC					CHB				
Moderate fibrosis (\geq F1)	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Significant fibrosis (\geq F2)	6.1	90	38	0.77	1.36	80.6	87.5	84.1	0.89	7.6	92.0	90.0	92.1	0.97
Advanced fibrosis (\geq F3)	7.1	90	50	0.8	1.45	90.3	87.5	88.5	0.94	9.2	91.6	96.7	93.1	0.96
Cirrhosis (F4)	10.9	90	70	0.89	1.7	90.9	90.3	90.4	0.95	10.4	94.6	94.9	94.7	0.98

Table 3 (Continued)

(C) Diagnostic modalities													
Index	MRI												
	MRE ⁴⁰				T1 ρ mapping ⁴⁸				DWI ⁴⁵				
	Cut-off (kPa)	Sensitivity (%)	Specificity (%)	AUROC	Cut-off (ms)	Sensitivity (%)	Specificity (%)	AUROC	Discrimination	Cut-off (mm ² / s × 10 ⁻³)	Sensitivity (%)	Specificity (%)	AUROC
Disease	NAFLD				CCL4-induced liver fibrosis rabbit models				PSC				
Moderate fibrosis (≥F1)	2 .99	58 .3	90.6	0.799	62.1	83.33	83.33	0.856	F1/0 from F2/3	1 .14	91 .7	82.1	0.926
Significant fibrosis (≥F2)	3 .62	66 .7	95.7	0.885	79.45	69.57	92.31	0.849	F1/0 from F4	1 .09	80	92.9	0.914
Advanced fibrosis (≥F3)	3 .62	90 .5	93.3	0.934	79.44	82.35	84.21	0.799					
Cirrhosis (F4)	4 .15	88 .9	91.4	0.882	92.43	80	69.23	0.692					

Sensitivity, specificity, accuracy and AUROC of contrast-enhanced CT detecting fibrosis in different stages. AUROC: receiver operating characteristic curve; CLD: chronic liver disease; N/A: not applicable.

Sensitivity, specificity and AUROC of TE and ARFI detecting fibrosis in different stages under different cut-off values. AUROC: receiver operating characteristic curve; NAFLD: non-alcoholic fatty liver disease; CHC: chronic hepatitis C; N/A: not applicable.

Sensitivity, specificity and AUROC of MRE, T1 ρ mapping and DWI detecting fibrosis in different stages under different cut-off values. AUROC: receiver operating characteristic curve; NAFLD: non-alcoholic fatty liver disease; PSC: primary sclerosing cholangitis; N/A: not applicable.

with a good interobserver agreement (intraclass correlation coefficient = 0.975). Importantly, its detection may not be affected by the presence of fatty liver.⁴⁹ However, the lack of robust clinical data cannot be ignored and optimization remains an open need, particularly since it has not shown any obvious superiority over the other imaging modalities, such as 2D-SWE.⁴⁸

Molecular imaging techniques

Liver fibrosis is widely recognized as a dynamic process, involving the turnover of ECM. Collagen overload is one of its hallmarks. Low molecular weight (<1 kDa)-based and peptide (<10 kDa)-based agents contributing to the process of ECM turnover have attracted the attention of molecular imaging researchers for their potential to provide better visualization, characterization and measurement of fibrosis process, and thereby to improve the technological approaches for diagnosing and staging of liver fibrosis. The techniques that utilized such agents include, but are not limited to, radiotracer imaging, MRI, MR spectroscopy and optical imaging.⁵⁰

Among these, MRI has been studied extensively in recent years. Studies using a rat model of liver fibrosis have validated the hypothesis that hepatic expression of integrin $\alpha\gamma\beta3$ reflects the activation of hepatic stellate cells and allowed for characterization of the radioiodinated cyclic RGDyk peptide high-affinity binding to both purified and membrane-bound integrin $\alpha\gamma\beta3$.⁵¹ These findings make it possible for subsequent researchers to use integrin $\alpha\gamma\beta3$ as an imaging tracer to visualize and stage liver fibrosis,⁵² and to monitor the progression of liver fibrosis and therapeutic response. Another fascinating probe is EP-3533, comprising a 10 amino acid cyclic peptide conjugated to three gadolinium moieties.⁵³ It has been clearly shown, through use of a rat model, that MRI-based molecular imaging is capable of distinguishing liver fibrosis stages and monitoring therapeutic effectiveness,⁵⁴ suggesting its potential for clinical utility.

