



Enfermedades Infecciosas y Microbiología Clínica

www.elsevier.es/eimc



Review article

Helminthosis and eosinophilia in Spain (1990–2015)[☆]



Cristina Carranza-Rodríguez^{a,b,1}, Miriam Escamilla-González^{c,1}, Isabel Fuentes-Corripio^c, María-Jesús Perteguer-Prieto^c, Teresa Gárate-Ormaechea^{c,2}, José-Luis Pérez-Arellano^{a,b,*,2}

^a Departamento de Ciencias Médicas y Quirúrgicas, Universidad de Las Palmas de Gran Canaria, Las Palmas de Gran Canaria, Spain

^b Unidad de Enfermedades Infecciosas, Complejo Hospitalario Universitario Insular Materno Infantil, Las Palmas de Gran Canaria, Spain

^c Servicio de Parasitología, Centro Nacional de Microbiología, Instituto de Salud Carlos III, Majadahonda, Madrid, Spain

ARTICLE INFO

Article history:

Received 7 September 2015

Accepted 30 November 2015

Available online 4 February 2018

Keywords:

Helminths

Eosinophilia

Spain

Imported diseases

ABSTRACT

The finding of blood eosinophilia in a patient is a relatively frequent reason to refer him/her to a Clinical Department of Infectious Diseases. The doctor usually intends to rule out a parasitic disease in the autochthonous population, travellers or immigrants. It is uncommon for an eosinophilia to be produced by protozoa infection, whereas helminth parasites are more frequently associated with an increase of eosinophil counts in the infected patient. Eosinophilia can be the only abnormal finding, or it could be part of more complex clinical manifestations suffered by the patient. Furthermore, many, but not all, helminth infections are associated with eosinophilia, and the eosinophil level (low, high) differs according to parasite stages, helminth species, and worm co-infections. The purpose of the present article is to carry out a systematic review of cases and case series on helminth infections and eosinophilia reported in Spain from 1990 to 2015, making a distinction between autochthonous and imported (immigrants and travellers) cases, and studying their relationship with immunodepression situations.

© 2015 Elsevier España, S.L.U. and Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. All rights reserved.

Helminthosis y eosinofilia en España (1990–2015)

RESUMEN

La detección de eosinofilia periférica es un motivo relativamente frecuente para la remisión de un paciente a una Unidad/Servicio de Enfermedades Infecciosas. En general, se pretende descartar una enfermedad parasitaria, tanto en personas autóctonas como en viajeros o inmigrantes. Excepcionalmente la eosinofilia relacionada con parásitos corresponde a una protozoosis, siendo los helmintos los principales agentes causales de este hallazgo hematológico. La eosinofilia puede ser el único hallazgo anormal o formar parte del cuadro clínico-biológico del paciente. Por otro lado, no todas las helmintosis se asocian de forma sistemática a eosinofilia, y el grado de la misma difiere entre las fases de la infección y el tipo de helminto. El propósito de esta revisión es un estudio sistemático de la relación entre helmintosis y eosinofilia en la literatura española, distinguiendo los casos autóctonos e importados, así como la relación con situaciones de inmunodepresión.

© 2015 Elsevier España, S.L.U. y Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. Todos los derechos reservados.

Palabras clave:

Helminthosis

Eosinofilia

España

Enfermedades importadas

DOI of original article: <http://dx.doi.org/10.1016/j.eimc.2015.11.019>

[☆] Please cite this article as: Carranza-Rodríguez C, Escamilla-González M, Fuentes-Corripio I, Perteguer-Prieto M-J, Gárate-Ormaechea T, Pérez-Arellano J-L. Helminthosis y eosinofilia en España (1990–2015). *Enferm Infecc Microbiol Clin.* 2018;36:120–136.

* Corresponding author.

E-mail address: jlperez@dcmq.ulpgc.es (J.-L. Pérez-Arellano).

¹ Both have contributed in a similar way to this work (first authors).

² Both have contributed in a similar way to this work (last authors).

Introduction

The term “eosinophilia” indicates the raising in the number or percentage of polymorphonuclear-eosinophil leukocytes in any solid or liquid tissue.¹ Although no limit has been established, it is considered that eosinophilia exists when blood values surpass 450 cells/ μ l.¹ Their detection in the blood requires an investigation of the cause responsible, as it may arise from highly diverse causes,

from mild conditions (e.g. allergic rhinitis) to severe processes (e.g. tumours of the hematopoietic and lymphoid tissues).¹ One of the main conditions leading to detection of eosinophilia is the presence of a parasitic disease. Furthermore, and with few exceptions (*Isospora belli*, *Dientamoeba fragilis*, *Sarcocystis* spp.), the protozoa are not the agents connected to the appearance of eosinophilia, and their presence suggests helminthosis.¹

In Spain extensive literature exists regarding the association between infection by helminthes and the presence of eosinophilia. The global map of this association in Spain is complex, since in several cases parasitism is detected in isolated cases, and in others, in form of outbreaks. Several parasitic diseases also only appear as imported diseases (travellers or immigrants) whilst others have a more cosmopolitan distribution.² Thirdly, the presence of eosinophilia depends on the life cycle stage of the parasite and even of response to treatment. Finally, several factors such as age, the patient's geographical origin, their immunological status and the presence of polyparasitism are determining factors in the detection of eosinophilia.

The aim of this study was to review the helminths associated with eosinophilia in Spain during the last 25 years. This study is based on a systematic search in PubMed, which included originals, brief originals, clinical notes and scientific letters (Fig. 1). The electronic search strategy was as follows: country (Spain) AND disease OR agent. The following MESH terms were considered in the inclusion of diseases: Helminthiasis, Taeniasis, Hymenolepiasis, Dipylidiasis, Cysticercosis, Echinococcosis, Sparganosis, Diphyllbothriasis, Schistosomiasis, Fascioliasis, Paragonimiasis Opisthorchiasis, Clonorchiasis, Dicrocoeliasis, Enterobiasis, Ancylostomiasis, Necatoriasis, Ascariasis, Trichuriasis, Strongyloidiasis, Dirofilariasis, Filariasis, Loiasis, Onchocerciasis, Mansonelliasis, Dracunculiasis, Trichinellosis, Anisakiasis, Toxocariasis, Gnathostomiasis. With regard to the agents, the following MESH terms were included: *Helminth*, *Taenia*, *Hymenolepis*, *Dipylidium*, *Cysticercus*, *Echinococcus*, *Spirometra*, *Sparganum*, *Schistosoma*, *Fasciola*, *Paragonimus*, *Opisthorchis*, *Clonorchis*, *Heterophyes*, *Metagonimus*, *Dicrocoelium*, *Ancylostoma*, *Necator*, Hookworm, *Ascaris*, *Trichuris*, *Strongyloides*, *Capillaria*, *Dirofilaria*, *Wuchereria*, *Brugia*, *Loa*, *Onchocerca*, *Mansonella*, *Dracunculus*, *Trichinella*, *Anisakis*, *Toxocara*, *Gnathostoma*. Results were restricted to studies carried out in humans. The search period lasted from January 1st 1990 to 31st August 2015.

Cestodiasis

Cestodiasis are diseases caused by flat worms (phylum *Plathelminthes*) with a segmented body (*Cestoda* classification). The main agents of the disease in humans include 2 types, *Pseudophyllidea* and *Cyclophyllidea*, and may cause the disease either through the adult form of the parasite or through the larva stage or both. Table 1 indicates the main types of cestodiasis. In general, we may state that the eosinophilia associated with the cestodiasis is mild or moderate, and often does not appear during the course of the disease. Eosinophilia is also more common in larval cestodiasis (with tissue compromise) than in those produced by adult worms (with isolated intestinal compromise). Finally, the rupture or surgical manipulation of larval forms (especially in cystic echinococcosis and to a lesser degree in alveolar echinococcosis) is associated with a notable raising in the number eosinophils in the bloodstream.

The cestodiasis diagnosed most frequently in Spain is, without a doubt, that of infections caused by the parasites of the order *Cyclophyllidea*. Among them, the lowest number of references corresponds to the *intestinal types*. Although the recording of intestinal infection caused by *Taenia* sp., is relatively frequent, as shown by indirect data (personal cases in the Hospital Universitario Insular de Gran Canaria) and samples sent to the Instituto de Salud

Carlos III, few cases have been published in the literature.³ Infection by *Hymenolepis nana* has been reported in Spain, mainly in cases of Sahrawi children from Tindouf who are spending their summer holidays in Spain. Prevalence is 6.5–7.5% and there is frequent co-parasitism with intestinal protozoa (*Giardia intestinalis*).^{4,5} A case of infection by this helminth has also been reported anecdotally in a child from Ecuador.⁶ We only found one case published on *Hymenolepis diminuta* in a child aged 5 in the province of Guadalajara.⁷ In the review carried out we did not locate cases of infection in humans by *Dipylidium caninum*. However, the detection of eggs from the parasite in dog faeces in several regions of Spain,^{8–11} suggests that this parasitism could be under-diagnosed.

In contrast, the *larval forms* of cestodiasis are common, both as autochthonous parasitism or imported. The 2 major ones are cystic hydatidosis and cysticercosis, and particularly in its neurological form (neurocysticercosis).

Cystic hydatidosis produced by *Echinococcus granulosus* is an autochthonous zoonotic disease, endemic in the Iberian peninsula, which had major socio-economic repercussions up to the end of the 20th century.^{12–15} Historically the most affected regions were the Northern communities (Basque Country, Navarre, Aragon, La Rioja, Cantabria) and the central regions (Castille and Leon, Extremadura, Castilla-La Mancha).^{15–22} Moreover, in recent years a large number of cases were reported in the Community of Valencia.²³ However, we found no published cases of cystic hydatidosis in the Canary Island Community or the Balearic Island communities. Hydatidosis was significantly reduced thanks to the control programmes introduced in the eighties and nineties, although direct and indirect data exist, such as the presence of new cases of infants or the maintenance of high levels of infection in young people in the last few years, which suggest a re-emergence of the disease.^{12,13} In Spain, just as in other parts of the world, the most common clinical manifestations of this disease are liver^{24–27} and bile duct^{28,29} compromise and secondly derive from respiratory system lesions.^{18,30–33} Furthermore, several Spanish groups have reported cases or series of “atypical” forms of cystic hydatidosis which were intra-abdominal (splenic,^{34–36} pancreatic,²⁰ renal,³⁷ ovarian³⁸ and other less common forms^{17,39–41}) such as in other organs (heart,^{42–45} muscles and skeleton,^{21,46,47} bone^{48,49} or skin and soft tissues⁵⁰). Several cases of imported cystic hydatidosis have also been reported, although they are scarce and not well characterised.^{51–53} The presence of eosinophilia is highly uncommon in the classic cystic hydatidosis, but a high rate of incidence has been published in several extra pulmonary forms, and particularly renal,³⁷ or after rupture of the cysts (spontaneous or during surgery).^{54–56}

Alveolar hydatidosis, cause by *Echinococcus multilocularis*, is exceptional in Spain, both in its autochthonous form and its imported form. There have only been 2 references in the literature in the last 25 years. These cases presented in the usual manner with similar symptoms to a primary liver tumour.^{57,58}

Cysticercosis is a disease caused by the larvae of *Taenia solium* (*Cysticercus cellulosae*). The 3 main locations of the larval forms are cutaneous, ocular and neurological. The most severe cases are logically those which affect the central nervous system and a few cases have been published in Spain, with mixed forms.⁵⁹ Table 2 displays the epidemiological data of the cases and the series of neurocysticercosis published in Spain.^{60–80} As maybe observed, and has already been reported in other references,^{60–82} there are 2 different patterns of the disease: (a) imported, which includes the greater part of cases detected in the last few years, and particularly those observed in immigrants with an age range from infancy to middle age, and (b) autochthonous, with rare cases, and reported in Spaniards over the age of 18. The main origin of the imported cases is Latin American (mostly Ecuador, Peru, Colombia and Bolivia), although a few cases of patients also

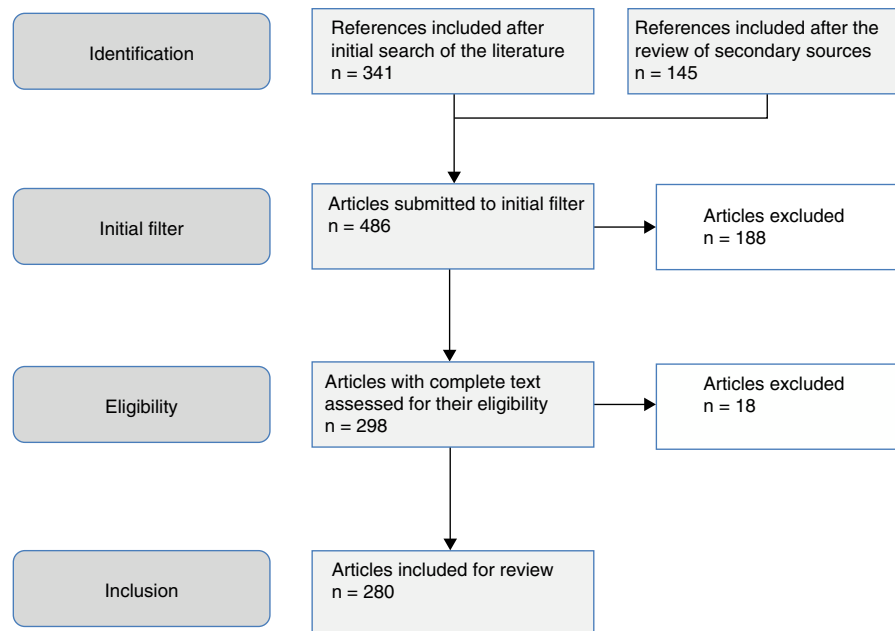


Fig. 1. Strategy for the selection of articles.

Table 1
Main cestodiasis.

Order	Genus	Species	Infective phase	Disease
<i>Pseudophyllidea</i>	<i>Diphyllobothrium</i>	<i>D. latum</i> <i>D. dendriticum</i> <i>D. pacificum</i> <i>D. nihonkaiense</i> <i>D. balenopterae</i>	Adults	Diphyllobothriasis
	<i>Spirometra</i>	<i>S. erinacei</i> <i>S. theileri</i> <i>S. mansonioides</i>	Larvae	Esparganosis
<i>Cyclophyllidea</i>	<i>Taenia</i>	<i>T. saginata</i>	Adults	Teniosis
		<i>T. solium</i>	Adults	Teniosis
		<i>T. asiatica</i>	Larvae	Cisticercosis
	<i>Echinococcus</i>	<i>E. granulosus</i>	Adults	Teniosis
		<i>E. multilocularis</i>	Larvae	Hydatidosis quística
	<i>Hymenolepis</i>	<i>H. nana</i>	Larvae	Hydatidosis alveolar
<i>Dipylidium</i>	<i>H. diminuta</i> <i>D. caninum</i>	Adults	Himenolepiosis	
			Adults	Dipilidiosis

come from Africa (Guinea Bissau, Cape Verde, the Ivory Coast) and Asia (India and China).^{64,66–69,74–80} Most autochthonous cases were described in Extremadura, La Rioja, Madrid and the Community of Valencia. In the imported cases there is a similar number of males and females, whilst in the autochthonous cases there is a clear predominance of males. In general, neurocysticercosis presents in immune-competent people although in Spain several cases have been reported in people infected with HIV⁷³ and those with transplants.⁷² From the clinical viewpoint, the most frequent manifestations of neurocysticercosis are epileptic crises (of different types) and headaches. However, in Spain other unusual presentations have been reported, such as blepharospasm,⁸³ Bruns syndrome (sudden headache associated with acute vestibular syndrome related to sudden movements of the head),^{71,78} medullar lesions,⁶⁰ psychiatric changes⁷⁹ and sudden death.⁶⁹ The presence of eosinophilia in patients with neurocysticercosis is exceptional and for the most part, poorly documented.^{84–86}

Infections by parasites of the *Pseudophyllidea* order are rare in Spain. Among these, most references correspond to cases of the diphyllobothriasis.^{87–91} Only one case of sparganosis (infection by different species of *Spirometra*) was recently reported.

This was an imported sparganosis by a male aged 29 from Bolivia, with symptoms of convulsions and multi-cystic cerebral lesion and who was diagnosed with suspected dysembryoplastic neuroepithelial tumour (DNET).⁹² With regard to diphyllobothriasis, the symptoms of this disease may not be present or may present non-specific abdominal upset which may or may not be associated with megaloblastic anaemia. Of the published cases in Spain, most are autochthonous, and diagnosed in a broad range of ages (3–71 years). The most frequently detected species is *Diphyllobothrium latum*, although cases caused by *Diphyllobothrium pacificum* and *Diplogonoporus balaenopterae*⁹⁰ have also been reported. These are possibly related to the consumption of imported fish or travel abroad. Eosinophilia is exceptional in the cases published in Spain.

Trematodosis

Trematodosis are diseases produced by flatworm (phylum *Plathelminthes*) with unsegmented foliaceous body (*Trematoda* class). Most are hermaphrodites, with the exception of the genus

Table 2
Neurocysticercosis in Spain.

