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SPECIAL ARTICLE

Nitric oxide and related factors linked to oxidation and inflammation as possible biomarkers of heart failure[☆]

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Abstract As a prevalent cardiovascular disease, heart failure is one of the leading causes of morbidity and premature mortality. Therefore, there is a special interest in the study of efficient markers associated with risk and/or prediction of cardiovascular events. Multiple candidates are proposed, especially those involved in oxidative and inflammatory processes typical of cardiovascular disease, such as superoxide anion, nitric oxide, and peroxynitrite. There is a lack of knowledge on the potential usefulness of these systems as biomarkers. This review aims to contribute to a better understanding of these systems, as well as an improved patient profile. Furthermore, a deep knowledge of these complex systems would also allow proposing new lines of research for the development of new therapeutic tools as a promising start for new approaches to this disease.

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PALABRAS CLAVE

Biomarcadores;
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Óxido nítrico y factores relacionados a oxidación e inflamación como posibles biomarcadores de insuficiencia cardíaca

Resumen Como enfermedad cardiovascular prevalente, la insuficiencia cardíaca es una de las principales causas de morbilidad prematura. Por ello, existe un especial interés sobre el estudio de marcadores eficientes asociados al riesgo y/o predicción de eventos cardiovasculares. En consecuencia se proponen a múltiples candidatos, pero sobresalen especialmente

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aquellos implicados en procesos oxidativos e inflamatorios propios de la enfermedad cardiovascular como el anión superóxido, óxido nítrico y peroxinitrito. En este sentido, existe una falta de conocimiento sobre las potenciales utilidades de estos sistemas como biomarcadores. La presente revisión procura contribuir a la mayor comprensión de estos sistemas para una mejor caracterización de pacientes. Por otra parte, un profundo conocimiento de estos complejos sistemas también permitiría proponer nuevas líneas de investigación para el desarrollo de inéditas herramientas terapéuticas como una auspiciosa frontera para el abordaje de esta patología.

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Introduction

With regard to cardiovascular disease (CVD), there is a special interest in the study and development of efficient markers associated with the risk and/or prediction of events, which in turn make timely intervention possible, finally validating their cardiovascular risk prediction potential. This led to the search for new biomarkers, with the proposal of multiple candidates. Nevertheless, those biomarkers involved in oxidative and inflammatory processes typical of CVD are especially noteworthy. In fact, reactive oxygen species (ROS), especially superoxide anion (O_2^-), and reactive nitrogen species (RNS), such as nitric oxide (NO) and peroxynitrite, show important functions (Fig. 1). In this sense, and with special emphasis on understanding the signalling pathways involved in the pathophysiology of heart failure (HF), there is a lack of knowledge on the potential usefulness of these systems as biomarkers. A better understanding would allow better characterisation of these systems and would also open up new lines of research for the development of novel therapeutic tools, which could mean a promising start for new approaches to this disease.

In particular, various neurohormonal systems have been studied in order to identify biomarkers with good predictive capacity; however, only a few meet all the criteria required to be useful in clinical practice. It is therefore necessary to continue searching for more and better substances that make an improved contribution.

In recent years, CVD has been recognised as a continuum that involves multiple entities, such as primary disease of the heart muscle (cardiomyopathy), hypertension (HTN), left ventricular hypertrophy (LVH), atherosclerotic heart disease, cardiac arrhythmias and diabetes mellitus; and HF is the final common pathway of all these entities with alteration of the signalling pathways involving NO, ROS/RNS, NADPH oxidase (Nox) and superoxide dismutase (SOD).

HF is often tackled from the perspective of the main mechanisms that induce ventricular damage and remodelling as a result of neurohormonal overstimulation by the renin-angiotensin-aldosterone (RAAS) and adrenergic systems. These alterations are classic and mark the progression of the disease. Of particular interest for this review is the fact that increased peripheral vascular resistance and cardiac remodelling constitute the main alterations and are associated with the signalling pathways mentioned above. Therefore, evaluation of HF development from a new perspective for clinical practice is a novel proposal, such as

remodelling via the signalling pathways involving NO, Nox and SOD. According to this suggestion, it has recently been reported that damage to the signalling pathways involving NO and related factors may be associated with prognosis and/or mortality during HF.

However, the use of classic biomarkers in HF and drugs for its treatment are expensive and hard to obtain, especially within the field of public health. Consequently, HF is a disease of epidemiological, health and economic importance. This justifies the support offered for major research efforts to ensure a better understanding, treatment and follow-up. The development of new, more accessible and more affordable methodologies for studying its progression/prognosis would therefore allow the natural history of HF to be positively altered. Implementation of these types of biomarker, which are used to improve life expectancy and quality of life, may also help studies on the evaluation of drugs and electrophysiological devices during HF.

Biomarkers in heart failure

In general terms, biomarkers provide useful prognostic information in HF patients and there is currently considerable interest in determining the ability of biomarkers to guide therapy in cases of acute and chronic HF. Under these precepts, Richards and Braunwald listed neurohormonal, inflammatory, oxidative stress, interstitial matrix remodelling, myocyte injury and other newer markers that reflect different pathophysiological aspects of HF.^{1,2}

Neurohormonal markers known as B-type natriuretic peptides (BNP, NT-proBNP and proBNP) are currently preferred due to their diagnostic/predictive value. These are secreted in both the left ventricle (LV) and right ventricle (RV) and specifically reflect LV filling pressure and parietal stress. However, they also increase when the patient has RV dysfunction in the absence of LV dysfunction, as is observed, for example, in respiratory diseases with chronic cor pulmonale and in group 1 patients with pulmonary arterial hypertension (Nice). These markers allow HF to be ruled out with reduced inaccuracies and costs and serial BNP testing is a useful tool for identifying the best discharge time and post-discharge progression.

In view of the above, biomarkers are generally useful, but they also have limitations at the time of use. Their clinical usefulness is related to the fact that testing allows for clinical management and improves prognosis for one or more situations and consequently improves diagnostic certainty

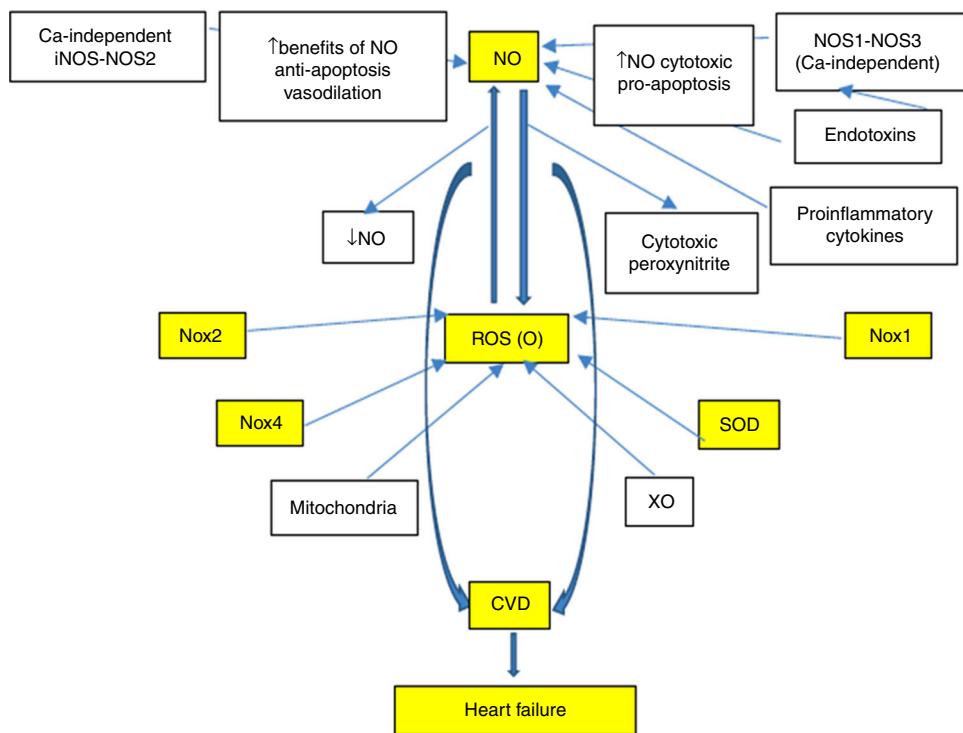


Figure 1 Main ROS and RNS-related mechanisms involved in cardiovascular disease.

associated with risk of onset or worsening of HF (the ideal thing is to have a response with a specific treatment). Monitoring through serial testing of markers should also improve the results obtained during patient follow-up, i.e. less acute decompensation, reduced mortality and/or improved quality of life.

