



INTERNATIONAL MEDICAL REVIEW ON DOWN'S SYNDROME

www.elsevier.es/sd



REVIEW

Oxidative stress, exercise and Down syndrome

C. Campos and A. Casado*

Department of Cell and Molecular Medicine, Spanish National Research Council Biological Research Centre (CSIC), Madrid, Spain

Received on July 31, 2014; accepted on September 25, 2014

KEY WORDS:

Antioxidants;
Down syndrome;
Exercise;
Oxidative stress

Abstract

An imbalance between free radical production and antioxidant defence leads to an oxidative stress state that can potentially result in a change in the intracellular redox balance towards a more oxidizing environment, with the possibility of causing ill-health and disease. Down's syndrome (DS) is the first clinically defined chromosomal syndrome, and is caused by the presence of three copies of chromosome 21. It is one of the most significant human congenital defects, with an incidence of 1 in every 700-1000 births. Regular physical activity is known as an important factor for health. However, exhaustive and/or intense physical activity increases free radical and reactive oxygen/nitrogen species (RONS) production. A minimal amount of RONS is necessary for muscular contraction. Nevertheless, oxidative stress, which results in muscle-increased RONS concentration, is associated with muscular fatigue during contraction and in post-exercise muscular damage suffering. The mitochondrial respiratory chain, the ischemia reperfusion phenomenon and the inflammatory reaction have been identified as the main sources of RONS during and after exercise. However, results reported in literature seem to show a beneficial effect of exercise on DS subjects, although the effect of exercise on oxidative stress parameters has been poorly studied in this condition.

PALABRAS CLAVE:

Antioxidantes;
Síndrome de Down;
Ejercicio;
Estrés oxidativo

Estrés oxidativo, ejercicio y síndrome de Down

Resumen

Un desequilibrio entre la producción de radicales libres y defensas antioxidantes puede conducir a un estado de estrés oxidativo que podría causar un cambio en el equilibrio redox intracelular generando un entorno más oxidante, con la posibilidad de causar

The authors confirm that it has not been published anywhere else, and that it is not under simultaneous consideration in any other publication.

This work is part of a project funded by the Fundación Inocente, Inocente.

*Author for correspondence.

E-mail: acasado@cib.csic.es (A. Casado).

enfermedad. El síndrome de Down (SD) es el primer síndrome cromosómico clínicamente definido y está causado por la presencia de tres copias del cromosoma 21. Es uno de los defectos congénitos humanos más importantes, con una incidencia de 1 de cada 700-1000 nacimientos. Se sabe que la actividad física, si se practica regularmente, es un factor importante para la salud; sin embargo, practicada de forma exhaustiva e intensa incrementa la formación de radicales libres y de especies reactivas de oxígeno y nitrógeno (ERON). Para la contracción muscular es necesaria una cantidad mínima de ERON. Por otra parte, el estrés oxidativo que resulta de la concentración de ERON en músculo está asociado con fatiga muscular durante la contracción y daño muscular después del ejercicio. Las principales fuentes de ERON durante y después del ejercicio son: cadena respiratoria mitocondrial, isquemia reperfusión y reacción inflamatoria. Sin embargo, los resultados recogidos en la bibliografía muestran efectos beneficiosos del ejercicio en el SD, aunque los efectos del ejercicio en parámetros de estrés oxidativo han sido muy poco estudiados en el SD.

Introduction

Regular physical activity, associated with a balanced diet, is known as an important factor for health¹. However, strenuous physical exercise is associated with a dramatic increase in oxygen uptake both by the whole body and particularly by the skeletal muscle. The production of reactive oxygen/nitrogen species (RONS) is believed to be the underlying mechanism for a series of biochemical and physiological changes that occur during exercise and are indicative of oxidative stress. In general, the body has adequate antioxidant reserves to cope with the production of reactive oxygen species ROS under physiological conditions. The system consists of antioxidant enzymes, antioxidant vitamins, glutathione and thiols. These antioxidant defence systems preserve homeostasis for normal cell functions at rest and perhaps during mild exercise. However, when ROS production is excessive, such as during prolonged aerobic exercise, or when antioxidant defence is severely hampered by nutritional deficiencies and pharmacological intervention, an inadequate defence may be overwhelmed by the RONS, leading to extensive cell and tissue damage². Antioxidant enzymes are endogenous; however their production can be modulated by certain factors.

