



Enfermedades Infecciosas y Microbiología Clínica

www.elsevier.es/eimc



Brief report

Trend and seasonality of community-acquired *Escherichia coli* antimicrobial resistance and its dynamic relationship with antimicrobial use assessed by ARIMA models[☆]

María Ángeles Asencio Egea^{a,*}, María Huertas Vaquero^a, Rafael Carranza González^a, Óscar Herráez Carrera^b, Olga Redondo González^c, Ángel Arias Arias^c

^a Laboratorio de Microbiología, Hospital General La Mancha Centro, Alcázar de San Juan, Ciudad Real, Spain

^b Unidad de Calidad del Laboratorio, Hospital General La Mancha Centro, Alcázar de San Juan, Ciudad Real, Spain

^c Unidad de Apoyo a la Investigación, Hospital General La Mancha Centro, Alcázar de San Juan, Ciudad Real, Spain

ARTICLE INFO

Article history:

Received 14 July 2017

Accepted 5 October 2017

Available online 10 July 2018

Keywords:

Trend

Seasonality

Antimicrobial resistance

Escherichia coli

Antimicrobial use

Primary healthcare

ABSTRACT

Introduction: We studied the trend and seasonality of community-acquired *Escherichia coli* resistance and quantified its correlation with the previous use of certain antibiotics.

Methods: A time series study of resistant community-acquired *E. coli* isolates and their association with antibiotic use was conducted in a Primary Health Care Area from 2008 to 2012. A Poisson regression model was constructed to estimate the trend and seasonality of *E. coli* resistance.

Results: A significant increasing trend in mean *E. coli* resistance to cephalosporins, aminoglycosides and nitrofurantoin was observed. Seasonal resistance to ciprofloxacin and amoxicillin-clavulanic acid was significantly higher in autumn-winter. There was a delay of 7, 10 and 12 months between the use of cotrimoxazole ($p < 0.038$), fosfomicin ($p < 0.024$) and amoxicillin-clavulanic acid ($p < 0.015$), respectively, and the occurrence of *E. coli* resistance.

Conclusions: An average delay of 10 months between the previous use of amoxicillin-clavulanic acid, cotrimoxazole and fosfomicin and the appearance of resistant community-acquired *E. coli* strains was detected.

© 2017 Elsevier España, S.L.U. and Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. All rights reserved.

Tendencia y estacionalidad de las resistencias de *Escherichia coli* comunitarios y su relación dinámica con el consumo de antimicrobianos mediante modelos ARIMA

RESUMEN

Introducción: Estudiamos la tendencia y estacionalidad de las resistencias en *Escherichia coli* comunitario y se cuantifica su asociación con el uso previo de determinados antibióticos.

Métodos: Estudio de series temporales de las resistencias de aislados comunitarios de *E. coli* y su relación con el consumo de antibióticos en un área de primaria durante 2008–2012. La tendencia y estacionalidad de las resistencias se estudiaron mediante regresión de Poisson.

Resultados: Se observó un aumento significativo de la resistencia promedio de *E. coli* a cefalosporinas, nitrofurantoína y aminoglucósidos. La estacionalidad de las resistencias fue significativa en otoño-invierno para amoxicilina-ácido clavulánico y ciprofloxacino. Observamos un retardo de 7, 10 y 12 meses entre el consumo de cotrimoxazol ($p < 0,038$), fosfomicina ($p < 0,024$) y amoxicilina-ácido clavulánico ($p < 0,015$), respectivamente, y la aparición de resistencias.

Palabras clave:

Tendencia

Estacionalidad

Resistencias

Escherichia coli

Consumo de antibióticos

Atención primaria

DOI of original article: <https://doi.org/10.1016/j.eimc.2017.10.013>

[☆] Please cite this article as: Asencio Egea MÁ, Huertas Vaquero M, Carranza González R, Herráez Carrera Ó, Redondo González O, Arias Arias Á. Tendencia y estacionalidad de las resistencias de *Escherichia coli* comunitarios y su relación dinámica con el consumo de antimicrobianos mediante modelos ARIMA. *Enferm Infecc Microbiol Clin.* 2018;36:502–506.