Year	Author/s	n	Immigrants (n)	Autoctonomous (n)	Origin/community	Age (years)	Gender
1996	Corral et al. ⁶⁰	1	0	1	La Rioja	20	W
1998	Fernández-Gómez et al. ⁶¹	1	1	0	Ecuador	39	M
1999	Font Puig et al. ⁶²	4	4	0	Latin America(3) India (1)	–	–
2000	Castellanos et al. ⁶³	2	0	2	Extremadura	64/72	M
2001	Terraza et al. ⁶⁴	10	9	1	Latin America (7) India (1) Guinea Bissau (1)	21–80	4 M/6 W
2002	Rodríguez-Sánchez et al. ⁶⁵	1	0	1	Extremadura	51	M
2003	Roca et al. ⁶⁶	23	23	0	Latin America (17) Africa (2) Asia (2)	5–65	13 M/10 W
2003	Cañizares et al. ⁶⁷	1	1	0	Ecuador	33	M
2004	Ortega-Herrera et al. ⁶⁸	1	1	0	Ecuador	26	W
2005	Llompert Pou et al. ⁶⁹	1	1	0	Peru	31	W
2005	Esquivel et al. ⁷⁰	20	–	–	–	–	–
2005	Jiménez-Caballero et al. ⁷¹	1	1	0	Ecuador	44	M
2007	Barra Valencia et al. ⁷²	1	1	0	Ecuador	49	M
2007	Ramos et al. ⁷³	2	1	1	Community of Valencia	Colombia	–
2007	Guerra del Barrio et al. ⁷⁴	1	1	0	Ecuador	56	W
2007	Sierra Bergua et al. ⁷⁵	1	1	0	Ecuador	47	W
2008	Más-Sesé et al. ⁷⁶	23	23	0	Ecuador (18) Bolivia (2)	29 ± 12	14 M/9 W
2011	Ruiz et al. ⁷⁷	35	35	0	Latin America	7–60	24 M/11 W
2011	Aguilar-Amat et al. ⁷⁸	1	1	0	Ecuador	29	W
2012	De Anta Tejado et al. ⁷⁹	1	1	0	Ecuador	15	M
2013	Friero-Dantas et al. ⁸⁰	2	2	0	Colombia Cape Verde	8/5	1 M/1 W

W: woman; M: man; –: no data.

Schistosoma, which present sexual dimorphism and characteristic morphology.

Schistosomiasis

All cases published in Spain on this disease correspond to imported forms. However, symptoms and biology differ notably between infections detected in travellers and those diagnosed in immigrants (Tables 3–5).

With regard to travellers, isolated cases and series have been reported in several Spanish regions.^{93–106} Practically all patients had travelled to Africa, with the most visited countries being Mali (particularly Dogon country), Burkina Fasso, Uganda (Sesé island), Malawi, Senegal and Madagascar. In general, the disease is more common in males and affects middle aged people (which are congruent with the common international traveller profile). The main species responsible is *Schistosoma haematobium*, followed by *Schistosoma mansoni* and *Schistosoma intercalatum*. Mixed forms of parasitism are also common. Symptoms are similar to acute schistosomiasis, with Katayama syndrome^{98,103–105} and bather's dermatitis⁹⁷ as the most common symptoms. Other major changes of the disease are genital-urinary (hematuria, prostatitis, hematospermia).^{93–95,98,99} We should note that the first description of hematospermia as a sign of schistosomiasis was reported by Spanish authors.¹⁰⁷ One interesting factors is the detection of asymptomatic patients, which together with the different interval between exposure and appearance of symptoms in people with the same outbreak, suggests the presence of host factors modulate the expression of the disease. With regard to eosinophilia, this is present in acute forms, generally high but variable, among people of the same outbreak.

Schistosomiasis in *Immigrants* presents similarities but also major differences with regard to the forms described in travellers.^{101,108–133} Thus most cases originate from sub-Saharan Africa, particularly West Africa and specifically the countries of

Mali, Equatorial Guinea and Mauritania. In immigrants, schistosomiasis also predominates in males, although the age interval is higher, with cases in children and older adults. *S. haematobium* is the most common agent, followed by *S. mansoni* and *S. intercalatum*. Symptoms are highly varied, with hematuria (micro or macroscopic) being the most common factor, and related to the infection by *S. haematobium* and secondly abdominal pain in infections by *S. mansoni* and *S. intercalatum*. Furthermore, in Spain there have been several cases with atypical symptoms and complications of the disease. Specifically, some of the most serious forms of the parasitosis (neuroschistosomiasis)¹³⁴ have been reported, and specifically transverse myelitis^{115,116} and a hemispheric focal lesion, with hemiparesis and convulsions.¹²⁵ Other complications described in immigrants in Spain are female infertility,¹⁰⁹ pulmonary hypertension¹²⁵ and acute appendicitis.¹²⁶ Also, in the population under study a large number of cases of schistosomiasis are asymptomatic. Eosinophilia in imported schistosomiasis in immigrants is highly variable. Absolute eosinophilia (mild-moderate) has been detected in several cases, relative eosinophilia in others and no identification of this change in the remainder. A screening for these disease in immigrants coming from areas under risk would be reasonable, bearing in mind the relevance of the long-term complications (e.g. portal hypertension, squamous cancer of the bladder).

A major problem of schistosomiasis is complex diagnosis. In acute cases it is very common for no eggs to be present in urine or faeces and in chronic cases the elimination of eggs is intermittent. Radiologic studies¹³⁵ and cytoscope¹²³ may help when there is suspected diagnosis, particularly in cases of infection by *S. haematobium*. Serology has been a standard test to detect several acute cases and other parasitological negative cases, although there are many limitations.^{136,137} Application of standard molecular biological techniques (PCR) has not been useful in clinical samples nor proven their usefulness in clinical practice.^{138,139} New diagnostic procedures (LAMP) are current under development, with promising results.¹⁴⁰

Table 3
Schistosomiasis imported by travellers (cases and series).

Author/s	Year	n	Destination	Age (years)	Gender	Species	Symptoms	Eosinophilia
Corachán et al. ⁹³	1992	43	Mali	22–47	21 M/22 W	<i>S. haematobium</i> (34) <i>S. mansoni</i> (4) <i>S. intercalatum</i> (10)	Prostatitis (3) Dermatitis (3)	–
Corachán et al. ⁹⁴	1997	80	Africa Brazil	21–54	43 M/37 W	<i>S. haematobium</i> (63) <i>S. mansoni</i> (14) <i>S. intercalatum</i> (13)	Prostatitis (13) Katayama S.(8) Vulvitis (6) Dermatitis(1) Cystitis (6) Diarrhoea (4) Asymptomatic (42)	Yes (>500 cells/μl)
Vilana et al. ⁹⁵	1997	9	–	25–35	M	<i>S. haematobium</i> (5) <i>S. mansoni</i> (1) <i>S. intercalatum</i> (2)	Hemospermia Perineal pain	–
Elcuaz et al. ⁹⁶	1998	20	Burkina Faso	19–43	M/W	<i>S. mansoni</i>	Katayama S. (14) Asymptomatic (6)	14 (800–10.500)
Bou et al. ⁹⁷	2001	25	Mali (23) Uganda (2)	23–37	11 M/14 W	<i>S. haematobium</i> (6) <i>S. mansoni</i> (3)	Katayama S. (25) Dermatitis (9)	yes (mean 3.500)
Roca et al. ⁹⁸	2002	80	Sub-Saharan Africa	–	–	<i>S. haematobium</i> (55) <i>S. mansoni</i> (9) <i>S. intercalatum</i> (6) Mixed forms (10)	Asymptomatic Katayama S. (14) Diarrhoea Prostatitis	12/14
Santos et al. ⁹⁹	2004	1	Mali	34	M	<i>S. haematobium</i>	Hematuria	Yes
Alonso et al. ¹⁰⁰	2006	2	Mali Senegal	26 32	M	<i>S. haematobium</i>	Hematuria Hemospermia	Yes (800–900 cel/μl)
de Górgolas et al. ¹⁰¹	2009	1	Mali	34	W	<i>S. haematobium</i>	Asymptomatic	Yes
Zamarrón et al. ¹⁰²	2010	44	Sub-Saharan Africa	–	–	<i>S. haematobium</i>	–	Yes
Rivas et al. ¹⁰³	2012	1	Madagascar	44	M	– ^a	Katayama S.	–
Pérez-Arellano et al. ¹⁰⁴	2012	2	Mali	30 34	M W	<i>S. intercalatum</i>	Katayama S.	Yes
Muñoz et al. ¹⁰⁵	2013	1	Senegal	25	M	– ^a	Katayama S.	Yes
Martínez-Calle et al. ¹⁰⁶	2013	1	Mali	26	M	<i>S. haematobium</i>	Asymptomatic	Yes

W: woman; M: man; –: no data.

^a Serological diagnosis.**Table 4**
Schistosomiasis imported by immigrants (isolated cases).

Author/s	Year	n	Origen	Age (years)	Gender	Species	Symptoms	Eosinophilia
Chaves et al. ¹⁰⁸	1992	1	Equatorial Guinea	24	M	<i>S. mansoni</i>	Itchiness	Yes (800 cells/μl)
Balasz et al. ¹⁰⁹	1995	1	Nigeria	26	W	<i>S. haematobium</i>	Infertility	–
Gairi Burgués et al. ¹¹⁰	2002	1	Gambia	13	M	<i>S. haematobium</i>	Hematuria	Yes
Lopez-Calleja et al. ¹¹²	2003	1	Gambia	25	W	<i>S. haematobium</i>	Abdominal pain	Yes
Donate et al. ¹¹⁴	2006	1	Mali	27	M	<i>S. haematobium</i>	Hematuria	Yes
Campo-Esquisabel et al. ¹¹⁵	2007	1	Benin	25	M	<i>S. mansoni</i>	Asymptomatic	Yes
Tarabini-Castellani et al. ¹¹⁶	2007	1	Mali	27	M	<i>S. haematobium</i>	Transverse myelitis	–
López López et al. ¹¹⁷	2007	1	Mali	28	M	<i>S. haematobium</i>	Hematuria	No
Rascarachi et al. ¹¹⁹	2009	1	Equatorial Guinea	51	W	<i>S. intercalatum</i>	Abdominal pain	–
Llenas-García et al. ¹²⁰	2009	1	Equatorial Guinea	25	M	<i>Schistosoma</i> sp	Epileptic episode	No
Landeyro et al. ¹²¹	2010	1	Guinea	11	M	<i>S. haematobium</i>	Rectal bleeding	Yes
Rodríguez-Guardado et al. ¹²²	2010	1	Equatorial Guinea	17	M	<i>S. intercalatum</i>	Diarrhoea Rectal bleeding	Yes
Carrión López et al. ¹²³	2010	1	Mali	19	M	<i>S. haematobium</i>	Hematuria	–
Álvarez Maestro et al. ¹²⁴	2010	1	Gambia	21	M	<i>S. haematobium</i>	Hematuria	No
Gran et al. ¹²⁵	2011	1	Equatorial Guinea	11	W	<i>S. intercalatum</i>	Right heart failure	–
López de Cenarruzabeitia et al. ¹²⁶	2012	1	Mauritania	45	M	<i>S. mansoni</i>	Abdominal pain	No
García Pérez et al. ¹²⁷	2014	1	Mali	11	M	<i>S. haematobium</i>	Hematuria	Yes (34%)

W: woman; M: man; –: no data.

Trematodosis by hermaphrodite species

The most common autochthonous trematodosis in Spain is *fasciolosis*, a disease caused by *Fasciola hepatica* in Spain and related to the ingestion of metacercariae which are present in vegetables

(mainly cress).^{141–154} With few exceptions, the cases reported are autochthonous and their analysis in the last 25 years (Table 6) has enabled us to made several generalisations: (1) Most published cases correspond to patients from the northern half of the Iberian peninsula (Basque country, Galicia, Cantabria, Navarre and Castille

Table 5
Schistosomiasis imported by immigrants (series).

Author/s	Year	n	Origen	Age (years)	Gender	Species	Symptoms	Eosinophilia
Roca et al. ⁹⁸	2002	200	Africa West Africa	– 35 ± 5	– 24 M/1 W	<i>S. haematobium</i> (175)	– Asymptomatic (10)	– Yes (85%)
Rotger et al. ¹¹¹	2004	15	Sub-Saharan Africa	6–48	14 M/1 W	<i>S. mansoni</i> (25) <i>S. haematobium</i> (10) <i>S. mansoni</i> (2)	Hematuria (5)	Yes (11/12)
Pardo et al. ¹¹³	2006	37	West Africa	–	–	<i>S. haematobium</i> (10) <i>S. mansoni</i> (6)	–	Yes (absolute) > 450/μl
Carranza et al. ¹¹⁸	2008	17	West Africa	–	–	<i>S. haematobium</i> (5) <i>S. mansoni</i> (5)	–	Yes (relative)
Barrio Muñoz et al. ¹²⁷	2013	8	Sub-Saharan Africa	27	–	<i>S. haematobium</i> (6)	Hematuria (6)	Yes (50%)
Salvador et al. ¹²⁹	2013	11 ^a	Sub-Saharan Africa (9) Latin America (2)	–	–	– ^b	Asymptomatic	Yes (7/11)
Cobo et al. ¹³⁰	2014	278	Sub-Saharan Africa Latin America	–	–	<i>S. haematobium</i> (165) <i>S. mansoni</i> (59)	–	–
Belhassen-García et al. ¹³¹	2015	22	Sub-Saharan Africa (19)	<18	–	– ^c	–	Yes
Monge-Maillo et al. ¹³²	2015	8	Sub-Saharan Africa	22	6 M/2 W	– ^b	–	–
Salas-Coronas et al. ¹³³	2015	133	Sub-Saharan Africa	–	–	<i>S. haematobium</i> (86) <i>S. mansoni</i> (30)	–	Yes

W: woman; M: man; –: no data.

^a Patients infected with HIV.^b Only serology.^c Serological diagnosis in most cases.**Table 6**
Fasciolosis in Spain.

Author/s	Year	n	Autonomous community	Age (years)	Gender	Symptoms	Eosinophilia
Pulpeiro et al. ¹⁴¹	1991	15	Galicia	16–64	7 M/8 W	Abdominal pain (6) Weight loss, asthenia (3) Right hypochondrium pain (2) Fever (1) Asymptomatic(3)	Yes (14/15)
López-Rosés et al. ¹⁴²	1993	1	Galicia	46	W	Fever, Abdominal pain, jaundice, dark urine, vomiting	No
Arjona et al. ¹⁴³	1995	20	Cantabria	11–62	14 M/6 W	Abdominal pain (13) Fever (12) Weight loss (7)	Yes (19/20)
Gómez Cerezo et al. ¹⁴⁴	1998	1	Castille and Leon	67	W	Fever, arthralgias, jaundice	Yes (2.860/μl)
Segado Soriano et al. ¹⁴⁵	1998	3	Madrid	34/39/48	1 M 2 W	Right hypochondrium pain, Fever	Yes (2.700–17.043/μl)
López-Vélez et al. ¹⁴⁶	1999	6	Madrid	24–53	2 M/4 W	Fever and Abdominal pain	Yes (1.440–14.400/μl)
Pérez et al. ¹⁴⁷	2000	1	Navarre	53	W	Eosinophilic panniculitis	Yes (20.300/μL)
Núñez Fernández et al. ¹⁴⁸	2001	2	Galicia	48 43	W M	Fever and Abdominal pain Asymptomatic	8.100/μl 15.900/μl
Cosme et al. ¹⁴⁹	2001	37	Basque country	19–71	23 M/14 W	Acute phase (32) Chronic phase (5)	Yes (34/37)
Cilla et al. ¹⁵⁰	2001	61	Basque country	20–81	34 M/27 W	Fever, Abdominal pain, hepatomegaly	Yes
González-Llorente et al. ¹⁵¹	2002	1	Castille and Leon	47	W	Abdominal pain	Yes (11%)
Cosme et al. ¹⁵²	2003	7	Basque country	29–69	4 M/3 W	Fever, hepatomegaly, weight loss	Yes
Cirera et al. ¹⁵³	2004	1	Catalonia	66	W	Constitutional Syndrome	5.100/μl
Echenique-Elizondo et al. ¹⁵⁴	2005	1	Basque country	31	W	Abdominal pain	Not defined

Leon). (2) During the reviewed period a notable reduction in cases was observed, thoroughly documented in Guipúzcoa.¹⁵⁰ In fact, we are unaware of cases published in Spain since 2005, which does not exclude the presence of isolated cases diagnosed in hospitals and benchmark centres. (3) Most cases are reported in adults, with similar prevalence in men and women. (4) The most common clinical symptoms correspond to the acute phase of the disease with constitutional syndrome associated with right hypochondriac pain, and chronic changes (compromise of bile duct) are less habitual. (5) Atypical symptoms are infrequent (subcutaneous nodules,

eosinophilic panniculitis, pulmonary infiltrates, pleuropericarditis, meningitis or swollen lymph nodes)^{141,143,147} and local complications (e.g., pancreatitis) or hepatic subcapsular abscess.^{151,154} (6) Eosinophilia is constant in the disease, usually with very high rates and is the main reason for suspicion of this disease.

Paragonimosis, caused by different species of *Paragonimus*, is an uncommon trematodosis in Spain and always diagnosed as an imported disease (2 patients in Equatorial Guinea and one in Ecuador).^{155–157} Clinical symptoms are most commonly respiratory, imitating tuberculosis, which is also frequently associated

with it. It is very common to find eosinophilia in patients with paragonimosis.