However, the use of biomarkers in HF questions the role of the laboratory and the value of a simple blood sample for diagnosis, prognosis, monitoring of progression and to guide therapy. Moreover, an improved knowledge of the importance of neurohormonal systems in HF progression has resulted in major therapeutic advances since the mid 1980s. This has been based on the study of circulating concentrations of myocyte stress markers, BNP, proBNP and NT-pro BNP, which were also closest to the typical ideal biomarker. Nevertheless, in addition to the aforementioned markers, special attention has been paid over recent years to investigating oxidative stress modulators and NO, Nox-ROS and SOD-mediated signalling pathways that exhibit great involvement in the development of CVD (Fig. 1). A profound knowledge of such matters will help us gain a better understanding of the mechanisms, neurohormonal changes and biomolecular damages that have not yet been well established and that occur during HF.

Nitric oxide as a biomarker in heart failure

Multiple studies have shown the involvement of NO in the pathophysiology of HF and that there is a marked NO/redox disequilibrium at the expense of increased free radical production by enzymatic pathways, including especially vascular Nox, cardiac XO, mitochondrial enzymes and haemoglobin oxidase in red blood cells. In general terms,

this oxidises proteins critical for excitation-contraction coupling and also diminishes NO bioavailability at the expense of altering the activity and/or site of producing enzymes (NOS and XO).^{3,4} This leads to the characteristic mechanoenergetic uncoupling since reduced contractility is not accompanied by a proportional reduction in energy consumption. There is also marked neurohormonal activation with increased proinflammatory cytokines that induce NOS2 expression⁵ and sometimes increase NOS1 and NOS3 activity, modifying the site of NOS1.^{6,7} However, the hypothesis that NO plays a major role in the aetiopathogenesis of HF does not match the finding that the same element exerts a cardioprotective effect during ischaemia or that there is reduced NOS expression in failing hearts and/or impaired bioavailability in patients with HF. In fact, in experimental models and in patients with HF, NOS3 activity may increase, decrease or remain the same,^{4,8,9} and NOS2 has been observed to increase dramatically in some studies, but not in all cases.^{5-7,10-14} This would suggest, at least in part, that changes in NO₂ and NOS3 expression may be due to an epiphenomenon that accompanies HF, without being determinants of its cause (Fig. 1).

In failing hearts, decreased NO would lead to reduced endothelium-dependent coronary vasodilation, impaired ventricular relaxation and increased maximal oxygen consumption (MVO₂).^{15,16} Therefore, NOS3 overexpression should improve these alterations as shown in some experimental studies. Also, after coronary artery ligation, there is enhanced contractile performance and reduced LV hypertrophy and mortality.¹⁷⁻¹⁹ However, NOS3 knock-out mice that also received coronary artery ligation showed LV hypertrophy and dilation, decreased fractional shortening, lower ejection fraction, greater end-diastolic volume, LV internal

diameter and vascular rarefaction and higher mortality.^{8,20} Enalapril and valsartan also had less beneficial effects in these mice.²¹ These results suggest that NOS3 reduces ventricular dysfunction and post-infarction remodelling and may also play some sort of role in angiotensin II-induced ventricular dysfunction.

However, it has been reported in patients with dilated cardiomyopathy (DCM) that decreased NO does not alter cardiac contraction, and with intracoronary infusion of sodium nitroprusside or substance P, it does not modify haemodynamic parameters.^{10,22–24} Nevertheless, one animal model with pacing-induced HF was able to show that the cardiac decompensation phase was characterised by a marked reduction in NO production, dP/dt_{max} and ventricular distensibility (which increases LV end-diastolic pressure with elevated MVO₂), while cardiac metabolism switched from using fatty acids to using glucose as an energy source¹⁵; these results suggest that NO may participate in the coupling of coronary flow, contractile performance and cardiac metabolism.

Patients with DCM have increased NOS1 expression, which is translocated from the sarcoplasmic reticulum (where it is coupled to XO) to the sarcolemma, where, as in the case of NOS3, it interacts with a calmodulin-regulated calcium-dependent ATPase.⁷ These alterations in expression and site may be beneficial during HF since NOS1: (a) inhibits the heart's response to β-receptor stimulation via inhibition of the Ca²⁺ current, and Ca²⁺ release from the sarcoplasmic reticulum may perhaps induce cardioprotective effects against catecholamine-induced cardiotoxicity²⁵; (b) increases cardiac vagal tone, decreasing heart rate²⁶; (c) restores central baroreceptor activity; and (d) may compensate for NOS3 inhibition.⁷ However, it is also possible that translocation of NOS1 to the sarcolemma may facilitate oxidative stress since its control on XO activity in the sarcoplasmic reticulum would be lost and this would alter the nitric oxide/redox balance at the sarcolemma.⁹

It is interesting that, during HF, reduced inotropic response to β-stimulation is observed as a result of alterations in receptor density (with decreased β1 and β2 receptors and increased β3 receptors which mediate negative inotropic responses) or in coupling of the receptor to its signalling pathways (with increased expression of β-adrenergic receptor-specific kinase or ARK and Gi proteins and decreased Gs proteins). The β3 receptors are more resistant to homologous desensitisation due to the increased sympathetic tone that is characteristic of HF and their stimulation may therefore facilitate continuous NO production in the presence of increased sympathetic tone in the setting of HF.²⁷ Therefore, NO synthesised through the β3-NOS3 pathways, NOS1 translocated to the sarcolemma and the induction of NOS2 may modulate response to catecholamines and antagonise their toxicity in the failing myocardium. In fact, inhibition of NOS potentiates increased contractility produced by β-adrenergic agonists in animal models¹⁶ and in patients with HF.^{9,28,29} However, it is logical to assume that NO is only one of the factors to regulate response to β-adrenergic agonists in HF patients.³⁰ Nevertheless, under certain circumstances, an increase in synthesised NO following induction of NOS2 may be beneficial since it improves ventricular relaxation,^{6,22} reduces

MVO₂³¹ and response to β-adrenergic stimulation^{8,11,15} and increases angiogenesis.³² In cardiomyocytes of heart transplant patients, isoproterenol produces a slight increase in contractility and heart rate, and, where NOS2 inhibition normalises both responses, it also increases Ca²⁺ transport. However, NOS2 inhibition has no effect on normal cardiomyocytes or on cardiomyocytes of HF patients in whom response to isoproterenol was preserved and in whom NOS2 expression is poor.¹⁴ In other words, in HF patients, NOS2 expression limits response to β-adrenergic agonists, an effect that may be mediated by inhibition of Ca²⁺ release from the sarcoplasmic reticulum via a cGMP-independent pathway and which is related to alterations in cellular redox status produced by peroxynitrite.³³ Cardiac-specific NOS2 overexpression in mice leads to increased production of peroxynitrite and dilatation, hypertrophy and cardiac fibrosis; moreover, although they rarely develop HF, they display a high incidence of sudden cardiac death due to bradyarrhythmia.³⁴ On the contrary, mice with NOS2 overexpression display a normal genotype since the NO produced is neutralised by cytoplasmic myoglobin; however, when these experiments are repeated in myoglobin-deficient mice, the animals develop signs of hypertrophy, interstitial fibrosis and ventricular dilatation.³⁵