Ultimately, this experimental imaging modality is attracting more and more attention of researchers, particularly in consideration of its cost-effectiveness, accuracy, convenience, and feasibility for human application. Yet, there's still a long way to go before the application of molecular imaging modalities in clinical diagnosis of liver fibrosis can be fully realized.

Conclusion

Admittedly, imaging modalities of today cannot rival liver biopsy in accuracy of diagnosing and staging liver fibrosis, and the ultimately definitive diagnosis of liver fibrosis still depends on liver biopsy. Despite myriad studies that have demonstrated superiorities of non-invasive imaging modalities for diagnosing liver fibrosis, there remain some problems. For example, almost all of the studies have been performed by skilled operators, which does not reflect the real-life heterogeneity in clinical practices, and it impacts the generalizability of the findings published and underlies the uncertainty as to whether a similarly good performance

of complex imaging modalities, such as DWI, could be realized in other clinical departments.

However, on the other hand, as compared with liver biopsy, the characteristics of higher cost-effectiveness, better compliance by patients, easier operation, better repeatability and so on, make non-invasive imaging modalities popular for use in diagnosing and evaluating liver fibrosis. The emergence of novel non-invasive techniques will provide more choices to both treating clinicians and patients for the diagnosis of liver fibrosis, particularly as they are being continually improved for better accuracy and greater practical value. Indexes of different techniques assess different fibrosis degrees are concisely described in Table 3.

Almost some of the newest (and promising) alternative techniques, such as molecular imaging techniques, have not yet been established in clinical practice, and their continued development may ultimately provide a foundation upon which even more techniques could be devised. In general, imaging techniques seem to have a great potential as a reliable alternative to liver biopsy. Tremendous efforts are being made to overcome their confounding factors and improve their accuracy in diagnosing liver fibrosis. Certainly, non-invasive imaging modalities represent a tangible hope for more convenient and accurate surveillance of patients with liver fibrosis and cirrhosis in the future.

Funding

This work was supported by the National Natural Science Foundation of China (Nos. 81270007 to Li F. and 81670513 to Li F.); the Shanghai Talent Development Funds (Grant No. 201304 to Li F.); the Shanghai Rising-Star Program (Grant No. 13QA1400700 to Li F.); and the Shanghai Outstanding Young Talent Plan in Health System (Grant No. 13 Y056 to Li F.).

Conflict of interest

None.

Acknowledgements

We are indebted to the National Natural Science Foundation of China (Nos. 81270007 to Li F. and 81670513 to Li F.); the Shanghai Talent Development Funds (Grant No. 201304 to Li F.); the Shanghai Rising-Star Program (Grant No. 13QA14007000 to Li F.); and the Shanghai Outstanding Young Talent Plan in Health System (Grant No. 13 Y056 to Li F.) for financial support. We also wish to thank Filipodia Publishing, LLC, for language editing service in English, making the expression in this manuscript more normative.