Infections by oriental varieties (*Opisthorchis* spp., *Clonorchis* spp., *Metagonimus* spp. or *Heterophyes* spp.) are exceptional in Spain, with the description of a single case in one south eastern Asiatic immigrant.¹⁵⁸

To conclude this section we should point out the presence of false parasitisms by *Dicrocoelium dendriticum* in immigrants (elimination of eggs in faeces without causing the disease). This trematode which parasitises the bile ducts of herbivores may cause disease in humans, although the most frequent occurrence is the expulsion of eggs in faeces after the ingestion of raw infected animal liver. The cases described in Spain principally correspond to people of sub-Saharan origin and more infrequently from North Africa.^{159–161}

Nematodosis

Nematodosis are helminthosis produced by parasites of the phylum *Nematoda*, characterised by the cylindrical shape of the worms and the presence of sexual dimorphism. From a clinical viewpoint, they may be classified into 3 large groups: intestinal, haematic/dermal/ocular (filariosis) and tissue (Table 7).

Intestinal

Parasitism by *Enterobius vermicularis* is one of the most common helminths. However, references in the Spanish literature, and especially recent references, are scarce and often discrepant.^{3,4,131,132,162–167} The first aspects to consider in the differences of prevalence reported are the *study design and scope of study*. In Gran Canaria, in a work based on the parasitological data of all health centres during one year, enterobiosis accounted for approximately a third of cases (31.5%; 301/957).³ Furthermore the prevalence of this parasitism depends on whether the study was conducted on asymptomatic peoples^{162–164} or in the presence of symptoms.¹⁶⁵ A further aspect of interest is the clear predominance of those infected at paediatric age, data which could be variable depending on the geographical region second (31.5% in Gran Canaria, 20.4% in the Guadalquivir valley, 10.8% in Valencia and 1.34% in Cuenca).^{3,162,163} In the wholes series described, co-parasitism is normal with other intestinal nematodes and/or protozoa. Data on this infection in immigrants is scarce, with it being more common in Maghreb immigrant children^{3,4} and exceptionally in sub-Saharan children^{130,131,166} and adults.¹⁶⁷ In addition

to pathogenic reasons, the low detection of cases could be related to the absence of a systematic use of the “Graham test”.¹⁶⁵ In general, enterobiosis is not a serious disease. It is characterised by anal and/or genital itchiness. However, several complications have been described such as eosinophilic colitis (related to the parasite larvae) or the reduction of serum concentration of metals (copper, zinc and magnesium).¹⁶⁸ Finally, eosinophilia is mild or does not exist in most cases, with the exception of invasive forms.

Standard intestinal nematodosis are caused by uncinarias (*Ancylostoma duodenale*, *Necator americanus*), *Ascaris lumbricoides* and *Trichuris* spp. (mainly *Trichuris trichiura* and exceptionally *Trichuris vulpis*).¹⁶⁹ These diseases were well represented throughout Spain in other times but incidence has since decreased considerably thanks to improvements in hygiene and health.¹⁷⁰ For this reason, with the exception of isolated cases and usually in people of advanced age, these diseases only appear in immigrants. Analysis of published cases in the last 25 years in Spain^{171–183} have led to several generalisations: (1) In the series where this type of parasitism has been studied in adults, most cases have corresponded to uncinarias, followed by *Trichuris* spp. and *A. lumbricoides*,^{3,113,118,133,167} with this pattern being inverted in series involving children.¹⁶⁶ (2) Most cases correspond to immigrants from Sub-Saharan Africa, although cases in patients from Latin America^{130,172,176,181,182} and Asia¹⁷⁹ have also been reported. (3) Co-parasitism is very common between the classical intestinal nematodes¹⁷³ and other helminths and protozoa.^{108,115,121} (4) Symptoms are very varied and include a large number of asymptomatic cases,^{2,178} either associated or not with absolute eosinophilia^{113,133} or relative eosinophilia,¹¹⁸ non-specific abdominal pains¹⁷⁸ or the “standard” disease symptoms. The latter are the least common form of the infection by intestinal nematodes, although they are over represented in the literature. One case of infection by *Trichuris* spp. with a rectal polyp was reported,¹²¹ several cases of iron-deficiency anaemia in infections by uncinarias^{172,175,178–180,184} and local or systemic complications in the infection by *A. lumbricoides* (e.g. intestinal obstruction,¹⁸² bile duct/pancreatic mass obstruction,^{174,181,183,185} and Löfller¹⁷⁶ syndrome or elimination of adult worm¹⁷⁷) were reported. (5) Detection of eosinophilia and its degree of severity is highly varied, although as a general rule, it is detected in approximately half of cases and is mild or moderate.

Strongyloidosis is a major parasitic disease in Spain. Analysis of isolated cases and patient series reveals several interesting characteristics.^{129,186–205,207–213} Firstly, there is the difficulty of completing precise diagnosis of the nematode. The standard

Table 7
Principal nematodosis in Spain.

Type	Genus	Main species	Epidemiological pattern	Disease
Intestinal	<i>Enterobius</i>	<i>E. vermicularis</i>	Cosmopolitan	Enterobiosis
	<i>Trichuris</i>	<i>T. trichiura</i>	Imported	Tricurosis
	<i>Ascaris</i>	<i>A. lumbricoides</i>	Imported	Ascariosis
	<i>Ancylostoma</i>	<i>A. duodenale</i>	Imported	Uncinariosis
	<i>Necator</i>	<i>N. americanus</i>	Imported	
	<i>Strongyloides</i>	<i>S. stercoralis</i>	Cosmopolitan	Strongyloidosis
	<i>Capillaria</i>	<i>C. philipinensis</i>	Imported	Capilariosis
Filariasis	<i>Loa</i>	<i>L. loa</i>	Imported	Loaosis
	<i>Onchocerca</i>	<i>O. volvulus</i>	Imported	Oncocercosis
	<i>Mansonella</i>	<i>M. perstans</i>	Imported	Mansonellosis
	<i>Dirofilaria</i>	<i>D. immitis</i>	Autoctonous	Dirofilariosis
	<i>Trichinella</i>	<i>T. spiralis</i>	Cosmopolitan	Triquinelosis
		<i>T. britovi</i>		
	<i>Anisakis</i>	<i>A. simplex</i>	Cosmopolitan	Anisakis
Tissue	<i>Trichinella</i>			
	<i>Toxocara</i>	<i>T. canis</i>	Cosmopolitan	Toxocarosis
		<i>T. cati</i>		
	<i>Gnathostoma</i>	<i>G. spinigerum</i>	Imported	Gnathostomosis

co-proparasitological study detects a minimum proportion of cases,^{3,132,167} a figure which increases on using more specific techniques (e.g. the Baermann concentration test, the Harada-Mori technique and the Koga agar plate culture). However, several Spanish strongyloidosis series are based on serological diagnosis, with inherent limitations to this technique.^{133,211} Strongyloidosis in Spain is a disease which mainly affects adults, although there have been isolated cases in children.¹⁹⁹ One essential aspect in strongyloidosis is the differentiation of clinical symptoms between immunocompetent and immunodepressed patients. In the immunocompetent person, this nematode is usually asymptomatic or the course of the disease is with one or several of the data of the eosinophilia-diarrhoea-skin lesion triad. However, in the immunodepressed patient the eosinophilia disappears and serious systemic symptoms may present, such as the systemic infection by intestinal microorganisms led by *Strongyloids*. We are interested in pointing out that the main forms of immunodepression associated with the hyperinfection syndrome correspond to the use of corticosteroids and other immunosuppressants, whilst the association with the HIV infection is infrequent and on many occasions linked to other risk factors (e.g. corticoid steroids or HTLV-I infection)²⁰⁸ (Table 8). From an epidemiological viewpoint in Spain there are 2 strongyloidosis patterns: the autochthonous and the imported forms. At present, strongyloidosis continues to be an autochthonous disease, which is why it is included in the differential diagnosis of any patient with digestive or “allergic” symptoms. Symptoms (Table 9).

Notwithstanding, most cases are sporadic,^{195,198,209} with the exception of a specific area in Valencia (Gandía and Oliva), where an accumulation of patients with a well defined profile arose: male adults with a compatible professional history.^{189,191,194,215} Imported strongyloidosis however is a disease mainly described in immigrants, and exceptionally in travellers^{203,212} (Table 10). Unlike other imported helminths, where sub-Saharan origin predominates, a high number of cases of strongyloidosis come from Latin America.^{129,193,196,200,206,208,212–214} Most cases are asymptomatic, and it therefore seems reasonable to include this disease in the screening of immigrants from the before-mentioned geographical areas. In the remainder of cases, the normal symptoms are digestive and to a lesser extent, cutaneous. We should point out the detection of several cases in patients with allergic symptoms,^{206,207} especially of Latin American origin since the use of corticoid steroids in this context leads to hyperinfection syndrome. Eosinophilia in immunocompetent patients (autochthonous or immigrants) is highly variable (Tables 9 and 10).

We only found one reference in the Spanish literature to the infection by *Capillaria philipinensis*.²¹⁶ This disease is not limited to the Philippines, but is also present in Far and Middle Eastern countries, or South American, and its detection is therefore possible in imported cases. Generally capilarosis is a diarrhoea process with vomiting, although patients without treatment for several months may die due to the loss of electrolytes, or the sepsis associated with secondary bacterial infection (auto-infection processes).

Table 8
Strongyloidosis and risk factors.

Author/s	Year	n	Risk factor	Age (years)	Gender	Symptoms	Eosinophilia
Batista et al. ¹⁸⁷	1992	1	HIV infection	35	M	Gastroenteritis	No
Cremades-Romero et al. ¹⁸⁸	1996	1	Use of corticosteroids	70	M	Pneumonia Gastritis/Abdominal pain	Yes
Olmos et al. ¹⁹⁷	2004	1	Infección HIV Use of immunosuppressants	58	M	Bilateral pneumonia	Yes
Beltrán Catalán et al. ²⁰¹	2009	2	Kidney trasplant	51/55	M	Diarrhoea	Yes
Rodríguez-Hernandez et al. ²⁰²	2009	1	Liver transplant	67	M	Bilateral pneumonia	Yes
Argelich et al. ²⁰⁴	2011	1	Use of corticosteroids	69	M	Meningitis by <i>E. coli</i>	No
Villena-Ruiz et al. ²⁰⁵	2012	1	HIV infection Use of corticosteroids	37	M	Bilateral pneumonia	No
Salvador et al. ¹²⁹	2013	35	HIV infection	–	–	Asymptomatic	9/35
Izquierdo et al. ²¹⁰	2013	1	Stem cell transplant	36	M	Bilateral pneumonia	No

W: woman; M: man; –: no data.

Table 9
Autochthonous strongyloidosis in Spain.

Author/s	Year	n	Location	Age (years)	Gender	Symptoms	Eosinophilia
Cremades-Romero et al. ¹⁸⁹	1997	37	Valencia	51–87	30 M/7 W	Asymptomatic (13) DigestivE (18) Hyper infection (2)	100%
Román Sánchez et al. ¹⁹¹	2001	152	Valencia	67 ± 10	120 M/32 W	Asymptomatic (77%) Digestive (11%) Cutaneous (4%)	82%
Pretel et al. ¹⁹²	2001	3	Murcia	77–82	3 M	Digestive (1) Hyperinfection (2)	100%
Román-Sánchez et al. ¹⁹⁴	2003	31	Valencia	68 ± 8	M	Digestive (70%) Cutaneous (22%)	83%
Martínez Vázquez et al. ¹⁹⁵	2003	1	Pontevedra	25	M	Abdominal pain	Yes
Oltra Alcaraz et al. ¹⁹⁶	2004	473	Valencia	51–91	342 M/131 W	–	–
Mayayo et al. ¹⁹⁸	2005	1	Zaragoza	79	M	Abdominal pain and dyspnoea	No
Valerio et al. ²⁰⁹	2013	2	Barcelona	–	–	–	–

W: woman; M: man; –: no data.

Table 10
Imported strongyloidosis in Spain.

Author/s	Year	n	Origen	Age (years)	Gender	Symptoms	Diagnostic method	Eosinophilia
Díaz et al. ¹⁹³	2002	21	Latin America	–	–	–	Direct	16/21
Oltra Alcaraz et al. ¹⁹⁶	2004	18	15 travellers (Africa, Latin America/Asia) 3 Immigrants	–	–	–	Direct	–
Velasco et al. ²⁰⁰	2006	1	Colombia	29	M	Abdominal pain/malabsorption syndrome	Direct	Yes
González et al. ²⁰³	2010	33	10 travellers 23 Immigrants	29–42	18 M/15 W	Digestive (16) Asymptomatic (17)	Direct	63%
De las Marinas et al. ²⁰⁶	2012	1	Bolivia	27	M	Cough and expectoration	Direct	Yes
Fernández Rodríguez et al. ²⁰⁷	2012	8	Latin America	35–61	2 M/6 M	Rhinitis/Asthma/jaundice	Serology	Yes
Salvador et al. ¹²⁹	2013	35	Latin America (22) sub-Saharan A. (11) North Africa (2)	–	–	–	Serology	26%
Valerio et al. ²⁰⁹	2013	68	Latin America (41) Africa (18) Asia (9)	–	–	Asymptomatic (64%) Digestive (23%) Cutaneous (12%)	Direct Serology	90%
Ramírez-Olivencia et al. ²¹¹	2014	178	Immigrants (120) Equatorial Guinea Bolivia Ecuador Travellers (58)	29–46	76 M/102 W	Asymptomatic (52%) Digestive (23%) Cutaneous (12%)	Direct Serology	49% Relative 30% Absolute
Cabezas-Fernández et al. ²¹³	2015	320	sub-Saharan A. (285) Latin America (20) North Africa (15)	13–71	271 M/49 W	Asymptomatic (58%) Digestive (40.7%) Cutaneous (2%)	Serology	45%
Ramos et al. ²¹⁴	2015	42	Latin America	30–53	23 M/19 W	Asymptomatic	Serology	28%

W: woman; M: man; –: no data.

Filariasis

Filariasis in Spain presents 2 different patterns: autochthonous (of cosmopolitan distribution) where there are no microfilariae, and imported forms, characterised by the presence of microfilariae in blood, skin or eyeball.

The cosmopolitan forms are caused mainly by 2 species of *Dirofilaria* (*Dirofilaria immitis* and *Dirofilaria repens*).^{217–220} In Spain, the 2 regions where this parasitism has been reported are Salamanca and Gran Canaria. The most common form of infection is vectoral transmission by different species of mosquitoes (*Aedes* spp., *Anopheles* spp. and *Culex* spp.) from infected mammals (mainly dogs). The infection takes place through immature worms, is generally asymptomatic and occasionally presents with subcutaneous, pulmonary (persistent or transitory) and ocular nodules. Eosinophilia is exceptional in these cases.

Filariasis associated with the presence of microfilariae in blood or tissues are in all cases imported diseases in Spain.^{102,113,131,166,221–238} Practically all of them are described in African patients, particularly sub-Saharan and with a clear predominance in West Africa (mainly Equatorial Guinea). The imported cases correspond mainly to immigrants, although they have also been described in travellers.^{102,237} Analysis of published cases (Tables 11 and 12) does not show any significant differences in the age of detection (1–80 years) or gender of patients. The 3 main imported filariasis are *mansonellosis* by *Mansonella perstans*, *loiasis* by *Loa loa* and *oncocercosis* by *Onchocerca volvulus*. Detection of *Wuchereria bancrofti* and *Mansonella streptocerca* is anecdotal,²²⁹ and there are no published case of infections by *Brugia malayi*, *Brugia timori* or *Mansonella ozzardi*. These data are subject to complexity in the diagnosis of these nematodes. Direct parasitological studies (blood smears and/or Knott test for the detection of microfilaremia, or “cutaneous pinching” in infection by *O. volvulus* and *M. streptocerca*) are very specific, but present limited sensitivity. Moreover, serological techniques are very sensitive but have inherent limitations (e.g. cross over reactions with other

helminths, no differentiation between active and past infections, etc.). In fact, the use of molecular biological techniques enables the detection of a large number of cases of undiagnosed loiasis from standard techniques.^{239,240} Furthermore, temporary evolution of imported filariasis presents a clear pattern, with a progressively lower number of cases of oncocercosis (very possibly related to control measures in endemic countries, such as Equatorial Guinea) and a progressively lower number of cases of mansonellosis (possibly linked to the screening of these entities in immigrants). Clinical symptoms are highly variable, with a large number of cases being asymptomatic. In symptomatic cases, common signs of these nematodes are: cutaneous (e.g. itching, exanthema, nodules) and ocular. It is therefore of interest to point out the presence of atypical manifestations, such as breast calcifications.^{227,232} The presence of eosinophilia is very common, although its absence does not exclude diagnosis.

Tissue

The 4 main tissue nematodosis reported in Spain are: triquinellosis, anisakidosis, toxocarosis and gnathostomosis.

Infections produced by tissue nematodes of the genus *Trichinella* are autochthonous parasites, well referenced in the Spanish literature and in exceptional cases as an imported pathology.^{241–256} The 2 main species involved in the clinical cases described in Spain are *Trichinella spiralis* and *Trichinella britovi*. The common form of contagion is ingestion of raw meat or not well cooked meat of infected pigs and wild boar, which implies that the cases published are grouped around outbreaks. After the control of the domestic cycle in Spain (pigs), wild animals, like wild boars, have been the origin for most of the recent outbreaks. In any case, the rate of cases of this parasitism is increasingly lower, probably due to veterinary surveillance prior to consumption of game. The main outbreaks (indicated in Fig. 2) are concentrated on mountainous areas: (1) Cantabrian and Pyrenean mountain ranges; (2) Iberian range; (3) Central range; (4) Toledo mountain range and (5) Baetic

Table 11
Imported Filariasis in Spain (cases).