As mentioned above, during HF, enzyme pathways that produce free radicals are upregulated and NO-producing enzymes (NOSs and XO) are altered, producing vasoconstriction and mechanoenergetic uncoupling. In fact, in animal models, the transition to decompensated HF implies NOS1 and NOS3 deficiency and consequently poor cardiac NO synthesis, plus increased XO activity.⁴ NO also regulates a Nox that inhibits Ca²⁺ release from the sarcoplasmic reticulum and, therefore, the balance between NO and oxidative stress also regulates cardiac function through its effects on intracellular Ca²⁺ cation signalling. XO, an important source of superoxide radical, increases in the myocardium and vessels of patients with HF,³⁶ producing endothelial dysfunction, depressed cardiac function, mechanoenergetic uncoupling and apoptosis.^{37,38} XO inhibition with allopurinol improves mechanical efficiency, post-infarction myocardial remodelling and response to catecholamines.³⁹ It is important to note that the effects of allopurinol are suppressed by blocking NOS with L-NAME (specific inhibitor) and the effects of this inhibitor are blocked with allopurinol,³⁸ which indicates that there may be interaction between the two signalling pathways.⁴ However, some drugs used in HF, such as statins⁴⁰ and ACE inhibitors,⁴¹ alter the NO/redox balance by increasing bradykinin levels and NO synthesis and reducing the production of free radicals (superoxide, peroxynitrite) upon inhibiting NADPH oxidase (Fig. 1).

NO also plays a major role in cardiac ion channels, the genesis of cardiac arrhythmias, cardiac apoptosis, ischaemic pre-conditioning, cardiac ischaemia and mitochondrial function. The damage or benefit modulated by NO will depend on cell type and condition. Therefore, high concentrations induce cell death during ischaemic injury leading to neurodegenerative diseases.^{42–44}

Cytotoxicity attributed to NO is due to peroxynitrite produced by the diffusion-controlled reaction between NO and the superoxide anion. Peroxynitrite interacts with lipids, DNA and proteins via direct oxidative reactions or indirect

radical-mediated mechanisms. These reactions trigger cellular responses ranging from subtle modulations of cell signalling to overwhelming oxidative injury leading to cell necrosis or apoptosis. Peroxynitrite is crucial in conditions such as stroke, myocardial infarction, chronic heart failure (CHF), diabetes, cardiogenic shock, chronic inflammatory diseases, cancer and neurodegenerative disorders.⁴⁵

The cardioprotective effect responds at least partly to the protein kinase G pathway (GMP/PKG),^{46,47} where NO activates a soluble guanylate cyclase that catalyses cGMP synthesis from GTP. Furthermore, NO participates in mechanisms with PKG-mediated anti-apoptotic actions and these are an active area of research as modulation of these pathways may have major therapeutic implications.

Nitric oxide-related factors as biomarkers in heart failure

In patients with chronic heart failure, the increased production of ROS, and in particular the importance of O⁻, have been clearly demonstrated. Evidence shows the role of both ROS and RNS in the pathophysiology of CVD. Consequently, multiple biomarkers of oxidative stress common to both systems are evaluated. Of these, Nox is the most efficient system and represents a major source of ROS in HF. Abnormal activation of the renin-angiotensin system (RAS), which significantly affects the induction of apoptosis and fibrosis, is a result of the production of ROS by Nox and is specific to CVD.^{48,49} In this respect, the emphasis is on changes in the ROS-Nox-dependent signalling pathways as significant factors responsible for the development of many cardiac diseases. NADPH oxidases (Nox) are major sources of O⁻ in vascular cells and myocytes, where they share some of the characteristics of the neutrophil enzymes. In response to growth factors and cytokines they produce O⁻, which may be metabolised to hydrogen peroxide. Both of these reactive oxygen species act as second messengers to activate multiple intracellular signalling pathways. In addition to Nox from myocytes, a vascular Nox has been found to be essential in the physiological response of vascular cells, including growth, migration and modification of the extracellular matrix. They have also been linked to HTN and disease states associated with uncontrolled growth and inflammation, such as atherosclerosis. In the case of pressure overload, myocardial Angiotensin II (Ang II) production is activated, which stimulates intracellular signalling pathways which in turn activate hypertrophic response, as is the case with Nox.⁵⁰

Nox has various constituents or subunits which are relevant for its relationship with CVD, shown in order of importance as Nox2, Nox4 and Nox1 and the subunits gp91phox, p22phox, p47phox and Rac1. These play an important role in cardiac damage, causing cardiac hypertrophy and opposite cellular responses, accelerating atherosclerosis, hypertension and myocardial remodelling and activated in HF and post-AMI.⁵⁰⁻⁵²

More specifically, Esposito et al.⁵³ evaluated mice from which Rac1 had been eliminated, and came to the conclusion that this prevented Ang II-induced hypertrophy. Rac1 initiated the ROS-dependent hypertrophic response generated by Nox in the heart,⁵⁴ confirming that the production of

O⁻ by the Nox2 subunit Rac1 initiated activation of protein kinase B (Akt) as a component of this signalling pathway and Ang II-induced cardiomyocyte hypertrophy.⁵⁵

Basic research provides evidence for Nox2 being responsible for the vascular production of ROS, lower NO bioavailability and the development of early lesions in mice aortas.⁵⁶ An additional finding combines the involvement of circulating transforming growth factor-beta (TGF-β) and apolipoprotein E, where the increase of TGF-β induced Nox activation and overproduction of ROS, accelerating atherosclerosis, hypertension and myocardial remodelling in apolipoprotein E-deficient mice.⁵⁷

However, elevated expression of Nox and O⁻ were also described in the carotid arteries of rabbits with CHF⁵⁸ and specifically in the activity of the Nox2 subunit p47phox in the LV of mice after myocardial infarction (MI).⁵⁹

While Nox2 is involved in Ang II-induced LVH, Nox4 is apparently also involved in pressure overload in mouse myocardium.⁶⁰ Accordingly, this isoform may also play an important, although controversial, role in CVD. It has been suggested that Nox4 was bound to the p22phox protein on the internal membrane of epithelial cells. It was also found—in contrast to other NADPH oxidase isoforms—that Nox4 mainly produced hydrogen peroxide and a very small quantity of O⁻. Cytosolic oxidase proteins or the GTPase Rac were not required for the activity of this enzyme.⁶¹ Accordingly, there is some evidence mentioning that although Nox4 produced hydrogen peroxide, it also generated O⁻ intracellularly.⁶² However, there is still considerable uncertainty regarding the production of Nox4-dependent ROS. Therefore, the current state of knowledge in this regard indicates that Nox4 primarily produces hydrogen peroxide and not O⁻, contradicting most other experimental data. A possible explanation would be the use of unreliable methods such as nitro blue tetrazolium reduction to detect O⁻.^{62,63} However, the use of specific and precise methods such as chemiluminescence to detect O⁻ provided different results.⁶⁴ For example, with the lucigenin chemiluminescence assay it was found that Nox4 and Nox2 produced approximately 75% of O⁻ in the coronary arteries of patients with coronary artery disease.⁶⁵

Nox4 is located at various sites within cardiac cells compared to other Nox. In particular, it is located in the mitochondria where it represents a major source of O⁻ production in myocytes. It can bring about mitochondrial dysfunction, cell death, left ventricular dysfunction in response to pressure overload and, paradoxically, it may also cause cardiac adaptation to chronic stress.⁶⁶ In aged mice under hypertrophic stimulation with pressure overload, Nox4 was stimulated and this increased the production of O⁻ and induced cardiac dysfunction with fibrosis and apoptosis.⁶⁷

Unlike other NADPH oxidase isoforms, Nox4 stimulation in cardiomyocytes caused protection against pressure overload-induced cardiac remodelling. The authors justify these results because of the preservation of Nox4-induced myocardial capillary density through activation of hypoxia-inducible factor-1 (Hif1) and the production of vascular endothelial growth factor. They also showed that the site of Nox4 in cardiomyocytes was not mitochondrial, but in the perinuclear endoplasmic reticulum. These findings contradict those of other authors.⁶⁶ There are contradictory results

concerning the effects on oxidative stress and its localisation. Further research is required regarding the origin of these differences⁶⁸ because they conflict with most findings on the effects of Nox in cardiomyocytes and other cells.