* Corresponding author.

E-mail address: marian.asencio@yahoo.es (M.Á. Asencio Egea).

Conclusiones: Detectamos un retardo medio de 10 meses entre la utilización de amoxicilina-ácido clavulánico, cotrimoxazol y fosfomicina, y la aparición de cepas resistentes de *E. coli* comunitarios.

© 2017 Elsevier España, S.L.U. y Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. Todos los derechos reservados.

Introduction

The increase in infections due to multidrug-resistant microorganisms, which is generating so much alarm due to their high morbidity and mortality as well as their high healthcare costs,¹ has often been linked to the high selective pressure of the antimicrobials used in human health.^{2,3}

So-called *autoregressive integrated moving average* (ARIMA) models are statistical tools for finding patterns to enable future estimates to be made. They have been applied to the management of nosocomial infection⁴ and more recently to the design of an efficient antimicrobial management programme following an outbreak of carbapenemase-producing bacteria.⁵ They also enable the relationships between the use of antibiotics and the development of resistance to be measured and estimate the delay to the development of increased resistance to an antibiotic following increased use of that antibiotic.^{6,7}

The objectives of this study are to examine trends and seasonal variations in the resistance of community-acquired *Escherichia coli* isolates to certain antimicrobials and to measure the dynamic relationship between prior use of antibiotics and the development of resistance for a 5-year period using ARIMA models.

Materials and methods

This was a retrospective study of time series of strains of *E. coli* isolated from patients from primary care (PC) in a healthcare area

Table 1
Average change in percentage of resistance (2008–2012).

	Incident rate	95% CI
Amoxicillin–clavulanic acid	1.01	0.97–1.06
Ampicillin	1.01	0.98–1.03
Cefoxitin	1.17 ^a	1.10–1.25
Cefotaxime	1.09 ^a	1.02–1.17
Cefuroxime	1.07 ^a	1.02–1.11
Ciprofloxacin	1.03	1.00–1.06
Co-trimoxazole	1.01	0.98–1.04
Fosfomicin	1.05	0.96–1.15
Gentamicin	1.11 ^a	1.05–1.17
Nitrofurantoin	1.11 ^a	1.04–1.18
Tobramycin	1.25 ^a	1.18–1.33

^a Significant increase in average resistance ($p < 0.05$).

Table 2
Average change in antibiotic consumption (2008–2012).

Antibiotic	DDD: Annual incident rate					Incident rate 2008–2012 Average change (95% CI)
	2008	2009	2010	2011	2012	
Ampicillin	1	0.87	1.16	0.84	0.88	0.95 (0.79–1.11)
Amoxicillin–clavulanic acid	1	1	1	1.07	1	1.01 (0.98–1.05)
Cefuroxime	1	0.88	0.79	0.84	0.78	0.86 (0.75–0.97) ^a
Ciprofloxacin	1	1	1.07	0.98	1	1.01 (0.97–1.05)
Co-trimoxazole	1	1.10	1.15	1.16	1.07	1.10 (1.02–1.18) ^a
Fosfomicin	1	1.22	1.30	1.31	1.42	1.25 (1.06–1.44) ^a
Nitrofurantoin	1	0.92	1.24	1.33	1.42	1.18 (0.92–1.45)
Gentamicin	1	1.56	1.14	1.09	1.15	1.19 (0.92–1.46)
Tobramycin	1	0.86	1.05	0.86	1.30	1.01 (0.79–1.24)
Overall DDD	24.31	24.00	23.37	24.27	23.70	–

Overall DDD: total DDD for therapeutic group J01 (antibacterials for systemic use) at the health centres in our setting by year of study.

^a Significant change in average use of antibiotics ($p < 0.05$).

with 211,533 inhabitants, use of antibiotics and development of resistance between 2008 and 2012.