References

1. Kisseleva T. The origin of fibrogenic myofibroblasts in fibrotic liver. *Hepatology*. 2017;65:1039–43, <http://dx.doi.org/10.1002/hep.28948>.
2. Matsuda M, Tsurusaki S, Miyata N, Saijou E, Okochi H, Miyajima A, et al. Oncostatin M causes liver fibrosis by regulating cooperation between hepatic stellate cells

- and macrophages in mice. *Hepatology*. 2018;67:296–312, <http://dx.doi.org/10.1002/hep.29421>.
3. James SL, Abate D, Abate KH, Abay SM, Abbafati C, Abbasi N, et al. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet*. 2018;392:1789–858, [http://dx.doi.org/10.1016/S0140-6736\(18\)32279-7](http://dx.doi.org/10.1016/S0140-6736(18)32279-7).
 4. Roth GA, Abate D, Abate KH, Abay SM, Abbafati C, Abbasi N, et al. Global, regional, and national age-sex-specific mortality for 282 causes of death in 195 countries and territories, 1980–2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet*. 2018;392:1736–88, [http://dx.doi.org/10.1016/S0140-6736\(18\)32203-7](http://dx.doi.org/10.1016/S0140-6736(18)32203-7).
 5. Schuppan D, Ashfaq-Khan M, Yang AT, Kim YO. Liver fibrosis: direct antifibrotic agents and targeted therapies. *Matrix Biol*. 2018;68–69:435–51, <http://dx.doi.org/10.1016/j.matbio.2018.04.006>.
 6. Younossi ZM, Loomba R, Anstee QM, Rinella ME, Bugianesi E, Marchesini G, et al. Diagnostic modalities for non-alcoholic fatty liver disease, nonalcoholic steatohepatitis, and associated fibrosis. *Hepatology*. 2018;68:349–60, <http://dx.doi.org/10.1002/hep.29721>.
 7. Obmann VC, Mertineit N, Berzigotti A, Marx C, Ebner L, Kreis R, et al. CT predicts liver fibrosis: prospective evaluation of morphology- and attenuation-based quantitative scores in routine portal venous abdominal scans. *PLoS ONE*. 2018;13:e0199611, <http://dx.doi.org/10.1371/journal.pone.0199611>.
 8. Dong J, He F, Wang L, Yue Z, Wen T, Wang R, et al. Iodine density changes in hepatic and splenic parenchyma in liver cirrhosis with dual energy CT (DECT): a preliminary study. *Acad Radiol*. 2018, <http://dx.doi.org/10.1016/j.acra.2018.08.018>.
 9. Choi KJ, Jang JK, Lee SS, Sung YS, Shim WH, Kim HS, et al. Development and validation of a deep learning system for staging liver fibrosis by using contrast agent-enhanced CT images in the liver. *Radiology*. 2018, <http://dx.doi.org/10.1148/radiol.2018180763>.
 10. Varenika V, Fu Y, Maher JJ, Gao D, Kakar S, Cabarrus MC, et al. Hepatic fibrosis: evaluation with semiquantitative contrast-enhanced CT. *Radiology*. 2013;266:151–8, <http://dx.doi.org/10.1148/radiol.12112452>.
 11. von Stillfried S, Apitzsch JC, Ehling J, Penzkofer T, Mahnken AH, Knuchel R, et al. Contrast-enhanced CT imaging in patients with chronic kidney disease. *Angiogenesis*. 2016;19:525–35, <http://dx.doi.org/10.1007/s10456-016-9524-7>.
 12. Liu L, You Z, Yu H, Zhou L, Zhao H, Yan X, et al. Mechanotransduction-modulated fibrotic microniches reveal the contribution of angiogenesis in liver fibrosis. *Nat Mater*. 2017;16:1252–61, <http://dx.doi.org/10.1038/nmat5024>.
 13. Ronot M, Asselah T, Paradis V, Michoux N, Dorvillius M, Baron G, et al. Liver fibrosis in chronic hepatitis C virus infection: differentiating minimal from intermediate fibrosis with perfusion CT. *Radiology*. 2010;256:135–42, <http://dx.doi.org/10.1148/radiol.10091295>.
 14. Suzuki T, Yamada A, Komatsu D, Kurozumi M, Fujinaga Y, Ueda K, et al. Evaluation of splenic perfusion and spleen size using dynamic computed tomography: usefulness in assessing degree of liver fibrosis. *Hepatol Res*. 2018;48:87–93, <http://dx.doi.org/10.1111/hepr.12900>.
 15. Lurie Y, Webb M, Cytter-Kuint R, Shteingart S, Lederkremer GZ. Non-invasive diagnosis of liver fibrosis and cirrhosis. *World J Gastroenterol*. 2015;21:11567–83, <http://dx.doi.org/10.3748/wjg.v21.i41.11567>.