Unlike the implications of Nox in cardiac diseases, the involvement of XO in ROS-dependent cardiac damage originally raised many questions. XO and xanthine dehydrogenase (XDH) are the oxidised and reduced forms of xanthine oxidoreductase (XOR). XO was considered the greatest source of O⁻ and hydrogen peroxide, and its mechanism was initially well established.⁶³ Paradoxically, subsequent studies discovered low XO activity in animal and human hearts.^{69,70}

Currently, however, there is consensus on the increase in XO levels and their activity in the cardiovascular system under pathological conditions, but that they cannot be detected easily in physiological conditions. Consequently, Thompson-Gorman and Zweier measured the production of XO-mediated ROS in isolated rat heart.⁷¹ They found that XO was an important factor of oxidative damage from ischaemia/reperfusion in the rat heart. Accordingly, it has also been demonstrated that the increased production of XO-catalysed ROS during ischaemia/reperfusion results from increased concentration of the substrate (xanthine and hypoxanthine) on account of ATP degradation during ischaemia.⁷² Ashraf and Samra also suggested that XO activity increases during ischaemia and was intensified following reperfusion.⁷³ XO is found in interstitial cells, coronary vascular endothelium and smooth muscle cells. De Jong et al.⁷⁴ showed that the production of ROS by xanthine oxidoreductase (XOR) increased in HF, but not in cardiac hypertrophy.

Like the NADPH oxidases, XO led to many ROS-dependent cardiac disorders such as endothelium-mediated deterioration of vasodilatation, increased XO activity in dilated cardiomyopathy, NO reduction in patients with coronary disease, coronary endothelial dysfunction and toxic effect of O⁻ caused by XO in the heart. Moreover, the increased activity of XO and the reduction of extracellular superoxide dismutase (ecSOD) caused deterioration of endothelium-mediated vasodilatation in patients with HF.⁷⁵ To that effect, later research established levels of protein XO and XO-dependent O⁻ induced by Ang II in endothelial cells of patients with coronary disease,⁷⁶ thereby suggesting that Ang II promotes overproduction of O⁻ by activation of redox-sensitive XO. In addition, basic studies confirmed the role of XO, as it was discovered that its activity would be elevated in DCM⁷⁷ and that chronic XO inhibition by allopurinol suppressed the progression of HF to DCM.

Rats with HF on account of spontaneous hypertension also showed increased mRNA expression and XOR activity, while XOR inhibition caused a reversal of remodelling in HF on account of spontaneous hypertension and dilated cardiomyopathy.⁷⁸ XOR and Nox also increased cardiac O⁻ formation in Dahl salt-sensitive hypertensive rats with diastolic HF.⁷⁹

XO-produced ROS reduced the bioavailability of coronary NO in patients with coronary artery disease (CAD).⁸⁰ Similarly, in a mouse model, myocardial ischaemia/reperfusion increased the expression of tumour necrosis factor-alpha (TNF- α) and induced XO activation and O⁻ production bringing about coronary endothelial dysfunction.⁸¹ Moreover, another toxic effect of XO-generated O⁻ was demonstrated in the hearts of spontaneously hypertensive rats with HF with

DCM. Specifically, O⁻ caused deterioration of S-nitrosylation of the ryanodine receptor (RyR) resulting in calcium leak from the sarcoplasmic reticulum in skeletal muscle.⁸²

Of particular interest and widely recognised, mitochondrial dysfunction is an essential source of ROS in CVD, and consequently mitochondria are the subject of ongoing study. It has been established that O⁻ generated by the mitochondria results in the release of transporter electrons from the respiratory chain (complexes I and III).⁸³ Many authors have confirmed the importance of the overproduction of mitochondrial ROS in the failing heart (myocardial insufficiency, ischaemia, ischaemia/reperfusion, coronary endothelial dysfunction caused by O⁻ in congestive HF, progression of LVH to pulmonary hypertension (PH) and right-sided HF, mitochondrial and LV damage and, finally, arrhythmias).

Originally, mitochondrial complex I was described as a potential source of ROS in the myocardium of dogs with HF.⁸⁴ Moreover, ischaemia increased ROS production in isolated mouse heart mitochondria.⁸⁵ The production of mitochondrial O⁻ in the failing heart can also induce complex II changes in the post-ischaemic myocardium of rats subjected to coronary ligation followed by reperfusion.⁸⁶ It should be noted that damage caused by ischaemia/reperfusion with the ablation of protein p66 (Shc) in mouse hearts plays a key role in the formation of mitochondrial ROS.⁸⁷ In that regard, mitochondrial depolarisation and the increased production of lipoxygenase-mediated ROS and arachidonic acid induced arrhythmias due to ischaemia/reperfusion.⁸⁸ In addition, the increased production of mitochondrial O⁻ was responsible for coronary endothelial dysfunction and reduced coronary flow in congestive HF.⁸⁹

It should be emphasised that studies performed during the progression of LVH to congestive HF demonstrated that mitochondrial Nox was the primary source of ROS—especially in hypertension-induced right ventricular failure. Surprisingly, the increased activity of mitochondrial complex II was the mechanism responsible and not of complexes I and III, acknowledged as the greatest sources of mitochondrial ROS. This contribution was particularly relevant for the production of ventricular ROS in HF.⁹⁰ However, Mariappan et al.⁹¹ demonstrated that the production of mitochondrial O⁻ induced by TNF- α increased complex I activity of the respiratory chain which caused mitochondrial damage in rat LV. All these findings suggest that the over-production of ROS by the mitochondria represents a causal origin in cardiac diseases.

Based on the above, it is clear that CVD is associated with a chronic state of oxidative stress and inflammation mediated by complex, interconnected signalling pathways. More specifically, the course of HF is characterised by mitochondrial dysfunction, overproduction of ROS, activation of RAS linked to greater activity of NADPH oxidase and NO reduction (Fig. 1). Interestingly, it has been demonstrated that lower NO bioavailability induces heat shock protein (Hsp70) expression, which causes beneficial effects against oxidative stress damage, inflammation and apoptosis. The induction of heat shock proteins as a response to damage by stimulation of the RAS system and/or NO deficiency was originally suggested by Bravo et al.⁹²

Currently, it is understood that heat shock proteins are elevated in the plasma of patients with CVD. However,

their physiological role is not yet fully understood and their value for predicting the development and/or progression of the disease is understood even less. In particular, it was suggested that circulating levels of Hsp70 could indicate the presence/progression of atherosclerosis in subjects with established hypertension; in addition, an intriguing possibility posed by the authors is that Hsp70 could protect against oxidative and inflammatory damage in this group of subjects.⁹³ Accordingly, it has recently been proposed that Hsp70 could be a potential biomarker and treatment objective in diseases such as cancer, CVD and neurological and hepatic diseases.⁹⁴

However, there is information indicating that low Hsp70 levels may be related to a healthy cardiovascular status, and it was suggested as a predictor of longevity.⁹⁵ Based on this, Hsp70 was investigated in patients with CHF in an attempt to re-establish a relationship between severity and survival. Elevated Hsp70 levels, particularly in those subjects with cardiac cachexia, were found and, therefore, could be related to disease severity, although not to survival.⁹⁶ Consequently, further study is required to understand the significance of the relationship between Hsp70 expression and CHF morbidity.