Author/s	Year	n	Origen	Age (years)	Gender	Microorganismo	Symptoms	Eosinophilia
Cuadros et al. ²²¹	1990	1	Equatorial Guinea	66	W	<i>Onchocerca volvulus</i>	Itching	Yes
Rodríguez et al. ²²²	1992	1	Guinea	15	W	<i>Onchocerca volvulus</i>	Itching and hyperpigmentation	Yes
Puente et al. ²²³	1995	1	Equatorial Guinea	27	W	<i>Onchocerca volvulus</i>	Hypopigmented lesions	Yes
Puente et al. ²²⁴	1995	1	Equatorial Guinea	38	W	<i>Loa loa</i>	Subcutaneous oedema	Yes
Molina et al. ²²⁵	1999	1	Senegal	30	M	<i>Mansonella perstans</i>	Asymptomatic	Yes
Daza et al. ²²⁶	2000	1	Equatorial Guinea	34	W	<i>Onchocerca volvulus</i>	Itching and subcutaneous oedema	Yes
Hernández et al. ²²⁸	2003	1	Equatorial Guinea	22	W	<i>Onchocerca volvulus</i>	Sowda	Yes
Saldarriaga et al. ²³⁰	2004	1	Nigeria	28	W	<i>Loa loa</i>	Angioedema	Yes
Arribas et al. ²³²	2005	1	Guinea ?	50	W	<i>Onchocerca volvulus</i>	Breast calcifications	–
López-Rodríguez et al. ²³³	2007	1	Cameroon	24	M	<i>Loa loa</i>	Ocular lava	–
Gil-Setas et al. ²³⁴	2010	1	Guinea ?	69	W	<i>Loa loa</i>	Itching/constitutional S.	Yes
Moliner et al. ²³⁵	2011	1	Equatorial Guinea	17	W	<i>Loa loa</i>	Meningoencephalitis	Yes
Iborra et al. ²³⁶	2011	1	Equatorial Guinea	20	W	<i>Loa loa</i>	Cutaneous lesions	Yes

W: woman; M: man; –: no data.

Table 12
Imported filariasis in Spain (series).

Author/s	Year	n	Origen	Age	Gender	Microorganismo	Symptoms	Eosinophilia
Bastarrika et al. ²²⁷	2001	4	Guinea ?	46 –64	W	–	Breast calcifications	–
Huerga et al. ¹⁶⁶	2002	39	Africa	0 –13	–	<i>Onchocerca volvulus</i> (30) <i>Mansonella perstans</i> (24) <i>Loa loa</i> (4)	Itching (19) Cutaneous lesions (7) Ocular lesion(6)	Yes (35/39)
López-Vélez et al. ²²⁹	2003	245	Africa (80%)	1 –80	–	<i>Onchocerca volvulus</i> (209) <i>Mansonella perstans</i> (85) <i>Loa loa</i> (13) <i>Mansonella streptocerca</i> (7) <i>Wuchereria bancrofti</i> (1) <i>Loa loa</i>	–	–
Carrillo Casas et al. ²³¹	2004	30	Sub-Saharan Africa Equatorial Guinea (57%)	39 ± 17	17 M/13 W	<i>Loa loa</i>	Cutaneous (itching, exanthema) ocular lava (1)	Yes (76%)
Pardo et al. ¹¹³	2006	63	Sub-Saharan Africa	–	–	<i>Mansonella perstans</i> (13) <i>Loa loa</i> (4) <i>Onchocerca volvulus</i> (1)	Asymptomatic (72%)	Yes (100%)
Zamarrón Fuertes et al. ¹⁰²	2010	50	Sub-Saharan Africa	50	–	<i>Loa loa</i> (26) <i>Onchocerca volvulus</i> (17) <i>Mansonella perstans</i> (13) <i>Wuchereria bancrofti</i> (1)	Cutaneous lesions (35/50)	Yes (38/50)
Monge-Maillo et al. ²³⁷	2014	18	Sub-Saharan Africa	25 –42	–	<i>Onchocerca volvulus</i> (16) <i>Mansonella perstans</i> (2) <i>Loa loa</i> (1)	Itching	–
Belhassem-García et al. ¹³¹	2015	40	Africa	12 ± 4	–	–	–	Yes (52%)
Cobo et al. ²³⁸	2015	97	West Africa	30 ± 10	86 M/11 W	<i>Mansonella perstans</i> (96) <i>Loa loa</i> (4)	Abdominal pain (37) Asymptomatic (17)	Yes (27%)

W: woman; M: man; –: no data.

rangea. During the last 25 years no autochthonous cases have been reported in Galicia, Murcia or the Canary island community. Clinical symptoms of the disease come from the tissue invasion of the parasite and the immunological response this triggers. The acute forms include, in variable proportions in each outbreak, the following data: myalgias, fever, exanthema, diarrhoea and palpebral oedema. Atypical forms have also been described, such as thoracic muscle calcification.²⁵⁴ Eosinophilia and the raising of quinase creatinine are common lab data in the described cases.

Since 1991, the description of isolated cases and clinical series of *anisakis* has been constant in Spain.^{257–287} Humans are infected by ingestion of fresh, raw or little cooked fish (e.g. by microwaves²⁶⁸), with the most common epidemiological background being the consumption of anchovies and in some cases, cebiche.²⁸⁴ Practically all cases correspond to acquired autochthonous infections and are mainly reported by autonomous communities, in the central area of the peninsula (Fig. 3). The most common agents are nematodes of the genus *Anisakis* (particularly *Anisakis simplex*) and to a lesser extent *Pseudoterranova decipiens*.²⁵⁸ The pathological

consequences of the infection by these nematodes is apparent in different patterns: digestive, allergic or mixed. The digestive forms may affect several parts of the same, leading to compromise of the gastroduodenal region, ileal region, caecum and colon. Clinical symptoms depend on 2 complementary mechanisms: direct aggression by the nematode and the response of local hypersensitivity to the same. In the upper digestive tract the most common signs are acute epigastric pain after the intake of fish, frequently associated with allergic symptoms. However, when the lower digestive tracts involved abdominal pain presents with characteristics which are indistinguishable from an acute appendicitis or intestinal obstruction. Other atypical manifestations have also been described, such as splenic rupture²⁸⁶ or the appearance of an abdominal mass.²⁷¹ Allergic signs are highly variables, both in their association with intestinal symptoms, and in their severity (from simple cutaneous forms to anaphylaxis).^{260,261,284} Other uncommon signs of hypersensitivity to these nematodes, described in Spain, are the appearance of a nephrotic syndrome²⁸¹ and gingivostomatitis.²⁷⁷ Eosinophilia is an inconstant finding in this

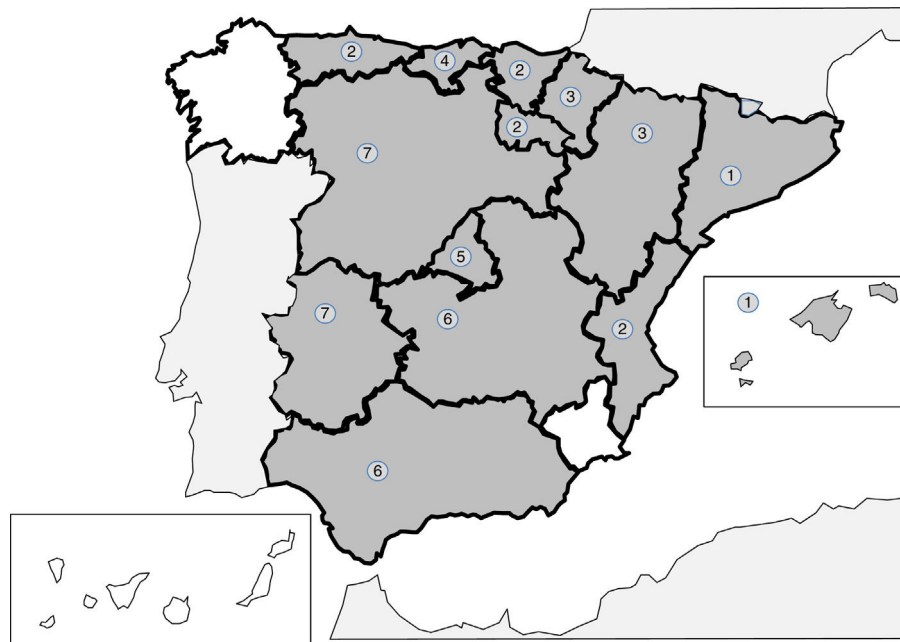


Fig. 2. Number of outbreaks of triquinelosis in Spain by autonomous community (1990–2015).

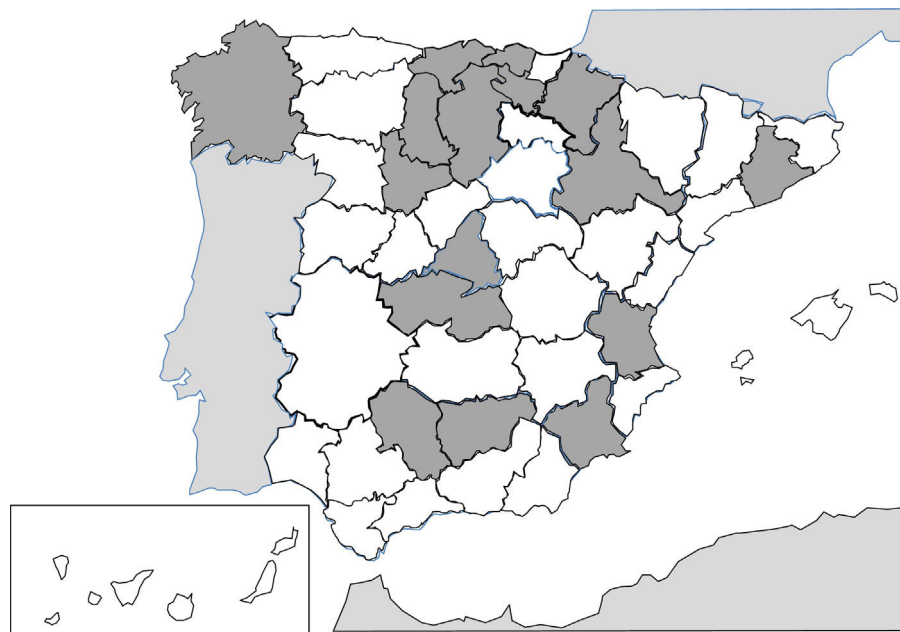


Fig. 3. Provinces with few published cases of anisakiasis (1990–2015).

parasitism (4–41%), and it depends on the before-mentioned clinical forms.

Data on *toxocarosis*, produced by *Toxocara* spp. are scarce in Spain and complex to interpret.^{287–295} The studies on seroprevalence of this infection, conducted in the nineties in several areas of the country, show a high rate of positivity, with figures of up to 66% in children from low socio-economic classes in Guipúzcoa, 3.4% in the general population of the Canary Island Community and between 17% and 32% in Galicia.^{288,292} These data should be further studied, since the etiological diagnosis of toxocarosis is based on serology, which displays cross-over reactivity with other nematodes such as *Anisakis* spp.²⁹⁴ The few reported clinical cases of toxocarosis correspond to both imported and autochthonous forms²⁹⁵ and within the latter,

to ocular toxocarosis²⁹⁰ and visceral *larva migrans*.²⁸⁹ In the cases with visceral manifestations the presence of eosinophilia is common.²⁹¹

Finally, *gnathostomiasis*, mainly produced by *Gnathostoma spinigerum*, is a rare tissue nematodosis, but occasionally reported in Spain.^{296–300} This helminthosis appears as a consequence of food consumption (e.g. raw fish, frogs, snakes) and usually the sign are cutaneous lesions (similar to the cutaneous *larva migrans*) and in severe cases, myeloradiculitis or a radiculomyelomeningoencephalitis.^{298,299} In general, it is a disease which is imported after travelling to Latin America^{297,298} and Asia (South East Asia and China),^{297,298,300} although 2 autochthonous cases have also been diagnosed in women from Granada who had not travelled to the tropics.²⁹⁶

Conclusions

To conclude, helminthosis (autochthonous or imported, in travellers or immigrants, with or without immunosuppression) is a major problem in the Spanish population, both with regards to its prevalence and medical consequences. Association with eosinophilia (absolute or relative) presents great variability and is dependent upon a number of factors. For this reason, knowledge of the current situation may help etiological diagnosis of helminthosis. An appropriate therapeutic approach is required, with the avoidance of “empirical” attitudes that may be unsatisfactory, inappropriate or even harmful.³⁰¹

Financing

The authors declare they have not received any financing for carrying out this study.

Conflict of interests

The authors have no conflict of interests to declare.