 16. Mazur R, Celmer M, Silicki J, Holownia D, Pozowski P, Miedzobrodzki K. Clinical applications of spleen ultrasound elastography – a review. *J Ultrasonogr*. 2018;18:37–41, <http://dx.doi.org/10.15557/JoU.2018.0006>.
 17. Polimeni L, Pastori D, Baratta F, Tozzi G, Novo M, Vicinanza R, et al. Spleen dimensions are inversely associated with lysosomal acid lipase activity in patients with non-alcoholic fatty liver disease. *Intern Emerg Med*. 2017;12:1159–65, <http://dx.doi.org/10.1007/s11739-017-1746-1>.
 18. Ferraioli G, Wong VW, Castera L, Berzigotti A, Sporea I, Dietrich CF, et al. Liver ultrasound elastography: an update to the world federation for ultrasound in medicine and biology guidelines and recommendations. *Ultrasound Med Biol*. 2018, <http://dx.doi.org/10.1016/j.ultrasmedbio.2018.07.008>.
 19. Hartl J, Denzer U, Ehken H, Zenouzi R, Peiseler M, Sebode M, et al. Transient elastography in autoimmune hepatitis: timing determines the impact of inflammation and fibrosis. *J Hepatol*. 2016;65:769–75, <http://dx.doi.org/10.1016/j.jhep.2016.05.023>.
 20. Papatheodoridis GV, Idilman R, Dalekos GN, Buti M, Chi H, van Boemmel F, et al. The risk of hepatocellular carcinoma decreases after the first 5 years of entecavir or tenofovir in Caucasians with chronic hepatitis B. *Hepatology*. 2017;66:1444–53, <http://dx.doi.org/10.1002/hep.29320>.
 21. Shen Y, Wu SD, Wu L, Wang SQ, Chen Y, Liu LL, et al. The prognostic role of liver stiffness in patients with chronic liver disease: a systematic review and dose-response meta-analysis. *Hepatol Int*. 2019;13:560–72, <http://dx.doi.org/10.1007/s12072-019-09952-5>.
 22. Eddowes PJ, Sasso M, Allison M, Tsochatzis E, Anstee QM, Sheridan D, et al. Accuracy of fibroscan controlled attenuation parameter and liver stiffness measurement in assessing steatosis and fibrosis in patients with nonalcoholic fatty liver disease. *Gastroenterology*. 2019;156:1717–30, <http://dx.doi.org/10.1053/j.gastro.2019.01.042>.
 23. D’Ambrosio R, Degasperis E, Iavarone M, Sangiovanni A, Aghemo A, Soffredini R, et al. Incidence and predictors of de novo hepatocellular carcinoma in HCV cirrhotic patients treated with direct-acting antivirals: a single-center prospective 3 year study. *J Hepatol*. 2018;68:5529, [http://dx.doi.org/10.1016/S0168-8278\(18\)31309-6](http://dx.doi.org/10.1016/S0168-8278(18)31309-6).
 24. Huang R, Gao ZH, Tang A, Sebastiani G, Deschenes M. Transient elastography is an unreliable marker of liver fibrosis in patients with portal vein thrombosis. *Hepatology*. 2018;68:783–5, <http://dx.doi.org/10.1002/hep.29893>.
 25. Bloom S, Kemp W, Nicoll A, Roberts SK, Gow P, Dev A, et al. Liver stiffness measurement in the primary care setting detects high rates of advanced fibrosis and predicts liver-related events in hepatitis C. *J Hepatol*. 2018;69:575–83, <http://dx.doi.org/10.1016/j.jhep.2018.04.013>.
 26. Zayed N, Darweesh SK, Mousa S, Atef M, Ramzy E, Yosry A. Liver stiffness measurement by acoustic radiation forced impulse and transient elastography in patients with intrahepatic cholestasis. *Eur J Gastroenterol Hepatol*. 2019;31:520–7, <http://dx.doi.org/10.1097/MEG.0000000000001327>.
 27. Alem SA, Abdellatif Z, Mabrouk M, Zayed N, Elsharkawy A, Khairy M, et al. Diagnostic accuracy of acoustic radiation force impulse elastography (ARFI) in comparison to other non-invasive modalities in staging of liver fibrosis in chronic HCV patients: single-center experience. *Abdom Radiol*. 2019;44:2751–8, <http://dx.doi.org/10.1007/s00261-019-02031-1>.
 28. Herrmann E, de Ledinghen V, Cassinotto C, Chu WC, Leung VY, Ferraioli G, et al. Assessment of biopsy-proven liver fibrosis by two-dimensional shear wave elastography: an individual patient data-based meta-analysis. *Hepatology*. 2018;67:260–72, <http://dx.doi.org/10.1002/hep.29179>.
 29. Taljanovic MS, Gimber LH, Becker GW, Latt LD, Klauser AS, Melville DM, et al. Shear-wave elastography: basic physics and