Conclusions and perspectives

HF is, in almost all cases, the end result of primary heart disease and any cause of structural damage to the heart, and is one of the most prevalent CVD. This makes it one of the main causes of premature morbidity and mortality in most developed and developing countries. Of particular interest is the fact that evidence obtained from sources such as prospective and interventional epidemiological cohort studies suggest that this disease is associated with alterations in endothelial function, oxidative metabolism, inflammation and apoptosis. Some of the most relevant determinants of such alterations include SOD activity, phosphorylation and expression of NO-producing enzymes, increased glutathione peroxidase activity, activation of NADPH oxidase and expression of p22phox.

As a result, the literature highlights that HF patients display alterations in signalling pathways that promote oxidative stress and inflammation. Such alterations include specifically elevated ROS with increased NADPH activity and reduced SOD activity. This was recently proven by our laboratory.⁹⁷ However, in this same context, low NO levels were also reported. These may be due to interactions with ROS, decreased NO production or a combination of the two. Although there are many basic research references on alterations of NO, Nox-ROS and SOD pathways, there are few references regarding their actual clinical impact. In spite of current knowledge and it now being more than 20 years since the functions of endogenous NO were identified, attempts to generate new therapeutic strategies have not been fruitful, reflecting slow progress in the study and understanding of potential related biomarkers.

Alteration and/or uncoupling of Nox, ROS and SOD conditions multiple signalling pathways involved in LVH, myocardial failure, progression of HF to right-sided heart failure, altered endothelial function and coronary artery vasomotility and reperfusion injury and arrhythmia, all

components of the cardiovascular continuum. In this context, NO is a key component for maintaining cardiovascular homeostasis, and impaired NO bioavailability is therefore clearly associated with CVD. There is substantial evidence, as outlined in this review, that ROS generated from NADPH oxidase may be primarily responsible for reduced NO bioavailability. Thus, O⁻ may interact with NO to produce peroxynitrite, which may in turn give rise to other reactive species. Specifically, increases in oxidative stress during HF may be the result of the functional uncoupling of the respiratory chain due to impaired antioxidant capacity (reduced SOD activity and/or stimulation of enzymatic sources, including NADPH oxidases).

The proposal to evaluate NO as a biomarker in HF, as proposed by our working group and others, responds to the assumption that this factor is involved in relevant pathophysiological aspects of HF. In fact, there is NO/redox disequilibrium in HF with increased enzymatic pathways that produce free radicals (NADPH oxidases, cardiac XO, mitochondrial enzymes, haemoglobin oxidase in red blood cells, etc.) which oxidise proteins involved in the excitation-contraction coupling and inactivate NO and alter the activity and site of NO-producing enzymes. This leads to functional uncoupling characterised by reduced contractility that is not accompanied by a similar reduction in energy consumption. This also conditions marked neurohormonal activation and increased proinflammatory cytokines. In this sense, it has been suggested very recently that low NO bioavailability induces the expression of oxidative stress proteins, such as Hsp70, which promotes protective effects against damage not only from oxidative stress, but also from inflammation and apoptosis. In fact, our laboratory suggested that Hsp70 may be used as a potential biomarker of oxidative-inflammatory damage.⁹⁸ Consequently, recent studies suggest that heat shock proteins play a key role in the pathogenesis of cardiovascular diseases, including HF. In this regard, it is unclear whether circulating Hsp70 levels are related to CVD risk factors, echocardiographic indexes of LV remodelling and/or prevalence of CVD.⁹⁹ However, the analysis of preliminary results from our laboratory suggests that patients with HF also have elevated inflammatory markers (C-reactive protein, IL-6 and TNF- α), while Hsp70 was reduced compared to healthy patients. As a result, the determination of oxidative stress markers, such as plasma inflammatory markers, allowed us to distinguish healthy patients from patients with HF. Such findings of lower plasma Hsp70 levels in patients with HF and with low NO levels were previously unpublished and controversial. Interpretation of our results has proven even more complex since learning about the opposite effects that Hsp70 has at extracellular versus intracellular locations upon activation of the inflammatory pathway by NF- κ B.¹⁰⁰ However, in relation to oxidative stress and inflammation, it is valid to consider differences in consequences of acute and chronic events on biological response capacity to these injuries (acute HF versus chronic HF). In this regard, earlier reports studied these differences and showed (in the chronic phase) that enhanced lipid peroxidation through higher TBARS levels and increased oxidative stress resulted in reduced total antioxidant activity and enhanced NADPH oxidase activity. This was demonstrated by Rinaldi Tosi et al. and was also accompanied by decreased inducible Hsp70 isoform

expression.¹⁰¹ Therefore, more studies should be conducted with special attention paid to HF and related models in order to gain a deeper understanding of these altered signalling pathways.

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Conflicts of interest

The authors declare that they have no conflicts of interest.