Exercise and training are well known to be potential factors for the increase of superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx) as shown by numerous studies³.

The aim of this article is to critically review the scientific literature concerning the link between oxidative stress and exercise in Down syndrome (DS). We review the extent of information available in scientific literature regarding oxidative stress and exercise in DS, providing evidence against and in favour of increased oxidative stress in this condition.

Down syndrome

Down syndrome is one of the most prevalent genetic disorders; it occurs at frequency of one in 700-1000 live births⁴ and results as a consequence of either a total or partial trisomy of chromosome 21⁵. The syndrome is characterized by chronicity and severity of abnormalities including intellectual disability, dysmorphic features, as well as immuno-

logical, hematological and endocrine defects⁶. The phenotypic expression of human trisomy 21 is presumed to result from the overexpression of certain genes residing on chromosome 21 at the segment 21q22—the Down locus^{7,8}.

The amount of studies reporting a number of dysfunctions associating oxidative stress with DS phenotype is increasing. Oxidative stress is part of the fundamental biology of DS. It has been suggested that the main source of ROS in DS patients is the excessive production of hydrogen peroxide (H₂O₂) through the action of Cu,Zn-superoxide dismutase (Cu,Zn-SOD)^{9,10}. As a result of the overexpression of Cu,Zn-SOD in DS patients, an imbalance between Cu,Zn-SOD and other antioxidant enzymes occurs, such as CAT and GPx, inducing a systemic oxidative damage⁹. SOD1 promotes the production of H₂O₂, an important precursor of hydroxyl radical (•OH), the most reactive and deleterious ROS, therefore it is able to react with important cellular components, oxidizing biomolecules such as amino acid residues, proteins, lipids and DNA¹¹.

Oxidative stress

Free radicals are reactive compounds that are naturally produced in the human body. They can exert positive effects (e.g. on the immune system) or negative effects (e.g. lipids, protein or DNA oxidation). A free radical is a molecule with one or more unpaired electrons in its outer orbital. These highly unstable molecules tend to react rapidly with adjacent molecules, donating, subtracting, or even sharing their outer orbital electron and being the most important ones those derived from either oxygen and nitrogen. ROS are substances that are released during oxidative metabolism. ROS include the superoxide anion (O₂•⁻), hydrogen peroxide (H₂O₂), and the hydroxyl radical (OH•)¹². The reaction of ROS with macromolecules can lead to DNA mutations, changes in the structure and function of proteins, and peroxidative cell-membrane lipids. Both the radicals and the non-radical species generated via interaction with free radicals, are referred to as RONS¹³.

To limit these harmful effects, an organism requires complex and protection—the antioxidant system. An antioxidant is a substance that, when present in low concentrations relative to oxidizable substrate, significantly delays or reduces oxidation of the substrate¹⁴. This system consists of

antioxidant enzymes (SOD, CAT, GPx, glutathione reductase [GR]) and non-enzymatic antioxidant (e.g. vitamins C, E, glutathione and uric acid [UA]). The antioxidant system efficiency depends on nutritional intakes (vitamins and micronutrients) and on endogenous antioxidant enzyme production, which can be modified by exercise, training nutrition and aging.

An imbalance between free radical production and antioxidant defence leads to an oxidative stress state that can potentially result in a change in the intracellular redox balance towards a more oxidizing environment, with the possibility to cause ill-health and disease. However, despite their excess leads to oxidative/nitrosative stress, free radical, and RONS are involved in several important biological processes, including cell signalling, redox regulation of gene transcription, cellular immunity and apoptosis, being essential for normal physiological function¹³.