- musculoskeletal applications. *Radiographics*. 2017;37:855–70, <http://dx.doi.org/10.1148/rg.2017160116>.
30. Zhuang Y, Ding H, Zhang Y, Sun H, Xu C, Wang W. Two-dimensional shear-wave elastography performance in the noninvasive evaluation of liver fibrosis in patients with chronic hepatitis B: comparison with serum fibrosis indexes. *Radiology*. 2017;283:873–82, <http://dx.doi.org/10.1148/radiol.2016160131>.
 31. Kim TK, Noh SY, Wilson SR, Kono Y, Piscaglia F, Jang HJ, et al. Contrast-enhanced ultrasound (CEUS) liver imaging reporting and data system (LI-RADS) 2017 – a review of important differences compared to the CT/MRI system. *Clin Mol Hepatol*. 2017;23:280–9, <http://dx.doi.org/10.3350/cmh.2017.0037>.
 32. Wu M, Li L, Wang J, Zhang Y, Guo Q, Li X, et al. Contrast-enhanced US for characterization of focal liver lesions: a comprehensive meta-analysis. *Eur Radiol*. 2018;28:2077–88, <http://dx.doi.org/10.1007/s00330-017-5152-x>.
 33. Qiu T, Wang H, Song J, Ling W, Shi Y, Guo G, et al. Assessment of liver fibrosis by ultrasound elastography and contrast-enhanced ultrasound: a randomized prospective animal study. *Exp Anim*. 2018;67:117–26, <http://dx.doi.org/10.1538/expanim.17-0098>.
 34. Quaia E. State of the art: LI-RADS for contrast-enhanced US. *Radiology*. 2019;293:4–14, <http://dx.doi.org/10.1148/radiol.2019190005>.
 35. Luetkens JA, Klein S, Traber F, Schmeel FC, Sprinkart AM, Kuetting DLR, et al. Quantification of liver fibrosis at T1 and T2 mapping with extracellular volume fraction MRI: preclinical results. *Radiology*. 2018;288:748–54, <http://dx.doi.org/10.1148/radiol.2018180051>.
 36. Eaton JE, Dzyubak B, Venkatesh SK, Smyrk TC, Gores GJ, Ehman RL, et al. Performance of magnetic resonance elastography in primary sclerosing cholangitis. *J Gastroenterol Hepatol*. 2016;31:1184–90, <http://dx.doi.org/10.1111/jgh.13263>.
 37. Moller S, Bendtsen F. The pathophysiology of arterial vasodilatation and hyperdynamic circulation in cirrhosis. *Liver Int*. 2018;38:570–80, <http://dx.doi.org/10.1111/liv.13589>.
 38. Bradley CR, Cox EF, Scott RA, James MW, Kaye P, Aithal GP, et al. Multi-organ assessment of compensated cirrhosis patients using quantitative magnetic resonance imaging. *J Hepatol*. 2018;69:1015–24, <http://dx.doi.org/10.1016/j.jhep.2018.05.037>.
 39. Chouhan MD, Taylor SA, Mookerjee RP. Multi-organ quantitative MRI for the assessment of liver disease – a whole much more than the sum of its parts. *J Hepatol*. 2018;69:996–8, <http://dx.doi.org/10.1016/j.jhep.2018.09.004>.
 40. Cui J, Heba E, Hernandez C, Haufe W, Hooker J, Andre MP, et al. Magnetic resonance elastography is superior to acoustic radiation force impulse for the diagnosis of fibrosis in patients with biopsy-proven nonalcoholic fatty liver disease: a prospective study. *Hepatology*. 2016;63:453–61, <http://dx.doi.org/10.1002/hep.28337>.