References

- Pérez Arellano JL, Pardo J, Hernández Cabrera M, Carranza C, Angel-Moreno A, Muro A. Manejo práctico de una eosinofilia. *An Med Intern (Madrid)*. 2004;21:244–52.
- Vilajeliu Balagué A, de Las Heras Prat P, Ortiz-Barreda G, Pinazo Delgado MJ, Gascón Brustenga J, Bardají Alonso A. Parasitosis importadas en población inmigrante en España. *Rev Esp Salud Publica*. 2014;88:783–802.
- Novo-Veleiro I, Martín Sánchez AM, Elcuaz-Romano R, Muro A, Afonso-Rodríguez O, García Bardeci D, et al. Parasitosis en Gran Canaria (España): estudio prospectivo multicéntrico durante un año. *Rev Ibero-Latinoam Parasitol*. 2012;71:34–41.
- Soriano JM, Doménech G, Martínez MC, Mañes J, Soriano F. Intestinal parasitic infections in hosted Saharawi children. *Trop Biomed*. 2011;28:557–62.
- Paricio Talayero JM, Santos Serrano L, Fernández Feijoo A, Ferriol Camacho M, Rodríguez Serrano F, Brañas Fernández P. Examen de salud de niños de la República Árabe Saharaui Democrática de vacaciones en España. *An Esp Pediatr*. 1998;49:33–8.
- Martínez-Peinado C, López-Perezagua MM, Arjona-Zaragozí FJ, Campillo-Gallego MM. Parasitosis en niña ecuatoriana. *Enferm Infecc Microbiol Clin*. 2006;24:207–8.
- Tena D, Pérez Simón M, Gimeno C, Pérez Pomata MT, Illescas S, Amondarain I, et al. Human infection with *Hymenolepis diminuta*: case report from Spain. *J Clin Microbiol*. 1998;36:2375–6.
- Martínez-Carrasco C, Berriatua E, Garijo M, Martínez J, Alonso FD, de Ybáñez RR. Epidemiological study of non-systemic parasitism in dogs in southeast Mediterranean Spain assessed by coprological and post-mortem examination. *Zoonoses Public Health*. 2007;54:195–203.
- Martínez-Moreno FJ, Hernández S, López-Cobos E, Becerra C, Acosta I, Martínez-Moreno A. Estimation of canine intestinal parasites in Córdoba (Spain) and their risk to public health. *Vet Parasitol*. 2007;143:7–13.
- Miró G, Mateo M, Montoya A, Vela E, Calonge R. Survey of intestinal parasites in stray dogs in the Madrid area and comparison of the efficacy of three anthelmintics in naturally infected dogs. *Parasitol Res*. 2007;100:317–20.
- Miró G, Montoya A, Jiménez S, Frisuelos C, Mateo M, Fuentes I. Prevalence of antibodies to *Toxoplasma gondii* and intestinal parasites in stray, farm and household cats in Spain. *Vet Parasitol*. 2004;126:249–55.
- Pardo J, Muro A, Galindo I, Cordero M, Carpio A, Siles-Lucas M. Hidatidosis en la provincia de Salamanca: ¿Debemos bajar la guardia? *Enferm Infecc Microbiol Clin*. 2005;23:266–9.
- Rojo-Vazquez FA, Pardo-Lledias J, Francos-von Hunefeld M, Cordero-Sanchez M, Alamo-Sanz R, Hernandez-Gonzalez A, et al. Cystic echinococcosis in Spain: current situation and relevance for other endemic areas in Europe. *PLoS Negl Trop Dis*. 2011;5:e893.
- Lopez-Bernus A, Belhassen-García M, Carpio-Perez A, Perez del Villar L, Romero-Alegria A, Velasco-Tirado V, et al. Is cystic echinococcosis re-emerging in western Spain? *Epidemiol Infect*. 2015;8:1–7.
- Armiñanzas C, Gutiérrez-Cuadra M, Fariñas MC. Hidatidosis: aspectos epidemiológicos, clínicos, diagnósticos y terapéuticos. *Rev Esp Quimioter*. 2015;28:116–24.
- Asencio MA, Herraes O, Tenias JM, Garduño E, Huertas M, Carranza R, et al. Seroprevalence survey of zoonoses in Extremadura, southwestern Spain, 2002–2003. *Jpn J Infect Dis*. 2015;68:106–12.
- Calvo AM, Cires JM, Montón S, Sarasibar H, Lasanta P, Artázcoz FJ. Tumoración inguinal: una forma de presentación de la hidatidosis ósea. A propósito de un caso. *An Sist Sanit Navar*. 2007;30:475–9.
- Hueto Pérez de Heredia J, Pérez de las Casas M, Domínguez del Valle J, Vila Mayo E, Urquía Braña M, Gómez Dorronsoro M. Hidatidosis torácica. Nuestra experiencia en los últimos quince años. *Rev Clin Esp*. 1999;199:13–7.
- Carabin H, Balsara-Rodríguez FJ, Rebollar-Sáenz J, Benner CT, Benito A, Fernández-Crespo JC, et al. Cystic echinococcosis in the Province of Álava, North Spain: the monetary burden of a disease no longer under surveillance. *PLoS Negl Trop Dis*. 2014;8:e3069.
- Echenique-Elizondo M, Amondarain Arratibel JA. Hydatid disease of the pancreas. *JOP*. 2004;5:51–2.
- Echenique Elizondo MM, Amondarain Arratibel JA. Muscular hydatid disease. *J Am Coll Surg*. 2003;197:162.
- Amado-Diago CA, Gutiérrez-Cuadra M, Armiñanzas C, Arnaiz de las Revillas F, Gómez-Fleitas M, Fariñas MC. Echinococcosis: epidemiología, clínica y resultados en un pano-rámica de 15 años. *Rev Clin Esp*. 2015;215:380–4.
- Moreno J, Téllez CJ, Pardo FJ. Casos de hidatidosis en el Departamento de Salud 2 de la Comunidad Valenciana. *Rev Esp Quimioter*. 2009;22:62–7.
- Ramía JM, de-la-Plaza R, Quiñones J, Adel F, Ramiro C, García-Parreño J. Frank intrabiliary rupture in liver hydatidosis located in the hilar plate: a surgical challenge. *Dig Surg*. 2013;30:439–43.
- Suárez Grau JM, Gómez Bravo MA, Alamo Martínez JM, Rubio Chaves C, Marín Gómez LM, Suárez Artacho G, et al. Giant hydatid cyst involving the right hepatic lobe. *Rev Esp Enferm Dig*. 2009;101:133–5.
- Hernando E, García Calleja JL, Córdoba E, Lahuerta L, del Río F, Ferreira V. Hidatidosis hepática. Revisión de una serie de 677 pacientes tratados quirúrgicamente. *Gastroenterol Hepatol*. 1996;19:140–5.
- Ramía JM, Poves I, Castellón C, Díez-Valladares L, Loinaz C, Serrablo A, et al. Radical laparoscopic treatment for liver hydatidosis. *World J Surg*. 2013;37:2387–92.
- Guarner-Argente C, Gomez-Oliva C, Poca M, Sainz S, Marinello FG, Villanueva C, et al. Cholangitis caused by biliary hydatidosis. *Gastrointest Endosc*. 2010;72:1264–5.
- Molina Infante J, Fernández Bermejo M, Martín Noguero E, Pérez Gallardo B. Biliary hydatidosis. *Rev Esp Enferm Dig*. 2009;101:136–8.
- García Ruiz de Gordejuela A, Lladó L, Torras J, Ramos E, Rafecas A. Hidatidosis hepática gigante con fistula quistopleural e hidatosis pleural. *Cir Esp*. 2007;82:177–9.
- Ramos G, Orduña A, García-Yuste M. Hydatid cyst of the lung: diagnosis and treatment. *World J Surg*. 2001;25:46–57.
- Burgos R, Varela A, Castedo E, Roda J, Montero CG, Serrano S, et al. Pulmonary hydatidosis: surgical treatment and follow-up of 240 cases. *Eur J Cardiothorac Surg*. 1999;16:628–34.
- Arroyo Pérez R, Echarte Pazos JL, Aguirre Tejedó A, Pijuan Andújar L, Lerma Chippirraz E, Terrer Galera E. Derrame pleural derecho, compatible con empiema, en mujer de 82 años. *Rev Clin Esp*. 2008;208:46–8.
- Ramía-Angel JM, Gasz A, de la Plaza-Llamas R, Quinones-Sampedro J, Sancho E, García Parreno J. Hidatidosis of the spleen. *Pol Przegl Chir*. 2011;83:271–5.
- Prieto M, Marquina T, Mifsut P, Moreno T. Hidatidosis esplénica: 5 casos de esta rara localización. *Enferm Infecc Microbiol Clin*. 2011;29:634–5.
- Fernández-Ruiz M, Guerra-Vales JM, Enguita-Valls AB, Vila-Santos J, García-Borda FJ, Morales-Gutiérrez C. Splenic hydatid cyst, a rare location of extrahepatic echino-coccosis: report of six cases. *Eur J Intern Med*. 2008;19:e51–3.
- Angulo JC, Sanchez-Chapado M, Diego A, Escribano J, Tamayo JC, Martín L. Renal echinococcosis: clinical study of 34 cases. *J Urol*. 1997;157:787–94.
- Alonso García ME, Suárez Mansilla P, Mora Cepeda P, Bayón Álvarez E, Alvarez Colomo C, González Martín JI. Ovarian hydatid disease. *Arch Gynecol Obstet*. 2014;289:1047–51.
- Ruiz-Rabelo JF, Gomez-Alvarez M, Sanchez-Rodriguez J, Rufian Peña S. Complications of extrahepatic echinococcosis: fistulization of an adrenal hydatid cyst into the intestine. *World J Gastroenterol*. 2008;14:1467–9.
- Senarriaga Ruiz de la Ila N, Loizaga Iriarte A, Iriarte Soldevilla I, Lacasa Viscasilas I, Unda Urzaiz M. Hidatidosis pelviana como ejemplo de masas pelvianas de etiología dudosa. *Actas Urol Esp*. 2009;33:1129–32.
- Colomina Pascual M, Giner Galvan V, Domenech Iglesias A, Planelles Asensio M. Hidatidosis abdominal diseminada. *Med Clin (Barc)*. 2011;136:182.
- Cecconi A, Maroto L, Vilacosta I, Luaces M, Ortega L, Escribano N, et al. Acute pericarditis secondary to hydatid cyst rupture: diagnosis by multimodality imaging. *Circulation*. 2013;128:2073–4.
- Díaz-Menéndez M, Pérez-Molina JA, Norman FF, Pérez-Ayala A, Monge-Maillou B, Fuentes PZ, et al. Management and outcome of cardiac and endovascular cystic echinococcosis. *PLoS Negl Trop Dis*. 2012;6:e1437.
- Perez-David E, Fernandez MA, Fernández-Avilés F. Isolated cardiac hydatidosis. *Eur Heart J*. 2007;28:2829.
- Tejada JG, Saavedra J, Molina L, Forteza A, Gomez C. Hydatid disease of the interventricular septum causing pericardial effusion. *Ann Thorac Surg*. 2001;71:2034–5.
- Seijas R, Catalán-Larracochea JM, Ares-Rodríguez O, Joshi N, de la Fuente JP, Pérez-Domínguez M. Primary hydatid cyst of skeletal muscle affecting the knee: a case report. *Arch Orthop Trauma Surg*. 2009;129:39–41.
- García-Alvarez F, Torcal J, Salinas JC, Navarro A, García-Alvarez I, Navarro-Zorraquino M, et al. Musculoskeletal hydatid disease: a report of 13 cases. *Acta Orthop Scand*. 2002;73:227–31.
- Belhassen-García M, Carpio-Perez A, Blanco JF, Velasco-Tirado V, Pardo-Lledías J. Recurrent spinal echinococcosis. *Int J Infect Dis*. 2011;15:e435–6.
- Herrera A, Martínez AA, Rodríguez J. Spinal hydatidosis. *Spine (Phila Pa 1976)*. 2005;30:2439–44.

50. Gómez-Senent S, Manceñido-Marcos N, Erdozain-Soa JC, Segura-Cabral J. Hidatidosis cutánea. *Med Clin (Barc)*. 2006;127:480.
51. Gironé G, Mateo C, Gaya V, Usó J, Mínguez S, Roca B, et al. Admissions for imported and non-imported parasitic diseases at a General Hospital in Spain: a retrospective analysis. *Travel Med Infect Dis*. 2015;13:322–8.
52. Arcos Machancoses JV, Parra Llorca A, Martín Benlloch J, Ortí Martín A. Waterlily sign. *BMJ Case Rep*. 2013, <http://dx.doi.org/10.1136/bcr-2012-008243>.
53. Herráiz Gastesi G, Bonnet Carrón C, Madariaga Ruiz B, Freile García E, Gracia Casanova M. Hidatidosis pulmonar bilateral en paciente inmigrante. *An Pediatr (Barc)*. 2012;76:361–2.
54. Jiménez-Mejías ME, Alarcón-Cruz JC, Márquez-Rivas FJ, Palomino-Nicás J, Montero JM, Pachón J. Orbital hydatid cyst: treatment and prevention of recurrences with albendazole plus praziquantel. *J Infect*. 2000;41:105–7.
55. Sanjuán Rodríguez S, Morán Penco JM, Rincón Rodera P, González Díez G. Abdomen agudo secundario a quiste hidatídico pancreático. *Cir Pediatr*. 2005;18:36–8.
56. Minciullo PL, Cascio A, David A, Pernice LM, Calapai G, Gangemi S. Anaphylaxis caused by helminths: review of the literature. *Eur Rev Med Pharmacol Sci*. 2012;16:1513–8.
57. Novo Alonso C, Garín Ferreira J, Rodríguez Fernández A, Hernández Antequera E, Zapata Blanco JA, Vaquero Gajate G. Hidatidosis alveolar humana. *Rev Esp Enferm Dig*. 1993;84:127–9.
58. Arcechea Irigoyen MA, Córdoba Iturriaga A, Tuñón Álvarez MA, Gómez Dorronsoro ML, Martínez-Peñuela Virseda JM. Equinococosis alveolar humana. Presentación de un caso. *Rev Esp Patol*. 2008;41:203–6.
59. Iglesias Oliva L, Pérez Rojí G, Pérez-Llantada Amunarriz E, González Mandly A, García-Castrillo Riesco L. Múltiples calcificaciones musculares en un varón de 68 años de edad. *Rev Clin Esp*. 2002;202:669–71.
60. Corral I, Quereda C, Moreno A, López-Vélez R, Martínez-San-Millán J, Guerrero A, et al. Intramedullary cysticercosis cured with drug treatment. A case report. *Spine (Phila Pa 1976)*. 1996;21:2284–7.
61. Fernández-Gómez JM, García-Garmendia JL, López-Domínguez JM, Casado-Chocán JL. Neurocisticercosis y crisis convulsivas. *Rev Neurol*. 1998;26:1072–3.
62. Font Puig C, Ruiz Postigo JA, Muñoz Batet C, Pardós Arnal F, Corachan Cuyás M. Neurocisticercosis en España. A propósito de 4 casos observados en pacientes inmigrados de países endémicos. *An Med Intern*. 1999;16:89–91.
63. Castellanos F, Montes I, Porras LF, Peragallo E, Ampuero J, Rueda R. Quistes subaracnoideos gigantes por neurocisticercosis: a propósito de dos casos observados en un área rural de Extremadura. *Rev Neurol*. 2000;30:433–5.
64. Terraza S, Pujol T, Gascón J, Corachán M. Neurocisticercosis: ¿una enfermedad importada. *Med Clin (Barc)*. 2001;116:261–3.
65. Rodríguez-Sánchez G, Castellanos-Pinedo F, Giménez-Pando J, Adeva-Bartolomé MT, Zancada-Díaz F. Hidrocefalia y quiste subaracnoideo por neurocisticercosis. Un nuevo caso en una zona rural de Extremadura. *Rev Neurol*. 2002;34:348–51.
66. Roca C, Gascón J, Font B, Pujol T, Valls ME, Corachán M. Neurocisticercosis and population movements: analysis of 23 imported cases in Spain. *Eur J Clin Microbiol Infect Dis*. 2003;22:382–4.
67. Cañizares R, Roig P, Esparica A, Zorraquino A, Ortiz de la Tabla V, Merino J. Crisis convulsiva en varón joven. *Rev Clin Esp*. 2003;203:601–3.
68. Ortega-Herrera R, Fernández-Segura ME, Gómez de Travecedo Y, Calvo I. Inmigrante ecuatoriana con cefalea. *Enferm Infecc Microbiol Clin*. 2004;22:248–9.
69. Llopart Pou JA, Gené A, Ayestarán JI, Saus C. Neurocisticercosis presenting as sudden death. *Acta Neurochir (Wien)*. 2005;147:785–6.
70. Esquivel A, Díaz-Otero F, Giménez-Roldán S. Growing frequency of neurocisticercosis in Madrid (Spain). *Neurología*. 2005;20:116–20.
71. Jiménez-Caballero PE, Mollejo Villanueva M, Marsal Alonso C, Álvarez Tejerina A. Síndrome de Bruns: descripción de un caso de neurocisticercosis con estudio anatomopatológico. *Neurología*. 2005;2:86–9.
72. Barra Valencia V, Moreno Elola-Olaso A, Fundora Suárez Y, Meneu Díaz JC, Jiménez de los Galanes SF, Pérez Saborido B, et al. Second case of neurocisticercosis in a patient with liver transplantation (first case in Spain): a case report. *Transplant Proc*. 2007;39:2454–7.
73. Ramos JM, Masia M, Padilla S, Bernal E, Martín-Hidalgo A, Gutiérrez F. Fatal infection due to larval cysts of cestodes (neurocisticercosis and hydatid disease) in human immunodeficiency virus (HIV) infected patients in Spain: report of two cases. *Scand J Infect Dis*. 2007;39:719–23.
74. Guerra del Barrio E, López Roger R. Lesión quística intracerebral en un paciente inmigrante. *Rev Clin Esp*. 2007;207:301–2.
75. Sierra Bergua B, Navarro Calzada J, Sanjoaquín Conde I, Santos Lasaosa S. Convulsión generalizada tónico-clónica en inmigrante. *Neurología*. 2007;22:253–5.
76. Más-Sesé G, Vives-Piñera I, Fernández-Barreiro A, Martínez-Lage JF, Martínez-Salcedo E, Alarcón-Martínez H, et al. Estudio descriptivo de neurocisticercosis en un hospital terciario. *Rev Neurol*. 2008;46:194–6.
77. Ruiz S, García-Vázquez E, Picazo R, Hernández A, Herrero JA, Gómez J. La neurocisticercosis en Murcia. *Rev Clin Esp*. 2011;211:133–8.
78. Aguilar-Amat MJ, Martínez-Sánchez P, Medina-Baez J, Díez-Tejedor E. Síndrome de Bruns causado por neurocisticercosis intraventricular. *Med Clin (Barc)*. 2011;137:43–6.
79. De Anta Tejado, Pozo KT, Palomino CB, de Dios de Vega JL. Psychiatric manifestations of neurocisticercosis in paediatric patients. *BMJ Case Rep*. 2012, <http://dx.doi.org/10.1136/bcr.03.2010.2840>.
80. Friero-Dantas C, Serramito-García R, Reyes-Santías RM, Rico-Cotelo M, Allut AG, Gelabert-González M. Neurocisticercosis pediátrica: a propósito de dos casos. *Rev Neurol*. 2013;56:86–90.
81. Giménez-Roldán S, Díaz F, Esquivel A. Neurocisticercosis e inmigración. *Neurología*. 2003;18:385–8.
82. Alvarez-Rodríguez E, Torres-Gárate R, Cabello J, Lozano Tonkin C. Neurocisticercosis en España. *Rev Clin Esp*. 2005;205:518.
83. Jiménez-Jiménez FJ, Molina-Arjona JA, Roldán-Montaud A, Agullá A, Santos J, Fernández-Ballesteros A. Blepharospasm associated with neurocisticercosis. *Acta Neurol (Napoli)*. 1992;14:56–9.
84. Accomando S, Caserta M, Trizzino A, Amato GM. Two strange cases of hyper-eosinophilia and child's relapsing angio-oedema. *Pediatr Med Chir*. 2003;25:367–9.
85. Singh RB, Pavithran NM, Bakshi N. Intraperitoneal rupture of cysticercosal cyst mimicking appendicular perforation. *Trop Doct*. 2006;36:180–1.
86. Minciullo PL, Spagnolo EV, Cascio A, Cardia G, Gangemi S. Fatal anaphylactic shock and *Taenia solium* infestation: a possible link? *Ann Allergy Asthma Immunol*. 2009;103:449–50.
87. Colomina J, Villar J, Esteban G. Parasitación asintomática por *Diphyllobothrium latum* en un niño español de 3 años. *Med Clin (Barc)*. 2002;118:279.
88. Gil-Setas A, Mazón A, Pascual P, Sagua H. Helminthiasis poco frecuente en nuestro medio en un varón de 71 años. *Enferm Infecc Microbiol Clin*. 2004;22:553–4.
89. Marcos Sánchez F, Albo Castaño I, Viana Alonso A, Caballero Sánchez-Robles L. Dolor abdominal en una mujer guineana de 21 años. *Rev Clin Esp*. 2005;205:459–60.
90. Pastor-Valle J, González LM, Martín-Clemente JP, Merino FJ, Gottstein B, Gárate T. Molecular diagnosis of diphyllobothriasis in Spain, most presumably acquired via imported fish, or sojourn abroad. *New Microbes New Infect*. 2014;2:1–6.
91. Esteban JG, Muñoz-Antoli C, Borrás M, Colomina J, Toledo R. Human infection by a 'fish tapeworm', *Diphyllobothrium latum*, in a non-endemic country. *Infection*. 2014;42:191–4.
92. Lo Presti A, Aguirre DT, de Andrés P, Daoud L, Fortes J, Muñoz J. Cerebral sparganosis: case report and review of the European cases. *Acta Neurochir (Wien)*. 2015;157:1339–43.
93. Corachan M, Ruiz L, Valls ME, Gascon J. Schistosomiasis and the Dogon country (Mali). *Am J Trop Med Hyg*. 1992;47:6–9.
94. Corachán M, Almeda J, Vinuesa T, Valls ME, Mejías T, Jou P, et al. Esquistosomiasis importada por viajeros españoles: estudio clínico-epidemiológico de 80 casos. *Med Clin (Barc)*. 1997;108:721–5.
95. Vilana R, Corachán M, Gascón J, Valls E, Bru C. Schistosomiasis of the male genital tract: transrectal sonographic findings. *J Urol*. 1997;158:1491–3.
96. Elcuaz R, Armas M, Ramírez M, Noguera FJ, Bolaños M, Quiñones I, et al. Brote de esquistosomiasis en un grupo de viajeros procedentes de Burkina Faso. *Enferm Infecc Microbiol Clin*. 1998;16:367–9.
97. Bou A, Gascón J, Valls ME, Corachán M. Fiebre de Katayama en turistas españoles: análisis de 25 casos. *Med Clin (Barc)*. 2001;116:220–2.
98. Roca C, Balanzó X, Gascón J, Fernández-Roure JL, Vinuesa T, Valls ME, et al. Comparative, clinico-epidemiologic study of *Schistosoma mansoni* infections in travellers and immigrants in Spain. *Eur J Clin Microbiol Infect Dis*. 2002;21:219–23.
99. Santos Y, Balliu E, Villán D. Hematuria en un varón de 34 años. *Med Clin (Barc)*. 2004;123:312–6.
100. Alonso D, Muñoz J, Gascón J, Valls ME, Corachan M. Failure of standard treatment with praziquantel in two returned travelers with *Schistosoma haematobium* infection. *Am J Trop Med Hyg*. 2006;74:342–4.
101. de Górgolas M, Casado V, Renedo G, Alen JF, Fernández Guerrero ML. Nodular lung schistosomiasis lesions after chemotherapy for dysgerminoma. *Am J Trop Med Hyg*. 2009;81:424–7.
102. Zamarrón Fuertes P, Pérez-Ayala A, Pérez Molina JA, Norman FF, Monge-Maíllo B, Navarro M, et al. Clinical and epidemiological characteristics of imported infectious diseases in Spanish travelers. *J Travel Med*. 2010;17:303–9.
103. Rivas P, Aguilar-Durán S, Lago M. Lung nodules, fever, and eosinophilia in a traveler returning from Madagascar. *Am J Trop Med Hyg*. 2012;86:2–3.
104. Perez-Arellano JL, Hernandez-Cabrera M, Malet-Pintos-Fonseca A, Carranza-Rodríguez C, Martín-Sánchez AM. Katayama's syndrome related to *Schistosoma intercalatum* in two travellers returning from Mali. *Am J Infect Dis*. 2012;8:128–31.
105. Muñoz J, Aldasoro E, Pinazo MJ, Arguis P, Gascon J. Pulmonary infiltrates and eosinophilia in a 25-year-old traveler. *PLoS Negl Trop Dis*. 2013;7:e2201.
106. Martínez-Calle N, Pascual I, Rubio M, Carías R, del Pozo JL, Yuste JR. Asymptomatic *Schistosoma haematobium* infection in a traveler with negative urine microscopy and late seroconversion presumably linked to artemisinin. *J Travel Med*. 2013;20:326–8.
107. Corachan M, Valls ME, Gascon J, Almeda J, Vilana R. Hematospermia: a new etiology of clinical interest. *Am J Trop Med Hyg*. 1994;50:580–4.
108. Chaves F, Dronda F, González-López A, López-Cabañas C, de los Ríos M. Prurito y eosinofilia en un nativo de Guinea Ecuatorial. *Enferm Infecc Microbiol Clin*. 1992;10:547–8.
109. Balasch J, Martínez-Román S, Creus M, Campo E, Fortuny A, Vanrell JA. Schistosomiasis: an unusual cause of tubal infertility. *Hum Reprod*. 1995;10:1725–7.
110. Gairi-Burgués Mf MA, Bosch Muñoz J, Llusá Parramon A, Gomá AR. *Schistosoma haematobium* como causa de hematuria. *An Esp Pediatr*. 2002;56:368–9.
111. Rotger M, Serra T, de Cárdenas MC, Morey A, Vicente MA. Increasing incidence of imported schistosomiasis in Mallorca, Spain. *Eur J Clin Microbiol Infect Dis*. 2004;23:855–6.