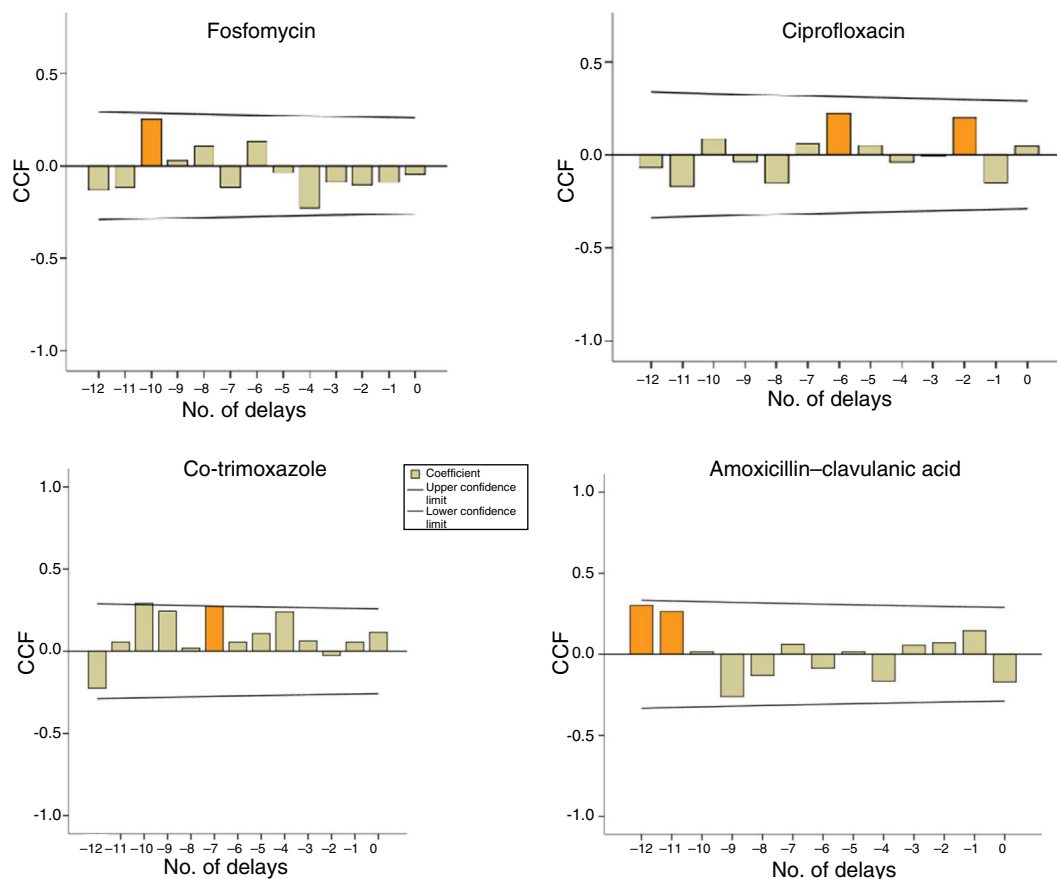
The antibiotic use data refer to the population studied and are expressed in terms of defined daily dose (DDD per 1000 inhabitants per day). They were provided by the Dirección General de Farmacia [Directorate General for Pharmacy] of the Autonomous Community. The study of susceptibility to ampicillin (AM), amoxicillin–clavulanic acid (AMC), cefoxitin (CFX), cefotaxime (CFT), cefuroxime (CFM), ciprofloxacin (CIP), co-trimoxazole (SXT), fosfomicin (F), nitrofurantoin (NF), gentamicin (G) and tobramycin (TO) was performed using the VITEK-2[®] system (bioMérieux, France) in the microbiology laboratory at the reference hospital. The data are expressed in terms of percentage of resistance (number of antibiotic-resistant *E. coli* isolates divided by total number of isolates). The month was used as a time aggregate both for use and for resistance.

Trends and seasonal variations in resistance were studied using a Poisson regression model where the percentage of resistance was used as a dependent variable. Trends were estimated using the variable representing the number of months elapsed in 5 years. Seasonal variations were estimated using the variable representing the month of the year, taking the month of January as a reference point. The result obtained was interpreted in terms of an incidence rate ratio (IRR) and its 95% confidence interval (95% CI). For trends, the IRR indicates the relative change in the series for each year elapsed. For seasonal variations, the IRR indicates the relative change in each month compared to the month taken as a reference point.