References

1. Richards AM. What we may expect from biomarkers in heart failure. *Heart Fail Clin.* 2009;5:463–70.
2. Braunwald E. Biomarkers in heart failure. *N Engl J Med.* 2008;358:2148–59.
3. Drexler H. Nitric oxide synthases in the failing human heart: a double-edged sword? *Circulation.* 1999;99:2972–5.
4. Hare JM, Stamer JS. NO/redox disequilibrium in the failing heart and cardiovascular system. *J Clin Invest.* 2005;115:509–17.
5. Prabhu SD, Chandrasekar B, Murray DR, Freeman GL. β -Adrenergic blockade in developing heart failure: effects on myocardial inflammatory cytokines, nitric oxide, and remodeling. *Circulation.* 2000;101:2103–9.
6. Damy T, Ratajczak P, Robidel E, Bendall JK, Oliviero P, Boczkowski J, et al. Up-regulation of cardiac nitric oxide synthase 1-derived nitric oxide after myocardial infarction in senescent rats. *FASEB J.* 2003;17:1934–6.
7. Damy T, Ratajczak P, Shah AM, Camors E, Marty I, Hasenfuss G, et al. Increased neuronal nitric oxide synthase-derived NO production in the failing human heart. *Lancet.* 2004;363:1365–7.
8. Massion PB, Feron O, Dessaix C, Balligand JL. Nitric oxide and cardiac function: ten years after, and continuing. *Circ Res.* 2003;93:388–98.
9. Hare JM, Loh E, Creager MA, Colucci WS. Nitric oxide inhibits the positive inotropic response to β -adrenergic stimulation in humans with left ventricular dysfunction. *Circulation.* 1995;92:2198–203.
10. Massion PB, Balligand JL. Modulation of cardiac contraction, relaxation and rate by the endothelial nitric oxide synthase (eNOS): lessons from genetically modified mice. *J Physiol.* 2003;546:63–75.
11. Hare JM. Nitric oxide and excitation-contraction coupling. *J Mol Cell Cardiol.* 2003;35:719–29.
12. Thoenes M, Forstermann U, Tracey WR, Bleese NM, Nussler AK, Scholz H, et al. Expression of inducible nitric oxide synthase in failing and non-failing human heart. *J Mol Cell Cardiol.* 1996;28:165–9.
13. De Belder AJ, Radomski MW, Why HJ, Richardson PJ, Martin JF. Myocardial calcium-independent nitric oxide synthase activity is present in dilated cardiomyopathy, myocarditis, and post-partum cardiomyopathy but not in ischaemic or valvular heart disease. *Br Heart J.* 1995;74:426–30.
14. Ziolo MT, Maier LS, Piacentino V 3rd, Bossuyt J, Houser SR, Bers DM. Myocyte nitric oxide synthase 2 contributes to blunted beta-adrenergic response in failing human hearts by decreasing Ca^{2+} transients. *Circulation.* 2004;109:1886–91.
15. Recchia FA, McConnell PI, Bernstein RD, Vogel TR, Xu X, Hintze TH. Reduced nitric oxide production and altered myocardial metabolism during the decompensation of pacing-induced heart failure in the conscious dog. *Circ Res.* 1998;83:969–79.
16. Heymes C, Vanderheyden M, Bronzwaer JG, Shah AM, Paulus WJ. Endomyocardial nitric oxide synthase and left ventricular preload reserve in dilated cardiomyopathy. *Circulation.* 1999;99:3009–16.
17. Janssens S, Pokreisz P, Schoonjans L, Pellens M, Vermeersch P, Tjwa M, et al. Cardiomyocyte-specific overexpression of nitric oxide synthase 3 improves left ventricular performance and reduces compensatory hypertrophy after myocardial infarction. *Circ Res.* 2004;94:1256–62.
18. Jones SP, Greer JJ, van Haperen R, Duncker DJ, de Crom R, Lefer DJ. Endothelial nitric oxide synthase overexpression attenuates congestive heart failure in mice. *Proc Natl Acad Sci U S A.* 2003;100:4891–6.
19. Jones SP, Greer JJ, Kakkar AK, Ware PD, Turnage RH, Hicks M, et al. Endothelial nitric oxide synthase overexpression attenuates myocardial reperfusion injury. *Am J Physiol Heart Circ Physiol.* 2004;286:H276–82.
20. Scherrer-Crosbie M, Ullrich R, Bloch KD, Nakajima H, Nasser B, Aretz HT, et al. Endothelial nitric oxide synthase limits left ventricular remodeling after myocardial infarction in mice. *Circulation.* 2001;104:1286–91.
21. Liu YH, Xu J, Yang JP, Yang F, Shesely E, Carretero OA. Effect of ACE inhibitors and angiotensin II type 1 receptor antagonists on endothelial NO synthase knockout mice with heart failure. *Hypertension.* 2002;39:375–81.
22. Paulus WJ, Bronzwaer JG. Nitric oxide's role in the heart: control of beating or breathing? *Am J Physiol Heart Circ Physiol.* 2004;287:H8–13.
23. Paulus WJ. The role of nitric oxide in the failing heart. *Heart Fail Rev.* 2001;6:105–18.
24. Kanai AJ, Mesaros S, Finkel MS, Oddis CV, Birder LA, Malinski T. β -Adrenergic regulation of constitutive nitric oxide synthase in cardiac myocytes. *Am J Physiol.* 1997;273:C1371–7.
25. Ashley EA, Sears CE, Bryant SM, Watkins HC, Casadei B. Cardiac nitric oxide synthase 1 regulates basal and β -adrenergic contractility in murine ventricular myocytes. *Circulation.* 2002;105:3011–6.
26. Takimoto Y, Aoyama T, Tanaka K, Keyamura R, Yui Y, Sasayama S. Augmented expression of neuronal nitric oxide synthase in the atria parasympathetically decreases heart rate during acute myocardial infarction in rats. *Circulation.* 2002;105:490–6.
27. Moniotte S, Kobzik L, Feron O, Trochu JN, Gauthier C, Balligand JL. Upregulation of β 3-adrenoceptors and altered contractile response to inotropic amines in human failing myocardium. *Circulation.* 2001;103:1649–55.
28. Varghese P, Harrison RW, Loftouse RA, Georgakopoulos D, Berkowitz DE, Hare JM. β 3-Adrenoceptor deficiency blocks nitric oxide-dependent inhibition of myocardial contractility. *J Clin Invest.* 2000;106:697–703.
29. Shinke T, Takaoka H, Takeuchi M, Hata K, Kawai H, Okubo H, et al. Nitric oxide spares myocardial oxygen consumption through attenuation of contractile response to β -adrenergic stimulation in patients with idiopathic dilated cardiomyopathy. *Circulation.* 2000;101:1925–30.
30. Ungureanu-Longrois D, Balligand JL, Simmons WW, Okada I, Kobzik L, Lowenstein CJ, et al. Induction of nitric oxide synthase activity by cytokines in ventricular myocytes is necessary but not sufficient to decrease contractile