112. López-Calleja AI, Torres L, Revillo MJ, Clavel A, Arazo P. Dolor cólico hipogástrico y eosinofilia en un paciente de Gambia. *Enferm Infecc Microbiol Clin.* 2003;21:315–6.
113. Pardo J, Carranza C, Muro A, Angel-Moreno A, Martín AM, Martín T, et al. Helminth-related eosinophilia in African immigrants, Gran Canaria. *Emerg Infect Dis.* 2006;12:1587–9.
114. Donate Moreno MJ, Pastor Navarro H, Giménez Bachs JM, Carrión López P, Segura Martín M, Salinas Sánchez AS, et al. Esquistosomiasis vesical, aportación de un caso y revisión de la literatura española. *Actas Urol Esp.* 2006;30:714–9.
115. Campo-Esquisabel AB, Ledesma-Castaño F, Salesa-Gutiérrez de Rozas R. Eosinofilia en inmigrante africano asintomático. *Enferm Infecc Microbiol Clin.* 2007;25:63.
116. Tarabini-Castellani P, González-Chinchón G, Aldamiz-Echebarría M, Portu-Zapirain J, Apraiz-Garmendia L, Alvarez de Arcaya A. Neuroesquistosomiasis: un reto para el diagnóstico. *Rev Neurol.* 2007;44:154–6.
117. López López AI, Cao Avellaneda E, Prieto González A, Ferri Níguez B, Maluff Torres A, Pérez Albacete M. Esquistosomiasis. Una parasitosis urinaria cada vez más frecuente. *Actas Urol Esp.* 2007;31:915–8.
118. Carranza-Rodríguez C, Pardo-Lledias J, Muro-Alvarez A, Pérez-Arellano JL. Cryptic parasite infection in recent West African immigrants with relative eosinophilia. *Clin Infect Dis.* 2008;46:e48–50.
119. Rascarachi G, Linares Torres P, Arias Rodríguez L, Pérez Andrada S, Sierra Ausin M, Díez Rodríguez R, et al. Esquistosomiasis intestinal. *Gastroenterol Hepatol.* 2009;32:131–2.
120. Llenas-García J, Guerra-Vales JM, Alcalá-Galiano A, Domínguez C, Pérez-Núñez A, Lizasoain M, et al. Cerebral neuroschistosomiasis: a rare clinical presentation and review of the literature. *BMJ Case Rep.* 2009, <http://dx.doi.org/10.1136/bcr.04.2009.1787>.
121. Landeyro J, Elguezabal A, Gené M, García-Fontgüell J, Mayayo E. Rectal inflammatory polyp in a child: an unusual case of polyparasitism. *J Trop Pediatr.* 2010;56:270–1.
122. Rodríguez-Guardado A, Miquel R, Pérez F, Fresno M, Corachán M. Colonic polyposis due to *Schistosoma intercalatum*. *Trans R Soc Trop Med Hyg.* 2010;104:443–5.
123. Carrión López P, Pastor Navarro H, Martínez Ruiz J, Martínez Sanchiz C, Donate Moreno MJ, Segura Martín M, et al. Cistoscopia en la bilharziasis vesical. *Arch Esp Urol.* 2010;63:85–6.
124. Alvarez Maestro M, Rios Gonzalez E, Dominguez Garcia P, Vallejo Herrador J, Díez Rodríguez J, Martínez-Piñeiro L. Esquistosomiasis vesical: a propósito de un caso y revisión de la literatura. *Arch Esp Urol.* 2010;63:554–8.
125. Gran F, Albert DC, Moreno A. Schistosomiasis and tropical endomyocardial fibrosis with pulmonary hypertension. *Rev Esp Cardiol.* 2011;64:713.
126. López de Cenarruzabeitia I, Landolfi S, Armengol Carrasco M. Intestinal schistosomiasis as unusual aetiology for acute appendicitis, nowadays a rising disease in Western countries. *Case Rep Infect Dis.* 2012, <http://dx.doi.org/10.1155/2012/896820>.
127. García Pérez N, Arístegui Fernández J, Garrote Llanos E, Sanchez Lafuente J. Esquistosomiasis vesical. Un nuevo caso importado. *An Pediatr (Barc).* 2014;80:e88–9.
128. Barrio Muñoz M, García Rojo D, González Sala JL, Prats López J. Esquistosomiasis urinaria: descripción de 8 casos. *Med Clin (Barc).* 2013;140:476–7.
129. Salvador F, Molina I, Sulleiro E, Burgos J, Curran A, van den Eynde E, et al. Tropical diseases screening in immigrant patients with human immunodeficiency virus infection in Spain. *Am J Trop Med Hyg.* 2013;88:1196–202.
130. Cobo F, Salas-Coronas J, Cabezas-Fernández MT, Vázquez-Villegas J, Cabeza-Barrera MI, Soriano-Pérez MJ. Infectious diseases in immigrant population related to the time of residence in Spain. *J Immigr Minor Health.* 2014 [Epub ahead of print].
131. Belhassen-García M, Pérez del Villar L, Pardo-Lledias J, Gutiérrez Zufiaurre MN, Velasco-Tirado V, Cordero-Sánchez M, et al. Imported transmissible diseases in minors coming to Spain from low-income areas. *Clin Microbiol Infect.* 2015;21, 370.e5–370.e8.
132. Monge-Maillo B, López-Vélez R, Norman F, Ferrere-González F, Martínez-Pérez Á, Pérez-Molina JA. Screening of imported infectious diseases among asymptomatic sub-Saharan African and Latin American immigrants: a public health challenge. *Am J Trop Med Hyg.* 2015;92:848–56.
133. Salas-Coronas J, Cabezas-Fernández MT, Vázquez-Villegas J, Soriano-Pérez MJ, Lozano-Serrano AB, Pérez-Camacho I, et al. Evaluation of eosinophilia in immigrants in Southern Spain using tailored screening and treatment protocols: a prospective study. *Travel Med Infect Dis.* 2015;13:315–21.
134. Carod Artal FJ. Cerebral and spinal schistosomiasis. *Curr Neurol Neurosci Rep.* 2012;12:666–74.
135. Salas-Coronas J, Vázquez-Villegas J, Villarejo-Ordóñez A, Sánchez-Sánchez JC, Espada-Chavarría J, Soriano-Pérez MJ, et al. Hallazgos radiológicos en pacientes con esquistosomiasis importada. *Enferm Infecc Microbiol Clin.* 2013;31:205–9.
136. Pardo J, Pérez-Arellano JL, López-Vélez R, Carranza C, Cordero M, Muro A. Application of an ELISA test using *Schistosoma bovis* adult worm antigens in travellers and immigrants from a schistosomiasis endemic area and its correlation with clinical findings. *Scand J Infect Dis.* 2007;39:435–40.
137. Pardo J, Carranza C, Turrientes MC, Pérez Arellano JL, López Vélez R, Ramajo V, et al. Utility of *Schistosoma bovis* adult worm antigens for diagnosis of human schistosomiasis by enzyme-linked immunosorbent assay and electroimmunotransfer blot techniques. *Clin Diagn Lab Immunol.* 2004;11:1165–70.
138. Sandoval N, Siles-Lucas M, Pérez-Arellano JL, Carranza C, Puente S, López-Abán J, et al. A new PCR-based approach for the specific amplification of DNA from different *Schistosoma* species applicable to human urine samples. *Parasitology.* 2006;133:581–7.
139. Fernández-Soto P, Velasco Tirado V, Carranza Rodríguez C, Pérez-Arellano JL, Muro A. Long-term frozen storage of urine samples: a trouble to get PCR results in *Schistosoma* spp. DNA detection? *PLOS ONE.* 2013;8:e61703.
140. Fernández-Soto P, Gandasegui Arahuetes J, Sánchez Hernández A, López Abán J, Vicente Santiago B, Muro A. A loop-mediated isothermal amplification (LAMP) assay for early detection of *Schistosoma mansoni* in stool samples: a diagnostic approach in a murine model. *PLoS Negl Trop Dis.* 2014;8:e3126.
141. Pulpeiro JR, Armesto V, Varela J, Corredoira J. Fascioliasis: findings in 15 patients. *Br J Radiol.* 1991;64:798–801.
142. López Rosés L, Alonso D, Iñiguez F, Mateos A, Bal M, Agüero J. Hepatic fascioliasis of long-term evolution: diagnosis by ERCP. *Am J Gastroenterol.* 1993;88:2118–9.
143. Arjona R, Riancho JA, Aguado JM, Salesa R, González-Macias J. Fascioliasis in developed countries: a review of classic and aberrant forms of the disease. *Medicine (Baltimore).* 1995;74:13–23.
144. Gómez Cerezo J, Ríos Blanco JJ, de Guevara CL, Barbado Hernández FJ, Vázquez Rodríguez JJ. Biliary aspiration after administration of intravenous cholecystokinin for the diagnosis of hepatobiliary fascioliasis. *Clin Infect Dis.* 1998;26:1009–10.
145. Segado Soriano A, López González-Cobos C, Muiño Míguez A, Pastor Gómez-Cornejo L, Merino Moreno J. Distomatosis por fasciola hepática: a propósito de tres casos en la Comunidad de Madrid. *An Med Interna.* 1998;15:427–9.
146. López-Vélez R, Domínguez-Castellano A, Garrón C. Successful treatment of human fascioliasis with triclabendazole. *Eur J Clin Microbiol Infect Dis.* 1999;18:525–6.
147. Perez C, Vives R, Montes M, Ostiz S. Recurrent eosinophilic panniculitis associated with *Fasciola hepatica* infection. *J Am Acad Dermatol.* 2000;42 5 Pt 2:900–2.
148. Núñez Fernández MJ, Anibarro García L, Piñeiro Gómez-Durán L. Fascioliasis en el sur de Galicia: presentación de dos casos. *An Med Interna.* 2001;18:280–1.
149. Cosme A, Ojeda E, Cilla G, Torrado J, Alzate L, Beristain X, et al. Fascioliasis hepatobiliar. Estudio de una serie de 37 pacientes. *Gastroenterol Hepatol.* 2001;24:375–80.
150. Cilla G, Serrano-Bengochea E, Cosme A, Abadía L, Pérez-Trallero E. Decrease in human fascioliasis in Gipuzkoa (Spain). *Eur J Epidemiol.* 2001;17:819–21.
151. González Llorente J, Herrero Domingo A, Carrero González P. Subcapsular abscess: an unusual CT finding in hepatic fascioliasis. *Am J Roentgenol.* 2002;178:514–5.
152. Cosme A, Ojeda E, Poch M, Bujanda L, Castiella A, Fernández J. Sonographic findings of hepatic lesions in human fascioliasis. *J Clin Ultrasound.* 2003;31:358–63.
153. Cirera I, Arguis P, Miquel R. Mujer de 66 años con astenia y eosinofilia. *Med Clin (Barc).* 2004;123:31–7.
154. Echenique-Elizondo M, Amondarain J, Lirón de Robles C. Fascioliasis: an exceptional cause of acute pancreatitis. *JOP.* 2005;6:36–9.
155. Añaños G, Trilla A, Graus F, Mas J, Corachán M, Soriano E. Paragonimiasis y tuberculosis pulmonar. *Med Clin (Barc).* 1992;98:257–9.
156. Requena A, Domínguez MA, Santin M. An African-born man with chronic recurrent hemoptysis. *Clin Infect Dis.* 2008;46:1253.
157. Gómez-Seco J, Rodríguez-Guzmán MJ, Rodríguez-Nieto MJ, Gómez-Escobar PF, Presa-Abos T, Fortes-Alen J. Paragonimiasis pulmonar. *Arch Bronconeumol.* 2011;47:610–2.
158. Esteban-Gutiérrez G, Rojo-Marcos G, Cuadros-González J, Bragado-Martínez L. Eosinofilia en un paciente de Tailandia-Laos. *Enferm Infecc Microbiol Clin.* 2011;29:629–30.
159. Cabeza-Barrera MI, Cabezas-Fernández MT, Vázquez-Villegas J, Salas-Corona J. Dolor abdominal en un paciente de origen magrebí. *Enferm Infecc Microbiol Clin.* 2007;25:213–4.
160. Zabala Martín-Gil I, Justel Pérez JP, Cuadros González J. Pseudoparasitismo por *Dicrocoelium dendriticum*. *Aten Primaria.* 2007;39:379–80.
161. Cabeza-Barrera I, Cabezas-Fernández T, Salas Coronas J, Vázquez Villegas J, Cobo F. *Dicrocoelium dendriticum*: an emerging spurious infection in a geographic area with a high level of immigration. *Ann Trop Med Parasitol.* 2011;105:403–6.
162. Jarabo MT, García-Morán NP, García-Morán JI. Prevalencia de parasitosis intestinales en una población escolar. *Enferm Infecc Microbiol Clin.* 1995;13:464–8.
163. Pérez Armengol C, Ariza Astolfí C, Ubeda Ontiveros JM, Guevara Benítez DC, de Rojas Alvarez M, Lozano Serrano C. Epidemiología del parasitismo intestinal infantil en el valle del Guadalquivir, España. *Rev Esp Salud Publica.* 1997;71:547–52.
164. Rustarazo S, Morales Suárez-Varela M, Antequera M, Esteban Sanchis JG. Enteroparasitosis en población escolar de Valencia. *Aten Primaria.* 2008;40:641–5.
165. González-Moreno O, Domingo L, Teixidor J, Gracenea M. Prevalence and associated factors of intestinal parasitisation: a cross-sectional study among outpatients with gastrointestinal symptoms in Catalonia, Spain. *Parasitol Res.* 2011;108:87–93.
166. Huerga H, Lopez-Velez R. Infectious diseases in sub-Saharan African immigrant children in Madrid, Spain. *Pediatr Infect Dis J.* 2002;21:830–4.
167. Martín Sánchez AM, Hernández García A, González Fernández M, Afonso Rodríguez O, Hernández Cabrera M, Pérez Arellano JL. Parasitosis intestinales en población inmigrante subsahariana asintomática. Gran Canaria 2000. *Rev Clin Esp.* 2004;204:14–7.