Different ARIMA models were built from time series according to the methodology reported by Box et al. in 1976⁸ and adapted by López-Lozano et al. in 2000.⁶ First, graphs of cross-correlations were used to identify the delays in which the association between use and resistance was most significant. Next, transfer models were built to quantify the relationship between, on the one hand, the use of AMC, CIP, SXT and F and, on the other hand, the development of resistance in *E. coli* isolates.

Results

A total of 9326 community-acquired *E. coli* isolates were studied. The average change in the percentage of resistance and the average change in the use of antibiotics are shown in [Tables 1 and 2](#), respectively. We found that the use of fosfomicin, co-trimoxazole and nitrofurantoin gradually increased, whereas



CCF: cross-correlation function

Antimicrobials	Delay (months)	Measurement of association	p
Amoxicillin-clavulanic acid	12	1.92%	0.015
Ciprofloxacin	2	0.21%	0.32
Co-trimoxazole	7	24.4%	0.038
Fosfomycin	10	24.5%	0.024

Fig. 1. ARIMA models and cross-correlation functions. Relationship between use of amoxicillin-clavulanic acid, ciprofloxacin, co-trimoxazole and fosfomycin and development of resistance in *E. coli* to these antibiotics.

the use of ciprofloxacin remained stable throughout the period studied. An ascending trend was observed in resistance to the cephalosporins studied, with an average increase in resistance of 7%, 9% and 17% for CFM, CFT and CFX, respectively. Average resistance to aminoglycosides (AMGs) also increased significantly: 11% for G and 25% for TO, as well as 11% for NF. There were two peak increases in AMC resistance, one in 2009 (IRR: 1.25; 95% CI: 1.06–1.47; $p=0.008$) and the other in 2011 (IRR: 3.55; 95% CI: 1.14–1.58; $p<0.001$). CIP resistance increased as of 2010 (IRR: 1.22; 95% CI: 1.06–1.40; $p=0.006$).

Regarding seasonal variations, resistance to AMC and CIP increased significantly in autumn and winter—CIP in December (IRR: 1.27; 95% CI: 1.03–1.57; $p=0.025$) and AMC in September (IRR: 1.30; 95% CI: 1.01–1.68; $p=0.039$), October (IRR: 1.41; 95% CI: 1.10–1.81; $p=0.007$) and December (IRR: 2.19; 95% CI: 1.03–1.70; $p=0.029$).

Figure 1 shows the ARIMA models for AMC, CIP, SXT and F. A positive relationship is seen between the prescription of these antimicrobials and the development of resistance in community-acquired *E. coli*, with a delay ranging from 2 to 12 months. This