Exercise and oxidative stress

Regular physical activity, together with a balanced diet, is known as an important factor for health. However, exhaustive or intense physical activity is associated with a dramatic increase in oxygen uptake both by the whole body and particularly by the skeletal muscle. The production of ROS is believed to be the underlying mechanism for a series of biochemical and physiological changes that occur during exercise and are indicative of oxidative stress. In general, the body has adequate antioxidant reserves to cope with the production of ROS under physiological conditions. The system consists of antioxidant vitamins, glutathione and thiols, and antioxidant enzymes¹⁵. These antioxidant defence systems preserve homeostasis for normal cell functions at rest and perhaps during mild exercise. However, when ROS production is excessive, such as during prolonged aerobic exercise, or when antioxidant defence is severely hampered by nutritional deficiencies and pharmacological intervention, an inadequate defence may be overwhelmed by the ROS, leading to extensive cell and tissue damage.

Since Dillard et al.¹⁶ first reported in 1978 that exercise increases lipid peroxidation, the link between exercise and oxidative stress has received considerable attention and the studies about this topic are increasing significantly in number. Thus, it is now well established that exercise of enough intensity and duration increases the formation of RONS which could result in increased oxidative stress¹⁷, as indicated by an increase in oxidized molecules in a variety of tissues and body fluids. However, whether or not an increase in RONS resulting from acute exercise causes oxidative damage remains still unclear.

The effect of exercise on the levels of oxidative stress biomarkers has been extensively studied for both healthy and diseased subjects¹⁸. These studies have reported inconsistent results concerning the effect of exercise on the levels of specific oxidative stress biomarkers, a fact that, at least partially, may be attributable to differences in the training status, dietary intake, age and gender of subjects, type of exercise test (mode aerobic or anaerobic, duration and intensity), sampling time points, tissue sampled and the assays used among the studies.

Exercise and Down syndrome

The mitochondrial respiratory chain is believed to be the major cellular source of free radical generation during exercise^{19,20}. Mitochondrial anomalies and dysfunctions are present in DS, which are directly related to the presence of oxidative stress²¹. Moreover, in addition to generation through mitochondrial electron transport, RONS can be produced through a number of alternative pathways during exercise. This is the case of the enzyme XO, whose activity has been found increased in DS²² and it has been suggested that this enzyme may be more important than mitochondria as a source of exercise-induced free radicals²³. Thus, mitochondrial dysfunction and the excess in XO activity found in DS lead to think that exercise may exacerbate oxidative stress and tissue damage in these subjects compared to patients without DS.

However, results reported in literature seem to show a beneficial effect of exercise on DS subjects, although the effect of exercise on oxidative stress parameters has been poorly studied in this condition. Monteiro et al.²⁴ found significant increases in plasma thiobarbituric acid reactive substance (TBARS) as well as in plasma and erythrocyte glutathione reduced (GSH) and glutathione oxidized (GSSG) in adults with DS after a 16-week training program. Besides, they also found that exercise did not significantly alter erythrocyte SOD1 activity, as it was also reported in a later study. Other antioxidant enzymes have been evaluated in DS subjects after an exercise program. Thus, increased activity of erythrocyte glucose-6-phosphate dehydrogenase (G6PDH), GPx²⁵ and GR²⁶ has been found in adolescents with DS, whereas not significant changes have been reported for erythrocyte CAT²⁶. Lipid peroxidation was also evaluated by Ordoñez et al.²⁷ in adolescents with DS and, in contrast to Monteiro et al.²⁴, they found decreased levels of methylene dioxy amphetamine (MDA) after a very similar exercise program, so discrepancies in results could be due to methodological differences. Supporting the results of Ordoñez et al.²⁷, it has been reported that lipid hydroperoxides decreased in saliva samples of DS subjects after an exercise program of slight intensity²⁸. Campos et al.²⁹ have found that the daily practice of physical exercise of moderate intensity reduces the urinary excretion of UA in DS individuals. On the other hand, it has been recently reported that exercise significantly reduced protein oxidation, measured as plasma protein carbonyl content, in adolescents with DS³⁰ and also plasma allantoin in the same population³¹. Since both plasma protein carbonyls and allantoin levels have been found increased in children with DS^{32,33}, exercise may improve health of this subjects.