168. Olivares JL, Fernández R, Flea J, Rodríguez G, Clavel A. Serum mineral levels in children with intestinal parasitic infection. *Dig Dis.* 2003;21:258–61.
169. Cortizo-Vidal S, Rodríguez-Otero LM, Villamil-Cajoto I, Llovo-Taboada J. Eosinofilia en niña adoptada procedente de Etiopía. *Enf Infecc Microbiol Clin.* 2010;28:60–1.
170. Rodríguez-Ocaña E, Menéndez-Navarro A. Hookworm in rural Spain, 1910–1960: shifting paradigms around the Civil War. *J Epidemiol Commun Health.* 2009;63:670–4.
171. Rodríguez Feijoo A, Alonso MP, Rey A, Corredoira J, Sayagues A. Eosinofilia en un paciente de Cabo Verde. *Enferm Infecc Microbiol Clin.* 1992;10:621–2.
172. García-Parra CJ, Cano-Ballesteros JC, Ortega-Sánchez E, Martín-Rabadán P. Varón brasileño de 19 años con anemia ferropénica grave. *Enferm Infecc Microbiol Clin.* 2015;33:211–2.
173. Alonso-Sanz M, Chaves F, Drona F, Catalán S, González-López A. Parasitosis intestinales en la población reclusa en el área de Madrid (1991–1993). *Enferm Infecc Microbiol Clin.* 1995;13:90–5.
174. de la Cruz Alvarez J, Pineda Mariño JR, Sánchez Míguez JR, Clofent Vilaplana J, Domínguez Rodríguez F, Hermo Brión JA, et al. Ascariasis biliopancreática: una enfermedad infrecuente en nuestro medio. *Gastroenterol Hepatol.* 1996;19:210–2.
175. Cabezas MT, Giménez MJ, Molina MA, Cruz G, Avivar C, Ortiz B. Anemia ferropénica y síndrome constitucional. *Enferm Infecc Microbiol Clin.* 2000;18:239–40.
176. Tordera P, Navarro A, la Fuente M, Cano J, la Cruz J, Pemán J. Un nuevo caso de Lóeffer. *Rev Clin Esp.* 2002;202:667–8.
177. Nuño Mateo FJ, Noval Menéndez J, Fonseca Aizpuru EM, Otero Guerra L. Varón de 25 años de edad con vómito con gusano e hinchazón facial. *Rev Clin Esp.* 2003;203:305–6.
178. Roca C, Balanzó X, Sauca G, Fernández-Roure JL, Boixeda R, Ballester M. Uncinariasis importada por inmigrantes africanos: estudio de 285 casos. *Med Clin (Barc).* 2003;121:139–41.
179. Fonseca Aizpuru EM, García Piney E, Nuño Mateo FJ, Sánchez González V. Varón con anemia ferropénica severa por uncinariasis. *An Med Interna.* 2006;23:451–2.
180. Alarcón-Fernandez O, Baudet JS, Sanchez del Rio A. Iron-deficiency anemia caused by hookworm infestation. *Clin Gastroenterol Hepatol.* 2006;4:A32.
181. Zapata E, Zubiaurre L, Salvador P, Castiella A, Alzate LF, López P, et al. Chole-cystopancreatitis due to *Ascaris lumbricoides*. *Endoscopy.* 2007;39 Suppl 1:e10–1.
182. Egea Valenzuela J, Sánchez Torres A, Carballo Alvarez F. Suboclusión intestinal por *Ascaris*. *Rev Esp Enferm Dig.* 2008;100:579–80.
183. Julián-Gómez L, Barrio J, de la Serna C, Pérez-Miranda M, Gil-Simón P, Gómez de la Cuesta S, et al. Infección intestinal y biliar por *Ascaris lumbricoides* en endoscopia digestiva. *Rev Esp Enferm Dig.* 2009;101:427–9.
184. Iborra MA, Carrilero B, Segovia M. Anquilostomiasis: una causa para considerar en anemias ferropénicas de pacientes procedentes de zonas endémicas. *Enferm Infecc Microbiol Clin.* 2009;27:427.
185. Casado Maestre MD, Alamo Martínez JM, Segura Sampedro JJ, Gómez Bravo MÁ, Padillo Ruiz FJ, Durán Izquierdo E, et al. Ascariasis pancreática que simula un tumor pancreático. *Gastroenterol Hepatol.* 2011;34:464–7.
186. Cuenca-Gómez JA, Salas-Coronas J, Cabezas-Fernández MT, Vázquez-Villegas J, Soriano-Pérez MJ, Cobo F. Uncinariasis importada en Almería. *Enferm Infecc Microbiol Clin.* 2013;31:599–601.
187. Batista N, Dávila MF, Gijón H, Pérez MA. Estrongiloidiasis en un paciente con síndrome de inmunodeficiencia adquirida. *Enferm Infecc Microbiol Clin.* 1992;10:431–2.
188. Cremades Romero MJ, Martínez García MA, Menéndez Villanueva R, Cremades Romero ML, Pemán García JP. Infección por *Strongyloides stercoralis* en un paciente corticodependiente con obstrucción crónica de la vía aérea. *Arch Bronconeumol.* 1996;32:430–1.
189. Cremades Romero MJ, Igual Adell R, Ricart Olmos C, Estellés Piera F, Pastor-Guzmán A, Menéndez Villanueva R. Infección por *Strongyloides stercoralis* en la comarca de La Safor (Comunidad Valenciana). *Med Clin (Barc).* 1997;109:212–5.
190. Cremades Romero MJ, Pellicer Ciscar C, Menéndez Villanueva R, Ricart Olmos C, Pastor-Guzmán A, Estellés Piera F, et al. Infección por *Strongyloides stercoralis* en pacientes con patología bronquial obstructiva. *Arch Bronconeumol.* 1997;33:384–8.
191. Román Sánchez P, Guzman AP, Moreno Guillen S, Adell RI, Estruch AM, Gonzalo IN, et al. Endemic strongyloidiasis on the Spanish Mediterranean coast. *QJM.* 2001;94:357–63.
192. Pretel Serrano L, Page del Pozo MA, Ramos Guevara MR, Ramos Rincón JM, Martínez Toldos MC, Herrero Huerta F. Infestación por *Strongyloides stercoralis* en pacientes con enfermedad pulmonar obstructiva crónica en la Vega del Segura (Murcia). Presentación de 3 casos. *Rev Clin Esp.* 2001;201:109–10.
193. Díaz J, Igual R, Alonso MC, Moreno MJ. Estudio del parasitismo intestinal en inmigrantes de la comarca de La Safor (Comunidad Valenciana). *Med Clin (Barc).* 2002;119:36.
194. Román-Sánchez P, Pastor-Guzmán A, Moreno-Guillén S, Igual-Adell R, Suñer-Generoso S, Tornero-Estébanez C. High prevalence of *Strongyloides stercoralis* among farm workers on the Mediterranean coast of Spain: analysis of the predictive factors of infection in developed countries. *Am J Trop Med Hyg.* 2003;69:336–40.
195. Martínez-Vázquez C, González Mediero G, Núñez M, Pérez S, García-Fernández JM, Gimena B. *Strongyloides stercoralis* en el sur de Galicia. *An Med Interna.* 2003;20:477–9.
196. Oltra Alcaraz C, Adell RI, Sánchez PS, Blasco MJ, Sánchez OA, Auñón AS, et al. Characteristics and geographical profile of strongyloidiasis in healthcare area 11 of the Valencian community (Spain). *J Infect.* 2004;49:152–8.
197. Olmos JM, Gracia S, Villoria F, Sales R, González-Macias J. Disseminated strongyloidiasis in a patient with acquired immunodeficiency syndrome. *Eur J Intern Med.* 2004;15:529–30.
198. Mayayo E, Gomez-Aracil V, Azua-Blanco J, Azua-Romeo J, Capilla J, Mayayo R. *Strongyloides stercoralis* infection mimicking a malignant tumour in a non-immunocompromised patient. Diagnosis by bronchoalveolar cytology. *J Clin Pathol.* 2005;58:420–2.
199. Díaz Conradi A, Tello Pérez LM, Clapés Ferran R, Massaguer Cabrera J, Delgado Pérez R, Alayeto Ortega J. Masa abdominal y eosinofilia en un niño de origen etíope. *An Pediatr (Barc).* 2006;64:399–400.
200. Velasco A, Sánchez F, de la Coba C, Fradejas P, Prieto V, Alvarez A, et al. Síndrome de malabsorción intestinal producido por *Strongyloides stercoralis* asociado con el síndrome de Loeffler en una mujer de 28 años de edad. *Gastroenterol Hepatol.* 2006;29:341–4.
201. Beltrán Catalán S, Crespo Albiach JF, Morales García AI, Gavela Martínez E, Górriz Tuérol JL, Pallardó Mateu LM. Infección por *Strongyloides stercoralis* en pacientes trasplantados renales. *Nefrología.* 2009;29:482–5.
202. Rodríguez-Hernandez MJ, Ruiz-Perez-Pipaon M, Cañas E, Bernal C, Gavilan F. *Strongyloides stercoralis* hyperinfection transmitted by liver allograft in a transplant recipient. *Am J Transplant.* 2009;9:2637–40.
203. González A, Gallo M, Valls ME, Muñoz J, Puyol L, Pinazo MJ, et al. Clinical and epidemiological features of 33 imported *Strongyloides stercoralis* infections. *Trans R Soc Trop Med Hyg.* 2010;104:613–6.
204. Argelich R, Pujol MT, Álvarez-Martínez MJ. Varón de 69 años con meningitis recidivante. *Med Clin (Barc).* 2011;136:167–74.
205. Villena-Ruiz MÁ, Arboleda-Sánchez JA, Del Arco-Jiménez A, Fernández-Sánchez F. Neumonía grave en paciente con infección por virus de la inmunodeficiencia humana. *Enferm Infecc Microbiol Clin.* 2012;30:209–11.
206. de las Marinas MD, Martorell A, Felix R, Cerdá JC, García A, Navalpoto D. Strongyloidiasis: an emerging infectious disease that simulates allergic diseases. *J Investig Allergol Clin Immunol.* 2012;22:286–7.
207. Fernández Rodríguez C, Enríquez-Matas A, Sánchez Millán ML, Mielgo Ballesteros R, Jukic Beteta KD, Valdez Tejeda M, et al. *Strongyloides stercoralis* infection: a series of cases diagnosed in an allergy department in Spain. *J Investig Allergol Clin Immunol.* 2012;22:455–7.
208. Llenas-García J, Fiorante S, Salto E, Maseda D, Rodríguez V, Matarranz M, et al. Should we look for *Strongyloides stercoralis* in foreign-born HIV-infected persons? *J Immigr Minor Health.* 2013;15:796–802.
209. Valerio L, Roure S, Fernández-Rivas G, Basile L, Martínez-Cuevas O, Ballesteros ÁL, et al. *Strongyloides stercoralis*, the hidden worm. Epidemiological and clinical characteristics of 70 cases diagnosed in the North Metropolitan Area of Barcelona, Spain, 2003–2012. *Trans R Soc Trop Med Hyg.* 2013;107:465–70.
210. Izquierdo I, Briones J, Lluch R, Arqueros C, Martino R. Fatal strongyloides hyperinfection complicating a gram-negative sepsis after allogeneic stem cell transplantation: a case report and review of the literature. *Case Rep Hematol.* 2013;2013:860976.
211. Ramírez-Olivencia G, Espinosa MÁ, Martín AB, Núñez NI, de Las Parras ER, Núñez ML, et al. Imported strongyloidiasis in Spain. *Int J Infect Dis.* 2014;18:32–7.
212. Salvador F, Sulleiro E, Sánchez-Montalvá A, Saugar JM, Rodríguez E, Pahissa A, et al. Usefulness of *Strongyloides stercoralis* serology in the management of patients with eosinophilia. *Am J Trop Med Hyg.* 2014;90:830–4.
213. Cabezas-Fernández MT, Salas-Coronas J, Lozano-Serrano AB, Vazquez-Villegas J, Cabeza-Barrera MI, Cobo F. Strongyloidiasis in immigrants in Southern Spain. *Enferm Infecc Microbiol Clin.* 2015;33:37–9.
214. Ramos JM, León R, Andreu M, de las Parras ER, Rodríguez-Díaz JC, Esteban Á, et al. Serological study of *Trypanosoma cruzi*, *Strongyloides stercoralis*, HIV, human T cell lymphotropic virus (HTLV) and syphilis infections in asymptomatic Latin-American immigrants in Spain. *Trans R Soc Trop Med Hyg.* 2015;109:447–53.
215. Martínez-Pérez A, Lopez-Velez R. Is strongyloidiasis endemic in Spain? *PLoS Negl Trop Dis.* 2015;9:e0003482.
216. Drona F, Chaves F, Sanz A, Lopez-Velez R. Human intestinal capillaritis in an area of nonendemicity: case report and review. *Clin Infect Dis.* 1993;17:909–12.
217. Perera L, Pérez-Arellano JL, Cordero M, Simón F, Muro A. Utility of antibodies against a 22 kD molecule of *Dirofilaria immitis* in the diagnosis of human pulmonary dirofilariasis. *Trop Med Int Health.* 1998;3:151–5.
218. Muro A, Genchi C, Cordero M, Simón F. Human dirofilariasis in the European Union. *Parasitol Today.* 1999;15:386–9.
219. Montoya-Alonso JA, Carretón E, Corbera JA, Juste MC, Mellado I, Morchón R, et al. Current prevalence of *Dirofilaria immitis* in dogs, cats and humans from the island of Gran Canaria, Spain. *Vet Parasitol.* 2011;176:291–4.
220. Simón F, Siles-Lucas M, Morchón R, González-Miguel J, Mellado I, Carretón E. Human and animal dirofilariasis: the emergence of a zoonotic mosaic. *Clin Microbiol Rev.* 2012;25:507–44.
221. Cuadros JA, Martínez R, Lizaolaín M, Alós JI. Prurito generalizado y eosinofilia en paciente procedente de Guinea Ecuatorial. *Enferm Infecc Microbiol Clin.* 1990;8:388–9.
222. Rodríguez R, Miranda C, Martínez M, Naval G, Domínguez LM, de Toro JM. Prurito e hiperpigmentación en una mujer africana. *Enferm Infecc Microbiol Clin.* 1992;10:169–70.