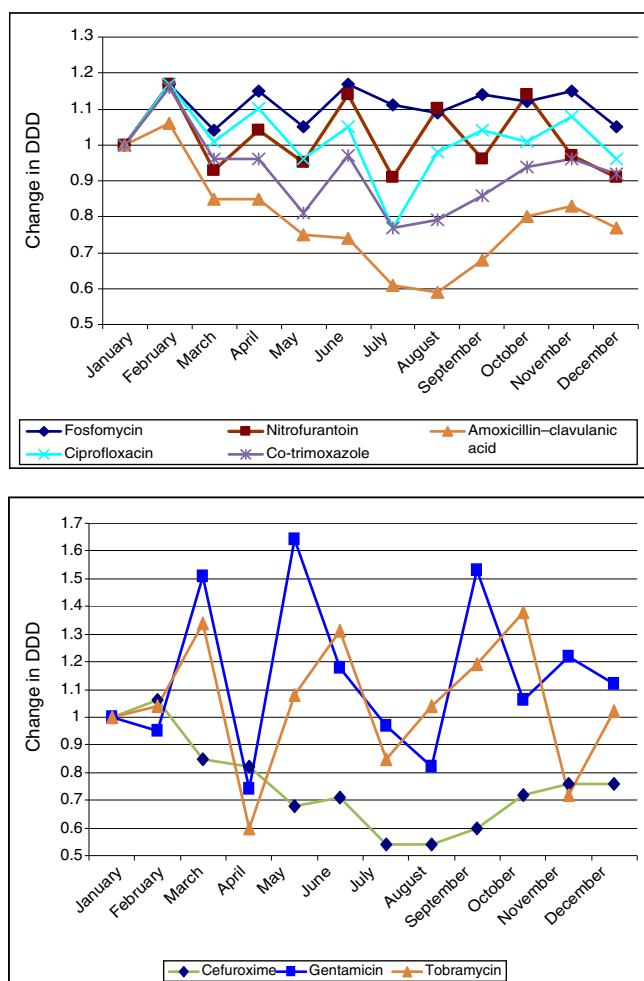


Fig. 2. Seasonal variations in average monthly use of the antibiotics studied (2008–2012). The antibiotics in the above graph are those that are most commonly used in primary care (fosfomicin, nitrofurantoin, amoxicillin–clavulanic acid, ciprofloxacin and co-trimoxazole). They show more uniform seasonal variations than those in the below graph (cefuroxime, gentamicin and tobramycin).

relationship was significant for AMC, SXT and F. Fig. 2 shows seasonal variations in average use.

Discussion

Our study has once again shown the solid relationship between the use of antimicrobials and the development of resistance for a microorganism as prevalent as *E. coli*. Analysis of time series enables prediction of the future behaviour of the series studied based on past behaviour. On this basis we sought to predict the development of resistance in community-acquired *E. coli* isolates based on series of resistance and use of CIP, AMC, SXT and F obtained in the past five years, as well as to estimate the delay in its presentation.

The ascending trend in the resistance of *E. coli* to cephalosporins, NF and AMG found in our study is consistent with European Antimicrobial Resistance Surveillance System report data.⁹ This could be related to the dissemination of broad-spectrum beta-lactamases in the community, growing since the 1980s,¹⁰ and often presenting co-resistance to other antibiotics such as fluoroquinolones and AMGs. Another possible cause is acquisition of cephalosporins in *E. coli* from animals.¹¹ Peak increases in AMC resistance could be related to greater seasonal use of this and other antibiotics due to the influenza A epidemic which started in Spain in April 2009, leading to higher rates of morbidity and superinfection.

Several studies have used ARIMA models to determine the relationship between the use of antibiotics and the development of resistance.^{6,7,12,13} Hsueh et al. found a significant relationship between increased resistance of *E. coli* to CFT and CIP and increased use of beta-lactam antibiotics, carbapenems, FQs and AMGs.¹² A French study has shown a significant association between the use of fluoroquinolones and increased resistance to ofloxacin and CIP in *E. coli* strains of urinary origin in hospitalised patients, with delays of less than six months.⁷ According to our study, each increase by one unit in the use of SXT, F, and AMC is expected to result in an increase in resistance with delays of seven, 10 and 12 months, respectively. We believe that widespread awareness of these data could guide physicians in empirical antibiotic prescription.

Our mean extrahospital antibiotic use in terms of DDD (23.93) was higher than the Spanish mean (20.30) and even higher than the European mean (21.16) in the same period.¹⁴ As antimicrobial use is far less managed in a community setting than in a specialised setting, using these predictive models in our setting would offer the option of active intervention to reduce the consequences of this deficiency in reasonable prescription of antibiotics and come closer to the target of the Optimisation Programmes for antibiotics in PC. The implementation of these programmes is a high priority in most Spanish communities.¹⁵ Therefore, it would be a good idea to conduct multi-centre studies evaluating the validity of the association and the delays found, as well as to broaden the study to other combinations of microorganisms and antimicrobials.

This study suffers from the limitations particular to retrospective studies: it has a higher risk of information and selection biases. Another limitation is its lack of application of DDDs to the child population.

In conclusion, we found a positive relationship between the use of the antimicrobials analysed and the development of resistance in community-acquired *E. coli*. This relationship was significant for AMC, SXT and F. The delay in the development of these forms of resistance ranged from seven to 12 months. Thus ARIMA models would enable forecasts to be established to make more reasonable use of antibiotics in a community setting.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Acknowledgement

We would like to thank Dr José María Tenías Burillo for his unconditional support.