The effect of exercise on total antioxidant capacity (TAC) has also been evaluated in DS and no significant effect was reported, in blood³⁴ or in saliva²⁸ samples. Moreover, levels of the ratio thiols/total proteins in plasma samples remained stable after exercise in DS, although these levels were lower over the entire physical activity test in DS compared with non-Down controls³⁴. In contrast, blood TAC was higher in DS subjects than in non-Down controls after exercise³⁴. Authors suggest that the greater TAC in DS participants during exercise might reflect a greater ability to mobilize blood-borne antioxidants due to chronic oxidative

stress. In our opinion, increased levels of UA during exercise could be another possible explanation for these results, even more when XO activity is increased in DS. In fact, it has been reported that high UA concentration increases serum antioxidant capacity and reduces exercise induced oxidative stress in healthy subjects³⁵. Campos et al. 2013²⁹ have observed that physical exercise performed daily decreases urine UA levels in DS. However, Zambrano et al.²⁸ found that exercise had no effect on salivary UA of DS participants. Therefore, more studies are necessary to clarify these hypotheses.

Data pointed to a greater ability to mobilize bloodborne antioxidants or a significant increase of antioxidant enzymes activity (G6PDH, GPx and GR) by exercise in DS subjects as the possible causes of the decrease of oxidative stress by exercise. Thus, exercise does not alter CAT and SOD1 activities, but induces increased GPx activity in erythrocytes of these subjects. Since erythrocyte SOD1 activity seems to be increased in this disorder⁴² and GPx is more efficient in scavenging H₂O₂ than CAT⁴³, exercise could compensate the imbalance of this enzymes protecting against oxidative stress in DS.

On the other hand, resistance exercise and endurance training cause adaptive responses of gene expression in nuclear and mitochondrial genomes in the skeletal muscle⁴⁴. However, changes of gene expression induced by physical activities and its relation with the gene-dosage imbalance in DS have not yet been studied. Therefore, more well-conducted studies which examine long-term physical outcomes, adverse effects and psychosocial outcomes are needed to reach firm conclusions.

Conclusions

Down's syndrome individuals generally tend to be sedentary, however physical activity has the potential to improve their health when is practised regularly. Thus the strength of publishing works like ours, that show the advantages that performing physical exercise may bring, should help raise awareness that it is a tool that could improve their quality of life.

Search strategy and selection criteria

A literature search was performed using the electronic databases PubMed, Scopus, Google Scholar and SciFinder (1866 onwards). We focused on recently publications, but did not exclude commonly referenced and highly regarded older publications. Further PubMed searching was performed by using the "see all related citations" function by manual scanning of the reference lists of several review articles, as well as original investigations. Articles from all languages were included. The following search terms were included in multiple combinations: "Down syndrome", "trisomy", "oxidative stress", "oxidative damage", "nitrosative stress", "lipid peroxidation", "antioxidant biomarker", "MDA", "TBARS", "4-hydroxy-2-nonenal", "carbonyls", "dityrosine", "homocysteine", "glutathione peroxidase", "catalase", "glutathione reductase", "antioxidant enzyme", "glu-

tathione", "uric acid", "vitamin", "thiol", "total antioxidant capacity", "allantoin", "xanthine oxidase", "xanthine oxidoreductase", "exercise", "physical activity". The search was conducted between September 2013 and March 2014.

Conflict of interests

Authors declare not to have any conflict of interests.

Acknowledgements

The authors would like to thank Dra. Noelia Cañas for her valuable advices, María Burgos for English correction of the manuscript, and Nieves Fonturbel for her helpful contribution with bibliographic compilation.

Funding

This study was supported in part by Fundación Inocente, Inocente.