223. Puente S, Subirats M, Martínez ML, Lago M, Bru F, González-Lahoz JM. Lesiones cutáneas despigmentadas en mujer ecuatoguineana. *Enferm Infecc Microbiol Clin.* 1995;13:313–4.
224. Puente S, Subirats M, González M, Lago M, Martínez ML, González-Lahoz JM. Tumoraciones subcutáneas recurrentes en mujer ecuatoguineana. *Rev Clin Esp.* 1995;195:716.
225. Molina MA, Cabezas MT, Giménez MJ. *Mansonella perstans* filariasis in a HIV patient: finding in bone marrow. *Haematologica.* 1999;84:861.
226. Daza RM, Gutiérrez J, Burkhardt P, Piédrola G. Lesión edematosa y pruriginosa en antebrazo izquierdo tras un viaje a Guinea Ecuatorial. *Med Clin (Barc).* 2000;114:791–2.
227. Bastarrica G, Pina L, Vivas I, Elorz M, San Julian M, Alberro JA. Calcified filariasis of the breast: report of four cases. *Eur Radiol.* 2001;11:1195–7.
228. Hernández Machín B, Borrego Hernando L, Báez Acosta B, Martín Sánchez AM, Hernández Hernández B, Pérez Arellano JL. Inmigrante con hiperpigmentación unila-teral en extremidades inferiores y eosinofilia. *Rev Clin Esp.* 2003;203:47–9.
229. López-Vélez R, Huerga H, Turrientes MC. Infectious diseases in immigrants from the perspective of a tropical medicine referral unit. *Am J Trop Med Hyg.* 2003;69:115–21.
230. Saldarrea A, García-Gil D, Soto-Cárdenas MJ, García-Tapia AM. Angioedema recurrente en mujer joven. *Enferm Infecc Microbiol Clin.* 2004;22:430–1.
231. Carrillo Casas E, Iglesias Pérez B, Gómez i Prat J, Guinovart Florensa C, Cabezas Otón J. Cribaje de microfilariosis sanguínea (*Loa Loa*) en la población inmigrante de zonas endémicas. *Rev Esp Salud Publica.* 2004;78:623–30.
232. Arribas J, Prieto A, Díaz AC, Calleja M, Nava E. Calcifications of the breast in *Onchocerca* infection. *Breast J.* 2005;11:507.
233. López-Rodríguez I, de-la-Fuente-Cid R, Carnero-López JM, Cordido-Carballido M, Zúñiga-Rodríguez C. Loaisis. Aproximación a una forma de parasitosis ocular. *Arch Soc Esp Otolmol.* 2007;82:55–7.
234. Gil-Setas A, Pérez Salazar M, Navascués A, Rodríguez Eleta F, Cebamanos JA, Rubio MT. Confección por dos filarias (*Loa loa* y *Mansonella perstans*) en una paciente proveniente de Guinea. *An Sist Sanit Navar.* 2010;33:227–31.
235. Moliner JV, Valverde AG, Sorolla JM. Meningoencefalitis por *Loa loa* secundaria a tratamiento con mebendazol. *Med Clin (Barc).* 2011;136:228–9.
236. Iborra MA, Carrilero B, Segovia M. Eosinofilia elevada y nódulo subcutáneo en paciente de Guinea Ecuatorial. *Enferm Infecc Microbiol Clin.* 2011;29:773–4.
237. Monge-Maillo B, Norman FF, Pérez-Molina JA, Navarro M, Díaz-Menéndez M, López-Vélez R. Travelers visiting friends and relatives (VFR) and imported infectious disease: travelers, immigrants or both? A comparative analysis. *Travel Med Infect Dis.* 2014;12:88–94.
238. Cobo F, Cabezas-Fernández MT, Salas-Coronas J, Cabeza-Barrera MI, Vázquez-Villagas J, Soriano-Pérez MJ. Filariasis in sub-Saharan immigrants attended in a health area of southern Spain: clinical and epidemiological findings. *J Immigr Minor Health.* 2015;17:306–9.
239. Jiménez M, González LM, Carranza C, Bailo B, Pérez-Ayala A, Muro A, et al. Detection and discrimination of *Loa loa*, *Mansonella perstans* and *Wuchereria bancrofti* by PCR-RFLP and nested-PCR of ribosomal DNA ITS1 region. *Exp Parasitol.* 2011;127:282–6.
240. Jiménez MI, González LM, Bailo B, Blanco A, García L, Pérez-González F, et al. Diagnóstico diferencial de filariasis importada mediante técnicas moleculares (2006–2009). *Enferm Infecc Microbiol Clin.* 2011;29:666–71.
241. Rodríguez-Osorio M, Gomez-García V, Rodríguez-Perez J, Gomez-Morales MA. Seroepidemiological studies on five outbreaks of trichinellosis in southern Spain. *Ann Trop Med Parasitol.* 1990;84:181–4.
242. Somalo J, Lamban MP, Sánchez Benítez ML. Encuesta epidemiológica a propósito de un caso de triquinosis. *Aten Primaria.* 1991;8:343–6.
243. Cobo J, Gómez Cerezo J, Medraño JC, Zapico R, Cruz Martínez A, Molina F, et al. Triquinosis. Estudio de un amplio brote en la Península Ibérica. *Med Interna.* 1991;8:441–4.
244. Nerín Sánchez C, Hermida Lazcano I, Arazo Garcés P, Sardaña Ferrer J. Brote de triquinosis por *T. britovi*. *Med Clin (Barc).* 1998;111:198–9.
245. De la Cruz de Julián I, Díaz García JM, Alvarez Lana P, García Colmenero C. Brote de triquinosis en Huerta del Marquesado (ZBS Cañete-Cuenca). Diciembre-92 a enero-93. *Rev Sanid Hig Publica (Madr).* 1994;68:513–20.
246. Tiberio G, Lanzas G, Galarza MI, Sanchez J, Quilez I, Martinez Artola V. Short report: an outbreak of trichinosis in Navarra, Spain. *Am J Trop Med Hyg.* 1995;53:241–2.
247. Tiberio G, Rivero M, Lanzas G, Redín D, Ardánaz E, Fernández C, et al. Triquinosis: estudio de dos brotes en Navarra. *Enferm Infecc Microbiol Clin.* 1997;15:151–3.
248. Rodríguez-Osorio M, Abad JM, de Haro T, Villa-Real R, Gómez-García V. Human trichinellosis in Southern Spain: serologic and epidemiologic study. *Am J Trop Med Hyg.* 1999;61:834–7.
249. López Hernández B, Velázquez de Castro MT, Galicia García MD, Sabonet JC. Brote epidémico por *Trichinella britovi* en Granada durante la primavera del 2000. *Rev Esp Salud Publica.* 2001;75:467–73.
250. Cortés-Blanco M, García-Cabañas A, Guerra-Peguero F, Ramos-Aceitero JM, Herrera-Guibert D, Martínez-Navarro JF. Outbreak of trichinellosis in Cáceres, Spain, December 2001–February 2002. *Euro Surveill.* 2002;7:136–8.
251. Gomez-García V, Hernandez-Quero J, Rodriguez-Osorio M. Short report: human infection with *Trichinella britovi* in Granada, Spain. *Am J Trop Med Hyg.* 2003;68:463–4.
252. Herráez García J, Leon García LA, Lanusse Senderos C, Cortés Blanco M, García Cabañas A. Brote de triquinosis en la comarca de la Vera (Cáceres) causado por *Trichinella britovi*. *An Med Interna.* 2003;20:63–6.
253. Rodríguez de las Parras E, Rodríguez-Ferrer M, Nieto-Martínez J, Ubeira FM, Gárate-Ormaechea T. Revisión de los brotes de triquinosis detectados en España durante 1990–2001. *Enferm Infecc Microbiol Clin.* 2004;22:70–6.
254. Valdés PV, Prieto A, Diaz A, Calleja M, Gomez JL. Microcalcifications of pectoral muscle in trichinosis. *Breast J.* 2005;11:150.
255. Gallardo MT, Mateos L, Artieda J, Wesslen L, Ruiz C, García MA, et al. Outbreak of trichinellosis in Spain and Sweden due to consumption of wild boar meat contaminated with *Trichinella britovi*. *Euro Surveill.* 2007;12:E070315.1.
256. Arévalo Velasco A, Bringas MJ, Rodríguez R, Menor A. Descripción de un brote de triquinosis en la provincia de Salamanca. *Rev Esp Quimioter.* 2009;22:115–6.
257. Arenal Vera JJ, Marcos Rodríguez JL, Borrego Pintado MH, Bowakin Dib W, Castro Lorenzo J, Blanco Alvarez JL. Anisakiasis como causa de apendicitis aguda y cuadro reumatológico: primer caso en la literatura médica. *Rev Esp Enferm Dig.* 1991;79:355–8.
258. López Vélez R, García A, Barros C, Manzarbeitia F, Oñate JM. Anisakiasis en España. Descripción de 3 casos. *Enf Infecc Microbiol Clin.* 1992;10:158–61.
259. Cruchaga S, Pascual J, Muñoz F, Guerra A, Ladrón de Guevara C. Hallazgo endoscópico de una larva de gusano en el fondo de una úlcera gástrica. *Enferm Infecc Microbiol Clin.* 1995;13:631–2.
260. Audicana M, Fernández de Corres L, Muñoz D, Fernández E, Navarro JA, del Pozo MD. Recurrent anaphylaxis due to ingestion and contact with *Anisakis simplex* parasitizing sea-fish. *J Allergy Clin Immunol.* 1995;96:558–60.
261. Fernández de Corres L, Audicana M, del Pozo MD, Muñoz D, Fernández E, Navarro JA. *Anisakis simplex* induces not only anisakiasis: report on 28 cases of allergy caused by this nematode. *J Investig Allergol Clin Immunol.* 1996;6:315–9.
262. Canut A, Labora A, López J, Romero JA. Anisakiasis gástrica aguda por cocción insuficiente en horno microondas. *Med Clin.* 1996;8:317–8.
263. Acebes Rey JM, Fernández Orcajo P, Díaz González G, Velicia Llamas R, González Hernández JM, Citores González R. Dos casos de anisakiasis en el Hospital del Río Hortega (Valladolid). *Rev Esp Enferm Dig.* 1996;88:59–60.
264. Romeo Ramírez JA, Martínez-Conde López AE, Olivares Galdeano U, Sancha Pérez A, López de Torre Ramírez de la Piscina J, Barros Ingerto J, et al. Anisakiasis gástrica diagnosticada por endoscopia. *Gastroenterol Hepatol.* 1997;20:306–8.
265. Louredo A, Acedo de la Rosa F, Ofiñas de Paz V, Sanz E, Quiros BL, Goyanes M. Anisakidosis del colon como causa de abdomen agudo. *Rev Esp Enf Dig.* 1997;89:403–6.
266. Gómez B, Tabar AI, Tuñón T, Larrínaga B, Alvarez MJ, García BE, et al. Eosinophilic gastroenteritis and *Anisakis*. *Allergy.* 1998;53:1148–54.
267. Del Olmo Escribano M, Cozar Ibáñez A, Martínez de Victoria JM, Ureña Tirao C. Anisakiasis a nivel ileal. *Rev Esp Enferm Dig.* 1998;90:120–3.
268. Rosales MF, Mascará C, Fernandez C, Luque F, Sanchez Moreno M, Parras L, et al. Acute intestinal anisakiasis in Spain: a fourth-stage *Anisakis simplex* larva. *Mem Inst Oswaldo Cruz.* 1999;94:823–6.
269. Oliveira A, Sánchez Rancaño S, Conde Gacho P, Moreno A, Martínez A, Comas C. Anisakiasis gastrointestinal. Siete casos en 3 meses. *Rev Esp Enferm Dig.* 1999;91:70–2.
270. López Peñas D, Ramírez Ortiz LM, del Rosal Palomeque R, López Rubio F, Fernández-Crehuet Navajas R, Miño Fugarolas G. Estudio de 13 casos de anisakiasis en la provincia de Córdoba. *Med Clin (Barc).* 2000;114:177–80.
271. Domínguez Ortega J, Cimarra M, Sevilla M, Alonso Llamazares A, Moneo I, Robledo Echarren T, et al. *Anisakis simplex*: una causa de pseudo-obstrucción intestinal. *Rev Esp Enferm Dig.* 2000;92:132–9.
272. Céspedes M, Saez A, Rodríguez I, Pinto JM, Rodríguez R. Chronic anisakiasis presenting as a mesenteric mass. *Abdom Imaging.* 2000;25:548–50.
273. Del Olmo Martínez L, González de Canales P, Sanjosé González G. Anisakiasis gástrica diagnosticada mediante endoscopia. *An Med Interna.* 2000;17:429–31.
274. Castán B, Borda F, Iñarrairaegui M, Pastor G, Vila J, Zozaya JM. Anisakiasis digestiva: clínica y diagnóstico según la localización. *Rev Esp Enferm Dig.* 2002;94:463–72.
275. Pérez-Naranjo S, Venturini-Díaz M, Colás-Sanz C, Aguilera-Diago V, Ruiz-de-Iglesia F, Pérez-Calvo JI. Intestinal anisakiasis mimicking intestinal obstruction. *Eur J Med Res.* 2003;8:135–6.
276. Repiso Ortega A, Alcántara Torres M, González de Frutos C, de Artaza Varasa T, Rodríguez Merlo R, Valle Muñoz J, et al. Anisakiasis gastrointestinal. Estudio de una serie de 25 pacientes. *Gastroenterol Hepatol.* 2003;26:341–6.
277. Eguía A, Aguirre JM, Echevarria MA, Martínez-Conde R, Pontón J. Gingivostomatitis after eating fish parasitized by *Anisakis simplex*: a case report. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2003;96:437–40.
278. González Quijada S, González Escudero R, Arias García L, Gil Martín AR, Vicente Serrano J, Corral Fernández E. Manifestaciones digestivas de la anisakiasis: descripción de 42 casos. *Rev Clin Esp.* 2005;205:311–5.
279. Ponferrada A, Matilla A, Borrego GM, Beceiro I, Núñez O, Lamónaca M, et al. Hemoperitoneo espontáneo secundario a yeyunoileítis por *Anisakis*. *Esp Enferm Dig.* 2005;97:292.
280. Ramos L, Alonso C, Guilarte M, Vilaseca J, Santos J, Malagelada JR. *Anisakis simplex*-induced small bowel obstruction after fish ingestion: preliminary evidence for response to parenteral corticosteroids. *Clin Gastroenterol Hepatol.* 2005;3:667–71.
281. Meseguer J, Navarro V, Sánchez-Guerrero I, Bartolomé B, Negro Alvarez JM. *Anisakis simplex* allergy and nephrotic syndrome. *Allergol Immunopathol (Madr).* 2007;35:216–20.
282. Puente P, Anadón AM, Rodero M, Romarís F, Ubeira FM, Cuéllar C. *Anisakis simplex*: the high prevalence in Madrid (Spain) and its relation with fish consumption. *Exp Parasitol.* 2008;118:271–4.

283. Jurado-Palomo J, López-Serrano MC, Moneo I. Multiple acute parasitization by *Anisakis simplex*. *J Investig Allergol Clin Immunol.* 2010;20:437–41.
284. Cabrera R. Anisakiasis outbreak by *Anisakis simplex* larvae associated to Peruvian food in Spain. *Rev Esp Enferm Dig.* 2010;102:610–1.
285. Hernández-Bel P, de la Cuadra J, García R, Alegre V. Dermatitis de contacto por proteínas. Revisión de 27 casos. *Actas Dermosifiliogr.* 2011;102:336–43.
286. Valle J, Lopera E, Sánchez ME, Lerma R, Ruiz JL. Spontaneous splenic rupture and *Anisakis appendicitis* presenting as abdominal pain: a case report. *J Med Case Rep.* 2012;6:114.
287. Carrascosa MF, Mones JC, Salcines-Caviedes JR, Román JG. A man with unsuspected marine eosinophilic gastritis. *Lancet Infect Dis.* 2015;15:248.
288. Cilla G, Pérez-Trallero E, Gutiérrez C, Part C, Gomáriz M. Seroprevalence of *Toxocara infection* in middle-class and disadvantaged children in northern Spain (Gipuzkoa, Basque Country). *Eur J Epidemiol.* 1996;12:541–3.
289. Roig J, Romeu J, Riera C, Texido A, Domingo C, Morera J. Acute eosinophilic pneumonia due to toxocaríasis with bronchoalveolar lavage findings. *Chest.* 1992;102:294–6.
290. López-Vélez R, Suárez de Figueroa M, Gimeno L, García-Camacho A, Fenoy S, Guillén JL, et al. Toxocaríasis ocular o retinoblastoma? *Enferm Infecc Microbiol Clin.* 1995;13:242–5.
291. Arias Irigoyen J, Senent Sánchez CJ. Toxocaríasis: a cause of hyper IgE and eosinophilia. *J Investig Allergol Clin Immunol.* 1995;5:232–4.
292. Jimenez JF, Valladares B, Fernandez-Palacios JM, de Armas F, del Castillo A. A serologic study of human toxocaríasis in the Canary Islands (Spain): environmental influences. *Am J Trop Med Hyg.* 1997;56:113–5.
293. Fenoy S, Cuéllar C, Guillén JL. Serological evidence of toxocaríasis in patients from Spain with a clinical suspicion of visceral larva *migrans*. *J Helminthol.* 1997;71:9–12.
294. Perteguer MJ, Cuéllar C, Guillén JL, Aguila C, Fenoy S, Chivato T, et al. Cross-reactivity between *Anisakis simplex* sensitization and visceral larva *migrans* by *Toxocara canis*. *Acta Trop.* 2003;89:85–9.
295. Turrientes MC, Perez de Ayala A, Norman F, Navarro M, Perez-Molina JA, Rodríguez-Ferrer M, et al. *Visceral larva migrans* in immigrants from Latin America. *Emerg Infect Dis.* 2011;17:1263–5.
296. Montero E, Montero J, Rosales MJ, Mascaró C. Human gnathostomiasis in Spain: first report in humans. *Acta Trop.* 2001;78:59–62.
297. Puente S, Gárate T, Grobusch MP, Janitschke K, Bru F, Rodríguez M, et al. Two cases of imported gnathostomiasis in Spanish women. *Eur J Clin Microbiol Infect Dis.* 2002;21:617–20.
298. Górgolas M, Santos-O'Connor F, Unzué AL, Fernández-Guerrero ML, Gárate T, Troyas Guarch RM, et al. Cutaneous and medullar gnathostomiasis in travelers to Mexico and Thailand. *J Travel Med.* 2003;10:358–61.
299. De Górgolas Hernández-Mora M, Fernández Guerrero ML. Gnathostomiasis: una enfermedad creciente en viajeros. *Med Clin (Barc).* 2005;125:190–2.
300. Oliván-Gonzalvo G. Gnathostomiasis tras un viaje a China para realizar una adopción internacional. *Med Clin (Barc).* 2006;126:758–9.
301. Norman FF, López-Vélez R. Immigration, helminths and eosinophilia: a complex triad. *Trav Med Infect Dis.* 2015;13:283–4.