References

- Schwaber MJ, Navon-Venezia S, Kaye KS, Ben-Ami R, Schwartz D, Carmeli Y. Clinical and economic impact of bacteremia with extended-spectrum β -lactamase-producing *Enterobacteriaceae*. *Antimicrob Agents Chemother.* 2006;50:1257–62.
- Allegranzi B, Luzzati R, Luzzani A, Girardini F, Antozzi L, Raiteri R, et al. Impact of antibiotic changes in empirical therapy on antimicrobial resistance in intensive care unit-acquired infections. *J Hosp Infect.* 2002;52:136–40.
- Aldeyab MA, Harbarth S, Vernaz N, Kearney MP, Scott MG, Darwish Elhajji FW, et al. The impact of antibiotic use on the incidence and resistance pattern of extended-spectrum beta-lactamase-producing bacteria in primary and secondary healthcare settings. *Br J Clin Pharmacol.* 2012;74:171–9.
- Aldeyab MA, Monnet DL, Lopez-Lozano JM, Hughes CM, Scott MG, Kearney MP, et al. Modelling the impact of antibiotic use and infection control practices on the incidence of hospital-acquired methicillin-resistant *Staphylococcus aureus*: a time-series analysis. *J Antimicrob Chemother.* 2008;62:593–600.
- Gharbi M, Moore LS, Gilchrist M, Thomas CP, Bamford K, Brannigan ET, et al. Forecasting carbapenem resistance from antimicrobial consumption surveillance: lessons learnt from an OXA-48-producing *Klebsiella pneumoniae* outbreak in a West London renal unit. *Int J Antimicrob Agents.* 2015;46:150–6.
- López-Lozano JM, Monnet DL, Yagüe A, Burgos A, Gonzalo N, Campillos P, et al. Modelling and forecasting antimicrobial resistance and its dynamic

- relationship to antimicrobial use: a time series analysis. *Int J Antimicrob Agents.* 2000;14:21–31.
7. Mahamat A, Lavigne JP, Fabbro-Peray P, Kinowski JM, Daures JP, Sotto A. Evolution of fluoroquinolone resistance among *Escherichia coli* urinary tract isolates from a French university hospital: application of the dynamic regression model. *Clin Microbiol Infect.* 2005;11:301–6.
 8. Box G, Jenkins G, Reinsel G. *Time series analysis: forecasting and control.* 1st ed. San Francisco, CA: Holden-Day; 1976.
 9. European Centre for Disease Prevention and Control. *Antimicrobial resistance surveillance in Europe 2012. Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net).* Stockholm: ECDC; 2015.
 10. Oteo J, Pérez-Vázquez M, Campos J. Extended-spectrum [beta]-lactamase producing *Escherichia coli*: changing epidemiology and clinical impact. *Curr Opin Infect Dis.* 2010;23:320–6.
 11. Fang LX, Sun J, Li L, Deng H, Huang T, Yang QE, et al. Dissemination of the chromosomally encoded CMY-2 cephalosporinase gene in *Escherichia coli* isolated from animals. *Int J Antimicrob Agents.* 2015;46:209–13.
 12. Hsueh PR, Chen WH, Luh KT. Relationships between antimicrobial use and antimicrobial resistance in Gram-negative bacteria causing nosocomial infections from 1991–2003 at a university hospital in Taiwan. *Int J Antimicrob Agents.* 2005;26:463–72.
 13. Monnet DL, López-Lozano JM, Campillos P, Burgos A, Yagüe A, Gonzalo N. Making sense of antimicrobial use and resistance surveillance data: application of ARIMA and transfer function models. *Clin Microbiol Infect.* 2001;7 Suppl 5:S29–36.
 14. European Centre for Disease Prevention and Control. *Surveillance of antimicrobial consumption in Europe 2012.* Stockholm: ECDC; 2014.
 15. Programa integral de prevención, control de las infecciones relacionadas con la asistencia sanitaria, y uso apropiado de los antimicrobianos (PIRASOA). Available from: <http://ws140.juntadeandalucia.es/piraso>