- responsiveness to β -adrenergic agonists. *Circ Res.* 1995;77:494–502.
31. Chen Y, Traverse JH, Du R, Hou M, Bache RJ. Nitric oxide modulates myocardial oxygen consumption in the failing heart. *Circulation.* 2002;106:273–9.
 32. Jones MK, Tsugawa K, Tarnawski AS, Baatar D. Dual actions of nitric oxide on angiogenesis: possible roles of PKC ERK, and AP-1. *Biochem Biophys Res Commun.* 2004;318:520–8.
 33. Ziolo MT, Katoh H, Bers DM. Positive and negative effects of nitric oxide on Ca^{2+} sparks: influence of β -adrenergic stimulation. *Am J Physiol Heart Circ Physiol.* 2001;281:H2295–303.
 34. Mungrue IN, Gros R, You X, Pirani A, Azad A, Csont T, et al. Cardiomyocyte overexpression of iNOS in mice results in peroxynitrite generation, heart block, and sudden death. *J Clin Invest.* 2002;109:735–43.
 35. Godecke A, Molajayyi A, Heger J, Flögel U, Ding Z, Jacoby C, et al. Myoglobin protects the heart from inducible nitric-oxide synthase (iNOS)-mediated nitrosative stress. *J Biol Chem.* 2003;278:21761–6.
 36. Landmesser U, Bahlmann F, Mueller M, Spiekermann S, Kirchhoff N, Schulz S, et al. Vascular oxidative stress and endothelial dysfunction in patients with chronic heart failure-role of xanthine-oxidase and extracellular superoxide dismutase. *Circulation.* 2002;106:3073–8.
 37. Saavedra WF, Paolocci N, St John ME, Skaf MW, Stewart GC, Xie JS, et al. Imbalance between xanthine oxidase and nitric oxide synthase signaling pathways underlies mechanoenergetic uncoupling in the failing heart. *Circ Res.* 2002;90:297–304.
 38. Cappola T, Kass DA, Nelson G, Berger RD, Rosas GO, Kobeissi Z, et al. Allopurinol improves myocardial efficiency in patients with idiopathic dilated cardiomyopathy. *Circulation.* 2001;104:2407–11.
 39. Doeher W, Schoene N, Rauchhaus M, Leyva-Leon F, Pavitt DV, Reaveley DA, et al. Effects of xanthine oxidase inhibition with allopurinol on endothelial function and peripheral blood flow in hyperuricemic patients with chronic heart failure: results from 2 placebo-controlled studies. *Circulation.* 2002;105:2619–24.
 40. Haendeler J, Hoffmann J, Zeiher AM, Dimmeler S. Antioxidant effects of statins via S-nitrosylation and activation of thioredoxin in endothelial cells: a novel vasculoprotective function of statins. *Circulation.* 2004;110:856–61.
 41. Wittstein IS, Kass DA, Pak PH, Maughan WL, Fetis B, Hare JM. Cardiac nitric oxide production due to angiotensin-converting enzyme inhibition decreases beta-adrenergic myocardial contractility in patients with dilated cardiomyopathy. *J Am Coll Cardiol.* 2001;38:429–35.
 42. Virág L, Szabó E, Gergely P, Szabó C. Peroxynitrite-induced cytotoxicity: mechanism and opportunities for intervention. *Toxicol Lett.* 2003;140–141:113–24.
 43. Duncan AJ, Heales SJR. Nitric oxide and neurological disorders. *Mol Aspects Med.* 2005;26:67–9.
 44. Ciani E, Guidi S, Bartesaghi R. Nitric oxide regulates cGMP-dependent cAMP-responsive element binding protein phosphorylation and bcl-2 expression in cerebellar neurons: implication for a survival role of nitric oxide. *J Neurochem.* 2002;1282–9.
 45. Fiscus RR. Involvement of cyclic GMP and protein kinase G in regulation of apoptosis and survival in neuronal cells. *Neurosignals.* 2002;11:175–90.
 46. Nagai-Kusuvara A, Nakamura M, Mukuno H, Kanamori A, Negi A, Seigel GM. cAMP-responsive element binding protein mediates a cGMP/protein kinase G-dependent antiapoptotic signal induced by oxide nitric in retinal-neuro-glial progenitor cells. *Exp Eye Res.* 2007;84:152–62.
 47. Tawfik HE, Cena J, Schulz R, Kaufman S. Role of oxidative stress in multiparity-induced endothelial dysfunction. *Am J Physiol Heart Circ Physiol.* 2008;295:H1736–42.
 48. Griendling KK, Sorescu D, Ushio-Fukai M. NAD(P)H oxidase: role in cardiovascular biology and disease. *Circ Res.* 2000;86:494–501.
 49. Murdoch CE, Zhang M, Cave AC, Shah AM. NADPH oxidase-dependent redox signalling in cardiac hypertrophy, remodelling and failure. *Cardiovasc Res.* 2006;71:208–15.
 50. Li JM, Gall NP, Grieve DJ, Chen M, Shah AM. Activation of NADPH oxidase during progression of cardiac hypertrophy to failure. *Hypertension.* 2002;40:477–84.
 51. Bendall JK, Cave AC, Heymes C, Gall N, Shah AM. Pivotal role of a gp91phox-containing NADPH oxidase in angiotensin II-induced cardiac hypertrophy in mice. *Circulation.* 2002;105:293–6.
 52. Nakagami H, Takemoto M, Liao JK. NADPH oxidase-derived superoxide anion mediates angiotensin II-induced cardiac hypertrophy. *J Mol Cell Cardiol.* 2003;35:851–9.
 53. Esposito G, Prasad SVN, Rapacciulo A, Mao L, Koch WJ, Rockman HA. Cardiac overexpression of a Gq inhibitor blocks induction of extracellular signal-regulates kinase and c-Jun NH2-terminal kinase activity in *In Vivo* pressure overload. *Circulation.* 2001;103:1453–8.
 54. Satoh M, Ogita H, Takeshita K, Mukai Y, Kwiatkowski DJ, Liao JK. Requirement of Rac1 in the development of cardiac hypertrophy. *Proc Natl Acad Sci U S A.* 2006;103:7432–7.
 55. Hingtgen SD, Tian X, Yang J, Dunlay SM, Peek AS, Wu Y, et al. Nox2-containing NADPH oxidase and Akt activation play a key role in angiotensin II-induced cardiomyocyte hypertrophy. *Physiol Genomics.* 2006;26:180–91.
 56. Judkins CP, Diep H, Broughton BRS, Mast AE, Hooker EU, Miller AA, et al. Direct evidence of a role for Nox2 in superoxide production, reduced nitric oxide bioavailability, and early atherosclerotic plaque formation in *ApoE*–/– mice. *Am J Physiol Heart Circ Physiol.* 2010;298:H24–32.
 57. Buday A, Orsy P, Godó M, Mózes M, Kókény G, Lacza Z, et al. Elevated systemic TGF- β impairs aortic vasomotor function through activation of NADPH oxidase-driven superoxide production and leads to hypertension, myocardial remodeling, and increased plaque formation in *apoE*–/– mice. *Am J Physiol Heart Circ Physiol.* 2010;299:H386–95.
 58. Li YL, Gao L, Zucker IH, Schultz HD. NADPH oxidase-derived superoxide anion mediates angiotensin II-enhanced carotid body chemoreceptor sensitivity in heart failure rabbits. *Cardiovasc Res.* 2007;75:546–54.
 59. Doerrers C, Grote K, Hilfiker-Klein D, Luchtefeld M, Schaefer A, Holland SM, et al. Critical role of the NAD(P)H oxidase subunit p47phox for left ventricular remodeling/dysfunction and survival after myocardial infarction. *Circ Res.* 2007;100:894–903.
 60. Byrne JA, Grieve DJ, Bendall JK, Li JM, Gove C, Lambeth JD, et al. Contrasting roles of NADPH oxidase isoforms in pressure-overload versus angiotensin II-induced cardiac hypertrophy. *Circ Res.* 2003;93:802–4.
 61. Martyn KD, Frederick LM, von Loehneysen K, Dinauer MC, Knaus UG. Functional analysis of Nox4 reveals unique characteristics compared to other NADPH oxidases. *Cell Signal.* 2006;18:69–82.
 62. Serrander L, Cartier L, Bedard K, Banfi B, Lardy B, Plastre O, et al. NOX4 activity is determined by mRNA levels and reveals a unique pattern of ROS generation. *Biochem J.* 2007;406:105–14.
 63. Denisov ET, Afanas'ev IB. Oxidation and antioxidants in organic chemistry and biology. Boca Raton, FL, USA: CRC Press/Taylor & Francis Group; 2005.
 64. Afanasev I. Detection of superoxide in cells, tissues and whole organisms. *Front Biosci (Elite Edition).* 2009;1:153–60.
 65. Guzik TJ, Sadowski J, Guzik B, Jopek A, Kapelak B, Przybylski P, et al. Coronary artery superoxide production and nox