 41. Park CC, Nguyen P, Hernandez C, Bettencourt R, Ramirez K, Fortney L, et al. Magnetic resonance elastography vs transient elastography in detection of fibrosis and noninvasive measurement of steatosis in patients with biopsy-proven nonalcoholic fatty liver disease. *Gastroenterology*. 2017;152:598–607, <http://dx.doi.org/10.1053/j.gastro.2016.10.026>, e592.
 42. Wagner M, Corcuera-Solano I, Lo G, Esses S, Liao J, Besa C, et al. Technical failure of MR elastography examinations of the liver: experience from a large single-center study. *Radiology*. 2017;284:401–12, <http://dx.doi.org/10.1148/radiol.2016160863>.
 43. Zhu L, Pan Z, Ma Q, Yang W, Shi H, Fu C, et al. Diffusion kurtosis imaging study of rectal adenocarcinoma associated with histopathologic prognostic factors: preliminary findings. *Radiology*. 2017;284:66–76, <http://dx.doi.org/10.1148/radiol.2016160094>.
 44. Wang QB, Zhu H, Liu HL, Zhang B. Performance of magnetic resonance elastography and diffusion-weighted imaging for the staging of hepatic fibrosis: a meta-analysis. *Hepatology*. 2012;56:239–47, <http://dx.doi.org/10.1002/hep.25610>.
 45. Keller S, Sedlacik J, Schuler T, Buchert R, Avanesov M, Zenouzi R, et al. Prospective comparison of diffusion-weighted MRI and dynamic Gd-EOB-DTPA-enhanced MRI for detection and staging of hepatic fibrosis in primary sclerosing cholangitis. *Eur Radiol*. 2018, <http://dx.doi.org/10.1007/s00330-018-5614-9>.
 46. Watanabe H, Kanematsu M, Goshima S, Kondo H, Onozuka M, Moriyama N, et al. Staging hepatic fibrosis: comparison of gadoxetate disodium-enhanced and diffusion-weighted MR imaging – preliminary observations. *Radiology*. 2011;259:142–50, <http://dx.doi.org/10.1148/radiol.10100621>.
 47. Papalavrentios L, Sinakos E, Chourmouzi D, Hytiroglou P, Drevelegas K, Drevelegas A, et al. 3 Tesla diffusion-weighted MRI for assessing liver fibrosis in nonalcoholic fatty liver disease. *Hepatology*. 2013;58:449–50, <http://dx.doi.org/10.1002/hep.26220>.
 48. Li RK, Ren XP, Yan FH, Qiang JW, Lin HM, Tao W, et al. Liver fibrosis detection and staging: a comparative study of T1rho MR imaging and 2D real-time shear-wave elastography. *Abdom Radiol*. 2018;43:1713–22, <http://dx.doi.org/10.1007/s00261-017-1381-3>.
 49. Xie S, Li Q, Cheng Y, Zhang Y, Zhuo Z, Zhao G, et al. Impact of liver fibrosis and fatty liver on T1rho measurements: a prospective study. *Korean J Radiol*. 2017;18:898–905, <http://dx.doi.org/10.3348/kjr.2017186.898>.
 50. Pomper MG, Lee S. Molecularly targeted MR imaging agent in liver fibrosis: high sensitivity and low gadolinium mean high translational potential. *Radiology*. 2018;287:590–1, <http://dx.doi.org/10.1148/radiol.2018180084>.