References

1. Finaud J, Lac G, Filaire E. Oxidative stress. Relationship with exercise and training. *Sports Med.* 2006;36:327-58.
2. Ji LL Oxidative stress during exercise: implication of antioxidant nutrients. *Free Radic Biol Med.* 1995;18:1079-86.
3. Clarkson PM. Antioxidants and physical performance. *Crit Rev Food Sci Nutr.* 1995;35:131-41.
4. Hook EB. Down's syndrome-frequency in human populations and factors pertinent to variation in rates. In: de la Cruz FF, Gerald PS, editors. *Trisomy 21 (Down syndrome): research perspectives.* Baltimore: University Park Press; 1981. p. 3-68.
5. Lejeune J, Gautier M, Turpin R. Etude des chromosomes somatiques de 9 enfants mongoliens. *CR Acad Sci Paris.* 1959;248:1721-8.
6. Pueschel SM. Clinical aspects of Down syndrome from infancy to adulthood. *Am J Med Genet.* 1990;7(Suppl.):52-6.
7. Groner Y, Elroy-Stein O, Avraham KB, Schickler M, Knobler H, Minc-Golomb D, et al. Cell damage by excess CuZnSOD and Down's syndrome. *Biomed Pharmacother.* 1994;48:231-40.
8. Pagano G, Castello G. Oxidative stress and mitochondrial dysfunction in Down syndrome. *Adv Exp Med Biol.* 2012;724:291-9.
9. Garlet TR, Parisotto EB, Medeiros GS, Pereira LCR, Moreira EAM, Dalmarco EM, et al. Systemic oxidative stress in children and teenagers with Down syndrome. *Life Sci.* 2013;93:558-63.
10. Lott IT. Antioxidants in Down syndrome. *Bioch Biophys Acta.* 2012;1822:657-63.
11. Halliwell B, Gutteridge JMC, editors. *Free Radicals in Biology and Medicine.* 4.^a ed. Oxford: Clarendon Press; 2006.
12. Halliwell B, Gutteridge JM. Oxygen toxicity, oxygen radicals, transition metals and disease. *Biochem J.* 1984;219:1-14.
13. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol.* 2007;39:44-842007.
14. Halliwell B. Antioxidant characterization. Methodology and mechanism. *Biochem Pharmacol.* 1995;49:1341-8.
15. Machlin LJ, Bendich A. Free radical tissue damage: Protective role of antioxidant nutrients. *FASEB J.* 1987;1:441-5.