- isoform expression in human coronary artery disease. *Arterioscler Thromb Vasc Biol.* 2006;26:333–9.
66. Kuroda J, Ago T, Matsushima S, Zhai P, Schneider MD, Sadoshima J. NADPH oxidase 4 (Nox4) is a major source of oxidative stress in the failing heart. *Proc Natl Acad Sci U S A.* 2010;107:15565–70.
67. Ago T, Kuroda J, Pain J, Fu C, Li H, Sadoshima J. Upregulation of Nox4 by hypertrophic stimuli promotes apoptosis and mitochondrial dysfunction in cardiac myocytes. *Circ Res.* 2010;106:1253–64.
68. Zhang M, Brewer AC, Schröder K, Santos CX, Grieve DJ, Wang M, et al. NADPH oxidase-4 mediates protection against chronic load-induced stress in mouse hearts by enhancing angiogenesis. *Proc Natl Acad Sci U S A.* 2010;107:18121–6.
69. Downey JM, Miura T, Eddy LJ, Chambers DE, Mellert T, Hearse DJ, et al. Xanthine oxidase is not a source of free radicals in the ischemic rabbit heart. *J Mol Cell Cardiol.* 1987;19: 1053–60.
70. Grum CM, Gallagher KP, Kirsh MM, Shlafer M. Absence of detectable xanthine oxidase in human myocardium. *J Mol Cell Cardiol.* 1989;21:263–7.
71. Thompson-Gorman SL, Zweier JL. Evaluation of the role of xanthine oxidase in myocardial reperfusion injury. *J Biol Chem.* 1990;265:6656–63.
72. Xia Y, Zweier JL. Substrate control of free radical generation from xanthine oxidase in the postischemic heart. *J Biol Chem.* 1995;270:18797–803.
73. Ashraf M, Samra ZQ. Subcellular distribution of xanthine oxidase during cardiac ischemia and reperfusion: an immunocytochemical study. *J Submicrosc Cytol Pathol.* 1993;25:193–201.
74. De Jong JW, Schoemaker RG, de Jonge R, Bernocchi P, Keijzer E, Harrison R, et al. Enhanced expression and activity of xanthine oxidoreductase in the failing heart. *J Mol Cell Cardiol.* 2000;32:2083–9.
75. Landmesser U, Spiekermann S, Dikalov S, Tatge H, Wilke R, Kohler C, et al. Vascular oxidative stress and endothelial dysfunction in patients with chronic heart failure: role of xanthine-oxidase and extracellular superoxide dismutase. *Circulation.* 2002;106:3073–8.
76. Landmesser U, Spiekermann S, Preuss C, Sorrentino S, Fischer D, Manes C, et al. Angiotensin II induces endothelial xanthine oxidase activation: role for endothelial dysfunction in patients with coronary disease. *Arterioscler Thromb Vasc Biol.* 2007;27:943–8.
77. Duncan JG, Ravi R, Stull LB, Murphy AM. Chronic xanthine oxidase inhibition prevents myofibrillar protein oxidation and preserves cardiac function in a transgenic mouse model of cardiomyopathy. *Am J Physiol Heart Circ Physiol.* 2005;289:H1512–8.
78. Minhas KM, Saraiva RM, Schulter KH, Lehrke S, Zheng M, Salaris AP, et al. Xanthine oxidoreductase inhibition causes reverse remodeling in rats with dilated cardiomyopathy. *Circ Res.* 2006;98:271–9.
79. Yamamoto E, Kataoka K, Yamashita T, Tokutomi Y, Dong YF, Matsuba S, et al. Role of xanthine oxidoreductase in the reversal of diastolic heart failure by candesartan in the salt-sensitive hypertensive rat. *Hypertension.* 2007;50: 657–62.
80. Baldus S, Köster R, Chumley P, Heitzer T, Rudolph V, Ostendorf MA, et al. Oxypurinol improves coronary and peripheral endothelial function in patients with coronary artery disease. *Free Radic Biol Med.* 2005;39:1184–90.
81. Zhang C, Xu X, Potter BJ, Wang W, Kuo L, Michael L, et al. TNF- α contributes to endothelial dysfunction in ischemia/reperfusion injury. *Arterioscler Thromb Vasc Biol.* 2006;26:475–80.
82. Gonzalez DR, Treuer AV, Castellanos J, Dulce RA, Hare JM. Impaired S-nitrosylation of the ryanodine receptor caused by xanthine oxidase activity contributes to calcium leak in heart failure. *J Biol Chem.* 2010;285:28938–45.
83. Afanas'ev IB. Signaling mechanisms of oxygen and nitrogen free radicals. Boca Raton, FL, USA: CRC Press/Taylor & Francis; 2009.
84. Ide T, Tsutsui H, Kinugawa S, Utsumi H, Kang D, Hattori N, et al. Mitochondrial electron transport complex I is a potential source of oxygen free radicals in the failing myocardium. *Circ Res.* 1999;85:357–63.
85. Chen Q, Moghaddas S, Hoppel CL, Lesnefsky EJ. Ischemic defects in the electron transport chain increase the production of reactive oxygen species from isolated rat heart mitochondria. *Am J Physiol Cell Physiol.* 2008;294:C460–6.
86. Chen YR, Chen CL, Pfeiffer DR, Zweier JL. Mitochondrial complex II in the post-ischemic heart: oxidative injury and the role of protein S-glutathionylation. *J Biol Chem.* 2007;282:32640–54.
87. Carpi A, Menabò R, Kaludercic N, Pelicci P, di Lisa F, Giorgio M. The cardioprotective effects elicited by p66Shc ablation demonstrate the crucial role of mitochondrial ROS formation in ischemia/reperfusion injury. *Biochim Biophys Acta.* 2009;1787:774–80.
88. Haworth RA, Potter KT, Russell DC. Role of arachidonic acid, lipoxygenase, and mitochondrial depolarization in reperfusion arrhythmias. *Am J Physiol Heart Circ Physiol.* 2010;299:H165–74.
89. Chen Y, Hou M, Li Y, Traverse JH, Zhang O, Salvemini D, et al. Increased superoxide production causes coronary endothelial dysfunction and depressed oxygen consumption in the failing heart. *Am J Physiol Heart Circ Physiol.* 2005;288:H133–41.
90. Redout EM, Wagner MJ, Zuidwijk MJ, Boer C, Musters RJ, van Hardeveld C, et al. Right-ventricular failure is associated with increased mitochondrial complex II activity and production of reactive oxygen species. *Cardiovasc Res.* 2007;75:770–81.
91. Mariappan N, Elks CM, Fink B, Francis J. TNF-induced mitochondrial damage: a link between mitochondrial complex I activity and left ventricular dysfunction. *Free Radic Biol Med.* 2009;46:462–70.
92. Bravo J, Quiroz Y, Pons H, Parra G, Herrera-Acosta J, Johnson RJ, et al. Vimentin and heat shock protein expression are induced in the kidney by angiotensin and by nitric oxide inhibition. *Kidney Int Suppl.* 2003:S46–51.
93. Pockley AG, Georgiades A, Thulin T, de Faire U, Frostegård J. Serum heat shock protein 70 levels predict the development of atherosclerosis in subjects with established hypertension. *Hypertension.* 2003;42:235–8.
94. Liu T, Daniels CK, Cao S. Comprehensive review on the HSC70 functions, interactions with related molecules and involvement in clinical diseases and therapeutic potential. *Pharmacol Ther.* 2012;136:354–74.
95. Terry DF, McCormick M, Andersen S, Pennington J, Schoenhofen E, Palaima E, et al. Cardiovascular disease delay in centenarian offspring: role of heat shock proteins. *Ann N Y Acad Sci.* 2004;1019:502–5.
96. Gentz-Zotz S, Bolger AP, Kalra PR, von Haehling S, Doehner W, Coats AJ, et al. Heat shock protein 70 in patients with chronic heart failure: relation to disease severity and survival. *Int J Cardiol.* 2004;96:397–401.
97. Bonafele RJ, Calvo JP, Fausti JMV, Puebla S, Gambarte AJ, Manucha W. Nitric oxide: a new possible biomarker in heart failure? Relationship with pulmonary hypertension secondary to left heart failure. *Clin Investig Arterioscler.* 2017;29:120–6.
98. Molina MN, Ferder L, Manucha W. Emerging role of nitric oxide and heat shock proteins in insulin resistance. *Curr Hypertens Rep.* 2016;18:1, <http://dx.doi.org/10.1007/s11906-015-0615-4>.
99. Dhingra R, Larson MG, Benjamin EJ, Lipska I, Gona P, Corey D, et al. Cross-sectional correlates of serum heat shock protein 70

- in the community. *Am J Hypertens.* 2006;19:227–31, discussion 232-3.
100. Mazzei L, Docherty NG, Manucha W. Mediators and mechanisms of heat shock protein 70 based cytoprotection in obstructive nephropathy. *Cell Stress Chaperones.* 2015;20:893–906.
101. Rinaldi Tosi ME, Bocanegra V, Manucha W, Gil Lorenzo A, Vallés PG. The Nrf2-Keap1 cellular defense pathway and heat shock protein 70 (Hsp70) response Role in protection against oxidative stress in early neonatal unilateral ureteral obstruction (UUO). *Cell Stress Chaperones.* 2011;16:57–68.