 51. Huang XW, Wang JY, Li F, Song ZJ, Xie C, Lu WY. Biochemical characterization of the binding of cyclic RGDyK to hepatic stellate cells. *Biochem Pharmacol*. 2010;80:136–43, <http://dx.doi.org/10.1016/j.bcp.2010.03.015>.
 52. Li F, Song Z, Li Q, Wu J, Wang J, Xie C, et al. Molecular imaging of hepatic stellate cell activity by visualization of hepatic integrin alphavbeta3 expression with SPECT in rat. *Hepatology*. 2011;54:1020–30, <http://dx.doi.org/10.1002/hep.24467>.
 53. Polasek M, Fuchs BC, Uppal R, Schuhle DT, Alford JK, Loving GS, et al. Molecular MR imaging of liver fibrosis: a feasibility study using rat and mouse models. *J Hepatol*. 2012;57:549–55, <http://dx.doi.org/10.1016/j.jhep.2012.04.035>.
 54. Zhu B, Wei L, Rotile N, Day H, Rietz T, Farrar CT, et al. Combined magnetic resonance elastography and collagen molecular magnetic resonance imaging accurately stage liver fibrosis in a rat model. *Hepatology*. 2017;65:1015–25, <http://dx.doi.org/10.1002/hep.28930>.
 55. Meng F, Zheng Y, Zhang Q, Mu X, Xu X, Zhang H, et al. Noninvasive evaluation of liver fibrosis using real-time tissue elastography and transient elastography (FibroScan). *J Ultrasound Med*. 2015;34:403–10, <http://dx.doi.org/10.7863/ultra.34.3.403>.
 56. Elsharkawy A, Alborai M, Fouad R, Asem N, Abdo M, Elmakhzangy H, et al. Establishing ultrasound based transient elastography cutoffs for different stages of hepatic fibrosis and cirrhosis in Egyptian chronic hepatitis C patients. *Arab J Gastroenterol*. 2017;18:210–5, <http://dx.doi.org/10.1016/j.ajg.2017.11.002>.
 57. Imajo K, Kessoku T, Honda Y, Tomeno W, Ogawa Y, Mawatari H, et al. Magnetic resonance imaging more accurately classifies steatosis and fibrosis in patients with nonalcoholic fatty liver disease than transient elastography. *Gastroenterology*. 2016;150:626–37, <http://dx.doi.org/10.1053/j.gastro.2015.11.048>, e7.
 58. Voican CS, Louvet A, Trabut JB, Njike-Nakseu M, Dharancy S, Sanchez A, et al. Transient elastography alone and in

- combination with FibroTest((R)) for the diagnosis of hepatic fibrosis in alcoholic liver disease. *Liver Int.* 2017;37:1697–705, <http://dx.doi.org/10.1111/liv.13440>.
59. Kaswala DH, Lai M, Afdhal NH. Fibrosis assessment in nonalcoholic fatty liver disease (NAFLD) in 2016. *Digest Dis Sci.* 2016;61:1356–64, <http://dx.doi.org/10.1007/s10620-016-4079-4>.
60. Wu WP, Chou CT, Chen RC, Lee CW, Lee KW, Wu HK. Non-invasive evaluation of hepatic fibrosis: the diagnostic performance of magnetic resonance elastography in patients with viral hepatitis B or C. *PLoS ONE.* 2015;10:e0140068, <http://dx.doi.org/10.1371/journal.pone.0140068>.