16. Dillard CJ, Litov RE, Savin WM, Dumelin EE, Tappel AL. Effects of exercise, vitamin E, and ozone on pulmonary function and lipid peroxidation. *J Appl Physiol*. 1978;45:927-32.
17. Bloomer RJ. Effect of exercise on oxidative stress biomarkers. *Adv Clin Chem*. 2008;46:1-50.
18. Fisher-Wellman K, Bloomer RJ. Acute exercise and oxidative stress: a 30 year history. *Dyn Med*. 2009;8:1.
19. Ji LL. Antioxidants and oxidative stress in exercise. *Proc Soc Exp Biol Med*. 1999;222:283-92.
20. Leeuwenburgh C, Heinecke JW. Oxidative stress and antioxidants in exercise. *Curr Med Chem*. 2001;8:829-38.
21. Pallardo FV, Lloret A, Lebel M, d'Ischia M, Cogger VC, Le Couteur DG, et al. Mitochondrial dysfunction in some oxidative stress related genetic diseases: Ataxia-Telangiectasia, Down Syndrome, Fanconi Anaemia and Werner Syndrome. *Biogerontology*. 2010;11:401-19.
22. Pallardo FV, Degan P, d'Ischia M, Kelly FJ, Zatteralle A, Calzone R. Multiple evidence for an early age pro-oxidant state in Down syndrome patients. *Biogerontology*. 2006;7:211-20.
23. Cooper CE, Vollaard NBJ, Choueiri T, Wilson MT. Exercise, free radicals and oxidative stress. *Biochem Soc Trans*. 2002;30:280-5.
24. Monteiro CP, Varela A, Pinto M, Neves J, Felisberto GM, Vaz C, et al. Effect of an aerobic training on magnesium, trace elements and antioxidant systems in a Down syndrome population. *Magnes Res*. 1997;10:65-71.
25. Francisco Javier O, Manuel R, Manuel RR. Regular physical activity increases glutathione peroxidase activity in adolescents with Down syndrome. *Clin J Sport Med*. 2006;16:355-6.
26. Rosety-Rodriguez M, Rosety M, Ordonez FJ. Influence of regular exercise on erythrocyte catalase activity in adolescents with Down syndrome. *Med Clin (Barc)*. 2006;127:533-4.
27. Ordoñez, FJ, Rosety-Rodriguez M. Regular exercise attenuated lipid peroxidation in adolescents with Down syndrome. *Clin Biochem*. 2007;40:141-2.
28. Zambrano JC, Marquina R, Sulbaran N, Rodriguez-Malaver AJ, Reyes RA. Aerobic exercise reduced oxidative stress in saliva of persons with Down syndrome. *Res Sports Med*. 2009;17:195-203.
29. Campos Vaquero C, Guzmán Martínez R, López-Fernández E, Casado Moragón A. Physical exercise and urinary uric acid levels in Down's syndrome. *Rev Med Int Sindr Down*. 2013;17:1-5.
30. Ordonez FJ, Rosety I, Rosety MA, Camacho-Molina A, Fornieles G, Rosety M, et al. Aerobic training at moderate intensity reduced protein oxidation in adolescents with Down syndrome. *Scand J Med Sci Sports*. 2012;22:91-4.
31. Rosety-Rodriguez M, Rosety I, Fornieles-Gonzalez G, Diaz A, Rosety M, Ordonez FJ. A 12-week aerobic training programme reduced plasmatic allantoin in adolescents with Down syndrome. *Br J Sports Med*. 2010;44:685-7.
32. Zitnanová I, Korytár P, Sobotová H, Horáková L, Sustrová M, Puschel S, et al. Markers of oxidative stress in children with Down syndrome. *Clin Chem Lab Med*. 2006;44:306-10.
33. Zitnanová I, Korytár P, Aruoma OI, Sustrová M, Garaiová I, Muchová J, et al. Uric acid and allantoin levels in Down syndrome: antioxidant and oxidative stress mechanism? *Clin Chim Acta*. 2004;341:139-46.
34. Flore P, Bricout VA, van Biesen D, Guinot M, Laporte F, Pépin JL, et al. Oxidative stress and metabolism at rest and during exercise in persons with Down syndrome. *Eur J Cardiovasc Prev Rehabil*. 2008;15:35-42.
35. Waring WS, Convery A, Mishra V, Shenkin A, Webb DJ, Maxwell SR. Uric acid reduces exercise induced oxidative stress in healthy adults. *Clin Sci (Lond)*. 2003;105:425-30.
36. Sinet PM, Allard D, Lejeune J, Jerome H. Augmentation d'activité de la superoxyde dismutase érythrocytaire dans la trisomie pour le chromosome 21. *CR Acad Sci (Paris)*. 1974;278:3267-70.
37. Brooksbank BW, Balazs R. Superoxide dismutase, glutathione peroxidase and lipoperoxidation in Down's syndrome fetal brain. *Brain Res*. 1984;318:37-44.
38. Anneren KG, Epstein CJ. Lipid peroxidation and superoxide dismutase-1 and glutathione peroxidase activities in trisomy 16 fetal mice and human trisomy 21 fibroblasts. *Pediatr Res*. 1987;21:88-92.
39. De La Torre R, Casado A, Lopez-Fernandez E, Carrascosa D, Ramirez V, Saez J. Overexpression of copper-zinc superoxide dismutase in trisomy 21. *Experientia*. 1966;52:871-3.
40. Sinet PM, Lavelle F, Michelson AM, Jerome H. Superoxide dismutase activities of blood platelets in trisomy 21. *Biochem Biophys Res Commun*. 1975;67:904-9.
41. Feaster WW, Kwok LW, Epstein CJ. Dosage effects for superoxide dismutase-1 in nucleated cells aneuploid for chromosome 21. *Am J Hum Genet*. 1977;29:563-70.
42. Sebastià J, Cristófol R, Pertusa M, Vilchez D, Torán N, Barambio S et al. Down's syndrome astrocytes have greater antioxidant capacity than euploid astrocytes. *Eur J Neurosci*. 2004;20:2355-66.
43. Antunes F, Han D, Cadenas E. Relative contributions of heart mitochondria glutathione peroxidase and catalase to H(2)O(2) detoxification in vivo conditions. *Free Radic Biol Med*. 2002;33:1260-67.
44. Radak Z, Chung HY, Koltai E, Taylor AW, Goto S. Exercise, oxidative stress and hormesis. *Ageing Res Rev*. 2008;7:34-42.